International Journal of Zoology and Applied Biosciences

Volume 1, Issue 6, pp: 257-261, 2016

Research Article





EFFECT OF SUBLETHAL TOXICITY OF ZINC CHLORIDE ON LIVER HISTOLOGY OF ESTUARINE EDIBLE FISH, *LIZA MACROLEPIS* (SMITH, 1846)

A. Senthil Anna and S. Raveendran*

P.G. and Research Department of Zoology, Khadir Mohideen College, Adirampattinam-614 701, Tamil Nadu, India.

Article History: Received 1st November 2016; Accepted 16th November 2016; Published 31st December 2016

ABSTRACT

In the present study, estuarine edible teleost, *Liza macrolepis* of size 9.6 ± 1.2 cm length were exposed to sublethal concentration ($^{1}/_{10}$ of LT₅₀/96 h) of heavy metal, Zinc chloride for a period of 15 days. The treated fish groups were compared with the control group for histopathological changes in the liver tissue. The histopathological changes occurred in the liver of L. *Macrolepis* suggest that histopathological studies are considered as direct evidence referring to any adverse effect on fish health. Moreover, the liver is considered as the principal target organ of detoxification in vertebrates and particularly in fish.

Keywords: Liza macrolepis, Zinc chloride, Liver tissue, Sublethal toxicity, Histopathology.

INTRODUCTION

The contamination of aquatic bodies with a range of heavy metal pollutants has become a global concern over the past few decades (Fostner and Wittman, 1979; Meyers and Hendricks, 1985; Wickland–Glynn, 1991; Bryan and Langston, 1992; Bucke, 1995; De Forest, *et al.*, 2007; Kaoud and El-Dahshan, 2010). Heavy metals contribute to anthropogenic contamination of aquatic resources such as rivers, estuaries and seas (Bryan and Langston, 1992; Moore *et al.*, 1999). Some of them are essential to the normal fish metabolism and others are toxic even at low concentrations (Pillai, 1983; Kalay and Canli, 2000; Sen *et al.*, 2011 Chavan and Muley, 2014).

Fish, having great economic importance are affected immensely by varied metals directly or indirectly in many ways (Rashed, 2001; Gaber, 2007; Al-Weher, 2008; Pourmoghaddas and Shahryari, 2010). The toxic effects of heavy metals have been studied by several workers in freshwater species of fishes and some of these studies indicate high mortality of juvenile fish and reduced breeding potentially of adults after acute and chronic

exposures to heavy metals (Besirovic *et al.*, 2010; Rambhare and Bakare 2012).

Lead (Pb) is one of the most toxic heavy metals in aquatic environs and its toxicity and accumulation have been reported on fish metabolism and histological responses. Absorption of lead occurs by different ways through the gills and skin or by ingestion of contaminated water and food (Aruldoss and Indra, 2005; De Forest *et al.*, 2007; Authman and Abbas, 2007; Khan *et al.*, 2011 Javeed and Usman 2011; Fatima and Usmani, 2013).

Histopathology of fish is not used as a standard tool for monitoring metal pollution but rather it is considered for inclusion as a viable tool for determining fish health in the laboratory and field studies (Weber and Gingerich, 1982; Bucke, 1993; Van Dyk *et al.*, 2007; Bhatkar, 2010; Deore and Wagh, 2012; Mokhtar *et al.*, 2013; Chavan and Muley, 2014). Perusal of the literature reveals that histopathological alterations in body tissues of freshwater fishes have been reported by metal toxicity (Ajmal *et al.*, 1985; Javeed, 2005; Akan *et al.*, 2009; Ambedkar and Muniyan, 2011). Very few studies are available on the

effects of metals on the histological responses of estuarine edible fishes (Avenimo *et al.*, 2005; Zhao *et al.*, 2012). With this view in mind, an attempt has been made to observe the acute effect of sublethal toxicity of lead nitrate on the liver histology of an edible teleost fish, *Liza macrolepis* of Thengaithittu estuary, Puducherry, South East Coast of India.

MATERIALS AND METHODS

Collection and maintenance of test animal

Liza macrolepis (n = 250) juveniles were collected from the estuary during low tide regime and brought to the laboratory. Fishes were acclimatized in fish tanks (50L cap) for one week. Fish were fed with fish food (goat meat) and the water in the tanks was renewed by freshly collected estuarine water at every 24h.

