# DICOMRTTool Tutorial

January 11, 2021

## 1 DICOM RT Tool Tutorial with Open-Access Data

This notebook demonstrates the various functions and utilities available in the Dicom RT tool Python package (https://github.com/brianmanderson/Dicom\_RT\_and\_Images\_to\_Mask) by Anderson et. al. It serves as supplementary information for the Technical Paper titled: "Simple Python Module for Conversions between DICOM Images and Radiation Therapy Structures, Masks, and Prediction Arrays". This notebook works through an example of publicly available brain tumor data of T1-w/FLAIR MRI sequences and corresponding RT structure files with multiple segmented regions of interest. Full information of the publicly available brain tumor data used in this notebook can be found at: https://figshare.com/articles/dataset/Data\_from\_An\_Investigation\_of\_Machine\_Learning\_Methods\_in\_Delt radiomics\_Feature\_Analysis/9943334. This notebook was written for easy accessibility for beginners to Python programming, medical imaging, and computational analysis. It should take no more than 10-15 minutes to run in it's entirety from scratch. The notebook generates about 10 GB worth of files, so ensure you have adequate space to run it.

The notebook covers the following topics (click to go to section): 1. Getting the data 2. Reading in DICOM and RT struct files and converting to numpy array format 3. Saving arrays to nifti format and reloading them 4. Saving and loading numpy array files 5. Calculating radiomic features 6. Predictions To RT-Structure Example

The notebook assumes you have the following nested directory structure after running cells that download necessary data:

```
[]: """

Top-level directory/

DICOMRTTool_manuscript.ipynb

Example_Data/ <- Generated when you run the cells below

| Image_Data/
| Structure/ <- These correspond to the Pre-RT scans

T1/
| Patient number/
| RT Struc file (.dcm)

T2FLAIR/
| Patient number/
| RT Struc file (.dcm)

| T1/
| Post1/
| Patient number/
```

```
DICOM image files (.dcm)
               Post2/
                   Patient number/
                       DICOM image files (.dcm)
               Pre/
                   Patient number/
                       DICOM image files (.dcm) <- The images we care about
           T2FLAIR/
               Post1/
                   Patient number/
                       DICOM image files (.dcm)
               Post2/
                   Patient number/
                       DICOM image files (.dcm)
               Pre/
                   Patient number/
                       DICOM image files (.dcm) <- The images we care about
  Data.zip <- Generated when you run the cells below, downloaded Figshare file
  Nifti_Data/ \leftarrow Generated \ when \ you \ run \ the \ cells \ below
     Image.nii
     \mathit{Mask.nii}
     MRN\_Path\_To\_Iteration.xlsx
     Overall_Data_Examples_(iteration)0.nii.gz
     Overall_mask_Examples_y(iteration)0.nii.gz
  Numpy_Data/ <- Generated when you run the cells below
     image.npy
     mask.npy
  RT_Structures/ <- Generated when you run the cells below
     RS\_Test\_UID.dcm
,, ,, ,,
```

```
[1]: %%capture

# Load or install the program, %%capture supresses print statements

!pip install DicomRTTool --upgrade

from DicomRTTool.ReaderWriter import DicomReaderWriter
```

```
# importing neccessary libraries

# file mangagment
import os
import zipfile
from six.moves import urllib

# array manipulation and plotting
import numpy as np
import pandas as pd
import matplotlib.pyplot as plt
```

```
# medical image manipulation
import SimpleITK as sitk
```

#### 1.1 Part 1: Getting the data.

The RT struc files and their corresponding DICOM images can be in the same directory or different directories. Here we show a case where structure files and images are located in different directories. This is a good dataset to work with since its somewhat messy but coherent enough to show power of DICOMRTTool. Many files (pre-RT, post-RT at 2 timepoints) but only pre-RT T1 and FLAIR images have associated RT structure files. Downloading and unzipping the necessary files will take about 10 minutes on most CPUs and takes up about 8 GB of storage. One may visualize these DICOM images using a free commercially available DICOM viewer, such as Radiant (https://www.radiantviewer.com/).

```
[3]: %%time
     data_path = os.path.join('.', 'Example_Data')
     if not os.path.isdir(data_path): # create Example_data directory if it doesn't_u
      \rightarrow exist
         os.mkdir(data_path)
     url_img = "https://ndownloader.figshare.com/files/20140100" # brain scans
     filename_img = os.path.join(data_path, 'Data.zip')
     if not os.path.exists(filename_img): # if zip file doesnt exist download
         print ("Retrieving zipped images...")
         print('Estimated download time is 5 minutes...')
         urllib.request.urlretrieve(url_img, filename_img)
         print('Finished downloading!')
     else:
         print ("Zipped images already downloaded.")
     if os.path.exists(filename_img): # If we downloaded the data
         if not os.path.exists(os.path.join(data_path, 'Image_Data')): # and it_
      \hookrightarrow hasn't been unzipped
             print ("Unzipping images...")
             print('Estimated unzip time is 2 minutes')
             z = zipfile.ZipFile(filename_img)
             z.extractall(data_path)
             print ("Done unzipping images.")
     print("All required files downloaded and unzipped!") # print when done
```

```
Zipped images already downloaded.
All required files downloaded and unzipped!
Wall time: 1.13 ms
```

```
[4]: def display_slices(image, mask, skip=1):
```

```
Displays a series of slices in z-direction that contains the segmented_
\rightarrow regions of interest.
   Ensures all contours are displayed in consistent and different colors.
       Parameters:
           image (array-like): Numpy array of image.
           mask (array-like): Numpy array of mask.
           skip (int): Only print every nth slice, i.e. if 3 only print every
\hookrightarrow 3rd slice, default 1.
       Returns:
           None (series of in-line plots).
   11 11 11
   slice_locations = np.unique(np.where(mask != 0)[0]) # get indexes for where
\rightarrow there is a contour present
   slice_start = slice_locations[0] # first slice of contour
   slice_end = slice_locations[len(slice_locations)-1] # last slice of contour
   counter = 1
   for img_arr, contour_arr in zip(image[slice_start:slice_end+1],__
→mask[slice_start:slice_end+1]): # plot the slices with contours overlayed_
\rightarrow ontop
       if counter % skip == 0: # if current slice is divisible by desired skip,
\rightarrow amount
           masked_contour_arr = np.ma.masked_where(contour_arr == 0,_
→contour arr)
           plt.imshow(img_arr, cmap='gray', interpolation='none')
           plt.imshow(masked_contour_arr, cmap='cool', interpolation='none', __
→alpha=0.5, vmin = 1, vmax = np.amax(mask)) # vmax is set as total number of
→contours so same colors can be displayed for each slice
           plt.show()
       counter += 1
```

# 1.2 Part 2: Reading in DICOM and RT struct files and converting to numpy array format.

The principal on which this set of tools operates on is based on the DicomReaderWriter object. It is instantiated with the contours of interest (and associations) and can then be used to create numpy arrays of images and masks of the format [slices, width, height].

