

Blood Module Preservation in ASD Cortical Tissue

Module Preservation ASD Blood Leukocytes, ASD Post-Mortem Cortical Tissue

Setup and read in data

```
library(easypackages)
libraries("WGCNA", "here")

## =====
## *
## *   Package WGCNA 1.63 loaded.
## *
## *   Important note: It appears that your system supports multi-threading,
## *   but it is not enabled within WGCNA in R.
## *   To allow multi-threading within WGCNA with all available cores, use
## *
## *       allowWGCNAThreads()
## *
## *   within R. Use disableWGCNAThreads() to disable threading if necessary.
## *   Alternatively, set the following environment variable on your system:
## *
## *       ALLOW_WGCNA_THREADS=<number_of_processors>
## *
## *   for example
## *
## *       ALLOW_WGCNA_THREADS=24
## *
## *   To set the environment variable in linux bash shell, type
## *
## *       export ALLOW_WGCNA_THREADS=24
## *
## *   before running R. Other operating systems or shells will
## *   have a similar command to achieve the same aim.
## *
## =====

options(stringsAsFactors=FALSE)

# Read in ASD brain data
load(here("data", "tidy", "asd_brain_data.Rdata"))

# Read in ASD blood data
load(here("data", "processed", "exprDataAdj.Rdata"))

# Read in WGCNA ASD blood results
wgcna_res = read.csv(here("WGCNAresults", "wgcna_results_summary.csv"))
blood_colors = wgcna_res$moduleColors
asd_blood_data = exprDataAdj
asd_blood_geneAnno = geneInfo
```

Find common genes amongst the two datasets

```
gene_mask = is.element(asd_blood_geneAnno$geneSymbol, asd_brain_geneAnno$hgnc_symbol)
genes2use = asd_blood_geneAnno$geneSymbol[gene_mask]
asd_blood_geneAnno_subset = asd_blood_geneAnno[gene_mask,]

asd_blood_data_subset = asd_blood_data[gene_mask,]
rownames(asd_blood_data_subset) = asd_blood_geneAnno_subset$geneSymbol
blood_colors_subset = blood_colors[gene_mask]

geneAnno2 = subset(asd_brain_geneAnno,
                   is.element(asd_brain_geneAnno$hgnc_symbol, genes2use))
asd_brain_data_subset = subset(asd_brain_data,
                               is.element(asd_brain_geneAnno$hgnc_symbol, genes2use))
rownames(asd_brain_data_subset) = geneAnno2$hgnc_symbol
asd_brain_data_subset = asd_brain_data_subset[rownames(asd_blood_data_subset),]
```

Run modulePreservation

```
setLabels = c("ASDBlood", "ASDBrain")
multiExpr = list(ASDBlood = list(data = t(asd_blood_data_subset)),
                 ASDBrain = list(data = t(asd_brain_data_subset)))
multiColor = list(ASDBlood = blood_colors_subset)

# Calculate module preservation stats
corFnc2use = "bicor"
networkType = "signed"
nperm = 1000
rand_seed = 1
mp = modulePreservation(multiExpr,
                        multiColor,
                        networkType = networkType,
                        corFnc = corFnc2use,
                        maxGoldModuleSize = 1000,
                        referenceNetworks = 1,
                        nPermutations = nperm,
                        randomSeed = rand_seed,
                        quickCor = 0,
                        verbose = 0)
```

Show module preservation results

```
ref = 1
test = 2
statsObs = cbind(mp$quality$observed[[ref]][[test]][, -1],
                 mp$preservation$observed[[ref]][[test]][, -1])
statsZ = cbind(mp$quality$Z[[ref]][[test]][, -1],
               mp$preservation$Z[[ref]][[test]][, -1])

# compute p-values from log10 pvalues in mp
```

```

Zsummary.log10pvals = mp$preservation$log.p$ref.ASDBlood$inColumnsAlsoPresentIn.ASDBrain$log.psummary.p
Zsummary.pvals = 10^-Zsummary.log10pvals
Zsummary.fdr = p.adjust(Zsummary.pvals, method = "fdr")

sumTable = cbind(statsObs[, c("medianRank.pres", "medianRank.qual")],
  signif(statsZ[, c("Zsummary.pres", "Zsummary.qual")], 2),
  Zsummary.pvals, Zsummary.fdr)

ModCols = c("black", "blue", "brown", "cyan", "darkred", "gold", "green",
  "greenyellow", "grey", "grey60", "lightcyan", "lightgreen",
  "lightyellow", "magenta", "midnightblue", "pink", "purple",
  "red", "royalblue", "salmon", "tan", "turquoise", "yellow")
ModNums = c("M7", "M2", "M3", "M14", "M21", NA, "M5", "M11", "M0", "M16", "M17",
  "M18", "M19", "M9", "M15", "M8", "M10", "M6", "M20", "M13", "M12",
  "M1", "M4")
modinfo = data.frame(moduleColors = ModCols, moduleLabels = ModNums)
sumTable = cbind(moduleLabels = modinfo$moduleLabels, sumTable)
sumTable

