# Package 'BayLum'

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Type Package
<b>Title</b> Chronological Bayesian Models Integrating Optically Stimulated Luminescence and Radiocarbon Age Dating
<b>Description</b> Bayesian analysis of luminescence data and 14C age estimate. Bayesian models are based on the following publications: Combes, B. & Philippe, A. (2017) <doi:10.1016 j.quageo.2017.02.003=""> and Combes et al (2015) <doi:10.1016 age="" amongst="" and="" application="" data="" export,="" import,="" j.cludes,="" model.<="" models="" of="" others,="" palaeodose="" td=""></doi:10.1016></doi:10.1016>
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### **Description**

A collection of various R functions for Bayesian analysis of luminescence data and C-14 age estimates. This includes, amongst others, data import, export, application of age and palaeodose models.

# **Details**

This package is based on the functions: Generate\_DataFile and Generate\_DataFile\_MG to import luminisecence data. These functions create a list containing all informations to compute age of Single-grain OSL measurements for the first function and Multi-grain OSL measurements for the second.

The functions: Age\_Computation and AgeS\_Computation use Bayesian analysis for OSL age estimation for one or various samples according to differents models (e.g. different dose-response curves and different equivalent dose distributions around the palaeodose).

It is possible to consider various BIN/BINX-files per sample, to compute ages of samples in stratigraphic constraints and to integrate systematic errors.

It is possible to calibrate C-14 age with the function AgeC14\_Computation. We can also estimate chronology containing 14C age and OSL samples with the function Age\_OSLC14.

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#### Author(s)

Claire Christophe, Anne Philippe, Sebastian Kreutzer, Guillaume Guérin

AgeC14\_Computation

Bayesian analysis for age estimation of 14C age of various samples

### **Description**

This function calibrate 14C age of samples to get an age (years). The user can choose one of the following radiocarbon calibration curve: Northern or Sourthen Hemisphere or marine atmospheric. It must be the same curve for all samples.

# Usage

```
AgeC14_Computation(Data_C14Cal, Data_SigmaC14Cal, SampleNames, Nb_sample,
    PriorAge = rep(c(10, 50), Nb_sample), SavePdf = FALSE,
    OutputFileName = c("MCMCplot", "HPD_Cal14CCurve", "summary"),
    OutputFilePath = c(""), SaveEstimates = FALSE,
    OutputTableName = c("DATA"), OutputTablePath = c(""),
    StratiConstraints = c(), sepSC = c(","), Model = c("full"),
    CalibrationCurve = c("AtmosphericNorth"), Iter = 50000, t = 5,
    Nb_chaines = 3, quiet = FALSE)
```

### **Arguments**

Data\_C14Cal numeric vector: corresponding to 14C age estimate.

Data\_SigmaC14Cal

numeric vector: correponding to the error of 14C age estimates.

SampleNames character vector: names of sample. The length of this vector is equal to Nb\_sample.

Nb\_sample integer: number of samples.

PriorAge numeric vector (with default): lower and upper bounds for age parameter of

each sample in years (not in ka). Note that, length(PriorAge)=2\*Nb\_sample and PriorAge[2i-1,2i] correponds to the lower and upper bounds of sample

whose number ID is equal to i.

SavePdf boolean (with default): if TRUE save graphs in pdf file named OutputFileName

in folder OutputFilePath.

OutputFileName character (with default): name of the pdf file that will be generated by the func-

 $tion\ if\ SavePdf=TRUE,\ length(OutputFileName)=3,\ see\ \textbf{PLOT}\ \textbf{OUTPUT}\ in$ 

Value section for more informations.

OutputFilePath character (with default): path to the pdf file that will be generated by the function

if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".

SaveEstimates boolean (with default): if TRUE save Bayes estimates, credible interval at level

68% and 95% and the result of the gelman en Rubin test of convergency, in a

csv table named OutputFileName in folder OutputFilePath.

OutputTableName

character (with default): name of the table that will be generated by the function

 $if \ {\tt SaveEstimates=} TRUE.$ 

#### OutputTablePath

character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".

#### StratiConstraints

numeric matrix or character(with default): input object for the statigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints; the user can refer to a matrix (numeric matrix) or to a csv file (character). If there is no stratigraphic relation default value is suitable.

sepSC

character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints csv file.

Model

character (with default): if "full", error on estimate calibration curve is taken account. If "naive" this error is not taken account in the age estimate.

### CalibrationCurve

character (with default): calibration curve choosen. Allowed inputs are

- "AtmosphericNorth" for Northern Hemisphere atmospheric radiocarbon calibration curve,
- "Marine" for Marine radiocarbon calibration curve,
- "AtmosphericSouth" for Southern Hemisphere atmospheric radiocarbon calibration curve
- a csv file, with tree columns, the first column is dedicated to "Cal.BP", the second to "X14C.age", the third to "Error". The decimal of this file must be a dot, and the separator must be a comma.

Iter integer (with default): number of iterations for the MCMC computation (for

more information see jags.model).

t integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see jags.model).

integer (with default): number of independent chains for the model (for more

information see jags.model).

quiet logical (with default): enables/disables rjags messages

# **Details**

Nb\_chaines

\*\* How to fill StratiConstraints? \*\*

If there is stratigraphic relations between samples, 14C age in Data\_C14Cal must be ordered by order of increasing ages.

The user can fill the StratiConstraints matrix as follow.

- 1. **Size of the matrix**: row number of StratiConstraints matrix is equal to Nb\_sample+1, and column number is equal to Nb\_sample.
- 2. **First line of the matrix**: for all i in {1,...,Nb\_Sample}, StratiConstraints[1,i]=1 that means the lower bound of the sample age (given in PriorAge[2i-1]) for the sample whose number ID is equal to i, is taken into account.
- 3. **Sample relations**: for all j in {2,...,Nb\_Sample+1} and all i in {j,...,Nb\_Sample}, StratiConstraints[j,i]=1 if sample age whose number ID is equal to j-1 is lower than sample age whose number ID is equal to i. Otherwise, StratiConstraints[j,i]=0.

Note that StratiConstraints\_{2:Nb\_sample+1,1:Nb\_sample} is a upper triangular matrix.

The user can also use SCMatrix or SC\_Ordered (if all samples are ordered) functions to construct the StratiConstraints matrix.

The user can also refer to a .csv file that containts the relation between samples as defined above. The user must take care about the separator used in the csv file using the argument sepSC.

### \*\* More precision on Model \*\*

We propose two models "full" or "naive". If Model='full' that means measurement error and error on calibration curve are taken account in the Bayesian model; if Model="naive" that means only error on measurement are taken account in the mode.

More precisely, the model considered here, as the one developped by Christen, JA (1994), assume multiplicative effect of errors to address the problem of outliers. In addition, to not penalyse variables that are not outliers and damage theirs estimation, we introduce a structure of mixture, that means only variable that are considered as outlier have in addition a multiplicative error.

