

Class 18: Pertussis Mini-project

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Background

Pertussis (a.k.a. whooping cough) is a common lung infection caused by the bacteria *B. Pertussis*.

The CDC tracks cases of Pertussis in the US: <https://tinyurl.com/pertussiscdc>

Examining cases of Pertussis by year

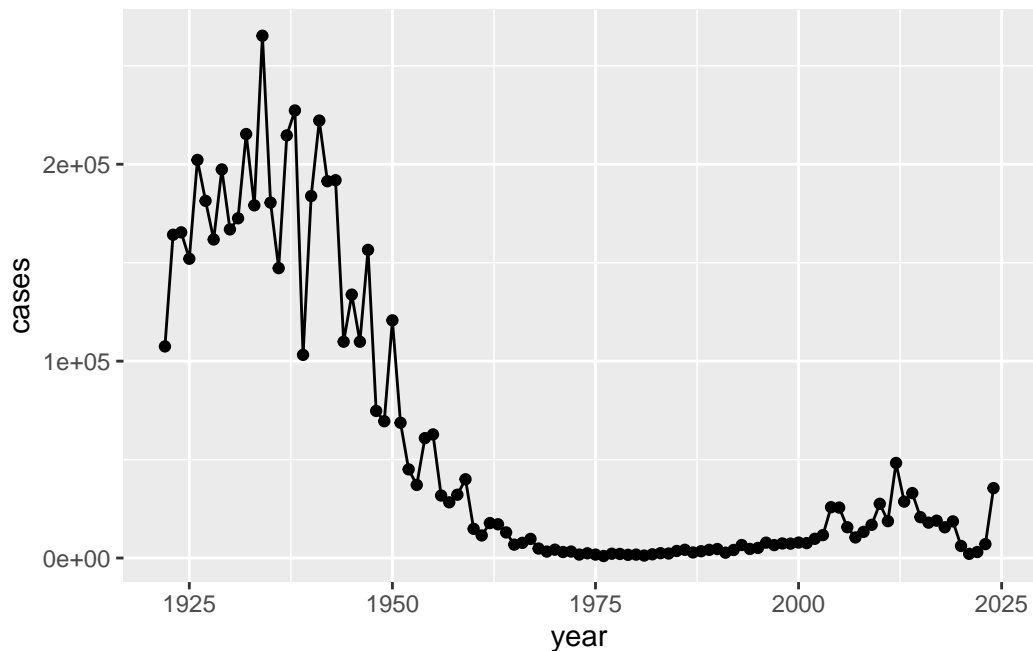
We can use the **datapasta** package to scrape case numbers from the CDC website.

Q1. With the help of the R “addin” package **datapasta** assign the CDC pertussis case number data to a data frame called `cdc` and use **ggplot** to make a plot of cases numbers over time.

```
library(ggplot2)

cases <- ggplot(cdc) +
  aes(year, cases) +
  geom_line() +
  geom_point()

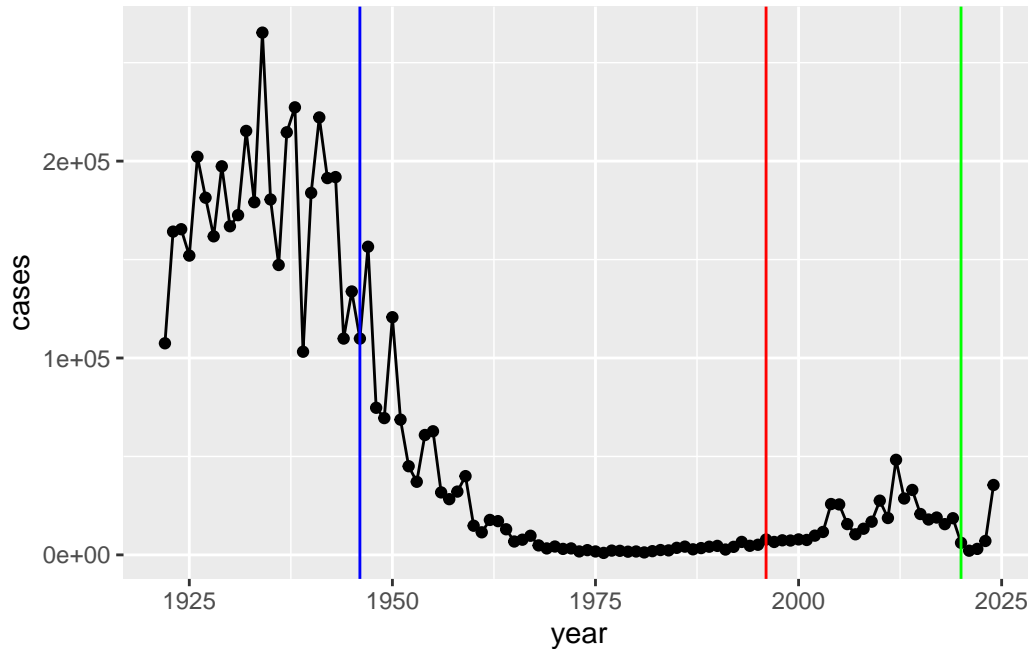
cases
```