The following code logic is used to demonstrate searching a path and returning indices for matched structures and images (by UID) for arbitrary directory structures (DICOM image files and RT Struct files not in the same folder). If all necessary structure files are in the same folder as the corresponding images (by UID), one can alternatively use an os.walk through directories of interest and call DicomReaderWriter each time a folder is discovered. For example, I normally use a folder structure MRN -> date of image (pre,mid,post-RT) -> type of scan (MRI, CT, etc.) -> files (DICOM images + RT Struct). However, this approach calls the DicomReaderWriter iteratively,

which can be computationally taxing.

```
[5]: DICOM path = os.path.join('.', 'Example Data', 'Image Data') # folder where
      \rightarrow downloaded data was stored
     print(DICOM_path)
    .\Example_Data\Image_Data
    This will walk through all of the folders, and using SimpleITK, will separate them based on Se-
    riesInstanceUIDs.
[6]: %%time
     Dicom_reader = DicomReaderWriter(description='Examples', arg_max=True)
     print('Estimated 30 seconds, depending on number of cores present in your ⊔
      Dicom reader.walk through folders(DICOM path) # need to define in order to use,
      \rightarrow all roi method
    Estimated 30 seconds, depending on number of cores present in your computer
    Loading from .\Example_Data\Image_Data\Structure\T1\006
    Loading from .\Example_Data\Image_Data\Structure\T1\001
    Loading from .\Example_Data\Image Data\Structure\T1\004Loading from
    .\Example_Data\Image_Data\Structure\T1\005
    Loading from .\Example_Data\Image_Data\Structure\T1\003
    Loading from .\Example Data\Image Data\Structure\T1\002
    Loading from .\Example_Data\Image_Data\Structure\T1\007
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    .\Example_Data\Image_Data\Structure\T1\009
    Loading from .\Example_Data\Image_Data\Structure\T1\011
    Loading from .\Example_Data\Image_Data\Structure\T1\012
    Loading from .\Example_Data\Image_Data\Structure\T2Flair\001
    Loading from .\Example_Data\Image_Data\Structure\T2Flair\002
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    Loading from .\Example_Data\Image_Data\Structure\T2Flair\008
    Loading from .\Example Data\Image Data\Structure\T2Flair\009Loading from
    .\Example_Data\Image_Data\Structure\T2Flair\010
    Loading from .\Example Data\Image Data\Structure\T2Flair\011Loading from
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.\Example\_Data\Image\_Data\Structure\T2Flair\012

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Loading from .\Example_Data\Image_Data\T1\002
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Loading from .\Example Data\Image Data\T2Flair\Pre\010
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Loading from .\Example_Data\Image_Data\T2Flair\Pre\012
Compiling dictionaries together...
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.\Example_Data\Image_Data\T2Flair\Pre\002
Index 1, description Ax T2Flair Propeller at
.\Example_Data\Image_Data\T2Flair\Post2\004
Index 2, description Ax T2Flair Propeller at
.\Example_Data\Image_Data\T2Flair\Pre\006
Index 3, description Ax T2Flair Propeller at
.\Example_Data\Image_Data\T2Flair\Post1\005
Index 4, description Ax T2Flair Propeller at
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Index 29, description Ax T2Flair Propeller at
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Index 30, description Ax T2Flair Propeller at
.\Example Data\Image Data\T2Flair\Pre\001
Index 31, description ax T1 3D 1MM fSPGR +C
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Index 32, description ax T1 3D 1MM fSPGR +C
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Index 33, description ax T1 3D 1MM fSPGR +C at
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.\Example_Data\Image_Data\T1\Post1\007
Index 45, description ax T1 3D 1MM fSPGR +C
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Index 46, description ax T1 3D 1MM fSPGR +C
.\Example_Data\Image_Data\T1\Pre\007
Index 47, description Ax T2Flair Propeller at
.\Example_Data\Image_Data\T2Flair\Post2\001
Index 48, description ax T1 3D 1MM fSPGR +C
.\Example Data\Image Data\T1\Post1\002
Index 49, description ax T1 3D 1MM fSPGR +C
.\Example Data\Image Data\T1\Pre\004
Index 50, description ax T1 3D 1MM fSPGR +C
.\Example_Data\Image_Data\T1\Post1\003
Index 51, description ax T1 3D 1MM fSPGR +C
. \verb|\Example_Data\\| Image_Data\\| T1\\| Post2\\| 004
Index 52, description ax T1 3D 1MM fSPGR +C
.\Example_Data\Image_Data\T1\Post2\001
Index 53, description ax T1 3D 1MM fSPGR +C
.\Example_Data\Image_Data\T1\Post1\004
Index 54, description Ax T2Flair Propeller at
.\Example_Data\Image_Data\T2Flair\Pre\011
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Index 55, description Ax T2Flair Propeller at

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    Index 56, description ax T1 3D 1MM +c at .\Example_Data\Image_Data\T1\Post2\011
    Index 57, description ax T1 3D 1MM +c at .\Example_Data\Image_Data\T1\Post2\008
    Index 58, description ax T1 3D 1MM +c at .\Example_Data\Image_Data\T1\Pre\010
    Index 59, description ax T1 3D 1MM +c at .\Example Data\Image Data\T1\Post2\010
    Index 60, description ax T1 3D 1MM +c at .\Example Data\Image Data\T1\Pre\011
    Index 61, description ax T1 3D 1MM
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    Index 62, description ax T1 3D 1MM
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    Index 65, description ax T1 3D 1MM
                                       +c at .\Example_Data\Image_Data\T1\Pre\012
    Index 66, description ax T1 3D 1MM +c at .\Example_Data\Image_Data\T1\Post2\009
    Index 67, description ax T1 3D 1MM +c at .\Example_Data\Image_Data\T1\Post2\012
    Index 68, description Ax T2Flair Propeller at
    .\Example_Data\Image_Data\T2Flair\Post2\002
    Index 69, description ax T1 3D 1MM fSPGR +C at
    .\Example_Data\Image_Data\T1\Post2\002
    Index 70, description Ax T2Flair Propeller at
    .\Example_Data\Image_Data\T2Flair\Post2\003
    Index 71, description ax T1 3D 1MM fSPGR +C at
    .\Example Data\Image Data\T1\Post2\003
    Index 72, description None at None
    Index 73, description None at None
    74 unique series IDs were found. Default is index 0, to change use
    set_index(index)
    Wall time: 3.5 s
[7]: all_rois = Dicom_reader.return_rois(print_rois=True) # Return a list of all_
     →rois present, and print them
    The following ROIs were found
    rttempglioma
    exprttempglioma
    brainstem
    dose 500[cgy]
    dose 1000[cgy]
    dose 1200[cgy]
    gtvplus2
    expltparrecgliom
    ltparrecglioma
    expltfrontrecao
    ltfrontrecao
    body
    expltfrparrecgbm
    ltfrparrecgbm
    explttempglioma
    lttempglioma
    exprtfrontrecgbm
```

```
rtfrontrecgbm
expinfrttemprecg
infrttempgbm
dose 2400[cgy]
expltfrontgbm
ltfrontgbm
exprttemprecglio
rttemprecglioma
rtfrontrecglioma
exprtfrontrecgli
brainstem1
eye, left
eye, right
chiasm
lens, left
lens, right
optic nerve, rig
optic nerve, lef
dose 2500[cgy]
exprttemprecgbm
rttemprecgbm
exprtfrparresxn
right_front_par_
abv
abv_roi
```

