Luminex Xponent: Plate- level Quality Control

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Aim

This code aims to do a plate- level quality control analysis for Luminex studies through bead counts, Background MFI, and Standard curve analysis.

R Markdown

This is an R Markdown document, areas that between the {r} frames that are require manual entries as described in the **bolded text**. When entries are completed, press the *knit* button at the top of the page.

Dependencies

```
• R version 4.3.1 (2023-06-16)

- tidyverse (v. 2.0.0)

- here (v. 1.0.1)
```

Required Inputs

1. Confirm working directory:

This is the project folder where the project is found. All folders and files added to the code will be within this folder. If needed you can set the working directory using the setwd() function.

Working directory: /Users/sahal/Documents/Research/R Projects/RTSS_Kisumu_Schisto

2. Choose file (input required):

Chose the raw plate file from working directory by adding what folder within (in the quotes). In the example below, the plate csv is in the Project/data/raw/luminex/ folder.

3. Set minimum bead count:

Set the minimum bead count as min_beadcount. In this example, the standard we have set here is 50. All bead counts <50 will be identified.

4 Define the file_path:

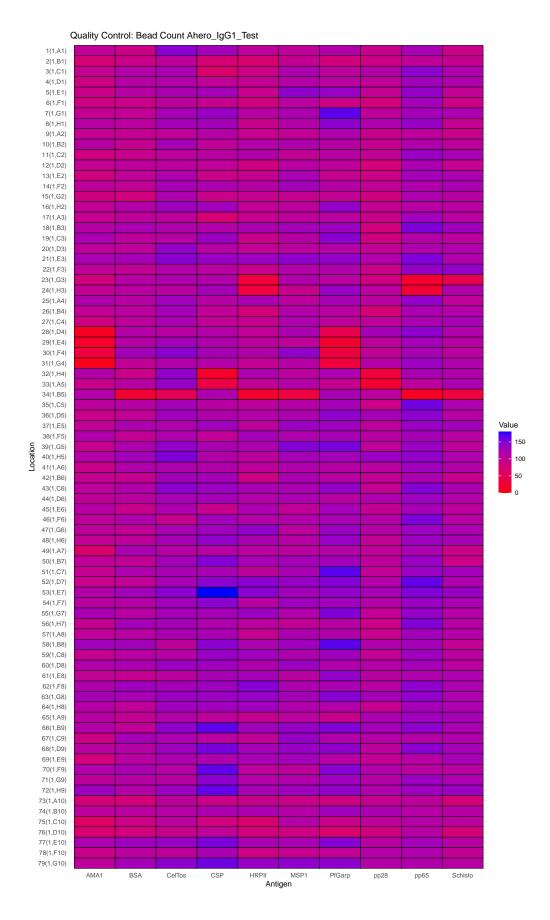
For example, here "Project/data/qc" folder is where the PlateName_beadqc_df.csv will be saved

```
# Choose the input file
input_file <- "data/raw/luminex/Ahero_IgG1_Test.csv"
# Minimum bead count for analysis
min_beadcount <- 50
#Output file path
file_path <- "data/qc"</pre>
```

Quality Control

Bead count (Input required)

 $\hbox{\tt \#\# The bead QC report has been exported to $\tt data/qc/Ahero_IgG1_Test_beadqc.png}$



List of wells/analytes <50 beads

This code results in a list of wells, sample IDs, and analytes that are less than the minimum bead count set. This file is exported to file_path (chosen above) as $PlateName_beadqc_list.csv$. Values in this list will be Na in the Median MFI dataframe. file_path: data/qc (this can be edited in the Bead Count section)

The bead QC list has been exported to data/qc/Ahero_IgG1_Test_beadqc_list.csv

```
##
      Location
                    Sample
                             Column
##
  1
      28(1,D4)
                   RAM-007
                               AMA1
##
  2
      29(1,E4) RAC-007-T01
                               AMA1
## 3
      30(1,F4) RAC-007-T07
                               AMA1
## 4
      31(1,G4) RAC-007-T08
                               AMA1
## 5
      32(1,H4)
                   RAM-008
                               pp28
## 6
      33(1,A5) RAC-008-T01
                               pp28
      23(1,G3) RAC-002-T07 Schisto
      34(1,B5) RAC-008-T07 Schisto
## 8
      23(1,G3) RAC-002-T07
                               pp65
## 10 24(1,H3) RAC-002-T08
                               pp65
## 11 34(1,B5) RAC-008-T07
                               pp65
## 12 34(1,B5) RAC-008-T07
                                BSA
## 13 34(1,B5) RAC-008-T07
                               MSP1
## 14 23(1,G3) RAC-002-T07
                              HRPII
## 15 24(1,H3) RAC-002-T08
                              HRPII
## 16 34(1,B5) RAC-008-T07
                              HRPII
## 17 34(1,B5) RAC-008-T07
                             CelTos
## 18 32(1,H4)
                   RAM-008
                                CSP
## 19 33(1,A5) RAC-008-T01
                                CSP
## 20 28(1,D4)
                   RAM-007
                             PfGarp
## 21 29(1,E4) RAC-007-T01
                             PfGarp
## 22 30(1,F4) RAC-007-T07
                             PfGarp
## 23 31(1,G4) RAC-007-T08
                             PfGarp
```