Q2. Add some key time points in our history of interaction with Pertussis. These include wP roll-out (the first vaccine) in 1946 and the switch to aP in 1996.

We can use `geom_vline()` for this.

```
cases +
  geom_vline(xintercept = 1946, col="blue") +
  geom_vline(xintercept = 1996, col="red") +
  geom_vline(xintercept = 2020, col="green")
```



Q3. Describe what happened after the introduction of the aP vaccine? Do you have a possible explanation for the observed trend?

The wP vaccine was very effective at reducing the number of Pertussis cases. After the aP vaccine, the number of cases increased. This may be due to evolution of the bacteria or a reduced number of people getting the vaccinations. The immune protection from the aP vaccine may also fade faster than the wP vaccine.

Mounting evidence suggests that the newer **aP** is less effective over the long term than the older **wP** vaccine that is replaced. In other words, vaccine protection wanes more rapidly with aP than with wP.

Enter the CMI-PB Project

CMI-PB (Computational Models of Immunity - Pertussis boost) major goal is to investigate how the immune system responds differently with aP vs wP vaccinated individuals and be able to predict this at an early stage.

CMI-PB makes all their collected data freely available and they store it in a database composed of different tables. Here we will access a few of these.

We can use the **jsonlite** package to read this data.

```
library(jsonlite)

subject <- read_json("https://www.cmi-pb.org/api/v5_1/subject",
                     simplifyVector = TRUE)

head(subject)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female	Not Hispanic or Latino	White
2	2	wP	Female	Not Hispanic or Latino	White
3	3	wP	Female	Unknown	White
4	4	wP	Male	Not Hispanic or Latino	Asian
5	5	wP	Male	Not Hispanic or Latino	Asian
6	6	wP	Female	Not Hispanic or Latino	White

	year_of_birth	date_of_boost	dataset
1	1986-01-01	2016-09-12	2020_dataset
2	1968-01-01	2019-01-28	2020_dataset
3	1983-01-01	2016-10-10	2020_dataset
4	1988-01-01	2016-08-29	2020_dataset
5	1991-01-01	2016-08-29	2020_dataset
6	1988-01-01	2016-10-10	2020_dataset

Q. How many subjects (i.e. enrolled people) are there in this dataset?

```
nrow(subject)
```

```
[1] 172
```

Q4. How many aP and wP infancy vaccinated subjects are in the dataset?

```
table(subject$infancy_vac)
```

```
aP wP
87 85
```

Q5. How many Male and Female subjects/patients are in the dataset?

```
table(subject$biological_sex)
```

Female	Male
112	60

Q6. What is the breakdown of race and biological sex (e.g. number of Asian females, White males etc...)?

```
table(subject$race, subject$biological_sex)
```

	Female	Male
American Indian/Alaska Native	0	1
Asian	32	12
Black or African American	2	3
More Than One Race	15	4
Native Hawaiian or Other Pacific Islander	1	1
Unknown or Not Reported	14	7
White	48	32

Q. Is this representative of the US population?

No, this is not representative. This is representative of the UCSD students population because the majority of the data was taken from the students.

Working with dates

```
library(lubridate)
```

Q7. Using this approach determine (i) the average age of wP individuals, (ii) the average age of aP individuals; and (iii) are they significantly different?