Test chemical: preparation of test concentrations

Zinc chloride (Qualigens AR Grade Mumbai) was procured and 10% stock solution using DDW. Varied concentrations viz. 0.10, 0.20, 0.30, 0.40, 0.50, 0.60, 0.70, 0.80, 0.90 (mg/L) were prepared by v/v method.

L_{T50} determination

Healthy juveniles of L. macrolepis with $(9.6 \pm 1.2 \text{ cm} \text{ length}; 10.3 \pm 0.78 \text{ g}$ weight) were chosen from the mass rearing to determine L_{T50} value. Three replicates of groups of 5 fishes were exposed to each one of the test concentrations of Zinc chloride. Dead fish were removed from the tanks and counted at 24, 48, 72 and 96h of exposures. Death was presumed when fishes were immobile and showed no response to touch with a glass rod. From the mortality values obtained to different concentrations of $Zncl_2$, a concentration rendering 50 percent population dead i.e., LT_{50} / 96h was calculated adopting Spearman–Karber Airithmetic mean method (Hamilton $et\ al.$, 1977).

Bioassay studies

Acclimated juvenile fish, *L. macrolepis* with uniform body length (9.6 \pm 1.2 cm) and mass weight (10.3 \pm 0.78 g) were chosen for bioassays. Two groups were formed, each consisting of five animals. One group was considered as the experimental and exposed to (10% of LT₅₀ / 96 h) sublethal toxicity of Zncl₂ for 15 days. The other groups was treated with Zncl₂ concentration and kept as control.

Both controls and exposed groups were maintained simultaneously.

Histological analysis

Liver samples from control and sublethal treated fishes were carefully removed at intervals of 5 days and fixed in 10% Neutral buffered formalin, dehydrated in ascending grades of alcohol and cleared in xylene. The fixed tissues were embedded in paraffin wax and sectioned into 5 to 7 micrometers thick (Steedman, 1960) and then stained with Haemotoxylin Eosin method according to Bucke (1972). Then the sections were examined on light microscope (Krempt–Wetzler) under 100x magnification and photographed by using a microscopic camera.

RESULTS

The liver of control fish is covered by a thin layer of fibrous layer of mesothelial cells. The parenchyma is dispersed randomly throughout the liver tissue. Within the parenchyma, the hepatocytes (H) were spread out as irregular cords.

The histopathological changes observed in the present study after chronic exposure to (10% of LT₅₀/96 h) sublethal concentration of zinc chloride in the liver of the estuarine fish, *L. macrolepis* have been depicted in Photoplate I (A to C). The liver of the fish exposed to ZnCl₂ metal sublethal toxicity exhibited marked histopathological alterations. The most alterations induced by Zncl₂ after 5 d sublethal exposure were the damage of lead characterized by cloudy swelling with large vacuoles (CSV) and cytoplasmic vacuolation (CV).

In addition, Degeneration of Pancreocytes (DP) and Necrosis (N) were observed in the liver tissue of the fish *L. macrolepis* after 10 d sublethal exposure (Plate. B). Hypertrophy of hepatocytes (HH) and psycnotic nuclei(PN) were the histopathological lesions also noticed in the liver tissue of the fish exposed to 15 days of sublethal toxicity of ZnCl₂.

Microphotograph showing transverse sections of liver tissue of the fish, *Liza macrolepis* exposed to sublethal (10% of LT50/96 h) concentration of the metal, Zinc chloride for 5(A), 10 and 15 days (CSV = Cloudy Swelling with large Vacuoles, CV = Cytoplasmic Vacuolation, DP = Degeneration of Pancreocytes, H = Hepatocytes, HH = Hypertrophy of Hepatocytes, N = Necrosis, PN = Psycnotic Nuclei).