As we can see, these ROIs correspond to a variety of structures. In particular, we can see many GBM and glioma structures. Note GBM denotes glioblastoma multiforme (a high grade glioma).

Contours of brainstem1 are located:

- .\Example\_Data\Image\_Data\Structure\T1\001\RS.CA1756\_T13D.dcm
- .\Example\_Data\Image\_Data\Structure\T1\011\RS.GF6065\_T13D.dcm
- .\Example\_Data\Image\_Data\Structure\T2Flair\001\RS.CA1756\_T2Flair.dcm
- .\Example\_Data\Image\_Data\Structure\T2Flair\011\RS.GF6065\_T2Flairdcm.dcm
- .\Example\_Data\Image\_Data\T1\001\RS.CA1756\_T13D.dcm
- [9]: Dicom\_reader.which\_indexes\_have\_all\_rois() # Check to see which indexes have\_\( \to \alpha \) all of the rois we want

  # Since we haven't defined anything yet, it prompts you to input a list of \( \to \cdot \) contour names

You need to first define what ROIs you want, please use .set\_contour\_names\_and\_associations(roi\_list)

From these ROIs, we will look for those that describe the following regions of interest: tumor

(glioblastoma multiforme only) and high-dose area of radiation therapy.

```
[10]: Contour_Names = ['tumor', 'high_dose']
      # Associations work as {'variant_name': 'desired_name'}
      associations = {'dose 1000[cgy]': 'high_dose', 'dose 1200[cgy]': 'high_dose', \#_{\square}
       →new high dose ROI through association
                     'exprtfrontrecgbm': 'tumor', 'rtfrontrecgbm': 'tumor',
       →'expltfrontgbm': 'tumor', # associating gbms to tumor
                      'ltfrontgbm': 'tumor', 'infrttempgbm': 'tumor', 'rttemprecgbm': u
       'exprttemprecgbm': 'tumor', 'expltfrparrecgbm': 'tumor',
       →'ltfrparrecgbm': 'tumor'}
[11]: Dicom_reader.set_contour_names_and_associations(Contour_Names=Contour_Names,_
       →associations=associations)
     Lacking ['tumor'] in index 0, location
     .\Example_Data\Image_Data\T2Flair\Pre\002. Found ['rttempglioma',
     'exprttempglioma', 'brainstem', 'dose 500[cgy]', 'dose 1000[cgy]', 'dose
     1200[cgy]', 'gtvplus2']
     Lacking ['tumor', 'high_dose'] in index 1, location
     .\Example Data\Image Data\T2Flair\Post2\004. Found []
     Lacking ['tumor'] in index 2, location
     .\Example Data\Image Data\T2Flair\Pre\006. Found ['brainstem', 'dose 500[cgy]',
     'dose 1200[cgy]', 'dose 1000[cgy]', 'expltparrecgliom', 'ltparrecglioma',
     'gtvplus2']
     Lacking ['tumor', 'high_dose'] in index 3, location
     .\Example_Data\Image_Data\T2Flair\Post1\005. Found []
     Lacking ['tumor', 'high_dose'] in index 4, location
     .\Example_Data\Image_Data\T2Flair\Post1\006. Found []
     Lacking ['tumor'] in index 5, location
     .\Example_Data\Image_Data\T2Flair\Pre\007. Found ['brainstem', 'dose 500[cgy]',
     'dose 1000[cgy]', 'dose 1200[cgy]', 'expltfrontrecao', 'ltfrontrecao', 'body',
     'gtvplus2']
     Lacking ['tumor', 'high dose'] in index 6, location
     .\Example_Data\Image_Data\T2Flair\Post2\006. Found []
     Lacking ['tumor', 'high dose'] in index 8, location
     .\Example_Data\Image_Data\T2Flair\Post2\010. Found []
     Lacking ['tumor', 'high_dose'] in index 9, location
     .\Example_Data\Image_Data\T2Flair\Post1\010. Found []
     Lacking ['tumor'] in index 10, location
     .\Example_Data\Image_Data\T2Flair\Pre\004. Found ['brainstem', 'dose 1000[cgy]',
     'dose 1200[cgy]', 'dose 500[cgy]', 'explttempglioma', 'lttempglioma',
     'gtvplus2']
     Lacking ['tumor', 'high_dose'] in index 12, location
     .\Example_Data\Image_Data\T2Flair\Post1\002. Found []
     Lacking ['tumor', 'high_dose'] in index 13, location
     .\Example_Data\Image_Data\T2Flair\Post2\005. Found []
```

```
Lacking ['tumor', 'high_dose'] in index 14, location
.\Example_Data\Image_Data\T2Flair\Post1\009. Found []
Lacking ['tumor', 'high_dose'] in index 15, location
.\Example_Data\Image_Data\T2Flair\Post2\008. Found []
Lacking ['tumor', 'high dose'] in index 16, location
.\Example_Data\Image_Data\T2Flair\Post2\009. Found []
Lacking ['tumor', 'high dose'] in index 17, location
.\Example_Data\Image_Data\T2Flair\Post2\012. Found []
Lacking ['tumor', 'high_dose'] in index 19, location
.\Example_Data\Image_Data\T2Flair\Post2\011. Found []
Lacking ['tumor', 'high_dose'] in index 20, location
.\Example_Data\Image_Data\T2Flair\Post1\011. Found []
Lacking ['tumor', 'high_dose'] in index 21, location
.\Example_Data\Image_Data\T2Flair\Post1\004. Found []
Lacking ['tumor', 'high_dose'] in index 22, location
.\Example_Data\Image_Data\T2Flair\Post1\007. Found []
Lacking ['tumor', 'high_dose'] in index 23, location