```
# subtract subject date of birth from today to find age in days
subject$age <- today() - ymd(subject$year_of_birth)
```

```
library(dplyr)
```

```
Attaching package: 'dplyr'
```

The following objects are masked from 'package:stats':

filter, lag

The following objects are masked from 'package:base':

intersect, setdiff, setequal, union

```
# find the age in years of the subjects in the aP vaccine group

ap <- subject %>%
  filter(infancy_vac == "aP")

round(summary(time_length(ap$age, "years")))
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
22	26	27	27	28	34

The average age of aP individuals is 27 years.

```
# find the age in years of the subjects in the wP vaccine group

wp <- subject %>%
  filter(infancy_vac == "wP")

round(summary(time_length(wp$age, "years")))
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
22	32	34	36	39	57

The average age of the wP individuals is 36 years.

Yes, the ages between the two groups are significantly different. The wP individuals are on average about 10 years older than the aP individuals.

Q8. Determine the age of all individuals at time of boost?

```
# subtract day of birth from day of boost to get age at boost

boost_age_days <- ymd(subject$date_of_boost) - ymd(subject$year_of_birth)

# convert the age into years

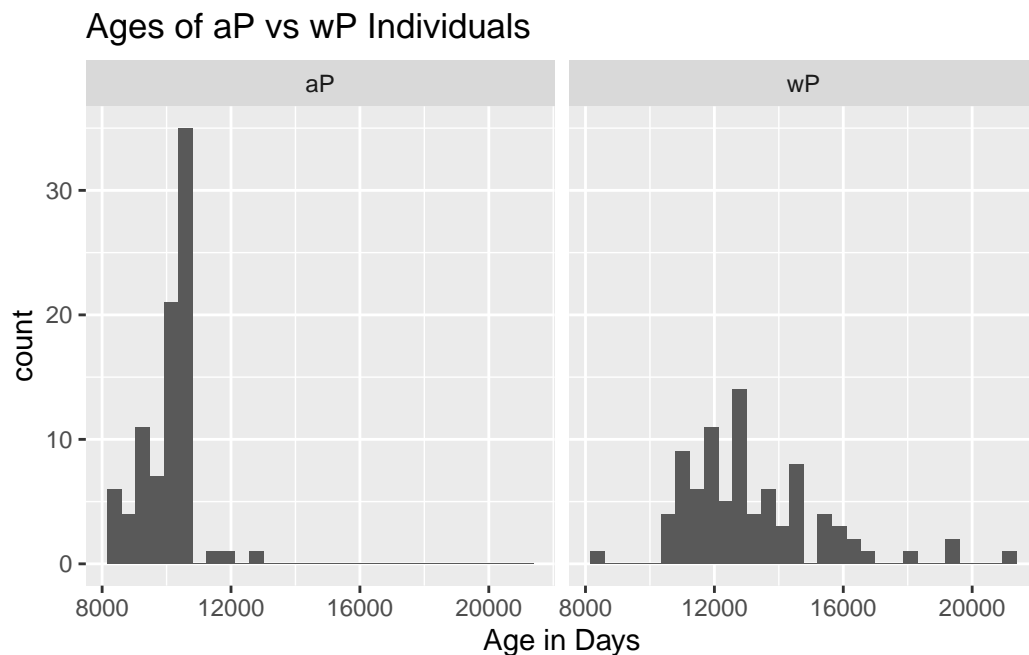
boost_age_years <- time_length(boost_age_days, 'years')
head(boost_age_years)
```

```
[1] 30.69678 51.07461 33.77413 28.65982 25.65914 28.77481
```

Q9. With the help of a faceted boxplot or histogram (see below), do you think these two groups are significantly different?

```
ggplot(subject) +
  aes(age) +
  geom_histogram() +
  facet_wrap(~infancy_vac) +
  labs(title='Ages of aP vs wP Individuals', x='Age in Days')
```

Don't know how to automatically pick scale for object of type <difftime>. Defaulting to continuous.
`stat_bin()` using `bins = 30`. Pick better value with `binwidth`.



Yes, there is a significant difference in the ages of the aP and wP groups.

Joining Multiple Tables

```
specimen <- read_json("https://www.cmi-pb.org/api/v5_1/specimen",
                      simplifyVector = TRUE)

ab_data <- read_json("https://www.cmi-pb.org/api/v5_1/plasma_ab_titer",
                     simplifyVector = TRUE)
```

Look at these data:

