Chisel Manual

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Installation

pip install poseydon-chisel

Requirements

(will automatically install if not satisfied):

```
'numpy>=1.20.0', 'pandas>=1.3.0', 'pysam>=0.18.0', 'pybigwig', 'biopython>=1.79', 'joblib>=1.1.0', 'matplotlib>=3.5.0', 'denseweight==0.1.2'
```

Usage

from poseydon_chisel import *

Notes

- Sections are not alphabetically sorted to preserve relationships. Use find (ctrl+f) or the navigation pane.
- All arrays are numpy arrays unless otherwise specified

Symbols

s = Sequence.

S = Collection of sequence(s).

z = Sub-sequence in S.

r = Region. sub-location in Z. rs = Regions

Arr = Array. A numpy array unless otherwise specified.

V = sub-array / slice

 $\mathbf{H} = \text{Signal}(\mathbf{s}).$

 $\mathbf{A} = \text{Area}(s)$.

C# = Column.

Abbreviations

B* Boolean (True or False)

IntIntegerseqSequence.LoLList of lists.Bw/bwBigwig.Bg/bgBedgraph.

RC Reverse Complement

OHE One Hot Encoded

OHERC OHE + RC.

PWM Position Weight Matrix.

np NumPy

Formats

Track List; each element (**sub-Track**) pertaining to a sub-sequence. $[z_1, z_2 ... z_N]$.

Tracks List of Tracks.

Sequence Track (SeqTrack) Track; each element a string.

Signal Track (SigTrack) Track; each element an Arr with the shape (len(z), H). Signal can be from one or

multiple sources. Each value an int or float.

Binary Track (BinTrack) SigTrack; values are either 0 or 1.

Filter Track (**FilterTrack**) BinTrack; 0s are positions to be excluded from analysis.

Profile A transposed slice of a SigTrack. An Arr with shape (H, len(r)).

Area Integration of a Profile. An Arr with shape (*H*, 1).

Markers: Pandas Arr with shape (L, 2-4+). Each row is a location in the sequence. C1 is the sub-

sequence identifier; C2 is the sub-sequence index; C3 (optional) is the strand where 0 is the positive (+) strand and 1 is the negative (-) strand; C-1 (the last column) (optional) is

the size of the region, centered on C2. Additional columns (optional) can include

genomic attributes.

Nucleotide Array (NucArr) Arr with shape (len(s), 4 or 8). Each row is a position in the sequence. C1-4 are 'A',

'C', 'G', 'T'/'U' respectively. C5-8 (optional) is the same as C1-4 but represents the

reverse complement.

Pack Arr with shape (rs, *V.shape).

SeqPack Pack of sequences. Arr with shape (rs,)

NucArrPack Pack of NucArr. Arr with shape (rs, len(s), 4 or 8).

Profile A transposed slice of a SigTrack. An Arr with shape (H, len(r)).

Area Integration of a Profile. An Arr with shape (*H*, 1).

ProfilePack Pack of Profiles. Arr with shape (rs, H, len(R)).

AreaPack Pack of Area(s). Arr with shape (rs, H, A).

TraitPack Pack of Trait(s).

FlattenedPack Arr with shape (rs, F), where F is the product of V shape.

Common Arguments

fileloc Location of the file.

^{*} all arguments that take in lists can also take in 1dim arrays.

seq String.

seqs List of seqs.

BaseSeq (BS) SeqTrack. Reference seq. E.g., a genome where each sub-seq is a chromosome.

BS_ids List of strings. Ids of each sub-seq. E.g., chromosome names.

select_BS_ids List of strings or None. BS_ids to include in output. If None, all BS_ids are used.

BS_sizes List of ints. Lengths of each sub-seq. I.e. chromosome lengths.

reso Int. The current resolution of the file/SigTrack.

newreso Int or None. New resolution of output track.

resomode Function to convert reso to newreso. Commonly either np.mean or np.max.

Msizes List or None. Marker size. If None, 1bp will be used. If length of Msizes is 1, this size

will be applied to all. Otherwise, must match length of Markers. Could use Markers[:, -1]

if sizes column present.

expand_dim Int or None. Dimension to expand output.

PWM NucArr.