.\Example_Data\Image_Data\T2Flair\Post2\007. Found []
Lacking ['tumor', 'high_dose'] in index 24, location
.\Example Data\Image Data\T2Flair\Post1\003. Found []
Lacking ['tumor', 'high_dose'] in index 25, location
.\Example Data\Image Data\T2Flair\Post1\012. Found []
Lacking ['tumor', 'high_dose'] in index 26, location
.\Example_Data\Image_Data\T2Flair\Post1\008. Found []
Lacking ['tumor', 'high_dose'] in index 27, location
.\Example_Data\Image_Data\T2Flair\Post1\001. Found []
Lacking ['tumor'] in index 29, location
.\Example_Data\Image_Data\T2Flair\Pre\008. Found ['brainstem', 'dose 1000[cgy]',
'dose 1200[cgy]', 'dose 500[cgy]', 'exprttemprecglio', 'rttemprecglioma',
'gtvplus2']
Lacking ['tumor'] in index 30, location
.\Example_Data\Image_Data\T2Flair\Pre\001. Found ['rtfrontrecglioma',
'exprtfrontrecgli', 'brainstem1', 'dose 1000[cgy]', 'dose 1200[cgy]', 'dose
500[cgy]', 'gtvplus2']
Lacking ['tumor', 'high dose'] in index 32, location
.\Example_Data\Image_Data\T1\Post1\001. Found []
Lacking ['tumor'] in index 33, location .\Example Data\Image Data\T1\Pre\002.
Found ['exprttempglioma', 'rttempglioma', 'brainstem', 'dose 500[cgy]', 'dose
1000[cgy]', 'dose 1200[cgy]', 'body', 'gtvplus2']
Lacking ['tumor', 'high_dose'] in index 34, location
.\Example_Data\Image_Data\T1\Post2\005. Found []
Lacking ['tumor', 'high_dose'] in index 36, location
.\Example_Data\Image_Data\T1\Post1\008. Found []
Lacking ['tumor', 'high_dose'] in index 37, location
.\Example_Data\Image_Data\T1\Post1\006. Found []
Lacking ['tumor', 'high_dose'] in index 38, location
.\Example_Data\Image_Data\T1\Post2\006. Found []
Lacking ['tumor'] in index 39, location .\Example_Data\Image_Data\T1\Pre\008.
```

```
Found ['eye, left', 'eye, right', 'brainstem', 'chiasm', 'lens, left', 'lens,
right', 'rttemprecglioma', 'exprttemprecglio', 'dose 500[cgy]', 'dose
1000[cgy]', 'dose 1200[cgy]', 'gtvplus2', 'body']
Lacking ['tumor', 'high_dose'] in index 40, location
.\Example Data\Image Data\T1\Post1\005. Found []
Lacking ['tumor'] in index 41, location .\Example_Data\Image_Data\T1\Pre\001.
Found ['exprtfrontrecgli', 'rtfrontrecglioma', 'brainstem1', 'dose 500[cgy]',
'dose 1200[cgy]', 'dose 1000[cgy]', 'body', 'gtvplus2']
Lacking ['tumor', 'high_dose'] in index 42, location
.\Example_Data\Image_Data\T1\Pre\009. Found []
Lacking ['tumor', 'high_dose'] in index 43, location
.\Example_Data\Image_Data\T1\Post1\009. Found []
Lacking ['tumor', 'high_dose'] in index 44, location
.\Example_Data\Image_Data\T1\Post1\007. Found []
Lacking ['tumor'] in index 45, location .\Example_Data\Image_Data\T1\Pre\006.
Found ['eye, left', 'lens, right', 'lens, left', 'ltparrecglioma', 'chiasm',
'optic nerve, rig', 'optic nerve, lef', 'expltparrecgliom', 'brainstem', 'eye,
right', 'dose 500[cgy]', 'dose 1000[cgy]', 'dose 1200[cgy]', 'gtvplus2']
Lacking ['tumor'] in index 46, location .\Example_Data\Image_Data\T1\Pre\007.
Found ['brainstem', 'chiasm', 'expltfrontrecao', 'eye, left', 'eye, right',
'lens, left', 'lens, right', 'ltfrontrecao', 'optic nerve, lef', 'optic nerve,
rig', 'dose 2500[cgy]', 'dose 500[cgy]', 'dose 1000[cgy]', 'dose 1200[cgy]',
'gtvplus2', 'body']
Lacking ['tumor', 'high_dose'] in index 47, location
.\Example_Data\Image_Data\T2Flair\Post2\001. Found []
Lacking ['tumor', 'high_dose'] in index 48, location
.\Example_Data\Image_Data\T1\Post1\002. Found []
Lacking ['tumor'] in index 49, location .\Example_Data\Image_Data\T1\Pre\004.
Found ['brainstem', 'dose 1000[cgy]', 'dose 1200[cgy]', 'dose 500[cgy]',
'explttempglioma', 'lttempglioma', 'gtvplus2']
Lacking ['tumor', 'high_dose'] in index 50, location
.\Example_Data\Image_Data\T1\Post1\003. Found []
Lacking ['tumor', 'high_dose'] in index 51, location
.\Example_Data\Image_Data\T1\Post2\004. Found []
Lacking ['tumor', 'high dose'] in index 52, location
.\Example Data\Image Data\T1\Post2\001. Found []
Lacking ['tumor', 'high dose'] in index 53, location
.\Example_Data\Image_Data\T1\Post1\004. Found []
Lacking ['tumor'] in index 55, location
.\Example_Data\Image_Data\T2Flair\Pre\012. Found ['brainstem', 'chiasm', 'dose
1000[cgy]', 'dose 1200[cgy]', 'dose 500[cgy]', 'exprtfrparresxn', 'gtvplus2',
'right_front_par_']
Lacking ['tumor', 'high_dose'] in index 56, location
.\Example_Data\Image_Data\T1\Post2\011. Found []
Lacking ['tumor', 'high_dose'] in index 57, location
.\Example_Data\Image_Data\T1\Post2\008. Found []
Lacking ['tumor', 'high_dose'] in index 59, location
.\Example_Data\Image_Data\T1\Post2\010. Found []
```

