Package 'RLowPCor'

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	s used to construct, integrate, refine and evaluate orks on large scale gene expression data.	
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Depends R (>= 3.2.3), ggplot2, plyr RoxygenNote 5.0.1	ed:	
adjmatrix2edgelist average.consensus confusion edgelist2adjmatrix RLowPCor		
		_
adj2rankadj Convert network matrix to network rank matrix		

Description

Convert network matrix to network rank matrix

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Usage

```
adj2rankadj(adjmatrix, directed = F)
```

Arguments

adjmatrix A network matrix

directed Logical. If TRUE, the network is considered as directed. If FALSE, the upper

triangular part of the matrix is used to calcuate the rank matrix

Value

an network connection rank matrix

adjmatrix2edgelist

Convert network matrix to edge list

Description

The inferred network matrix is converted to edge list

Usage

```
adjmatrix2edgelist(adjmatrix, cutoff = 0, directed = F)
```

Arguments

adjmatrix a network matrix

cutoff threshod to cut the edge list

directed logical, if FLASE the adjmatrix is transformed to symmetric matrix

Value

a edge list

```
##load data
library(networkBMA)
library(RLowPCor)
data(dream4)
data.exp<-dream4ts10[[1]][,-c(1:2)]
genes<-colnames(data.exp)
##build correlation network
inf.cor<-abs(cor(data.exp))
diag(inf.cor)<-0
##convert matrix to edge list
adjmatrix2edgelist(inf.cor)</pre>
```

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average.consensus

Consensus network from average rank

Description

Consensus network is built of taking the average ranks of the edges from multiple network predictions.

Usage

```
average.consensus(adjmatrix.list, directed = F)
```

Arguments

adjmatrix.list A list of network prediciton matrices with same rownames and colnames.

directed

Logical. If TRUE, the networkis considered as directed. If FALSE, the upper triangular part of the matrix is used to calcuate the rank matrix

Value

a network with rank weighted edges. The weights are rescale to 0-1 and hihger values indicate higher ranks.

Examples

```
##create two random networks
library(RLowPCor)
set.seed(4)
net1<-abs(matrix(rnorm(16),4,4))</pre>
net1<-pmax(net1,t(net1))</pre>
diag(net1)<-0
set.seed(5)
net2<-abs(matrix(rnorm(16),4,4))</pre>
net2<-pmax(net2,t(net2))</pre>
diag(net2)<-0
dimnames(net1)<-dimnames(net2)<-list(letters[1:4],letters[1:4])</pre>
net.list<-list(net1=net1,net2=net2)</pre>
inf.consensus<-average.consensus(adjmatrix.list = net.list,directed = F)</pre>
adj2rankadj(net1)
adj2rankadj(net2)
inf.consensus
```

confusion

Derivations of confusino

Description

Calculate statistical measures of the performance of binary classification test from the output confusion matrix table.evaluate

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Usage

```
confusion(input.table)
```

Arguments

input.table the output confusion table from table.evaluate

Details

```
true positive: tp; false positive: fp; true negative: tn; false negative: fn; positives in reference network: p; negatives in reference network: n; true positive rate: tpr = recall = \frac{tp}{tp+fn}; false positive rate: fpr = \frac{fp}{fp+tn}; true negative rate: tnr = \frac{tn}{tn+fp}; false negative rate: fpr = \frac{fn}{fn+tp}; precision: precision = \frac{tp}{tp+fp}; negative predictive value: npv = \frac{tn}{tn+fn}; false discovery rate: fdr = \frac{fp}{fp+tp}; accuracy: accuracy = \frac{tp+tn}{p+n}; f1 scaore: f1 = \frac{2tp}{2tp+fp+fn}; Matthews correlation coefficient:
```

$$mcc = \frac{tp \times tn - fp \times fn}{\sqrt{(tp + fp) \times (tp + fn) \times (tn + fp) \times (tn + fn)}}$$

Value

a table of of statistical measures of performance, see @details

References

Powers DMW: Evaluation: From Precision, Recall and F-Factor to ROC, Informedness, Markedness & Correlation. In. Adelaide, Australia; 2007.

```
library(networkBMA)
library(RLowPCor)
##load DREAM4 size100_1 datasets
data(dream4)
data.exp<-dream4ts10[[1]][,-c(1:2)]
genes<-colnames(data.exp)
ref.edge<-dream4gold10[[1]]
ref.adj<-edgelist2adjmatrix(ref.edge,genes)
inf.cor<-abs(cor(data.exp))
diag(inf.cor)<-0
table.cor<-table.evaluate(inf.adj = inf.cor,ref.adj = ref.adj)
head(table.cor)</pre>
```

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edgelist2adjmatrix Convert edge list to network matrix

Description

The function is to convert edge list to network matrix.

Usage

```
edgelist2adjmatrix(edgelist, genes, cutoff = 0, directed = F)
```

Arguments

edgelist a edge list of network

genes gene names

cutoff the threshold to cut the edge list

directed logical, to create directed or undirected network matrix

Value

a network matrix

Examples

```
library(networkBMA)
library(RLowPCor)
##load DREAM4 size100_1 datasets
data(dream4)
data.exp<-dream4ts10[[1]][,-c(1:2)]
genes<-colnames(data.exp)
ref.edge<-dream4gold10[[1]]
ref.adj<-edgelist2adjmatrix(ref.edge,genes)</pre>
```

RLowPCor

Relevance low order partial correlation

Description

Consensus network is built of taking the average ranks of the edges from multiple network predictions.

Usage

```
RLowPCor(data.exp, edgelist, estimator = "pearson", pc.estimator = "shrink")
```

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Arguments

data.exp gene expression matrix. Columns are variables and rows are samples.

edgelist edge list. First column are the name of regulators, second coloumn are the target genes and the third column are the edge weights.

estimator a character string indicating which correlation coefficient (or covariance) is to be computed. Options are "pearson", "spearman" and "kendall". If shrinkage method is used to estimate PC, the estimator is set to "pearson".

pc.estimator a character string indicating which method is used to estimate the PC of nodes connected to shared neigbours. Options are "shrink" and "pc", correspoinding to the item (c) and (d) in Details: Step 2, respectively.

Details

Step 1: Extract a sparse and scale-free topology from pre-inferred networks as an indirect edge search space for RLowPCor. Unlike a fully connected network, the nodes in the sparse network are assumed to connect to their more relevant neighbours. For example, correlation network can be cut with a range of thresholds until it most fit to scale-free topology. Step 2: Calculate relevance low order partial correlation. For each pair of nodes connected by an edge in the searching space, the edge weight is redefined as (a) Pearson correlation if they do not connect to the same set of neighbour nodes, (b) PC by removing shared neighbours and (c) shrink PC if the covariance matrix used to estimate PC in (b) is not positive definite or invertible. If the searching space is very large, there might still be a number of irrelevant controls involved in shrink PC procedure (c). An alternative is (d) deleting less connected neighbours of the nodes until the covariance matrix in (b) is positive definite and invertible.

Value

a network matrix

References

Sch\"afer J, Strimmer K: A Shrinkage Approach to Large-Scale Covariance Matrix Estimation and Implications for Functional Genomics. Statistical Applications in Genetics and Molecular Biology, The Berkeley Electronic Press 2005, 4(1).

```
##load size 100_1 network DREAM4 datasets
library(networkBMA)
library(RLowPCor)
data(dream4)
data.exp<-dream4ts100[[1]]
#create edge list
edgelist<-dream4gold100[[1]]
edgelist<-edgelist[edgelist[,3]>0,]
##infer RLowPCor network
inf.net=RLowPCor(data.exp = data.exp[,-c(1:2)],edgelist = edgelist)
```

table.evaluate 7

table.evaluate	Evaluate inferred network to refence network	

Description

The inferred network is evaluated by comparing to the reference network. The output is a tables of TP, FP, TN and FP with different edge weight cut-offs

Usage

```
table.evaluate(inf.adj, ref.adj, directed = F)
```

Arguments

inf.adj	the inferred network matrix. Column names and row names match to the reference network.
ref.adj	the reference network matrix with 1 inidating connected edge and 0 unconnected edge.
directed	logical, to compare as directed or undirected networks. In a undirected network, only the upper triangular of the network matrix is used for evaluation.

Value

a confusion table of TP, FP, TN and FN

References

Meyer PE, Lafitte F, Bontempi G: minet: A R/Bioconductor package for inferring large transcriptional networks using mutual information. BMC Bioinformatics 2008, 9:461.

```
library(networkBMA)
library(RLowPCor)
##load DREAM4 size100_1 datasets
data(dream4)
data.exp<-dream4ts10[[1]][,-c(1:2)]
genes<-colnames(data.exp)
ref.edge<-dream4gold10[[1]]
ref.adj<-edgelist2adjmatrix(ref.edge,genes)
inf.cor<-abs(cor(data.exp))
diag(inf.cor)<-0
table.cor<-table.evaluate(inf.adj = inf.cor,ref.adj = ref.adj)
head(table.cor)</pre>
```

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