#### Value

### **NUMERICAL OUTPUT**

- 1. A list containing the following objects:
  - Sampling: that corresponds to a sample of the posterior distributions of the age parameters:
  - Outlier: stating the names of samples that are considered as outliers;
  - Model: stating which model was chosen ("full" or "naive");
  - CalibrationCurve: stating which radiocarbon calibration curve was chosen;
  - **PriorAge**: stating the priors used for the age parameter;
  - **StratiConstraints**: stating the stratigraphic relations between samples considered in the model.
- 2. The Gelman and Rubin test of convergency: print the result of the Gelman and Rubin test of convergency for the age estimate for each sample. A result close to one is expected. In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see PLOT OUTPUT for more informations). If both convergencies (Gelman and Rubin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC interations (Iter) or being more precise if it is possible on the PriorAge parameter to reach convergency.
- 3. **Credible intervals and Bayes estimates**: prints the Bayes estimates, the credible intervals at 95% and 68% for the age parameters for each sample.

#### PLOT OUTPUT

- 1. **MCMC trajectories**: A graph with the MCMC trajectories and posterior distributions of the age parameter is displayed.
  - On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.
- 2. **Summary of sample age estimates**: plot credible intervals and Bayes estimate of each sample age on one graph.

To give the results in a publication, we recommend to give the Bayes estimate of the parameters as well as the credible interval at 95% or 68%.

#### Note

Please note that the initial values for all MCMC are currently all the same for all chains since we rely on the automatic initial value generation of JAGS. This is not optimal and will be changed in future. However, it does not affect the quality of the age estimates if the chains have converged.

### Author(s)

Claire Christophe, Anne Philippe, Guillaume Guérin

#### References

Christen, JA (1994). Summarizing a set of radiocarbon determinations: a robust approach. Applied Statistics, 489-503.

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

### See Also

```
rjags, plot_MCMC, SCMatrix
```

### **Examples**

```
## Load data
data(DATA_C14,envir = environment())
C14Cal=DATA_C14$C14[,1]
SigmaC14Cal=DATA_C14$C14[,2]
Names=DATA_C14$Names
nb_sample=length(Names)

## Age computation of samples without stratigraphic relations
Age <- AgeC14_Computation(
Data_C14Cal = C14Cal,
Data_SigmaC14Cal = SigmaC14Cal,
SampleNames = Names,
Nb_sample = nb_sample,
PriorAge = rep(c(20000,60000),nb_sample),
Iter = 500,
quiet = TRUE)</pre>
```

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AgeS	Output of AgeS_Computation function for the samples: "GDB5" and "GDB3"

### **Description**

Output of AgeS\_Computation function for the samples: "GDB5" and "GDB3", there is no stratigraphic relation neither systematic errors.

### Usage

```
data("AgeS")
```

#### **Format**

A list containing

Sampling MCMC.list that corresponds to a sample of the posterior distributions of the ages (in ka), palaeodoses (in Gy) and equivalent dose dispersions (in Gy) parameters of samples "GDB5" and "GDB3";

Model\_GrowthCurve stating which dose response fitting option was chosen to run the function

Distribution stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;

PriorAge stating the priors used for the age parameter (in ka);

StratiConstraints stating the matrix of stratigraphic relations between samples considered in the model;

CovarianceMatrix stating the covariance matrix of error used in the model, highlighting not common errors between samples in our cases (diagonal matrix).

# References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L, Pleurdeau, D, Pearson, O, Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

### **Examples**

```
data(AgeS)
str(AgeS)
```

AgeS\_Computation

Bayesian analysis for the OSL age estimation of various samples

### **Description**

This function computes the age (in ka) of at least two samples according to the model developed in Combes and Philippe (2017), based on outputs of Generate\_DataFile or Generate\_DataFile\_MG or both of them using combine\_DataFiles.

Samples, for which data is avalilable in several BIN files, can be analysed.

Single-grain or Multi-grain OSL measurements can be analysed simultaneouly.

# Usage

```
AgeS_Computation(DATA, SampleNames, Nb_sample, PriorAge = rep(c(0.01, 100), Nb_sample), BinPerSample = rep(1, Nb_sample), SavePdf = FALSE, OutputFileName = c("MCMCplot", "summary"), OutputFilePath = c(""), SaveEstimates = FALSE, OutputTableName = c("DATA"), OutputTablePath = c(""), THETA = c(), sepTHETA = c(","), StratiConstraints = c(), sepSC = c(","), LIN_fit = TRUE, Origin_fit = FALSE, distribution = c("cauchy"), Iter = 50000, t = 5, Nb_chaines = 3, quiet = FALSE)
```

#### **Arguments**

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement,
	provided by the function Congrete DataFile or Congrete DataFile MC or

provided by the function Generate\_DataFile or Generate\_DataFile\_MG or combine\_DataFiles. DATA contains informations for more than one sample. If there is stratigraphic relations between samples, informations in DATA must be ordered by order of incresing ages. See the details section to for more informa-

tions.

SampleNames character vector: names of samples. The length of this vector is equal to Nb\_sample.

Nb\_sample integer : number of samples, Nb\_sample>1.

PriorAge numeric vector (with default): lower and upper bounds for age parameter of each

sample (in ka). Note that, length(PriorAge)=2\*Nb\_sample and PriorAge[2i-1, 2i] correponds to the lower and upper bounds of sample whose number ID is equal

to:

to i.

BinPerSample integer vector (with default): vector with the number of BIN files per sample.

The length of this vector is equal to Nb\_sample. BinPerSample[i] correponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in Generate\_DataFile

or in Generate\_DataFile\_MG.

SavePdf boolean (with default): if TRUE save graphs in pdf file named OutputFileName

in folder OutputFilePath.

OutputFileName character (with default): name of the pdf file that will be generated by the func-

tion if SavePdf=TRUE; length(OutputFileName)=2, see PLOT OUTPUT in

Value section for more informations.

OutputFilePath character (with default): path to the pdf file that will be generated by the function

if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".

SaveEstimates boolean (with default): if TRUE save Bayes estimates, credible interval at level

68% and 95% and the result of the gelman en Rubin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.

OutputTableName

character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.

OutputTablePath

character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".

THETA numeric matrix or character (with default): input object for systematic and in-

dividual error. If systematic errors are considered, see the details section for instructions regarding how to correctly fill THETA; the user can refer to a matrix (numeric matrix) or to a csv file (character). Otherwise, default value is suitable,

and only individual errors are considered.

sepTHETA character (with default): if THETA is character, indicate column separator in

THETA csv file.

StratiConstraints

numeric matrix or character(with default): input object for the statigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints; the user can refer to a matrix (numeric matrix) or to a csv file (character). If there

is no stratigraphic relation default value is suitable.

 ${\tt sepSC} \qquad \qquad {\tt character} \ ({\tt with} \ {\tt default}) ; \ {\tt if} \ {\tt StratiConstraints} \ {\tt is} \ {\tt character}, \ {\tt indicate} \ {\tt column}$ 

separator in StratiConstraints .csv file.

LIN\_fit logical (with default): if TRUE (default) allows a linear component, on top of

the (default) saturating exponential curve, for the fitting of dose response curves. See details section for more informations on the proposed dose response curves.