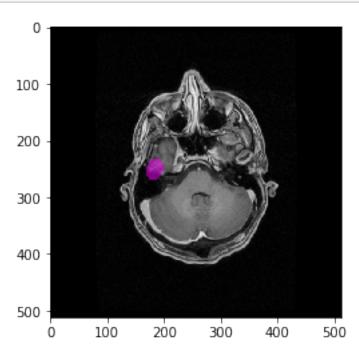
```
Lacking ['tumor', 'high_dose'] in index 61, location
     .\Example_Data\Image_Data\T1\Post1\011. Found []
     Lacking ['tumor', 'high_dose'] in index 62, location
     .\Example_Data\Image_Data\T1\Post1\010. Found []
     Lacking ['tumor', 'high dose'] in index 63, location
     .\Example_Data\Image_Data\T1\Post2\007. Found []
     Lacking ['tumor', 'high dose'] in index 64, location
     .\Example_Data\Image_Data\T1\Post1\012. Found []
     Lacking ['tumor'] in index 65, location .\Example Data\Image Data\T1\Pre\012.
     Found ['brainstem', 'chiasm', 'exprtfrparresxn', 'right_front_par_', 'body',
     'dose 500[cgy]', 'dose 1000[cgy]', 'dose 1200[cgy]', 'abv', 'abv_roi',
     'gtvplus2']
     Lacking ['tumor', 'high_dose'] in index 66, location
     .\Example_Data\Image_Data\T1\Post2\009. Found []
     Lacking ['tumor', 'high_dose'] in index 67, location
     .\Example_Data\Image_Data\T1\Post2\012. Found []
     Lacking ['tumor', 'high_dose'] in index 68, location
     .\Example_Data\Image_Data\T2Flair\Post2\002. Found []
     Lacking ['tumor', 'high_dose'] in index 69, location
     .\Example Data\Image Data\T1\Post2\002. Found []
     Lacking ['tumor', 'high dose'] in index 70, location
     .\Example Data\Image Data\T2Flair\Post2\003. Found []
     Lacking ['tumor', 'high_dose'] in index 71, location
     .\Example_Data\Image_Data\T1\Post2\003. Found []
     Note: The module is printing "Found []" because many of the scans (post-1 and post-2 RT) do
     not have associated structure files. The module recognizes these images exist (unique UIDs) but
     associated structure files cannot be located for them.
[12]: indexes = Dicom_reader.which_indexes_have_all_rois() # Check to see which_
       →indexes have all of the rois we want, now we can see indexes
     The following indexes have all ROIs present
     Index 7, located at .\Example Data\Image Data\T2Flair\Pre\009
     Index 11, located at .\Example_Data\Image_Data\T2Flair\Pre\003
     Index 18, located at .\Example_Data\Image_Data\T2Flair\Pre\010
     Index 28, located at .\Example_Data\Image_Data\T2Flair\Pre\005
     Index 31, located at .\Example_Data\Image_Data\T1\Pre\003
     Index 35, located at .\Example_Data\Image_Data\T1\Pre\005
     Index 54, located at .\Example_Data\Image_Data\T2Flair\Pre\011
     Index 58, located at .\Example_Data\Image_Data\T1\Pre\010
```

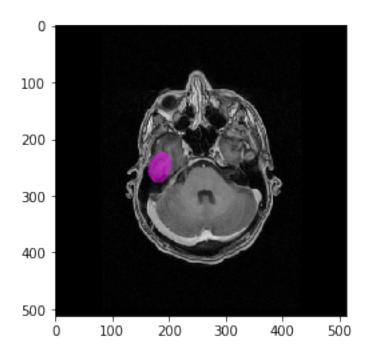
Index 60, located at .\Example\_Data\Image\_Data\T1\Pre\011

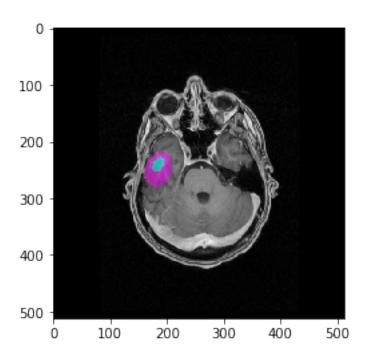
Loading images for ax T1 3D 1MM +c at
 .\Example\_Data\Image\_Data\T1\Pre\011

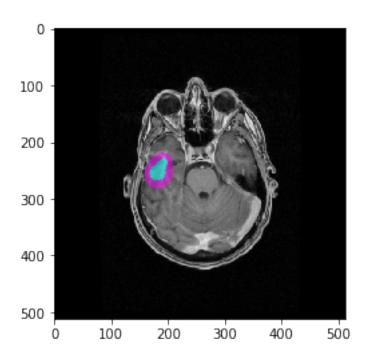
[14]: image = Dicom\_reader.ArrayDicom # image array
 mask = Dicom\_reader.mask # mask array
 dicom\_sitk\_handle = Dicom\_reader.dicom\_handle # SimpleITK image handle
 mask\_sitk\_handle = Dicom\_reader.annotation\_handle # SimpleITK mask handle

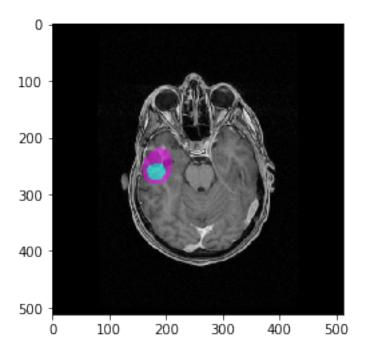
[15]: n\_slices\_skip = 4
display\_slices(image, mask, skip = n\_slices\_skip) # visualize that our\_
segmentations were succesfully convereted