PWMScores SigTrack of scores for each PWM window in sub-seqs in BaseSeq.

window Int or None. Window size.

stride Int or None. Stride size. Positions to skip for every slide of a window.

Vsize Int. size of slice to retrieve centered at Marker.

exact B*. When reso > 1, retrieving a slice can be from a rounded to reso Marker (False) or

exact (True). The latter involves interpolating the signal.

Vpad Int or None. Padding added to slice for exact = True.

stranded B*. If True, will take strand information to retrieve sequence. Markers must have strand

column.

segmode None or function used on SegPack. Can be Seg2OHE or Seg2OHERC.

xlabels None or List of strings. Labels for x axis.

labels None or List of strings. Labels for y axis or subplot title.

suptitle None or string. Title of plot.figsize Tuple of Ints. Figure size.

fontsize Int. font size.

cmap String. Matplotlib color palette.

threshold Int or None. If int, values equal to or greater than are taken.

threshsign Function where the values of are compared to the threshold; threshsign(values,

threshold). Can be np.greater, np.greater_equal, np.less, np.less_equal, np.equal,

np.not_equal.

mode Function.

center Int or None. If int, will pad the beginning window//2 by the int value such that values

from windowing represent the center of the window instead of the start.

extend Int or None. If int, will extend subTrack after windowing to match the template length by

the int value.

skiprows Int. Number of rows to skip in file.

sep String. File delimiter. Default is: '\t'.

addcols List of int(s) or None. Additional columns: columns to add to Markers.

addcols_names List of string(s) or None. Names for additional columns.

Functions

In the following format:

Function

Description

(<u>Argument</u> [= Default])

<u>Argument</u> description. Refer to Common Arguments if not included here.

Return Product(s)

Fasta2Seqs

Converts a fasta file into SeqTrack and ids (optional).

 $(\underline{\text{fileloc}}, \underline{\text{idents}} = \text{False})$

<u>idents</u> Output fasta description if True.

SeqTrack; a list of ids if idents = True.

TrackSelect

Selects sub-Track(s) based on sub-seq identifiers.

(Track, BS_ids, select_BS_ids)

Track

ListLengths

Gets the length of each element in a list.

(x)

 $\underline{\mathbf{x}}$ List.

List.

Rounder

 $\underline{\mathbf{X}}$

Rounds a number/numbers to the nearest base.

 $(\underline{x}, \underline{base} = 5)$

Int or 1dim NumPy Arr

➤ Int or 1dim NumPy Arr with dtype = int.

Sequence Functions

Seqs2kmer

Finds regions in SeqTrack that match k-mer.

 $(\underline{\text{seqs}}, \underline{\mathbf{k}} = 6)$

 $\underline{\mathbf{k}}$ Int. Size of k-mers.

 \triangleright List of *k*-mers.

KmerFinder

Finds regions in SeqTrack that match kmer.

 $(\underline{SeqTrack}, \underline{kmer}, \underline{RC} = None, \underline{center} = None, \underline{extend} = None)$

<u>kmer</u> String. The pattern to search SeqTrack for. It is case sensitive.

 \underline{RC} B*. If True, searches the reverse complement of each k-mer.

➤ BinTrack.

Seqs2OHE

Converts list of seqs to OHE.

 $(\underline{Seqs}, \underline{expand} \underline{dim} = None, \underline{customdict} = None)$

<u>customdict</u> dictionary that maps (capital) letter to one hot encoding. If None, default is used.

> OHE.

Seqs2OHERC

Converts a list of seqs to OHERC.

 $(\underline{Seqs}, \underline{expand_dim} = None, \underline{customdict} = None)$

<u>customdict</u> see Seqs2OHE.

> OHERC.

CM2PWM

Converts a count matrix to a PWM.

 $(\underline{CM}, \underline{pseudo} = 1, \underline{bkg} = [0.25, 0.25, 0.25, 0.25], \underline{base} = 2)$:

CM NucArr. Count Matrix.

<u>pseudo</u> Int. Pseudo count to be added to each element of the CountMatrix.

<u>bkg</u> List. Background probabilities of each nucleotide.

<u>base</u> Int. Base of log for log scaled normalized frequency values.

> PWM.

Seq2PWMScore

Scores PWM fit on seq.