Origin\_fit logical (with default): if TRUE, forces the dose response curves to pass through

the origin. See details section for more informations on the proposed growth

curves.

distribution character (with default): type of distribution that defines how individual equiv-

alent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal\_A" and "lognormal\_M", see details sec-

tion for more informations.

Iter integer (with default): number of iterations for the MCMC computation (for

more information see jags.model).

t integer (with default): 1 every t iterations of the MCMC is considered for sam-

pling the posterior distribution (for more information see jags.model).

Nb\_chaines integer (with default): number of independent chains for the model (for more

information see jags.model).

quiet logical (with default): enables/disables rjags messages

### **Details**

\*\* How to fill StratiConstraints? \*\*

If there is stratigraphic relations between samples, **informations in DATA must be ordered by order of increasing ages**. To do this the user can either fill right Names in Generate\_DataFile or in

Generate\_DataFile\_MG (as it is indicated in Details section of these function), or ordered by order of increasing ages outputs of Generate\_DataFile or Generate\_DataFiles.

The user can fill the StratiConstraints matrix as follow.

- 1. **Size of the matrix**: row number of StratiConstraints matrix is equal to Nb\_sample+1, and column number is equal to Nb\_sample.
- 2. **First line of the matrix**: for all i in {1,...,Nb\_Sample}, StratiConstraints[1,i]=1 that means the lower bound of the sample age (given in PriorAge[2i-1]) for the sample whose number ID is equal to i, is taken into account.
- 3. **Sample relations**: for all j in {2,...,Nb\_Sample+1} and all i in {j,...,Nb\_Sample}, StratiConstraints[j,i]=1 if sample age whose number ID is equal to j-1 is lower than sample age whose number ID is equal to i. Otherwise, StratiConstraints[j,i]=0.

Note that StratiConstraints\_{2:Nb\_sample+A,1:Nb\_sample} is a upper triangular matrix.

The user can also use SCMatrix or SC\_Ordered (if all samples are ordered) functions to construct the StratiConstraints matrix.

The user can also refer to a csv file that containts the relation between samples as defined above. The user must take care about the separator used in the csv file using the argument sepSC.

# \*\* How to fill THETA covariance matrix concerning common and individual error? \*\*

If systematic errors are considered, the user can fill the THETA matrix as follow.

- row number of THETA is equal the column number, equal to Nb\_sample.
- For all i in {1,...,Nb\_sample}, THETA[i,i] containts individual error plus systematic error of the sample whose number ID is equal to i.
- For all i, j in {1,...,Nb\_sample} and i different from j, THETA[i,j] containts common error between samples whose number ID are equal to i and j.

Note that THETA[i, j] is a symetric matrix.

The user can also refer to a .csv file that containts the errors as defined above.

### \*\* Option on growth curves \*\*

As for Age\_Computation and Palaeodose\_Computation, the user can choose from 4 dose response curves:

• Saturating exponential plus linear growth (AgesMultiCS2\_EXPLIN):

```
for all x in IR+, f(x)=a(1-exp(-x/b))+cx+d; select
```

- LIN\_fit=TRUE
- Origin\_fit=FALSE
- Saturating exponential growth (AgesMultiCS2\_EXP):

```
for all x in IR+, f(x)=a(1-exp(-x/b))+d; select
```

- LIN\_fit=FALSE
- Origin\_fit=FALSE
- Saturating exponential plus linear growth and fitting through the origin (AgesMultiCS2\_EXPLINZ0): for all x in IR+, f(x)=a(1-exp(-x/b))+cx; select

```
- LIN_fit=TRUE
```

- Origin\_fit=TRUE
- Saturating exponential growth and fitting through the origin (AgesMultiCS2\_EXPZ0):

for all x in IR+, f(x)=a(1-exp(-x/b)); select

- LIN fit=FALSE
- Origin\_fit=TRUE

### \*\* Option on equivalent dose distribution around the palaeodose \*\*

The use can choose between:

- cauchy: a Cauchy distribution with location parameter equal to the palaeodose of the sample;
- gaussian: a Gaussian distribution with mean equal to the palaeodose of the sample;
- lognormal\_A: a log-normal distribution with mean or Average equal to the palaeodose of the sample:
- lognormal\_M: a log-normal distribution with Median equal to the palaeodose of the sample.

#### Value

### **NUMERICAL OUTPUT**

- 1. A list containing the following objects:
  - **Sampling**: that corresponds to a sample of the posterior distributions of the age (in ka), palaeodose (in Gy) and equivalent dose dispersion (in Gy) parameters for each sample;
  - Model\_GrowthCurve: stating which dose response fitting option was chosen;
  - **Distribution**: stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
  - **PriorAge**: stating the priors used for the age parameter (in ka);
  - **StratiConstraints**: stating the stratigraphic relations between samples considered in the model;
  - CovarianceMatrix: stating the covariance matrix of error used in the model, highlighting common errors between samples or not.
- The Gelman and Rubin test of convergency: prints the result of the Gelman and Rubin test of convergency for the age, palaeodose and equivalent dose dispersion parameters for each sample. A result close to one is expected.
  - In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).
  - If both convergencies (Gelman and Rubin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC interations (Iter) or being more precise on the PriorAge parameter (for example specify if it is a young sample c(0.01,10) an old sample c(10,100)), or changing the parameter distribution or the growth curve, to reach convergency.
- 3. **Credible intervals and Bayes estimates**: prints the Bayes esitmates, the credible intervals at 95% and 68% for the age, palaeodose and equivalent dose dispersion parameters for each sample.

# PLOT OUTPUT

 MCMC trajectories: A graph with the MCMC trajectories and posterior distributions of the age, palaeodose and equivalent dose dispersion parameters is displayed, there is one page per sample.

The first line of the figure correponds to the age parameter, the second to the palaeodose parameter and the third to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

2. **Summary of sample age estimates**: plot credible intervals and Bayes estimate of each sample age on a same graph.

To give the results in a publication, we recommend to give the Bayes estimate of the parameters as well as the credible interval at 95% or 68%.

#### Note

Please note that the initial values for all MCMC are currently all the same for all chains since we rely on the automatic initial value generation of JAGS. This is not optimal and will be changed in future. However, it does not affect the quality of the age estimates if the chains have converged.