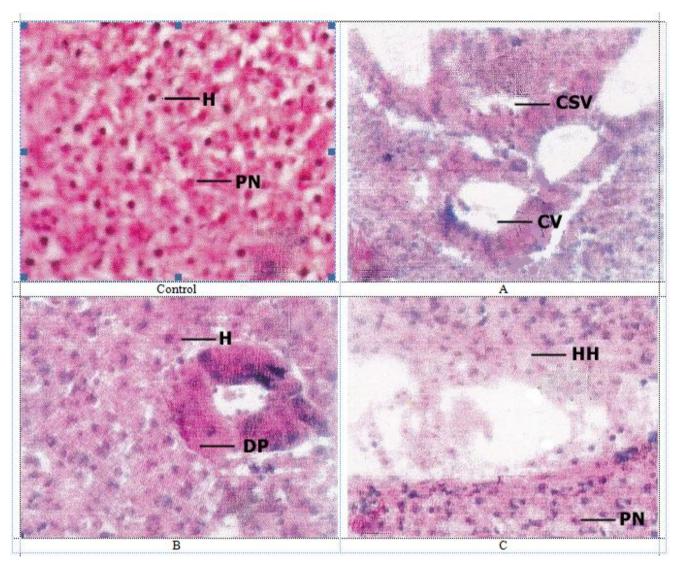


Plate I. Microphotograph showing transverse sections *of* liver tissue of the fish, *Liza macrolepis* exposed to sublethal (10% of LT50/96 h) concentration of the metal, Zinc chloride for 5(A), 10 and 15 days (CSV = Cloudy Swelling with large Vacuoles, CV = Cytoplasmic Vacuolation, DP = Degeneration of Pancreocytes, H = Hepatocytes, HH = Hypertrophy of Hepatocytes, N = Necrosis, PN = Psycnotic Nuclei).

DISCUSSION

Fishes have the ability to accumulate toxicants such as heavy metals in their tissues viz. gills and liver (Mobarak and Sharaf, 2011). The present study revealed that there is a strong link between liver damage and sublethal toxicity of heavy metals (Weber and Gingerich, 1982; Radaiah and Jayantha Rao, 1992). Pathological changes in the liver of the fish, *L. macrolepis* exposed to $^1/_{10}^{\rm th}$ of LT $_{50}/$ 96 of Znll $_2$ were depicted in the photoplate (A–C). The liver of control fish is covered by a thin layer of mesothelial cells. Hepatocytes and pancreocytes were randomly dispersed throughout the liver tissue Marked variations in the liver architecture were observed by the $5^{\rm th}$ days, $10^{\rm th}$ days and $15^{\rm th}$ days (Photoplate: A, B and C).

The most alterations of the liver exposed to Zinc sublethal 5d exposure were cloudy swelling of cells with large vacuoles (CSV) and cytoplasmic vacuolation (VC).

These observation were in agreement with the reports of Aruldoss and Indra (2005) in tilapia, Oreochromis mossambicus Gaber (2007) in nile tilapia, Oreochromis niloticus; Deore and Singh (2012) in the carp, Channa gaticua (Photoplate-A). In addition marked variations in liver tissue histology were noticed after 10d exposure and they were degeneration of pancreocytes (DP) and hepatocyte hypertrophy (HH). Similar alterations in the liver tissue were observed in several species exposed to heavy metals and these alterations were described by Kaoud and El- Dahshan (2010) in nile tilapia, O. niloticus; Pourmoghaddas and Shahryari (2011) in three carnivorous fishes; Mokhtar et al., (2013) in O. niloticus; Histopathological lesions such as Necrosis degeneration of Pancreocytes (DP) and Pycnotic Nuclei (PN) were recorded in the liver tissue of the fish exposed for 15 days. Such findings on liver pathological lesions were studied by Van Dyk et al.(2007) in O. mossoambicus; El-Naggar, et al. (2009) in the liver of O. niloticus due to the toxicity of Fe, Mn, Cu, Zn, Cd and Pb and Gaber et al., (2014) in two fishes, Sparus aurata and Dicentrarockus labrax.

ACKNOWLEDGEMENTS

The first author wishes to express sincere thanks to the authorities of Khadir Mohideen College, Adirampattinam for facilities provided to carry out this work. In addition, the suggestions and guidance in histological profiles offered by Dr. D.S.M. SHAH, Former HOD, P.G. and Research Department of Zoology, Khadir Mohideen College, Adirampittinam is gratefully acknowledged.