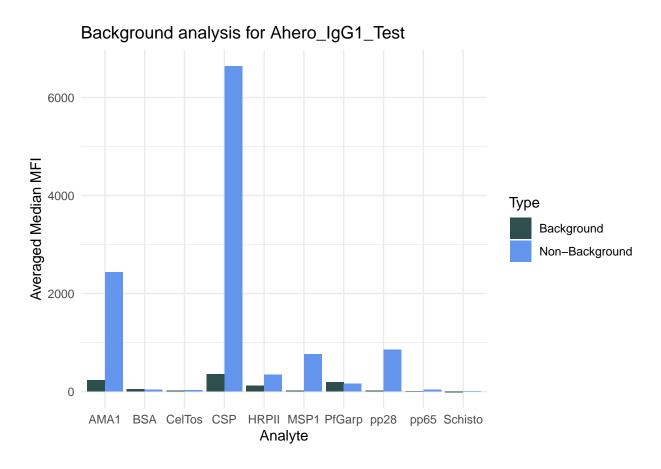
QC of median MFI data

This extracts the median MFI data, with all well-analyte combinations <50 beads/well replaced with NA. It completes the BSA MFI subtraction as well

note: negative values are kept

Background MFI

This code averages the median MFI values for all background and non-background samples. This requires all Background samples to have "Background" within the characters in the Sample column



Standard curve

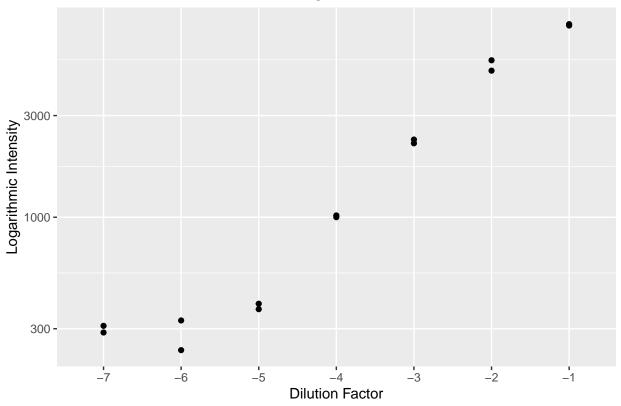
This code will attempt to create standard curves. This requires the samples have "Standard" within the characters in the Sample column. This will extraction Dilution_Factor as the number within the name (x -1).

For example: *Standard 2* will have a Dilution_factor of -2

This code will run for all analyte

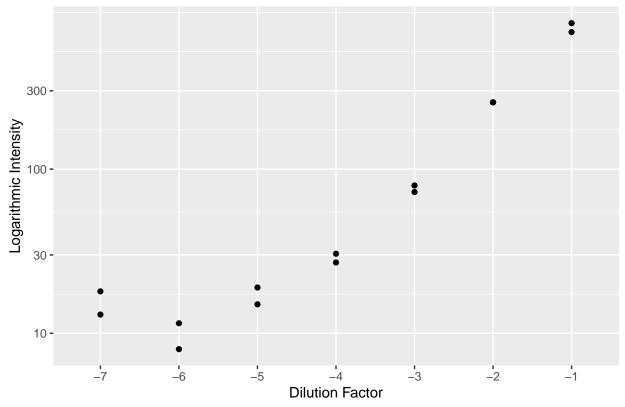
[[1]]

AMA1 standard curve for Ahero_lgG1_Test



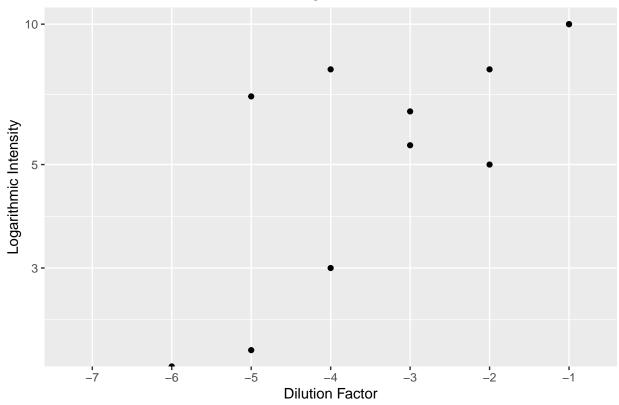
[[2]]

pp28 standard curve for Ahero_lgG1_Test

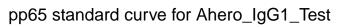


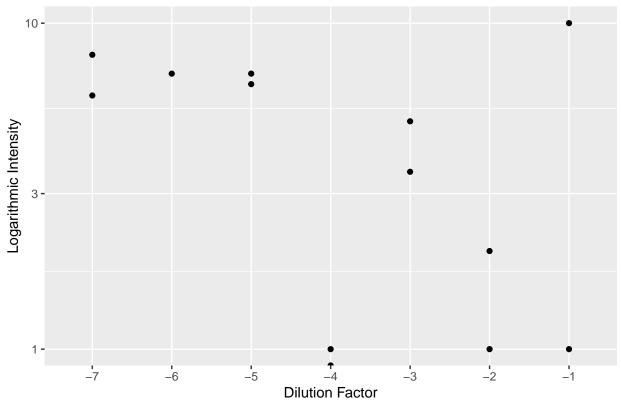
[[3]]