### Author(s)

Claire Christophe, Anne Philippe, Guillaume Guérin

### References

Combes, Benoit and Philippe, Anne, 2017. Bayesian analysis of multiplicative Gaussian error for multiple ages estimation in optically stimulated luminescence dating. Quaternary Geochronology (39, 24-34)

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. Quaternary Geochronology 28, 62-70. doi:10.1016/j.quageo.2015.04.001

# See Also

Generate\_DataFile, Generate\_DataFile\_MG, rjags, plot\_MCMC, SCMatrix Age\_Computation, Palaeodose\_Computation

### **Examples**

```
## load data
data(DATA1,envir = environment())
data(DATA2,envir = environment())
Data <- combine_DataFiles(DATA2,DATA1)

## Age computation of samples GDB5 and GDB3,
priorage=c(1,10,20,60) # these samples are not young
## without common error and without stratigraphic constraints
Age <- AgeS_Computation(
   DATA = Data,
   Nb_sample = 2,
   SampleNames = c("GDB5","GDB3"),
   PriorAge = priorage,
   Iter = 50,
   Nb_chaines = 2,
   quiet = TRUE
   )</pre>
```

## Age computation of samples GDB5 and GDB3,

```
## without common error, assuming GDB5 age younder than GDB3 age
## Not run:
Nb_sample <- 2
SC <- matrix(</pre>
  data = c(1,1,0,1,0,0),
  ncol = 2,
  nrow = (Nb_sample+1),byrow = T)
Age <- AgeS_Computation(
 DATA = Data,
 Nb_sample = Nb_sample,
 SampleNames = c("GDB5", "GDB3"),
 PriorAge = priorage,
 StratiConstraints = SC,
 Iter = 10000,
 quiet = FALSE)
## End(Not run)
```

Age\_Computation

Bayesian analysis for the OSL age estimation of one sample

### **Description**

This function computes the age (in ka) of a sample according to the model developed in Combes and Philippe (2017), based on an output of Generate\_DataFile or Generate\_DataFile\_MG. A sample, for which data is available in several BIN files, can be analysed.

# Usage

```
Age_Computation(DATA, SampleName, PriorAge = c(0.01, 100),
BinPerSample = c(1), SavePdf = FALSE, OutputFileName = c("MCMCplot"),
OutputFilePath = c(""), SaveEstimates = FALSE,
OutputTableName = c("DATA"), OutputTablePath = c(""), LIN_fit = TRUE,
Origin_fit = FALSE, distribution = c("cauchy"), I = 1, Iter = 50000,
t = 5, Nb_chaines = 3, quiet = FALSE)
```

# **Arguments**

DATA list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J, K, Nb\_measurement,

provided by the function Generate\_DataFile or Generate\_DataFile\_MG. DATA

can contain informations for more than one sample.

SampleName character: name of the sample.

PriorAge numeric vector (with default): lower and upper bounds for the sample age pa-

rameter (in ka). Note that, length(PriorAge)=2.

BinPerSample integer vector (with default): vector with the number of BIN files per sample.

If in DATA there is more than one sample, the BinPerSample vector must be the

same as that used to run the function Generate\_DataFile or in Generate\_DataFile\_MG

for generating the DATA object.

SavePdf boolean (with default): if TRUE save graph in pdf file named OutputFileName

in folder OutputFilePath.

OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE; length(OutputFileName)=2, see <b>PLOT OUTPUT</b> in <b>Value</b> section for more informations.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".
SaveEstimates	boolean (with default): if TRUE save Bayes estimates and credible interval at level 68% and 95% and the result of the gelman en Rubin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	
	character (with default): name of the table that will be generated by the function if $SaveEstimates=TRUE$ .
OutputTablePath	1
	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. See details section for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. See details section for more informations on the proposed growth curves.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M", see details section for more informations.
I	integer (with default): if DATA contains data from more than one sample, I indicates the ID number of the sample to be analysed.
Iter	integer (with default): number of iterations for the MCMC computation (for more information see ${\tt jags.model}$ ).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see $jags.model$ ).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see ${\tt jags.model}$ ).
quiet	logical (with default): enables/disables rjags messages

# **Details**

# \*\* Option on growth curves \*\*

As for  $AgeS\_Computation$  and  $Palaeodose\_Computation$ , the user can choose from 4 dose response curves:

 $\bullet \ \ \textbf{Saturating exponential plus linear growth} \ (\texttt{AgeMultiBF\_EXPLIN}) :$ 

```
for all x in IR+, f(x)=a(1-exp(-x/b))+cx+d; select
```

- LIN\_fit=TRUE
- Origin\_fit=FALSE
- **Saturating exponential growth** (AgeMultiBF\_EXP):

```
for all x in IR+, f(x)=a(1-exp(-x/b))+d; select
```

- LIN\_fit=FALSE

- Origin\_fit=FALSE
- $\bullet \ \ \textbf{Saturating exponential plus linear growth and fitting through the origin (Age \texttt{MultiBF\_EXPLINZO}):} \\$

for all x in IR+, f(x)=a(1-exp(-x/b))+cx; select

- LIN fit=TRUE
- Origin\_fit=TRUE
- Saturating exponential growth and fitting through the origin (AgeMultiBF\_EXPZO):

for all x in IR+, f(x)=a(1-exp(-x/b)); select

- LIN\_fit=FALSE
- Origin\_fit=TRUE

### \*\* Option on equivalent dose distribution around the palaeodose \*\*

The use can choose between:

- cauchy: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- gaussian: a Gaussian distribution with mean equal to the palaeodose of the sample
- lognormal\_A: a log-normal distribution with mean or Average equal to the palaeodose of the sample
- lognormal\_M: a log-normal distribution with Median equal to the palaeodose of the sample

#### Value

### NUMERICAL OUTPUT

- 1. A list containing the following objects:
  - **Sampling** that corresponds to a sample of the posterior distributions of the age (in ka), palaeodose (in Gy) and equivalent dose dispersion (in Gy) parameters.
  - Model\_GrowthCurve, stating which dose response fitting option was chosen;
  - **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
  - **PriorAge**, stating the priors used for the age parameter (in ka).
- 2. **The Gelman and Rubin test of convergency**: prints the result of the Gelman and Rubin test of convergency for the age, palaeodose and equivalent dose dispersion parameters. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

- If both convergencies (Gelman and Rubin test and plot checking) are satisfactory, the user can consider the printed estimates as valid. Otherwise, the user may try increasing the number of MCMC interations (Iter), or being more precise on the PriorAge parameter (for example specify if it is a young sample c(0.01,10) an old sample c(10,100)), or changing the parameter distribution or the growth curve, to reach convergency.
- 3. **Credible intervals and Bayes estimates**: prints the Bayes estimates, the credible intervals at 95% and 68% for the age, palaeodose and equivalent dose dispersion parameters of the sample.

### PLOT OUTPUT

A graph with the MCMC trajectories and posterior distributions of the age, palaeodose and equivalent dose dispersion parameters is displayed.

The first line of the figure correponds to the age parameter, the second to the palaeodose parameter

and the third to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

To give the results in a publication, we recommend to give the Bayes estimate of the parameter as well as the credible interval at 95% or 68%.

#### Note

Please note that the initial values for all MCMC are currently all the same for all chains since we rely on the automatic initial value generation of JAGS. This is not optimal and will be changed in future. However, it does not affect the quality of the age estimates if the chains have converged.

#### Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guérin

#### References

Combes, Benoit and Philippe, Anne, 2017. Bayesian analysis of multiplicative Gaussian error for multiple ages estimation in optically stimulated luminescence dating. Quaternary Geochronology (39, 24-34)

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. Quaternary Geochronology 28, 62-70. doi:10.1016/j.quageo.2015.04.001

#### See Also

Generate\_DataFile, Generate\_DataFile\_MG, rjags, plot\_MCMC AgeS\_Computation, Palaeodose\_Computation

### **Examples**

```
## load data file generated by the function Generate_DataFile
data(DATA1,envir = environment())
priorage <- c(10,60) # GDB3 is an old sample
Age <- Age_Computation(
DATA = DATA1,
    SampleName = "GDB3",
    PriorAge = priorage,
    Iter = 100,
    quiet = TRUE)</pre>
```

Age\_OSLC14

Bayesian analysis for age estimation of OSL measurments and 14C ages of various samples

# Description

This function compute an age of OSL data of at least two samples and calibrate 14C ages of samples to get an age (in ka).