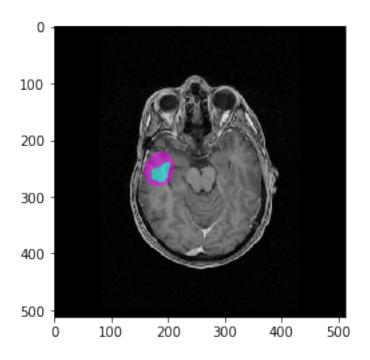


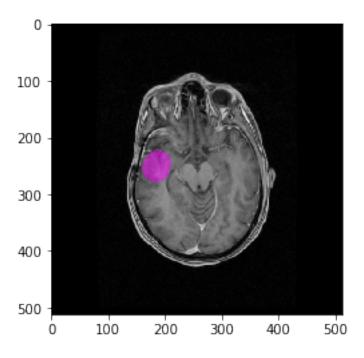


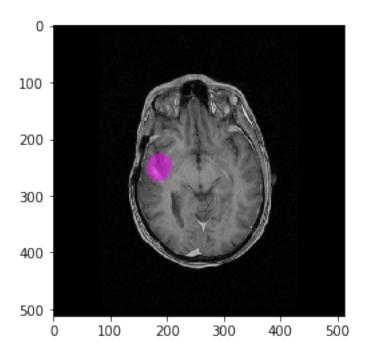


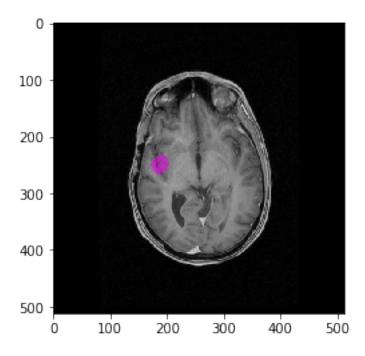












Note: Cyan color denotes tumor while magenta denotes surrounding area of high-dose radiation. Only displaying 7 slices.

#### 1.3 Part 3: Saving arrays to nifti format.

If you want to use a manual approach, you can view the nifti files easily after running get\_images\_and\_mask(). Saving files as nifti is advisable since spacing information is preserved.

```
[16]: nifti_path = os.path.join('.', 'Example_Data', 'Nifti_Data') # nifti subfolder
if not os.path.exists(nifti_path):
    os.makedirs(nifti_path)
```

```
[17]: dicom_sitk_handle = Dicom_reader.dicom_handle # SimpleITK image handle
mask_sitk_handle = Dicom_reader.annotation_handle # SimpleITK mask handle
sitk.WriteImage(dicom_sitk_handle, os.path.join(nifti_path, 'Image.nii'))
sitk.WriteImage(mask_sitk_handle, os.path.join(nifti_path, 'Mask.nii'))
```

One can also use the built in .write\_parallel attribute to generate nifti files for all relevant pairs the DicomReaderWriter object has found/generated. In this case there are 9 image/mask pairs for unique UIDs that contain all contours we are interested in. Note a corresponding log excel file in the specified output path. The nifti files are written in the following format: "Overall\_Data\_{description}\_ {iteration}.nii.gz" (image) or "Overall\_mask\_{description}\_ y{iteration}.nii.gz" (mask).

Wall time: 305 ms

We can now reload the nifti files and disaply them to check that nothing went wrong. You can inspect the other converted files by changing the numerical suffix as per the excel log file ('MRN Path To Iteration.xlsx').

```
[19]: nifti_image = sitk.ReadImage(os.path.join(nifti_path,"Overall_Data_Examples_8.

→nii.gz")) # reload image

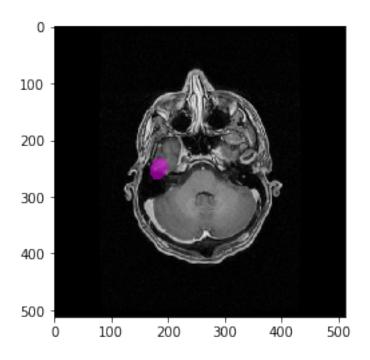
image = sitk.GetArrayFromImage(nifti_image)

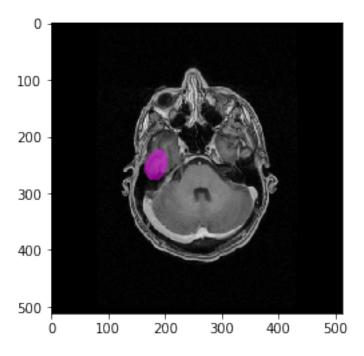
nifti_mask = sitk.ReadImage(os.path.join(nifti_path,"Overall_mask_Examples_y8.

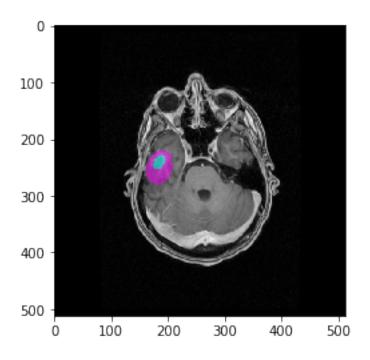
→nii.gz")) # reload mask

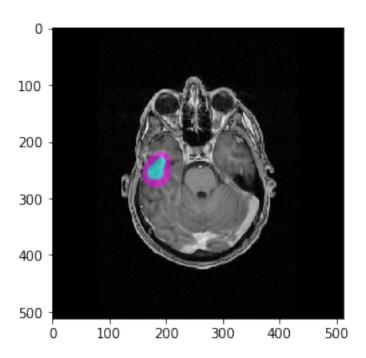
mask = sitk.GetArrayFromImage(nifti_mask)
```

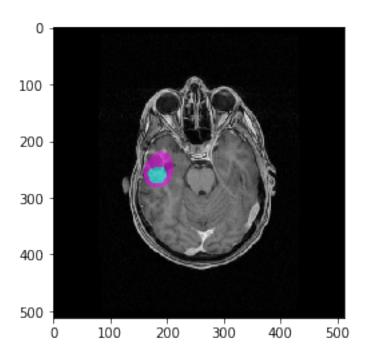
```
[20]: display_slices(image, mask, skip = n_slices_skip) # visualize that our_segmentations were succesfully convereted from nifti
```