 $(\underline{\text{seq}}, \underline{\text{PWM}}, \underline{\text{pieces}} = \text{None}, \underline{\text{RC}} = \text{True}, \underline{\text{center}} = \text{None}, \underline{\text{extend}} = \text{None})$

pieces Int or None. Breaks up seq into multiple equal length pieces. They are overlapping so to

provide a score for every position in the seq as if 1 piece. Useful for memory

consumption.

RC B*. If True, will score the sequence and its reverse complement and keeps the greater

score.

List of scores for each PWM window in seq.

PWMScorer

Scores PWM on BaseSeq.

(<u>PWM</u>, <u>BaseSeq</u>, <u>BS_ids</u>, <u>select_BS_ids</u> = None, <u>parallel</u> = None, <u>pieces</u> = None, <u>RC</u> = True, <u>center</u> = None, <u>extend</u> =

None):

<u>parallel</u>: None or int. Number of processors to use for parallel multiprocessing. If None, no

parallel processing.

<u>pieces</u> See Seq2PWMScore.

<u>RC</u> See Seq2PWMScore.

➤ SigTrack.

Track Functions

Bw2Track

Converts a .bigwig file into a SigTrack.

 $(\underline{fileloc}, \underline{BS_sizes}, \underline{BS_ids} \text{ , } \underline{select_BS_ids} = None, \underline{newreso} = 20, \underline{resomode} = np.mean, \underline{window} = None)$

window Int or None. maximum size of interval to include score from.

➤ SigTrack.

Bg2Track

Converts a .bedgraph file into a SigTrack.

(<u>fileloc</u>, <u>BS sizes</u>, <u>BS ids</u>, <u>select BS ids</u> = None, <u>newreso</u> = 20, <u>resomode</u> = np.mean, <u>window</u> = None, <u>sep</u> = '\t', <u>skiprows</u> = None)

window See Bw2Track.

➤ SigTrack.

Bam2Track

Converts a .bedgraph file into a SigTrack.

(<u>fileloc</u>, <u>BS sizes</u>, <u>BS ids</u>, <u>select BS ids</u> = None, <u>newreso</u> = 20, <u>resomode</u> = np.mean, <u>stranded</u> = True, <u>read length</u> = None, <u>paired</u> = False, <u>make_index</u> = False)

stranded B*. Reads are mapped to respective strand on a SigTrack. If False, will sum signals to a

single signal SigTrack.

read_length Int or None. If None, read_length is inferred from bam file. If int, read will be extended

to read_length strand specifically.

<u>paired</u> B*. If True, mapped mates will be used to extend the read of the first mate. If False, reads

are treated independently and the size by read_length.

<u>make index</u> B*. If True, will create a index file for bam file in source directory.

➤ SigTrack.

TrackInterpolator

Interpolates each individual signal of SigTrack by values that pass a threshold.

(<u>SigTrack</u>, <u>reso</u>, <u>window</u> = None, <u>threshold</u> = 0, <u>threshsign</u> = np.greater)

window Int or None. If gap between indices greater than threshold is greater than window, gap is

not interpolated.

➤ SigTrack.

Tracks2Track

Stacks a list of SigTracks to a single SigTrack,

(SigTracks)

➤ SigTrack.

TrackMerger

Merges multi signal SigTrack into a single signal SigTrack.

 $(\underline{SigTrack}, \underline{mode} = np.mean)$

➤ SigTrack.

TrackModifier

Applies a function with a Modifier to SigTrack. Most used for multiplying a SigTrack with a FilterTrack to get a filtered SigTrack.

(<u>SigTrack</u>, <u>modifier</u>, <u>mode</u> = np.multiply)

Modifier a list that should match length of SigTrack. Each sub-modifier is applied to each

subTrack of a SigTrack. Sub-modifiers must be able to broadcast over the subTrack.

➤ SigTrack.

TrackTransformer

Min-max normalizes or standardizes each individual signal of a SigTrack.

 $(\underline{SigTrack}, \underline{minmax} = (0, None), \underline{standardize} = False)$

minmax

a tuple where the first and second position are minimum and maximum values for

normalization. If a value is None, it is found from the SigTrack. Is not used if standard is

True.

standardize

B*. If True, values will be standardized instead of minmax normalized.