Age of OSL data are computed according to the model given in Combes and Philippe (2017). Single-grain or Multi-grain OSL measurements can be analysed simultaneouly (with output of Generate\_DataFile or Generate\_DataFile\_MG or both of them using combine\_DataFiles).

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Samples, for which data is avalilable in several BIN files, can be analysed.

For C14 data, the user can choose one of the following radiocarbon calibration curve: Northern or Sourthen Hemisphere or marine atmospheric.

### Usage

```
Age_OSLC14(DATA, Data_C14Cal, Data_SigmaC14Cal, Nb_sample, SampleNames,
   SampleNature, PriorAge = rep(c(10, 60), Nb_sample), SavePdf = FALSE,
   OutputFileName = c("MCMCplot", "HPD_Cal14CCurve", "summary"),
   OutputFilePath = c(""), SaveEstimates = FALSE,
   OutputTableName = c("DATA"), OutputTablePath = c(""),
   StratiConstraints = c(), sepSC = c(","), BinPerSample = rep(1,
   sum(SampleNature[1, ])), THETA = c(), sepTHETA = c(","), LIN_fit = TRUE,
   Origin_fit = FALSE, distribution = c("cauchy"), Model_C14 = c("full"),
   CalibrationCurve = c("AtmosphericNorth"), Iter = 50000, t = 5,
   Nb_chaines = 3, quiet = FALSE)
```

### **Arguments**

DATA list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J, K, Nb\_measurement,

provided by the function Generate\_DataFile or Generate\_DataFile\_MG or combine\_DataFiles. DATA contains information for more than one sample. If there is stratigraphic relations between samples, informations in DATA must be ordered by order of incresing ages. See the details section to for more informa-

tions.

Data\_C14Cal numeric vector: corresponding to 14C age estimate (in years, conversion in ka is

automatically donne in the function). If there is stratigraphic relations between

samples,  $Data\_C14Cal$  must be ordered by order of incresing ages.

Data\_SigmaC14Cal

numeric vector: correponding to the error of 14C age estimates.

Nb\_sample integer: number of samples (OSL data and 14C age), (Nb\_sample>3, at least to

sample of OSL data and one sample of 14C age).

SampleNames character vector: sample names for both OSL data and C14 data. The length

of this vector is equal to Nb\_sample. If there is stratigephic relation, this vector must be ordered by increasing order (to mix OSL samples and 14C ages if it is

needed).

SampleNature numeric matrix: states the nature of the sample. Row number of SampleNature

matrix is equal to 2 and column number is equal to Nb\_sample. First line of the matrix: SampleNature[1,i] states if sample whose number ID is equal to i, is an OSL sample 1 or not 0. Second line of the matrix: SampleNature[2,i]

states if sample whose number ID is equal to i, is an 14C sample 1 or not 0.

PriorAge numeric vector (with default): lower and upper bounds for age parameter of each

sample (in ka). Note that, length(PriorAge)=2\*Nb\_sample and PriorAge[2i-1,2i]

correponds to the lower and upper bounds of sample whose number ID is equal

to i.

SavePdf boolean (with default): if TRUE save graphs in pdf file named OutputFileName

in folder OutputFilePath.

OutputFileName character (with default): name of the pdf file that will be generated by the func-

tion if SavePdf=TRUE, length(OutputFileName)=3, see PLOT OUTPUT in

Value section for more informations.

OutputFilePath character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".

SaveEstimates boolean (with default): if TRUE save Bayes estimates, credible interval at level

68% and 95% and the result of the gelman en Rubin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.

OutputTableName

character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.

OutputTablePath

character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".

StratiConstraints

numeric matrix or character(with default): input object for the statigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints, the user can refer to a matrix (numeric matrix) or to a csv file (character). Otherwise, default value is suitable.

sepSC character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints csv file.

BinPerSample integer vector (with default): vector with the number of BIN files per OSL sample. The length of this vector is equal to the number of OSL samples. BinPerSample[i] correponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to

detatils in Generate\_DataFile or in Generate\_DataFile\_MG.

THETA numeric matrix or character (with default): input object for systematic and individual error for OSL samples. If systematic errors are considered, see the details section for instructions regarding how to correctly fill THETA; the user can refer to a matrix (numeric matrix) or to a csv file (character). Otherwise, default value

is suitable, and only individual error is considered.

sepTHETA character (with default): if THETA is character, indicate column separator in

THETA csv file.

LIN\_fit logical (with default): if TRUE (default) allows a linear component, on top of the

(default) saturating exponential curve, for the fitting of dose response curves, for OSL samples. See details for more informations on the proposed dose response

curves.

Origin\_fit logical (with default): if TRUE, forces the dose response curves to pass through

the origin. See details for more informations on the proposed growth curves, for

OSL samples.

distribution character (with default): type of distribution that defines how individual equiv-

alent dose values are distributed around the palaeodose, for OSL samples. Allowed inputs are "cauchy", "gaussian", "lognormal\_A" and "lognormal\_M",

see details for more informations.

Model\_C14 character (with default): if "full", error on estimate calibration curve is taken

account, for 14C samples. If "naive" this error is not taken account in the age

estimate.

CalibrationCurve

character (with default): calibration curve choosen, for 14C samples. Allowed inputs are

 "AtmosphericNorth" for Northern Hemisphere atmospheric radiocarbon calibration curve,

- "Marine" for Marine radiocarbon calibration curve,
- "AtmosphericSouth" for Southern Hemisphere atmospheric radiocarbon calibration curve.

• a csv file, with tree columns, the first column is dedicated to "Cal.BP", the second to "X14C.age", the third to "Error". The decimal of this file must be a dot, and the separator must be a comma.

Iter integer (with default): number of iterations for the MCMC computation (for

more information see jags.model).

t integer (with default): 1 every t iterations of the MCMC is considered for sam-

pling the posterior distribution (for more information see  ${\tt jags.model}$ ).

Nb\_chaines integer (with default): number of independent chains for the model (for more

information see jags.model).

quiet logical (with default): enables/disables rjags messages

### **Details**

Note that there is tree type of arguments in the previous list. There are arguments for informtations concerning only OSL samples: DATA, BinPerSample, THETA, sepTHETA, LIN\_fit, Origin\_fit, distribution.

There are arguments for informtations concerning only C14 samples: Data\_C14Cal, Data\_SigmaC14Cal, Model\_C14, CalibrationCurve.

There are arguments for informtaions concerning all the samples: Nb\_sample, SampleNames, SampleNature, PriorAge, SavePdf, OutputFileName, OutputFilePath, SaveEstimates, OutputTableName, OutputTablePath, StratiConstraints, sepSC.