Schisto standard curve for Ahero_lgG1_Test



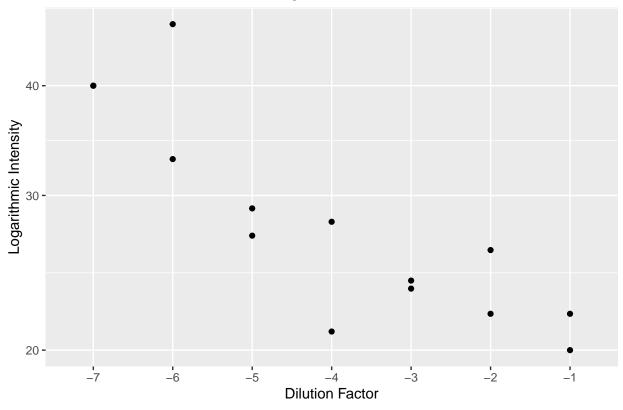
[[4]]





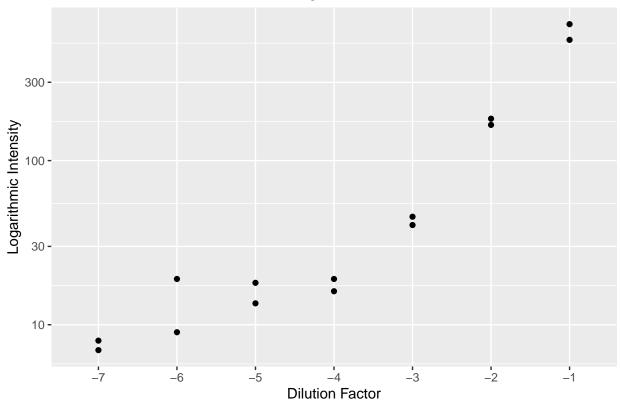
[[5]]

BSA standard curve for Ahero_lgG1_Test



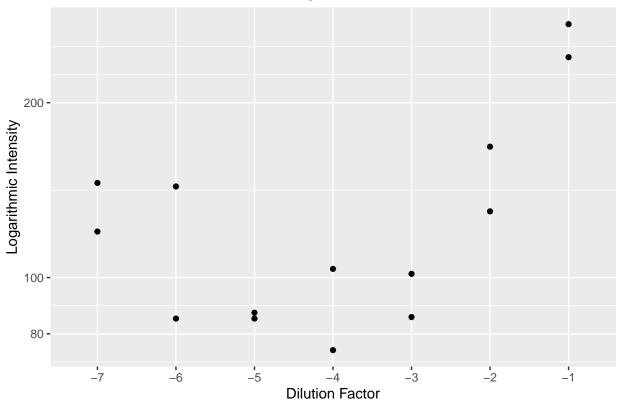
[[6]]

MSP1 standard curve for Ahero_lgG1_Test



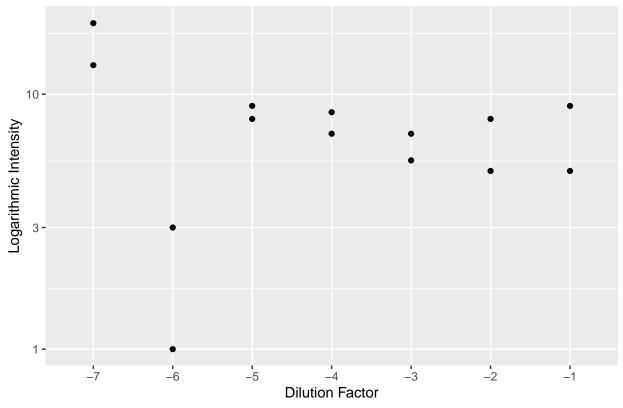
[[7]]

HRPII standard curve for Ahero_lgG1_Test



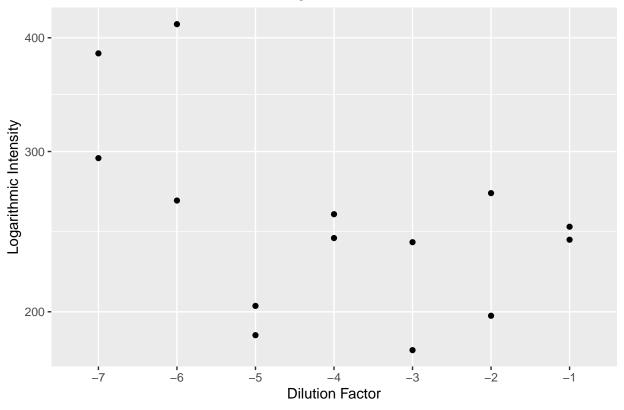
[[8]]

CelTos standard curve for Ahero_lgG1_Test



[[9]]

CSP standard curve for Ahero_lgG1_Test



[[10]]

PfGarp standard curve for Ahero_lgG1_Test