➤ SigTrack.

LowerResTrack

Lowers the resolution of a SigTrack.

 $(\underline{SigTrack}, \underline{reso}, \underline{newreso}, \underline{resomode} = np.mean)$

➤ SigTrack.

TrackExpander

Extends a single signal SigTrack to size of reference sequence by repeating values and adding filler at ends.

(SigTracks, reso, BS_size, BS_ids, select_BS_ids = None, filler = 0)

filler

Int. Adds this value to the end to match reference sequence size.

➤ SigTrack.

BinTrackInverter

Inverts binary values; 0s become 1s and vice-versa.

(BinTrack)

➤ BinTrack.

TrackWindower

Applies a function over windows of a SigTrack.

(SigTrack, reso = 1, mode = np.mean, window = None, center = True, extend = True)

window

Int or None. size of window to apply *mode* over. If None, window is equal to reso.

➤ SigTrack.

TrackThresholder

Finds values in SigTrack that pass a threshold. Can return either a BinTrack or a SigTrack that includes those values that passed while others are zeroed. .

(SigTrack, threshold = 0, threshsign = np.greater, binary = False)

binary

B*. If True, will return a BinTrack else will return a SigTrack.

➤ SigTrack.

TrackPadder

Finds a value in SigTrack and expands it to surrounding values. Useful for expanding 0s or 1s in BinTracks.

 $(\underline{\text{SigTrack}}, \underline{\text{reso}} = 1, \underline{\text{pad}} = 10, \underline{\text{shift}} = 0, \underline{\text{value}} = 1, \underline{\text{base}} = 0)$

<u>pad</u> Int. number of surrounding bp on either side to expand value to.

shift Int. can be negative. Number of bp to shift pad window. Useful if padding is not centered.

<u>value</u> Int or float. Value to be expanded over base.

<u>base</u> Int or float. Background values.

SigTrack.

TrackFlagger

Identifies flags in SigTrack.

(SigTrack, reso = 5, window = 1000, stride = 50, flagmode = np.argmax, double = False)

<u>flagmode</u> Function to be applied to window to find index (*flag*) within window.

double B*. If True, only keeps best *flag* within a window.

➤ SigTrack.

Marker Functions

Bed2Markers

Converts a bed file to Markers.

(<u>fileloc</u>, <u>sep</u> = '\t', <u>header</u> = None, <u>skiprows</u> = None, <u>strandcol</u> = None, <u>addcols</u> = None, <u>addcols_names</u> = None)

<u>strandcol</u> Int or None. Column with strand information.

Markers

MarkersFilter

Filters Markers by SigTrack values.

(<u>Markers</u>, <u>SigTrack</u>, <u>BS_ids</u>, <u>select_BS_ids</u> = None, <u>reso</u> = 1, <u>exact</u> = False, <u>Msizes</u> = None, <u>threshold</u> = None,

threshmode = np.sum, threshsign = np.greater)

<u>reso</u> Int. SigTrack reso.

Markers.

Markers2BinTrack

Converts Markers to BinTrack

(Markers, BS_sizes, BS_ids, select_BS_ids = None, Msizes=None, reso = 1, inverse = False)

inverse B*. If False, Markers are 1s and rest are 0s, else inversed.

<u>reso</u> Int. reso of output BinTrack.

BinTrack.

SigTrack2Markers

Produces Markers from SigTrack.

(SigTrack, BS_ids, select_BS_ids = None, reso = 1, select = None, select_mode = None, select_mode_args = None)

select Int or None. If None, will make Markers from all non-zero values. If specified, will

return that number of Markers.

select_mode Function. If None and select is an int, will randomly select from non-zero values.

Currently can be either Top, RevDistro, or Harpoon.

<u>mode_args</u> Dictionary or None. Arguments for select_mode function, only used if select_mode is not

None. If None, will use select_mode defaults.

Markers.

Top

Returns indices of the largest or smallest values.

 $(\underline{\text{vals}}, \underline{\text{select}}, \underline{\text{smallest}} = \text{False})$

<u>vals</u>
List or 1-dim array of int or floats.
select
Int. Number of indices to return.

smallest B*. If False, larger values are taken for top/threshold; if True, smaller values.