### \*\* How to fill StratiConstraints? \*\*

If there is stratigraphic relations between samples, 14C estimate age in Data\_C14Cal must be ordered by order of increasing ages, as informations in DATA. Names in SampleNames must be ordered and correponds to the order in Data\_C14Cal and in DATA, also if it is needed to mix names of OSL samples and 14C samples.

The user can fill the StratiConstraints matrix as follow.

- 1. **Size of the matrix**: row number of StratiConstraints matrix is equal to Nb\_sample+1, and column number is equal to Nb\_sample.
- 2. **First line of the matrix**: for all i in {1,...,Nb\_Sample}, StratiConstraints[1,i]=1 that means the lower bound of the sample age (given in PriorAge[2i-1]) for the sample whose number ID is equal to i, is taken into account.
- 3. **Sample relations**: for all j in {2,...,Nb\_Sample+1} and all i in {j,...,Nb\_Sample}, StratiConstraints[j,i]=1 if sample age whose number ID is equal to j-1 is lower than sample age whose number ID is equal to i. Otherwise, StratiConstraints[j,i]=0.

Note that  $StratiConstraints_{2:Nb\_sample+1,1:Nb\_sample}$  is a upper triangular matrix.

The user can also use SCMatrix or SC\_Ordered (if all samples are ordered) function to construc the StratiConstraints matrix.

The user can also refer to a csv file that containts the relation between samples as defined above. The user must take care about the separator used in the csv file using the argument sepSC.

### \*\* How to fill THETA covariance matrix concerning common and individual error? \*\*

If systematic errors are considered, the user can fill the THETA matrix as follow.

- row number of THETA is equal the column number, equal to Nb\_sample.
- For all i in {1,...,Nb\_sample}, THETA[i,i] containts individual error plus systematic error of the sample whose number ID is equal to i.
- For all i, j in {1,...,Nb\_sample} and i different from j, THETA[i,j] containts common error between samples whose number ID are equal to i and j.

Note that THETA[i,j] is a symetric matrix.

The user can also refer to a .csv file that containts the errors as defined above.

### \*\* Option on growth curves \*\*

As for Age\_Computation and Palaeodose\_Computation, the user can choose from 4 dose response curves:

• Saturating exponential plus linear growth (AgesMultiCS2\_EXPLIN):

```
for all x in IR+, f(x)=a(1-exp(-x/b))+cx+d; select
```

- LIN\_fit=TRUE
- Origin\_fit=FALSE
- Saturating exponential growth (AgesMultiCS2\_EXP):

```
for all x in IR+, f(x)=a(1-exp(-x/b))+d; select
```

- LIN\_fit=FALSE
- Origin\_fit=FALSE
- Saturating exponential plus linear growth and fitting through the origin (AgesMultiCS2\_EXPLINZO):

```
for all x in IR+, f(x)=a(1-exp(-x/b))+cx; select
```

- LIN\_fit=TRUE
- Origin\_fit=TRUE
- Saturating exponential growth and fitting through the origin (AgesMultiCS2\_EXPZ0):

```
for all x in IR+, f(x)=a(1-exp(-x/b)); select
```

- LIN\_fit=FALSE
- Origin\_fit=TRUE

# \*\* Option on equivalent dose distribution around the palaeodose \*\*

The use can choose between:

- cauchy: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- gaussian: a Gaussian distribution with mean equal to the palaeodose of the sample
- lognormal\_A: a log-normal distribution with mean or Average equal to the palaeodose of the sample
- lognormal\_M: a log-normal distribution with Median equal to the palaeodose of the sample

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#### \*\* More precision on Model \*\*

We propose two models "full" or "naive". If Model='full' that means measurement error and error on calibration curve are taken account in the Bayesian model; if Model="naive" that means only error on measurement are taken account in the mode.

More precisely, the model considered here, as the one developped by Christen, JA (1994), assume multiplicative effect of errors to address the problem of outliers. In addition, to not penalyse variables that are not outliers and damage theirs estimation, we introduce a structure of mixture, that means only variable that are considered as outlier have in addition a multiplicative error.

#### Value

#### NUMERICAL OUTPUT

#### 1. A list containing the following objects:

- **Sampling**: that corresponds to a sample of the posterior distributions of the age parameters (in ka for both C14 samples and OSL samples);
- **PriorAge**: stating the priors used for the age parameter;
- **StratiConstraints**: stating the stratigraphic relations between samples considered in the model:
- Model\_OSL\_GrowthCurve: stating which dose response fitting option was chosen;
- **Model\_OSL\_Distribution**: stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
- Model\_C14: stating which model was chosen ("full" or "naive");
- CalibrationCurve: stating which radiocarbon calibration curve was chosen;
- Outlier: stating the names of samples that must be outliers.
- 2. The Gelman and Rubin test of convergency: prints the result of the Gelman and Rubin test of convergency for the age estimate for each sample. A result close to one is expected. In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see PLOT OUTPUT for more informations). If both convergencies (Gelman and Rubin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC interations (Iter) or be more precise on the PriorAge parameter to reach convergency.
- 3. **Credible intervals and Bayes estimates**: prints the Bayes estimates, the credible intervals at 95% and 68% for the age parameters for each sample.

# PLOT OUTPUT

- 1. **MCMC trajectories**: A graph with the MCMC trajectories and posterior distributions of the age parameter is displayed.
  - On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.
- 2. **Age estimate and HPD at 95% of 14C samples on calibration curve**: plot age estimate and HPD on calibration plot.
- 3. **Summary of sample age estimates**: plot credible intervals and Bayes estimate of each sample age on a same graph.

#### Note

Please note that the initial values for all MCMC are currently all the same for all chains since we rely on the automatic initial value generation of JAGS. This is not optimal and will be changed in future. However, it does not affect the quality of the age estimates if the chains have converged.

#### Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

### References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine 13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

### See Also

```
rjags, plot_MCMC, SCMatrix
```

### **Examples**

```
## Load data
# OSL data
data(DATA1,envir = environment())
data(DATA2,envir = environment())
Data <- combine_DataFiles(DATA2,DATA1)</pre>
# 14C data
C14Cal <- DATA_C14$C14[1,1]
SigmaC14Cal <- DATA_C14$C14[1,2]</pre>
Names <- DATA_C14$Names[1]</pre>
# Prior Age
prior=rep(c(1,60),3)
samplenature=matrix(data=c(1,0,1,0,1,0),ncol=3,nrow=2,byrow=TRUE)
SC <- matrix(data=c(1,1,1,0,1,1,0,0,1,0,0,0),ncol=3,nrow=4,byrow=TRUE)
## Age computation of samples
Age <- Age_OSLC14(DATA=Data,Data_C14Cal=C14Cal,Data_SigmaC14Cal=SigmaC14Cal,
   SampleNames=c("GDB5", Names, "GDB3"), Nb_sample=3, SampleNature=samplenature,
   PriorAge=prior,StratiConstraints=SC,Iter=50,Nb_chaines=2)
```

AtmosphericNorth\_CalC14

Atmospheric North data for calibration of 14C age

### **Description**

As 14C years is not equal to calendar years because atmospheric 14C concentration varies through time. Hence, data in AtmosphericNorth\_CalC14 allows a calibration for mid-latitude Northern Hemisphere atmospher reservoir.