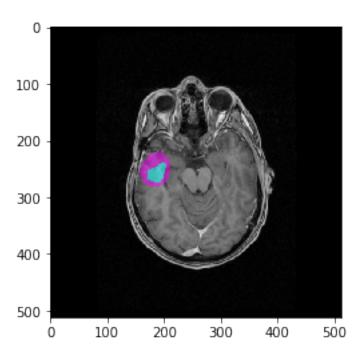


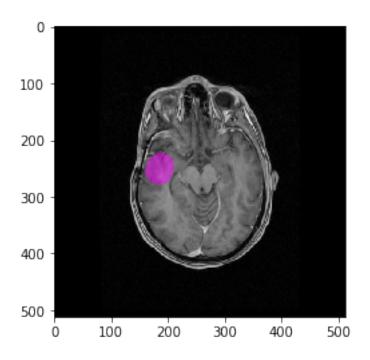


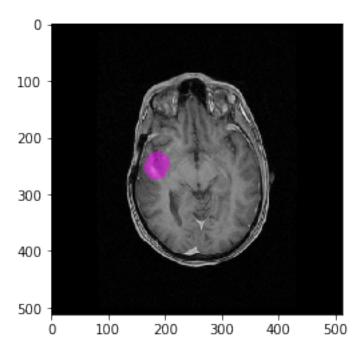


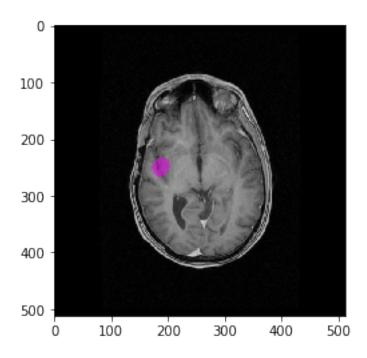












#### 1.4 Part 4: Saving and loading numpy files for later use.

Finally we can save the numpy arrays themselves to files for later use (so you don't have to reinstantiate the computationally expensive DicomReaderWriter object) and subsequently re-load the numpy arrays.

```
[21]: numpy_path = os.path.join(data_path, 'Numpy_Data') # go into numpy subfolder
    if not os.path.exists(numpy_path):
        os.makedirs(numpy_path)

[22]: np.save(os.path.join(numpy_path, 'image'), image) # save the arrays
        np.save(os.path.join(numpy_path, 'mask'), mask)

[23]: image = np.load(os.path.join(numpy_path, 'image.npy')) # load the arrays
        mask = np.load(os.path.join(numpy_path, 'mask.npy'))
```

### 1.5 Part 5: Radiomics Use-case Example.

Here we use the popular open-source radiomics library PyRadiomics (https://pyradiomics.readthedocs.io/en/latest/) to calculate radiomic features for our ROIs. In this case, we only calculate a limited number features from the tumor as an illustrative example.

```
[24]: try:
    from radiomics import featureextractor
    except:
    !pip install pyradiomics
```

```
[25]: pd.set_option('display.max_columns', None) # show all columns
[26]: %%time
      # note: need sitk images (sitk.ReadImage(nifti file)) to plug into PyRadiomics, \Box
      →preserves spacing
      ROI_index = 1 # index for tumor
      nifti_mask_tumor = sitk.BinaryThreshold(nifti_mask, lowerThreshold=ROI_index,__
       →upperThreshold=ROI_index) # select only ROI of interest
      params = {} # can edit in more params as neccessary
      extractor = featureextractor.RadiomicsFeatureExtractor(**params) # instantiate_
      → extractor with parameters
      extractor.disableAllFeatures() # in case where only want some features, can_
      → delete disable/enable lines if you want deafult
      extractor.enableFeatureClassByName('firstorder')
      extractor.enableFeatureClassByName('glcm')
      features = {} # empty dictionary
      features = extractor.execute(nifti_image, nifti_mask_tumor) # unpack results_
       → into features dictionary
      df = pd.DataFrame({k: [v] for k, v in features.items()}) # put dictionary into_
       \rightarrow a dataframe
     GLCM is symmetrical, therefore Sum Average = 2 * Joint Average, only 1 needs to
     be calculated
     Wall time: 4.06 s
[27]: df # display dataframe to inspect features
[27]:
       diagnostics_Versions_PyRadiomics diagnostics_Versions_Numpy \
                                  v3.0.1
        diagnostics_Versions_SimpleITK diagnostics_Versions_PyWavelet \
      0
        diagnostics_Versions_Python \
      0
                              3.6.8
                        diagnostics_Configuration_Settings \
      O {'minimumROIDimensions': 2, 'minimumROISize': ...
        diagnostics_Configuration_EnabledImageTypes \
      0
                                   {'Original': {}}
                  diagnostics_Image-original_Hash \
```

from radiomics import featureextractor

```
adcd47a617bf0a6906a361ba78d7e86388bc2fc9
  diagnostics_Image-original_Dimensionality \
0
              diagnostics_Image-original_Spacing \
   (0.5859000086784363, 0.5859000086784363, 1.0)
  diagnostics Image-original Size diagnostics Image-original Mean \
0
                  (512, 512, 192)
                                                        138.125106
   diagnostics_Image-original_Minimum diagnostics_Image-original_Maximum \
0
             diagnostics_Mask-original_Hash \
   ca99975f7f6ce5b9da272d5b065bb0fd8d40810e
               diagnostics_Mask-original_Spacing \
   (0.5859000086784363, 0.5859000086784363, 1.0)
  diagnostics_Mask-original_Size diagnostics_Mask-original_BoundingBox \
0
                 (512, 512, 192)
                                            (167, 225, 68, 37, 50, 16)
   diagnostics_Mask-original_VoxelNum diagnostics_Mask-original_VolumeNum
0
                                10905
         diagnostics_Mask-original_CenterOfMassIndex \
   (184.79257221458047, 254.31728564878497, 75.06...
              diagnostics_Mask-original_CenterOfMass \
   (-41.43703106818862, 1.4954971831525938, -7.49...
  original_firstorder_10Percentile original_firstorder_90Percentile \
0
                                                             1042.0
                             659.0
  original_firstorder_Energy original_firstorder_Entropy \
                                       4.503961941913401
               8464051347.0
  original_firstorder_InterquartileRange original_firstorder_Kurtosis \
0
                                   188.0
                                                    3.056529328502632
  original_firstorder_Maximum original_firstorder_MeanAbsoluteDeviation \
                                                     114.14490366028481
                       1292.0
  original_firstorder_Mean original_firstorder_Median \
         869.2987620357634
                                                888.0
```

```
original_firstorder_Minimum original_firstorder_Range
0
                                                  981.0
  original_firstorder_RobustMeanAbsoluteDeviation \
                                80.00459220139915
  original_firstorder_RootMeanSquared original_firstorder_Skewness \
                     881.000814067669
                                              -0.5230388203250016
0
  original_firstorder_TotalEnergy original_firstorder_Uniformity \
                2905529560.251311
                                           0.051218440447186646
0
  original_firstorder_Variance original_glcm_Autocorrelation \
           20482.096710984642
                                           579.312440764934
0
  original_glcm_ClusterProminence original_glcm_ClusterShade
                                         -693.8616497595558
               42908.67954319103
0
  original_glcm_ClusterTendency original_glcm_Contrast
                                     8.74050977809287
            118.47853188255739
  original_glcm_Correlation original_glcm_DifferenceAverage
        0.8617988560957099
                                          2.128732075933059
  original_glcm_DifferenceEntropy original_glcm_DifferenceVariance \
               2.6510141072718554
                                                3.9429016300371384
                      original_glcm_Idm original_glcm_Idmn \
    original glcm Id
0 0.4588122607941779 0.3890481464071379 0.9946965137645681
    original_glcm_Idn
                        original_glcm_Imc1 original_glcm_Imc2 \
0 0.9515499470890418 -0.25514812891031574 0.9358524694499407
  original_glcm_InverseVariance original_glcm_JointAverage
            0.37803667377261313
                                       23.491920944502194
  original_glcm_JointEnergy original_glcm_JointEntropy original_glcm_MCC \
      0.006641162131651515
                                    7.807733854014718 0.8696978190928191
  original_glcm_MaximumProbability original_glcm_SumAverage
               0.01625971801846341
                                         46.98384188900442
  original_glcm_SumEntropy original_glcm_SumSquares
          5.42170370906024
                                  31.80476041516255
```