REFERENCES

- Ajmal, M., Khan, M.A., and Nomani, A.A., 1985. Distribution of heavy metals in plants and fish of the Yamuna river (India). *Environ. Monit. Assess.*, 5, 361-367
- Akan, J.C., Abdulrahman, F.I., Sudipo, O.A., and Akandu, P.I., 2009. Bioaccumulation of some heavy metals of six freshwater fishes caugh from river Chad, Maiguguri, Borno state, Nigeria. J. Appl. Sci. Environ. Sanit., 4, 103-114.
- Al-Weher, S.M., 2008. Levels of heavy metals Cd, Cu and Zn in three fish species collected from the Northern Jordan valley Jordan. Jordan *J. Biol. Sci.*, 1, 41-46.
- Ambedkar, G., and Muniyan, M., 2011. Bioaccumulation of some heavy metals in the selected five freshwater fish from Kollidam river, Tamil Nadu, India. *Adv. Appl. Sci. Res.*, 2, 221-225.
- Aruldoss, K. and N. Indira, 2005. Impact of lead on the histology of liver tissue of the freshwater fish, *Oreochromis mossambicus. J. Exp. Zool. India*, 8(2), 389-391.
- Authman, M.M., and Abbas, H.H., 2007. Accumulation and distribution of copper and zinc in both water and some vital tissues of two fish species (*Tilapia zillii* and *Mugil cephalus*) of river qarum, Egypt. *Pak. J. Biol. Sci.*, 10, 2106-2122.
- Avenimo, J.G., Adeeyinwo, C.E., and Amoo, I.A., 2005. Heavy metal pollutants in Warri River estuary, Nigeria kragujevac. *J. Sci.*, 27, 43-50.
- Besirovic, H.A., Alie, S., Prasovic, S., and Drommer, W., 2010. Histopathological effects of chronic exposure to Cadmium and Zinc on kidneys and gills of brown trout, *Salino trutta* Turk. *J. Fish. Aquat. Sci.*, 10, 255-262.

- Bhaktar, N.V., 2010. Chromium, Nickel and Zinc induced histopathological alterations in the gill of Indian common carp, *Labeo rohita* (Ham.) *J. Appl. Nat. Sci.*, 2(2), 234-238.
- Bryan, G.W., and Langston, W.J., 1992. Bio-availability, accumulation and affects of heavy metals in water with special reference to U.K. estuaries A review. *Environ. Pollut.*, 76(2), 89-131.
- Bucke, D., 1993. Aquatic pollution effects on the health of fish and shellfish. *Parasitology*, 103, 825-837.
- Chavan, V.R., and Muley, D.V., 2014. Effect of heavy metals on liver and gill of fish *Cirrhinus mrigala Int. J. Curr. Microbiol. Appl. Sci.*, 3(5), 277-288.
- De Forest, D.K., Brux, K.V. and Adams, W.J., 2007. Assessing metal bioaccumulation in aquatic environments. *Aquat. Toxicol.*, 84, 236-246.
- Deore, S.V. and Wagh, S.B., 2012. Heavy metal induced histopathological alterations in liver of *Channa gactiua* (Ham.). *J. Exp. Sci.*, 2(2), 35-38.
- El-Nagger, A.M., Mohmoud, S.A., and Tayel, S.I., 2009. Bioaccumulation of some heavy metals and histopathological alterations in liver of *Oreochromis niloticus* in relation to water quality at different locations along the river nile, Egypt. *World J. fish Mar. Sci.*, 1(2), 105-114.
- Fatima, M., and N. Usmani, 2013. Histopathology and bioaccumulation of heavy metals (Cr, Ni and Pb) in fish *Channa striatus* and *Heteropneustes fossilis* tissues. A study for toxicity and ecological impacts *Pak. J. Biol. Sci.*, 16, 412-424.
- Fostner, U. and Wittman, G.T.W., 1979. Metal pollution in aquatic environment IInd Edn. Spainger Verlag, Berlin., pp. 99-118.
- Gaber, H.S., 2007. Impact of certain metals on the gill and liver of the nile tilapia (*Oreochromis niloticus*) Egypt. *J. Aquat. Biol. Fish.*, 11(2), 79-100.
- Gaber, H. S., Abbas, Mohammad, W.T., Authman, M.N., and Gaber, S.A., 2014. Histological studies on some organs of two fish species in Bardawill Lagoon, North Sinai, *Egypt. Global Veterin.*, 12,(1), 1-11.
- Hamilton, M.A., Russo, R.C., and Thurston, R.V., 1977. Trimmed Spearman-Karber method for estimating median lethal concentration in toxicity bioassays. *Env. Sci. Tech.*, 11, 714-719.
- Javeed, M., 2005. Heavy metal contamination in fresh water fish and led sediments in the river Ravi stretch and related tributaries. Pak. *J. Biol. Sci.*, 8, 1337-1341.
- Javeed, M., and Usmani, N., 2011. Accumulation of heavy metals in fishes. A human health concern. *Int. J. Env. Sci.*, 3, 659-670.
- Kalay, M. and Canli, M., 2000. Estimation of essential (Cu and Zn) and non-essential (Cd and Pb) metals from