```

##	moduleLabels	medianRank.pres	medianRank.qual	Zsummary.pres
## black	M7	17	18.0	0.240
## blue	M2	2	11.0	8.100
## brown	M3	19	10.0	-0.940
## cyan	M14	9	15.0	0.900
## darkred	M21	16	5.0	-0.190
## gold	<NA>	14	22.0	5.500
## green	M5	9	18.0	1.300
## greenyellow	M11	3	1.0	4.100
## grey	M0	18	23.0	-0.054
## grey60	M16	20	7.0	-1.200
## lightcyan	M17	18	17.0	-0.380
## lightgreen	M18	9	4.0	0.810
## lightyellow	M19	14	12.0	-0.190
## magenta	M9	1	12.0	6.500
## midnightblue	M15	14	3.5	-0.160
## pink	M8	4	6.0	5.400
## purple	M10	11	20.0	1.700
## red	M6	13	9.5	-0.940
## royalblue	M20	14	2.0	0.340
## salmon	M13	15	18.0	-0.240
## tan	M12	5	8.0	2.500
## turquoise	M1	12	13.5	1.500
## yellow	M4	10	21.0	2.400
##	Zsummary.qual	Zsummary.pvals	Zsummary.fdr	
## black	25	1.588561e-01	2.810530e-01	
## blue	56	3.402800e-19	7.826441e-18	
## brown	80	6.023294e-01	6.297080e-01	
## cyan	23	1.225806e-01	2.563049e-01	
## darkred	48	4.955699e-01	6.297080e-01	
## gold	1	2.483746e-10	1.904205e-09	
## green	22	7.677112e-02	1.765736e-01	
## greenyellow	84	4.107692e-07	1.889538e-06	
## grey	-14	1.767056e-01	2.903021e-01	
## grey60	41	8.646665e-01	8.646665e-01	

```

## lightcyan          15  5.928797e-01 6.297080e-01
## lightgreen         55  1.413842e-01 2.709863e-01
## lightyellow        19  5.709222e-01 6.297080e-01
## magenta            48  1.341995e-13 1.543294e-12
## midnightblue       62  5.573078e-01 6.297080e-01
## pink               78  6.224280e-09 3.578961e-08
## purple             19  2.293958e-02 5.862338e-02
## red                53  4.692526e-01 6.297080e-01
## royalblue          58  3.449277e-01 5.288892e-01
## salmon             21  5.660540e-01 6.297080e-01
## tan                47  6.357133e-03 2.088772e-02
## turquoise          86  1.643211e-02 4.724230e-02
## yellow             20  6.254077e-03 2.088772e-02

# Plot results
modColors = rownames(mp$preservation$observed[[ref]][[test]])
moduleSizes = mp$preservation$Z[[ref]][[test]][,1]
plotMods = !(modColors %in% c("grey", "gold"));
text = modColors[plotMods]
plotData = cbind(mp$preservation$observed[[ref]][[test]][,2],
                  mp$preservation$Z[[ref]][[test]][,2])
mains = c("Preservation Median rank", "Preservation Zsummary")
par(mfrow = c(1,2))
par(mar = c(4.5,4.5,2.5,1))
for (p in 1:2)
{
  min = min(plotData[, p], na.rm = TRUE)
  max = max(plotData[, p], na.rm = TRUE)
  # Adjust plotting ranges appropriately
  if (p==2)
  {
    if (min > -max/10) min = -max/10
    # ylim = c(min - 0.1 * (max-min), max + 0.1 * (max-min))
    ylim = c(-2,12)
  } else
  {
    ylim = c(max + 0.1 * (max-min), min - 0.1 * (max-min))
    # ylim = c(-2,12)
  }
  plot(moduleSizes[plotMods],
        plotData[plotMods, p],
        col = 1,
        bg = modColors[plotMods],
        pch = 21,
        main = mains[p],
        cex = 2.4,
        ylab = mains[p],
        xlab = "Module size",
        log = "x",
        ylim = ylim,
        xlim = c(10, 2000),
        cex.lab = 1.2,
        cex.axis = 1.2,
        cex.main = 1.4)
  # labelPoints(moduleSizes[plotMods],
  #              plotData[plotMods, p],

```

```

#           text,
#           cex = 1,
#           ofs = 0.08);

# For Zsummary, add threshold lines
if (p==2) {
  abline(h=0)
  abline(h=2, col = "blue", lty = 2)
  abline(h=10, col = "darkgreen", lty = 2)
}
}

```