> Indices.

RevDistro

Returns indices of weighted (DenseWeight) random selections.

($\underline{\text{vals}}$, $\underline{\text{select}}$, $\underline{\text{alpha}} = 1.0$)

<u>vals</u>
List or 1-dim array of int or floats.

<u>select</u>
Int. Number of indices to return.

<u>alpha</u> Int or float. The alpha parameter in DenseWeights (Steininger et al., 2021).

> Indices.

Harpoon

Returns indices of a uniformized distribution.

($\underline{\text{vals}}$, $\underline{\text{select}}$, $\underline{\text{bins}} = 100$)

<u>vals</u>
List or 1-dim array of int or floats.

<u>select</u>
Int. Number of indices to return.

bins Int. Number of bins to use for binning values.

> Indices.

IdxFlat2Track

Converts list of indices from flattened SigTrack to lists of indices per sub-Track.

(SigTrack, flatidx)

<u>flatidx</u> List or 1-dim array of int.

> Track Indices.

Txt2Markers

Produces Markers from txt file.

(fileloc, addcols, sep = '\t', header = None, skiprows = None, addcols_names = None)

Markers.

AddTraits

Transfers Traits from one Markers to another.

 $(\underline{Markers}_A, \underline{Markers}_B, \underline{Msizes}_A = None, \underline{Msizes}_B = None)$

Markers A Markers to transfer traits to.

<u>Markers B</u> Markers to transfer traits from.

Msizes A Msizes for Markers_A.

Msizes_B Msizes for Markers_B.

Markers.

Markers4Packs

Edits Markers for Pack production.

(<u>Markers</u>, <u>select_BS_ids</u> = None, <u>opposite</u> = None, <u>ends</u> = None, <u>BS_ids</u> = None, <u>BS_sizes</u> = None)

opposite B*. If True, will include the opposite strand for each Marker then removes duplicates.

ends Int or None. Markers falling within the ends of sub-seq are removed.

Markers.

Slice, Pack and Split Functions

SeqTrack2Slice

Gets a Slice at Marker of a SegTrack.

(SeqTrack, Marker, BS_ids, select_BS_ids, Vsize = 1000, seqmode = None, stranded = False)

> Slice.

SeqTrack2Pack

Gets a Pack at Markers of a SigTrack.

(SeqTrack, Markers, BS_ids, select_BS_ids = None, Vsize = 1000, seqmode = None, stranded = False)

Markers (Filtered and modified), Pack.

SigTrack2Slice

Gets a Slice at Marker of a SigTrack.

 $(\underline{SigTrack}, \underline{Marker}, \underline{BS_ids}, \underline{select_BS_ids} = None, \underline{Vsize} = 1000, \underline{reso} = 5, \underline{newreso} = None, \underline{resomode} = np.mean, \underline{exact} = 1000, \underline{reso} = 1000, \underline{reso} = 1000, \underline{resomode} = 10000, \underline{resomode} = 1000, \underline{resomode} = 10000, \underline{resomode} = 10000,$

= True, $\underline{Vpad} = 2$, $\underline{stranded} = False$)

Vpad Int. Adds Vpad*reso to the Slice for interpolation (when exact = True).

> Slice.

SigTrack2Pack

Gets a Pack at Markers of a SigTrack.

(<u>SigTrack</u>, <u>Markers</u>, <u>BS_ids</u>, <u>select_BS_ids</u>, <u>Vsize</u> = 1000, <u>reso</u> = 5, <u>newreso</u> = None, <u>resomode</u> = np.mean, <u>areas</u> = None, exact = False, stranded = False)

areas list of ints. Sizes to integrate the signal over. Largest must be smaller or equal to the

Vsize.

> Markers (Filtered and modified), Pack.

LowerResProfilePack

Lowers the resolution of a ProfilePack.

($\underline{ProfilePack}$, \underline{reso} , $\underline{newreso}$, $\underline{resomode} = np.mean$)

ProfilePack.

Traits2Pack

Gets a Pack from traits of a Markers.

($\underline{Markers}$, \underline{cols} = None, $\underline{vectorize}$ =None)

cols list of ints or None. Columns of Markers to retrieve traits from.