### Usage

```
data("AtmosphericNorth_CalC14")
```

#### **Format**

A data frame with 5 variables.

CAL.BP a numeric vector correpondig to calendar years befor present

X14C. age a numeric vector correponding to 14C age

Error a numeric vector correponding to error arround 14C age measurement

### References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

### **Examples**

```
data(AtmosphericNorth_CalC14)
## maybe str(AtmosphericNorth_CalC14) ; head(AtmosphericNorth_CalC14) ...
```

AtmosphericSouth\_CalC14

Atmospheric South data for calibration of 14C age

# **Description**

As 14C years is not equal to calendar years because atmospheric 14C concentration varies through time. Hence, data in AtmosphericSouth\_CalC14 allows a calibration for mid-latitude Southern Hemisphere atmospher reservoir.

### Usage

```
data("AtmosphericSouth_CalC14")
```

24 combine\_DataFiles

#### **Format**

A data frame with 3 variables.

CAL.BP a numeric vector correpondig to calendar years befor present

X14C.age a numeric vector correponding to 14C age

Error a numeric vector correponding to error arround 14C age measurement

#### References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine 13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

### **Examples**

### **Description**

```
Combine objects generated by Generate_DataFile and Generate_DataFile_MG Old function Concat_DataFile()
```

### Usage

```
combine_DataFiles(...)
Concat_DataFile(...)
```

### Arguments

. list objects generated by Generate\_DataFile or Generate\_DataFile\_MG

### **Details**

The function allows to combine data already generated by Generate\_DataFile or Generate\_DataFile\_MG. The number of input objects is not limited and the function works similar to the standard base R function c(), but preserves the particular structure of the objects imported and generated by 'Bay-Lum'. The elements are combined by list element names.

Combining such data is rather useful in two scenarious:

- The data have been already imported and treated and then stored in RData-files. Using the function combine\_DataFiles() will significantly speed up the processing time,
- simultaneous analysis of single and multi-grain OSL measurements.

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#### Value

A nested list combining the input objects.

#### **Function version**

0.1.0

#### Author(s)

Sebastian Kreutzer, IRAMAT-CRP2A, UMR 5060, CNRS - Université Bordeaux Montaigne (France), adapting the idea from the function 'Concat\_DataFile()' by Claire Christophe.

#### See Also

```
Generate_DataFile, Generate_DataFile_MG
```

# **Examples**

```
# load data files
data(DATA1,envir = environment())
data(DATA2,envir = environment())
#combine objects
DATA3 <- combine_DataFiles(DATA1, DATA2)
str(DATA3)</pre>
```

DATA1

DATA of sample named GDB3

### **Description**

list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J,K,Nb\_measurement obtained using  ${\tt Generate\_DataFile} \ function \ with \ single-grain \ OSL \ measurements l \ of \ the \ sample \ GDB3.$ 

### Usage

```
data("DATA1")
```

# Format

A list containing:

LT: (one list per sample): each list contains all L/T values for the corresponding sample;

sLT: (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes: (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab= a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot\_env: a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose: (one list per sample): each list contains all regenerated doses;

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J: a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K: a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb\_measurement: a vector giving, for each BIN file, the number of measurements;

#### References

For more informations on this sample we refer to the following publication:

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L, Pleurdeau, D, Pearson, O, Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

### **Examples**

```
data(DATA1)
str(DATA1)
```

DATA2

DATA on sample named GDB5

### **Description**

list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J,K,Nb\_measurement obtained using Generate\_DataFile function with single-grain OSL measurementsl of the sample GDB5.

### Usage

```
data("DATA2")
```

### **Format**

A data frame containing:

LT: (one list per sample): each list contains all L/T values for the corresponding sample;

sLT: (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes: (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab: a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot\_env: a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose: (one list per sample): each list contains all regenerated doses;

J: a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K: a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb\_measurement: , a vector giving, for each BIN file, the number of measurements;

DATA3

#### References

For more informations on this sample we refer to the following publication:

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L, Pleurdeau, D, Pearson, O, Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

### **Examples**

data(DATA2)
str(DATA2)

DATA3

DATA of sample named FER1

### **Description**

list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J,K,Nb\_measurement obtained using Generate\_DataFile function with multi-grain OSL measurementsl of the sample FER1.

#### Usage

data("DATA3")

#### **Format**

A list containing:

LT: (one list per sample): each list contains all L/T values for the corresponding sample;

sLT: (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes: (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab= a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot\_env: a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose: (one list per sample): each list contains all regenerated doses;

J: a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K: a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb\_measurement: a vector giving, for each BIN file, the number of measurements;

### References

For more informations on this sample we refer to the following publication:

Guerin, G., Frouin, M., Talamo, S., Aldeias, V., Bruxelles, L., Chiotti, L., Goldberg, P., Hublin, J.J., Jain, M., Lahaye, C., Madelaine, S., Maureille, B., McPherron, S., Mercier, N., Murray, A., Sandgathe, D., Steele, T., Thomsen, K., Turq, A. (2015). A multi-method luminescence dating of the Palaeolithic sequence of La Ferrassie based on new excavations adjacent to the La Ferrassie 1 and 2 skeletons. Journal of Archaeological Science, 58, 147-166.

#### **Examples**

data(DATA3)
str(DATA3)

DATA\_C14

C14 cal age estiamte and its error

# **Description**

C14 cal age estiamtes and theirs error of samples S-EVA-26510, S-EVA-26506, S-EVA-26507, S-EVA-26508.

### Usage

```
data("DATA_C14")
```

### **Format**

A list containing:

Names: character vector of the sample names;

C14: numeric matrix, in the first column the 14C Cal age of the samples, and in the second column theirs errors.

#### References

For more informations on this sample we refer to the following publication:

Guerin, G., Frouin, M., Talamo, S., Aldeias, V., Bruxelles, L., Chiotti, L., Goldberg, P., Hublin, J.J., Jain, M., Lahaye, C., Madelaine, S., Maureille, B., McPherron, S., Mercier, N., Murray, A., Sandgathe, D., Steele, T., Thomsen, K., Turq, A. (2015). A multi-method luminescence dating of the Palaeolithic sequence of La Ferrassie based on new excavations adjacent to the La Ferrassie 1 and 2 skeletons. Journal of Archaeological Science, 58, 147-166.

# **Examples**

```
data(DATA_C14)
(DATA_C14)
```

Generate\_DataFile

Generates, from one (or several) BIN-file(s) of Single-grain OSL measurements, a list of luminescence data and information before statistical analysis

### **Description**

This function is used to generate, from the BIN file(s), a list of values of: **Single-grain** OSL intensities and associated uncertainties, regenerative doses, etc., which will be the input of the Bayesian models. To be easy-to-use, this function requires a rigorous organisation - all needed files should be arranged in one folder - of informations concerning each BIN file.

It is possible to process data for various samples simultaneously and to consider more than one BIN file per sample.