Numerical results for radiomic features shown here are consistent with importing nifti files as image and label map in 3D Slicer (https://www.slicer.org/) and using Radiomics extension (https://www.slicer.org/wiki/Documentation/Nightly/Extensions/Radiomics).

#### 1.6 Part 6: Predictions To RT-Structure Example

Here we will provide a simple example for converting a predicted NumPy array of a square into a Dicom RT-Structure file

```
[28]: RT_path = os.path.join('Example_Data', 'RT_Structures')
if not os.path.exists(RT_path):
    os.makedirs(RT_path)
```

First, we will create a fake prediction, it will be the same size as the image NumPy array

```
[29]: image = Dicom_reader.ArrayDicom
```

Now, deep learning model typically create segmentations in the format of (z\_images, rows, cols, # of classes)

```
[31]: predictions = np.zeros(image.shape + (4,)) # Four classes: background, square, we circle, target

predictions.shape

predictions[75:80, 250:350, 100:200, 1] = 1 # Here we are drawing a square

predictions[75:80, 250:350, 300:400, 2] += create_circular_mask(100, 100, u)

where we center=None, radius=50).astype('int')

predictions[75:80, 100:200, 200:300, 3] += create_circular_mask(100, 100, u)

where we are drawing a square

predictions[75:80, 250:350, 300:400, 2] += create_circular_mask(100, 100, u)

where we are drawing a square

predictions[75:80, 100:200, 200:300, 3] += create_circular_mask(100, 100, u)

where we are drawing a square

predictions[75:80, 100:200, 200:300, 3] += create_circular_mask(100, 100, u)

where we are drawing a square

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where we are drawing a square

predictions[75:80, 100:200, 200:300, 3] += create_circular_mask(100, 100, u)

where we are drawing a square

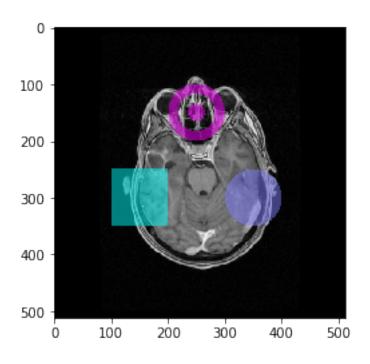
predictions[75:80, 100:200, 200:300, 3] += create_circular_mask(100, 100, u)

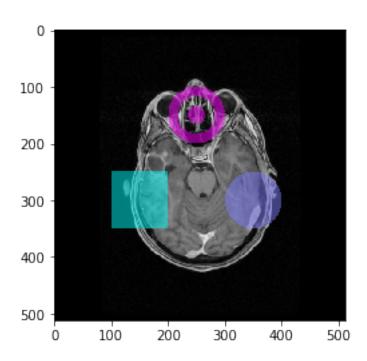
where we are drawing a square

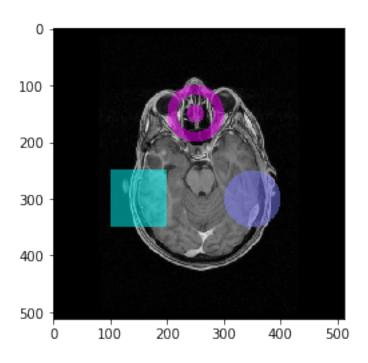
predictions[75:80, 100:200, 200:300, 3] += create_circular_mask(100, 100, u)

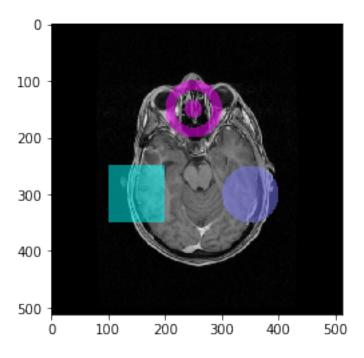
where we
```

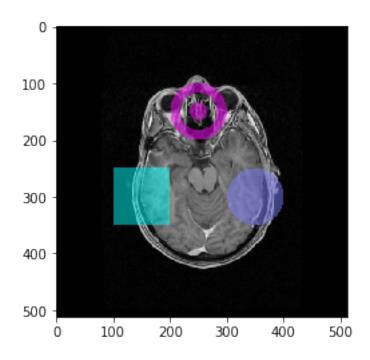
```
[32]: display_slices(image, np.argmax(predictions, axis=-1), skip = 1) # visualize<sub>□</sub> → our square on the image
```











Convert the NumPy arrays into RT-Structure

Running off a template
Writing data for square
Writing data for circle
Writing data for target
Writing out data...Example\_Data\RT\_Structures
Finished!

#### 2 Final notes

2.0.1 I hope that this code has been useful, if you have any suggestions or problems, please open an issue ticket or merge request on the Github: https://github.com/brianmanderson/Dicom\_RT\_and\_Images\_to\_Mask

Thank you!

[]: