Class 11: Alphafold

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Here we read the results from Alphafold and try to interpret all the models and quality score metrics:

```
pth <- "dimer_23119/"
pdb.files <- list.files(path = pth, full.names = TRUE, pattern = ".pdb")</pre>
```

Align and superpose all these models

```
file.exists(pdb.files)
```

[1] TRUE TRUE TRUE TRUE TRUE

```
pdbs <- pdbaln(pdb.files, fit = TRUE, exefile = "msa")</pre>
```

```
Reading PDB files:
```

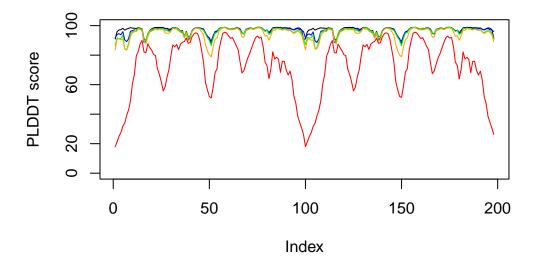
```
dimer_23119/dimer_23119_unrelaxed_rank_001_alphafold2_multimer_v3_model_2_seed_000.pdb dimer_23119/dimer_23119_unrelaxed_rank_002_alphafold2_multimer_v3_model_4_seed_000.pdb dimer_23119/dimer_23119_unrelaxed_rank_003_alphafold2_multimer_v3_model_1_seed_000.pdb dimer_23119/dimer_23119_unrelaxed_rank_004_alphafold2_multimer_v3_model_5_seed_000.pdb dimer_23119/dimer_23119_unrelaxed_rank_005_alphafold2_multimer_v3_model_3_seed_000.pdb .....
```

Extracting sequences

```
pdb/seq: 1 name: dimer_23119/dimer_23119_unrelaxed_rank_001_alphafold2_multimer_v3_model_2 pdb/seq: 2 name: dimer_23119/dimer_23119_unrelaxed_rank_002_alphafold2_multimer_v3_model_4 pdb/seq: 3 name: dimer_23119/dimer_23119_unrelaxed_rank_003_alphafold2_multimer_v3_model_1 pdb/seq: 4 name: dimer_23119/dimer_23119_unrelaxed_rank_004_alphafold2_multimer_v3_model_5 pdb/seq: 5 name: dimer_23119/dimer_23119_unrelaxed_rank_005_alphafold2_multimer_v3_model_3
```

```
library(bio3dview)
# view.pdbs(pdbs)
```

```
plot(pdbs$b[1,], typ="l", ylim=c(0,100), ylab="PLDDT score")
lines(pdbs$b[2,], typ="l", col="blue")
lines(pdbs$b[3,], typ="l", col="green")
lines(pdbs$b[4,], typ="l", col="orange")
lines(pdbs$b[5,], typ="l", col="red")
```



```
pdbs$sse
```

NULL

Score Residue conservation from alignment file

Alphafold returns it's large alignment file used for analysis. Here we read this file and score conservation per position

[1] "dimer_23119/dimer_23119.a3m"

```
aln <- read.fasta(aln_file[1], to.upper = TRUE)</pre>
```

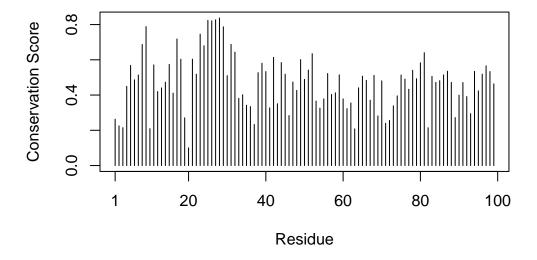
```
[1] " ** Duplicated sequence id's: 101 **"
[2] " ** Duplicated sequence id's: 101 **"
```

dim(aln\$ali)

[1] 5378 132

```
sim <- conserv(aln)</pre>
```

```
plotb3(sim[1:99],
     ylab="Conservation Score")
```



```
con <- consensus(aln, cutoff = 0.9)
con$seq</pre>
```

The sequence above shows the conserved residues which are D, T, G, and A.

Predicting Alignment Error for Domains

```
library(jsonlite)
# Listing of all PAE JSON files
pae_files <- list.files(path=pth,</pre>
                         pattern=".*model.*\\.json",
                         full.names = TRUE)
pae1 <- read_json(pae_files[1],simplifyVector = TRUE)</pre>
pae5 <- read_json(pae_files[5],simplifyVector = TRUE)</pre>
attributes(pae1)
$names
               "max_pae" "pae"
[1] "plddt"
                                    "ptm"
                                               "iptm"
# Per-residue pLDDT scores
# same as B-factor of PDB..
head(pae1$plddt)
```

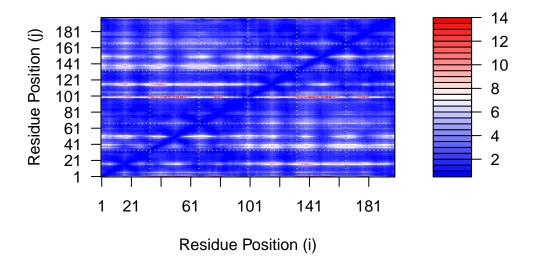
[1] 90.88 95.88 97.06 97.25 98.19 96.94

pae1\$max_pae

[1] 13.86719

```
pae5$max_pae
```

[1] 30



Heatmap of RMSD values

```
rd <- rmsd(pdbs, fit=T)
```

Warning in rmsd(pdbs, fit = T): No indices provided, using the 198 non NA positions

range(rd)

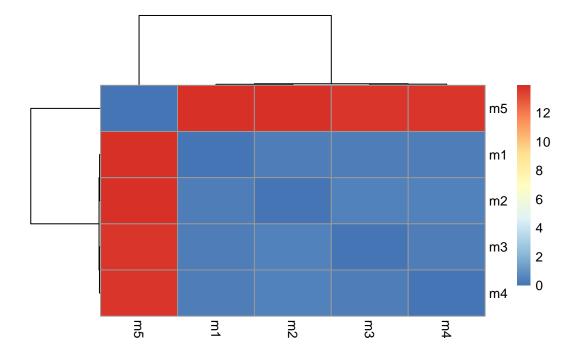
[1] 0.000 13.904

```
library(pheatmap)

colnames(rd) <- paste0("m", 1:5)

rownames(rd) <- paste0("m", 1:5)

pheatmap(rd)</pre>
```



The heatmap shows that models 1, 2, 3, and 4 are most similar to each other, while model 5 is the most different to all of the other models.