```
head(specimen)
```

	specimen_id	subject_id	actual_day_relative_to_boost	
1	1	1	-3	
2	2	1	1	
3	3	1	3	
4	4	1	7	
5	5	1	11	
6	6	1	32	

	planned_day_relative_to_boost	specimen_type	visit
1	0	Blood	1
2	1	Blood	2
3	3	Blood	3
4	7	Blood	4
5	14	Blood	5
6	30	Blood	6

We want to “join” these tables to get all our information together. For this, we will use the **dplyr** package and the `inner_join()` function.

Q9. Complete the code to join specimen and subject tables to make a new merged data frame containing all specimen records along with their associated subject details:

```
meta <- inner_join(subject, specimen)
```

Joining with ``by = join_by(subject_id)``


```
head(meta)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female Not Hispanic or Latino	White	
2	1	wP	Female Not Hispanic or Latino	White	
3	1	wP	Female Not Hispanic or Latino	White	
4	1	wP	Female Not Hispanic or Latino	White	
5	1	wP	Female Not Hispanic or Latino	White	
6	1	wP	Female Not Hispanic or Latino	White	

	year_of_birth	date_of_boost	dataset	age	specimen_id
1	1986-01-01	2016-09-12	2020_dataset	14394 days	1
2	1986-01-01	2016-09-12	2020_dataset	14394 days	2
3	1986-01-01	2016-09-12	2020_dataset	14394 days	3
4	1986-01-01	2016-09-12	2020_dataset	14394 days	4
5	1986-01-01	2016-09-12	2020_dataset	14394 days	5
6	1986-01-01	2016-09-12	2020_dataset	14394 days	6

	actual_day_relative_to_boost	planned_day_relative_to_boost	specimen_type
1	-3	0	Blood
2	1	1	Blood
3	3	3	Blood
4	7	7	Blood
5	11	14	Blood
6	32	30	Blood

	visit
1	1
2	2
3	3
4	4
5	5
6	6

Q10. Now using the same procedure join meta with titer data so we can further analyze this data in terms of time of visit aP/wP, male/female etc.

One more “join” to get ab_data and meta all together

```
abdata <- inner_join(ab_data, meta)
```

Joining with `by = join_by(specimen_id)`

```
head(abdata)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection	subject_id	infancy_vac	biological_sex
1	UG/ML	2.096133	1	wP	Female
2	IU/ML	29.170000	1	wP	Female
3	IU/ML	0.530000	1	wP	Female
4	IU/ML	6.205949	1	wP	Female
5	IU/ML	4.679535	1	wP	Female
6	IU/ML	2.816431	1	wP	Female

	ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
4	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
5	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset
6	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset

	age	actual_day_relative_to_boost	planned_day_relative_to_boost
1	14394 days	-3	0
2	14394 days	-3	0
3	14394 days	-3	0
4	14394 days	-3	0
5	14394 days	-3	0
6	14394 days	-3	0

	specimen_type	visit
1	Blood	1
2	Blood	1
3	Blood	1
4	Blood	1
5	Blood	1
6	Blood	1

```
dim(abdata)
```

```
[1] 61956    21
```

Q11. How many specimens (i.e. entries in abdata) do we have for each isotype?

```
table(abdata$isotype)
```

```

IgE   IgG  IgG1  IgG2  IgG3  IgG4
6698  7265 11993 12000 12000 12000

```

Q. How many different antigens are measured in the dataset?

```
table(abdata$antigen)
```

```

ACT    BETV1      DT    FELD1      FHA  FIM2/3  LOLP1      LOS Measles      OVA
1970    1970    6318    1970    6712    6318    1970    1970    1970    6318
PD1     PRN      PT     PTM    Total      TT
1970    6712    6712    1970    788     6318

```

Q12. What are the different \$dataset values in abdata and what do you notice about the number of rows for the most “recent” dataset?

```
table(abdata$dataset)
```

```

2020_dataset 2021_dataset 2022_dataset 2023_dataset
      31520         8085         7301         15050

```

The different values for \$dataset are from the years 2020-2023. The most recent dataset has half the number of rows from the 2020 dataset, but about double the rows than the 2021 and 2022 datasets.

Q. Make a boxplot of antigen levels across the whole dataset (MFI vs antigen)