<u>vectorize</u> list of ints or None. Columns of Markers to vectorize. Must be in cols. Vectorize applies

pandas get_dummies and returns alphabetically sorted vectorizations.

TraitPack.

PackShaper

Modifies shape of a Pack.

 $(\underline{Pack}, \underline{expand_dim} = None, \underline{squeeze} = False, \underline{twoD} = False, \underline{flatten} = False)$

<u>squeeze</u> B*. If True, will call np.squeeze to remove 1 length dimensions.

twoD B*. If True, will reshape pack into 2 dimensions of (observations, features).

<u>flatten</u> B*. If True, will call np.flatten to convert pack to 1 dimension.

Pack.

PackShaper

Concatenates Packs of the same dimensions and the same first dimension.

(Packs)

<u>Packs</u> List of packs. Must be of the same dimensions and the same first dimension.

Pack.

Splitter

Generates Splits.

 $(X, \underline{\text{num splits}} = 3, \underline{\text{proportions}} = [0.7, 0.3], \underline{\text{random}} = \text{False}, \underline{\text{cut}} = \text{True})$

X Int. Length of Markers.

<u>num_splits</u> Int. Number of Split to generate.

<u>proportions</u> List of int or float. Proportion of indices in each sub-split.

<u>random</u> B*. When True, will randomize the distribution of indices.

cut B*. When True, will apply cut method like card shuffling.

> Splits.

PackSplit

Divides a Pack based on a Split.

(Pack, Split)

➤ List of Packs for each sub-split.

Visuals Functions

GridPlot

(<u>Arr</u>, <u>bounds</u> = (None, None), <u>xlabels</u> = None, <u>labels</u> = None, <u>suptitle</u> = None, <u>figsize</u> = (15,5), <u>fontsize</u> = 10, <u>cmap</u> = 'viridis')

<u>bounds</u>

➤ GridPlot

Tuple of Int or None. Sets the lower and upper limit for coloring respectively.

FilledLinePlot

(<u>Arr</u>, <u>bounds</u> = (None, None), <u>xlabels</u> = None, <u>labels</u> = None, <u>suptitle</u> = None, <u>figsize</u> = (15,5), <u>fontsize</u> = 10, <u>cmap</u> = 'viridis')

<u>bounds</u> Tuple of Int or None. Sets the lower and upper limit y-axis.

➤ FilledLinePlot

LinePlot

(<u>Arr</u>, <u>bounds</u> = (None, None), <u>xlabels</u> = None, <u>labels</u> = None, <u>suptitle</u> = None, <u>figsize</u> = (15,5), <u>fontsize</u> = 10, <u>cmap</u> = 'viridis')

bounds Tuple of Int or None. Sets the lower and upper limit y-axis.

➤ LinePlot

Track2Visual

Produces visual of a Slice in SigTrack.

(SigTrack, Marker, BS_ids, select_BS_ids = None, reso = 5, Vsize = 2000, exact = False, vismode = GridPlot, bounds = (None, None), xlabelsnum = 3, labels = None, suptitle = None, figsize = (15,5), fontsize = 10, cmap = 'viridis')

> vismode

ProfilePack2Visual

Creates a visual from ProfilePack.

 $(\underline{ProfilePack}, \underline{combomode} = np.mean, \underline{vismode} = GridPlot, \underline{Vsize} = 1000, \underline{bounds} = (None, None), \underline{xlabelsnum} = 3, \underline{labels} = None, \underline{suptitle} = None, \underline{figsize} = (15,5), \underline{fontsize} = 10, \underline{cmap} = 'viridis')$

combomode

Function to use to collapse profile signals to a 2D array. Either np.mean or np.sum.

> vismode

Common Internal Variables

resoratio = newreso // reso

z = sub-seq

ind = index

 $\mathbf{m} = \text{mask}$, ms = masks

T = Track, nT = new Track

I = interpolated

 $\mathbf{r} = \text{region}$

 $\mathbf{st} = \mathbf{strand}$

l* = len(*)

si = list of sizes

 $\mathbf{sp} = \text{list of spacing}$

 $\mathbf{t} = \text{threshold}$

V = slice

ud = updown

 $\mathbf{c} = \text{cent} = \text{center}$

iro, ro = index, row for pandas iterrows