- tissues of a freshwater fish, *Tilapia zilli. Trop. J. Zool.*, 24, 429-436.
- Kaoud, H.A. and El-Dahshan, A.R., 2010. Bioaccumulation and histopathological alterations of the heavy metals in *Oreochromis niloticus* fish. *Nat. Sci.*, 8(4), 147-156.
- Khan, H.A., Sikkandar, B., Kamalesh, B., Wani, A.A., and Parveez, P.A., 2011. Lead nitrate induced histopathological changes in the gills of the African catfish, *Clarias batrachus J. Appl. Sci. Res.*, 7(7), 1081-1086.
- Meyers, T.R., and Hendricks, J.D., 1985. Histopathology, In: Fundamentals of Aquatic Toxicology Methods and Applications (Eds. Rand, G.M., and Petrocelli, S.R.,) Hemisphere Publishing Corp., Washington, DC, pp. 283-331.
- Mobarak, Y.M.S. and M.M., Sharaf, 2011. Lead acetate induced histopathological changes in the gills and digestive system of silver Sailfin molly *Poecilia latipinna* int. *J. Zool. Res.*, 7, 1-18.
- Mokhtar, D.M., Hanan and Abd-Elhafeez, H., 2013. Histological changes in selected organs of *Oreochromis niloticus* exposed to doses of lead acetate. *J. Life Sci. Biomed.*, 3(3), 256-263.
- Pillai, K.C., 1983. Heavy metals in aquatic environment. In: Water pollution and Management (Ed. Varshney, C.K.) Wiley Eastern Ltd., New Delhi, pp. 74-93.
- Pourmoghaddas, H. and Shahryani, A., 2010. The concentration of lead, chromium, cadmium, nickel and mercury in three species of consuming fishes of Isfahan city. *Health Syst. Res.*, 6, 30-35.

- Radhaiah, V., and Jayantha Rao, K., 1992. Fenvallerate toxicity to the liver in a freshwater teleost, *Tilapia* mossambica (Peters) Comp. *Physiol. Ecol.*, 17(2), 48-53.
- Rambhare, V.S., and Bakare, R.V.G., 2012. Effect of heavy metal pollution on freshwater fishes. *Proc. Int. Conf. Shivaji Univ. Kolhpur*, pp. 170-172.
- Rasheed, M.N., 2001. Cadmium and Lead levels in fish *Tilapia nilotica* tissues as biological indicator for river water pollution. *Envt. Monit. Assess.*, 68(1), 75-89.
- Sen, I., Shandil, A., and V. B., Shrivastava, 2011. Study for determination of heavy metals in fish species of the river Yamuna (Delhi) by (ICP OES). *Adv. Appl. Sci. Res.*, 2, 161-166.
- Steedman, H. E., 1960. Section cutting microscopy, Oxford Blackwell Scientific. PP. 172.
- Van Dyk, J.C., Piesterse, G.M., and Van Huren, J.H.J., 2007. Histological changes in the liver of *Oreochromis mossambicus* after exposure to cadmium and zinc. *J. Ecotox. Environ. Saf.*, 66, 432-440.
- Weber, I.J., and Gingerich, W.H., 1982. Hepatic toxicology of fishes. In: Aquatic Toxicology (Ed. Weber, L.J.) Raven Press, New York, pp. 55-105.
- Wickland-Gynna, A., 1991. Cd and Zn kinetics fish: studies on water borne Cd and Zn turnover and intracellular distribution in minnows, *Phoxinus phoxinus*. *Pharm. Toxicol.*, 69, 485-491.
- Zhao, S., Feng, C., Quan, W., Chen, X., Niu, J., and Shen, Z., 2012. Role of living organisms in the accumulation characteristics of heavy metals in fishes and Crabs in the Yangtce Estuary, China. *Mar. Pollut. Bull.*, 64: 1163-1171.