```

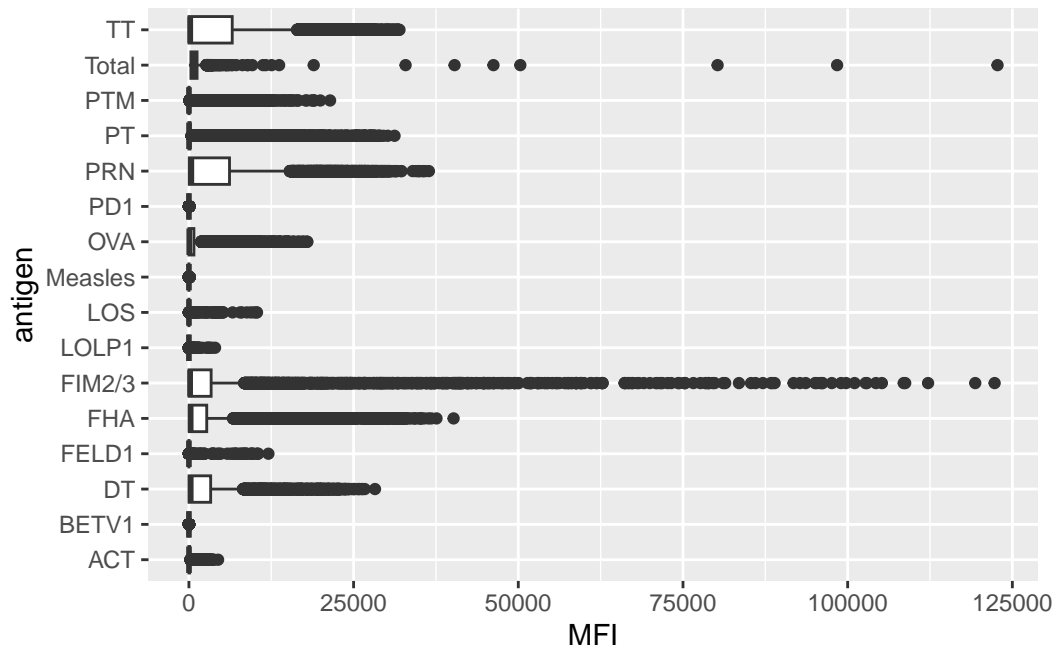
ggplot(abdata) +
  aes(MFI, antigen) +
  geom_boxplot()

```

```

Warning: Removed 1 row containing non-finite outside the scale range
(`stat_boxplot()`).

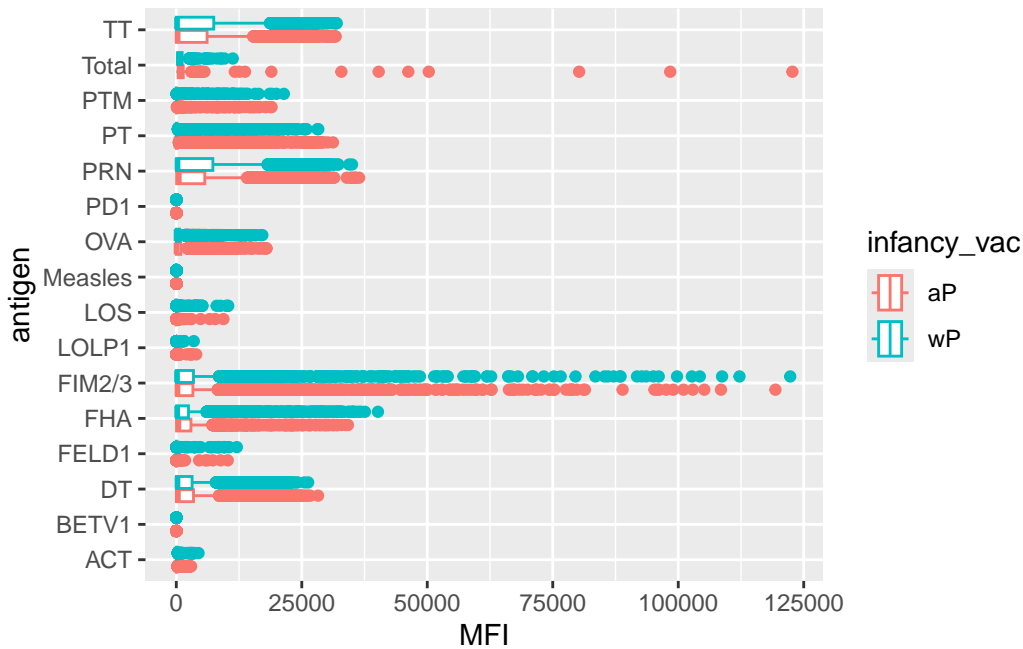
```



Q. Are there obvious differences between aP and wP values?

```
ggplot(abdata) +
  aes(MFI, antigen, col=infancy_vac) +
  geom_boxplot()
```

Warning: Removed 1 row containing non-finite outside the scale range (`stat_boxplot()`).



Focus on IgG levels

IgG is the most abundant antibody in blood. With four sub-classes (IgG1 to IgG4) crucial for long-term immunity and responding to bacterial and viral infections.

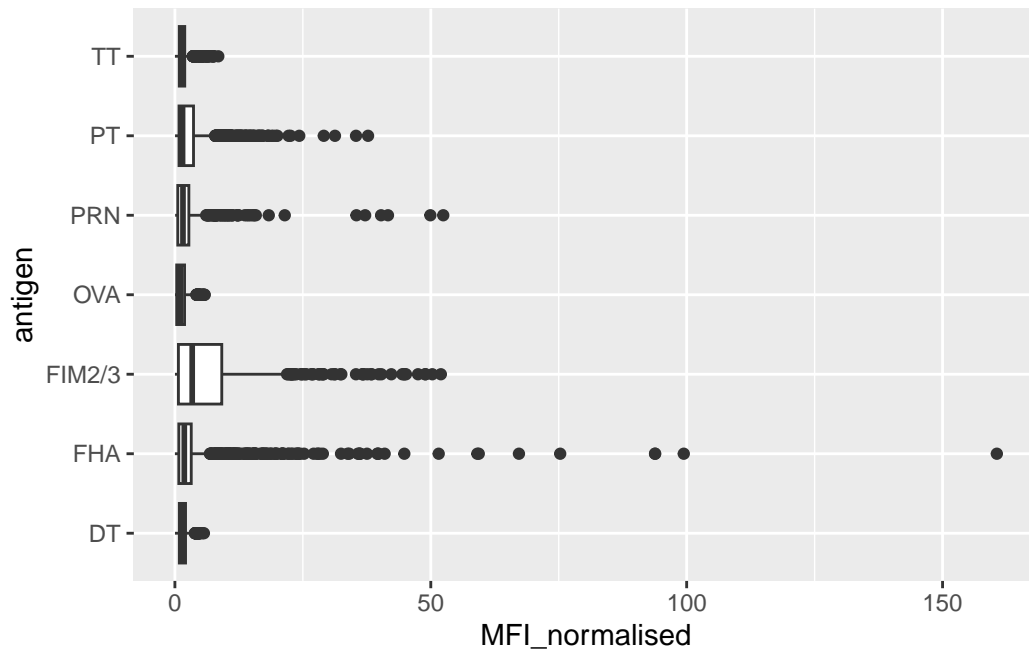
```
igg <- abdata |> filter(isotype == "IgG")
head(igg)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgG	TRUE	PT	68.56614	3.736992
2	1	IgG	TRUE	PRN	332.12718	2.602350
3	1	IgG	TRUE	FHA	1887.12263	34.050956
4	19	IgG	TRUE	PT	20.11607	1.096366
5	19	IgG	TRUE	PRN	976.67419	7.652635
6	19	IgG	TRUE	FHA	60.76626	1.096457
	unit	lower_limit_of_detection	subject_id	infancy_vac	biological_sex	
1	IU/ML	0.530000	1	wP	Female	
2	IU/ML	6.205949	1	wP	Female	
3	IU/ML	4.679535	1	wP	Female	
4	IU/ML	0.530000	3	wP	Female	
5	IU/ML	6.205949	3	wP	Female	

6	IU/ML		4.679535	3	wP	Female
		ethnicity	race	year_of_birth	date_of_boost	dataset
1	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset	
2	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset	
3	Not Hispanic or Latino	White	1986-01-01	2016-09-12	2020_dataset	
4		Unknown	White	1983-01-01	2016-10-10	2020_dataset
5		Unknown	White	1983-01-01	2016-10-10	2020_dataset
6		Unknown	White	1983-01-01	2016-10-10	2020_dataset
		age	actual_day_relative_to_boost	planned_day_relative_to_boost		
1	14394 days		-3			0
2	14394 days		-3			0
3	14394 days		-3			0
4	15490 days		-3			0
5	15490 days		-3			0
6	15490 days		-3			0
		specimen_type	visit			
1		Blood	1			
2		Blood	1			
3		Blood	1			
4		Blood	1			
5		Blood	1			
6		Blood	1			

Same boxplot of antigens as before

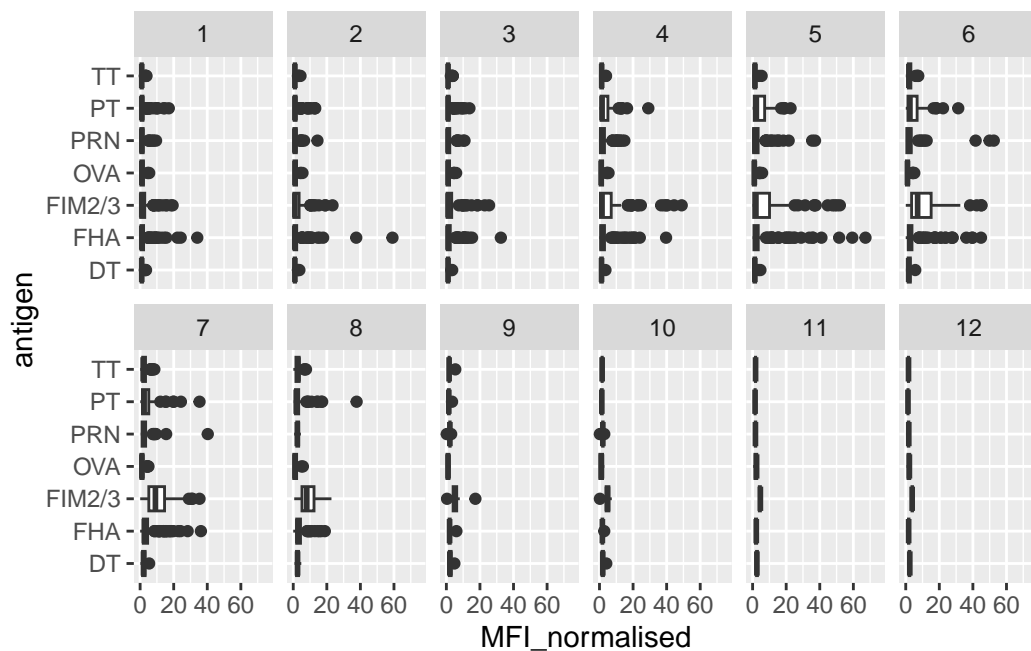
```
ggplot(igg) +
  aes(MFI_normalised, antigen) +
  geom_boxplot()
```



Q13. Complete the following code to make a summary boxplot of Ab titer levels (MFI) for all antigens:

```
ggplot(igg) +
  aes(MFI_normalised, antigen) +
  geom_boxplot() +
  xlim(0,75) +
  facet_wrap(vars(visit), nrow=2)
```

Warning: Removed 5 rows containing non-finite outside the scale range (`stat_boxplot()`).

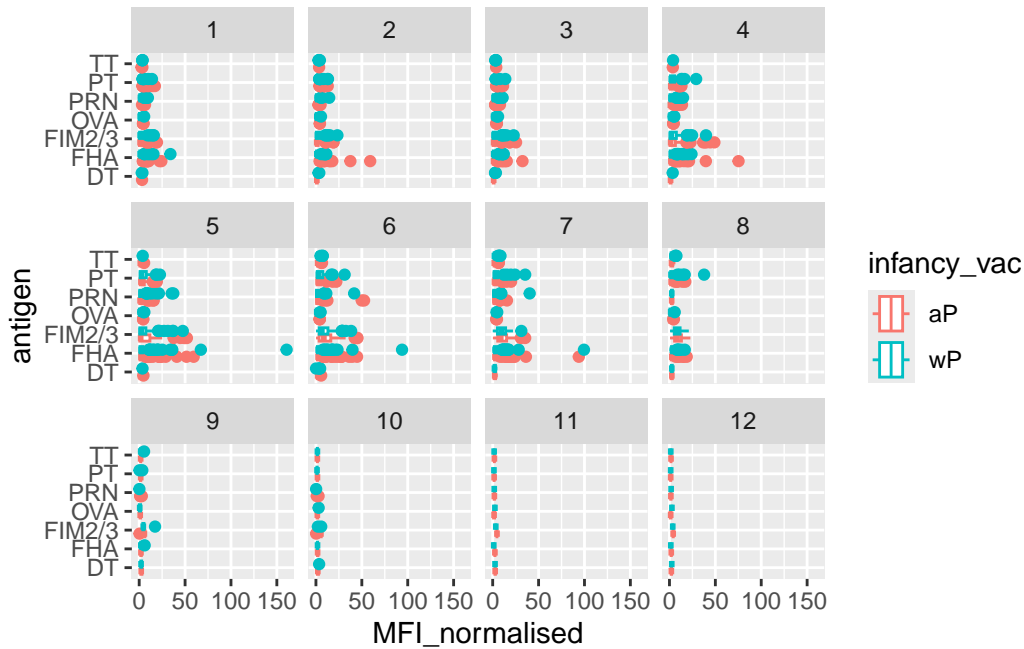


Q14. What antigens show differences in the level of IgG antibody titers recognizing them over time? Why these and not others?

PT, PRN, FIM2/3, and FHA show changes in IgG antibody titers recognizing them over time. These antigens change because these antigens are also found on the bacteria that causes Pertussis.

Look at the differences of the antigens between the aP and wP groups:

```
ggplot(igg) +
  aes(MFI_normalised, antigen, col=infancy_vac) +
  geom_boxplot() +
  facet_wrap(~visit)
```

Focus in further in just one of these antigens - let's pick **PT** (Pertussis Toxin, one of the main toxins of the bacteria) in the **2021_dataset** again for **IgG** antibody isotypes.

```
table(igg$dataset)
```

```
2020_dataset 2021_dataset 2022_dataset 2023_dataset
      1182       1617       1456       3010
```

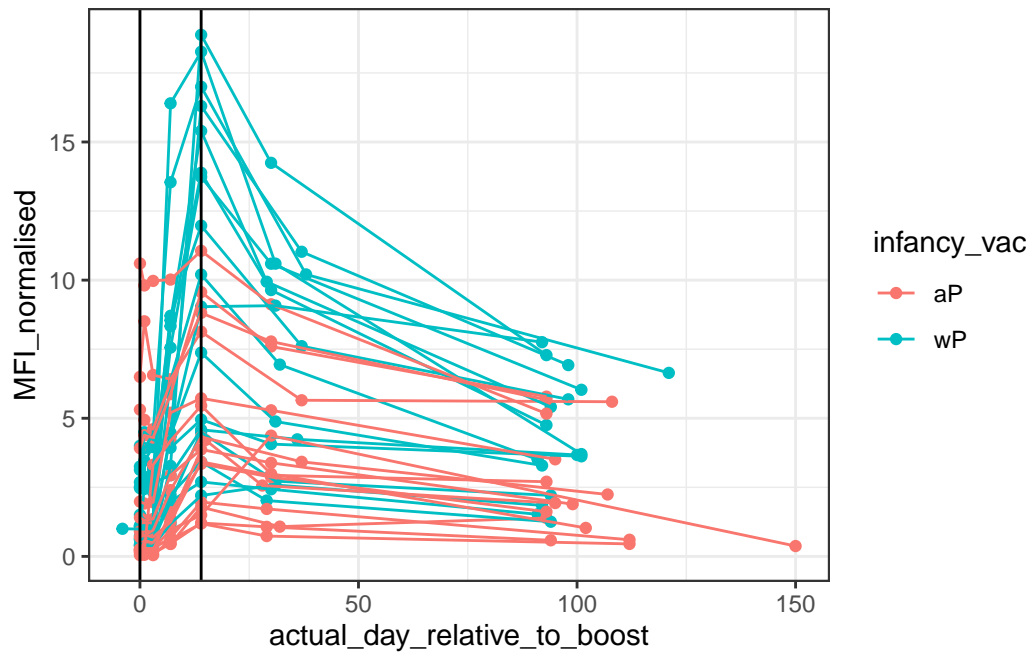
```
pt_igg <- abdata |>
  filter(isotype=="IgG", antigen=="PT", dataset=="2021_dataset")
```

```
dim(pt_igg)
```

```
[1] 231  21
```

```
ggplot(pt_igg) +
  aes(actual_day_relative_to_boost,
       MFI_normalised,
       col=infancy_vac,
       group = subject_id) +
```

```
geom_point() +
geom_line() +
theme_bw() +
geom_vline(xintercept = 0) +
geom_vline(xintercept = 14)
```



On day 14, you get peak levels in both aP and wP individuals.