#### Usage

```
Generate_DataFile(Path, FolderNames, Nb_sample,
  Nb_binfile = length(FolderNames), BinPerSample = rep(1, Nb_sample),
  sepDP = c(","), sepDE = c(","), sepDS = c(","), sepR = c("="),
  verbose = TRUE, ...)
```

### Arguments

Path character (required): the path to the project folder, containing one or more

subfolders in which the BIN files are located. If it is not equal to "", it must be

terminated by "/".

FolderNames character (required): list of names of the sub-folders containing the BIN files

each subfolder must contain a BIN file and associated csv files. See details
for more informations on associated csv files required in the subfolders.
If there is more than one BIN file per sample, see the details section for
instructions regarding how to correctly fill the FolderNames vector.

Nb\_sample integer (**required**): number of samples.

Nb\_binfile integer (with default): number of BIN files. It must be equal to, or greater than

Nb\_sample.

BinPerSample integer vector (with default): vector with the number of BIN files per sample.

The length of this vector must be equal to Nb\_sample and the sum of entries of this vector must be equal to Nb\_binfile. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill

BinPerSample vector. Otherwise, this vector must contain a list of 1 values.

sepDP character (with default): column separator in the DiscPose.csv files.
sepDE character (with default): column separator in the DoseEnv.csv files.
sepDS character (with default): column separator in the DoseLab.csv files.
sepR character (with default): column separator in the Rule.csv files.

verbose logical (with default): enable/disable verbose mode

... further arguments that can be passed to Luminescence::read\_BIN2R.

# **Details**

With Path and FolderNames, this function goes to the subfolders containing the BIN files and associated information to compute the luminescence data.

# \*\* What are the required files in each subfolder? \*\*

Each subfolder can be named, for example, as the sample name followed by a number; it must contain:

- bin.BIN: the bin file renamed as bin.BIN (note: the name of all files matters);
- **DiscPos.csv**: a two columns csv file containing the list of disc and grain position number of the previously selected grains (typically this list will include the position of grains based on their sensitivity, recycling or other properties);
- **DoseEnv.csv**: a two columns file containing the observation of the natural (or environmental), dose rate, and its non-shared variance (i.e. after removing all shared errors), both in Gy. Note: the user shall provide the squared value of the error associated with the dose rate experienced by the sample grains in nature;

• **DoseSourve.csv**: a two columns file containing the observation of the laboratory dose rate, and its variance (squared error) both in Gy;

- rule.csv: a csv file containing information on
  - beginSignal= the first channel for summing the natural or regenerative OSL signal (typically 1 or 6);
  - endSignal= the last channel for summing the natural or regenerative OSL signal (typically 5 or 10);
  - beginBackground= the first channel for background estimation of the natural or regenerative OSL signal (typically 76 or 81);
  - endBackground= the last channel for background estimation of the natural or regenerative
     OSL signal (typically 95 or 100);
  - beginTest=,
  - endTest=,
  - beginTestBackground=,
  - endTestBackground= same values as above, for the test dose response (typically the same values should be used);
  - inflatePercent= uncertainty arising from the instrument reproducibility (typically 0.02, i.e. 2%);
  - nbOfLastCycleToRemove= number of cycles at the end of the SAR protocol which should not be included in the dose response curve fitting (typically 1 if only a recycling test is performed, or 2 if both recycling and IR depletion are tested).

#### \*\* How to fill the FolderNames vector? \*\*

FolderNames is a vector of length Nb\_binfile. FolderNames[i] is the name (e.g., Sample1-File1, or successive names separated by "/" signs, if BIN files are in subfolders, e.g. Sample1/File1) of the subfolder containing all informations on the BIN file of ID number i. The names in FolderNames are ordered following two rules:

- The names in the FolderNames vector must be ordered following the sample order (the names of subfolders containing BIN files for the same sample should follow each other in the FolderNames vector, e.g. Sample1, Sample2-File1, Sample2-File2, etc.).
- If stratigraphic constraints apply to samples, and so a **Bayesian model with stratigraphic constraints** is implemented, then the names in the FolderNames vector must be ordered by order of increasing ages.

For example, FolderNames=c(noun1, noun2), in which case noun1 (respectively, noun2) corresponds to the subfolder name containing the BIN file of sample 1 (respectively of sample 2). In addition, if we know that sample 1 is younger than sample 2, then FolderNames vector is correctly filled.

If conversely, FolderNames=c(noun2, noun1), the analysis performed by  $AgeS\_Computation$  would not be consistent.

# \*\* How to fill the BinPerSample vector? \*\*

BinPerSample[i] correponds to the number of BIN files for the sample whose number ID is equal to i.

For example, let us consider a case with two samples (Sample1 and Sample2), with 2 BIN files for Sample1 and 1 for Sample2. In this case, Nb\_binfile=3 and Nb\_sample=2. The user may then set FolderNames=c("Sample1-File1", "Sample1-File2", "Sample2-File1"), in which

```
case "Sample1-File1" is the name of the subfolder containing the first BIN file for Sample1, "Sample1-File2" the name of the subfolder for the second BIN file of Sample1; eventually, "Sample2-File1" is the name of the subfolder containing the BIN file for the second sample. In this case, BinPerSample=c(2,1).
```

For the general BIN-file structure, the reader is referred to the following website: http://www.nutech.dtu.dk/ The function read\_BIN2R developed in Luminescence package is used to read the BIN files.

#### Value

A list containing the following objects:

- LT (one list per sample); each list contains all L/T values for the corresponding sample;
- sLT (one list per sample); each list contains all uncertainties on L/T values for the corresponding sample;
- **ITimes** (one list per sample); each list contains irradiation time values for the corresponding sample;
- dLab, a matrix containing in line i, the laboratory dose rate and its variance for sample i;
- **ddot\_env**, a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;
- regDose (one list per sample); each list contains all regenerated doses;
- J, a vector giving, for each BIN file, the number of aliquots selected for the analysis;
- K, a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;
- **Nb\_measurement**, a vector giving, for each BIN file, the number of measurements.

### \*\* How to save this list \*\*

You can save this list in a .RData object. To do this, you can use the fonction save. Then, to load this list you can use the function load (see example section fore more details).

### Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guerin

# See Also

```
read_BIN2R, combine_DataFiles, Generate_DataFile_MG, LT_RegenDose Age_Computation, AgeS_Computation, Palaeodose_Computation
```

#### **Examples**

```
## Example for one sample with one Bin File
path<- system.file("extdata/samp1", "", package="BayLum")
folder=""
nbsample=1 # give the number of sample
Data <- Generate_DataFile(
  Path = path,
  FolderNames = folder,
  Nb_sample = nbsample,
  verbose = FALSE)
str(Data)</pre>
```

```
## to save information in RData object in folder containing bin file
# save(Data,file=c(paste(path,folder,'Data.RData',sep="")))
## to load information containing Data.RData object
# load(file=c(paste(path,folder,"Data.RData",sep="")))
```

Generates, from one (or several) BIN file(s) of Multi-grain OSL measurements, a list of luminescence data and information before statistical analysis

### **Description**

This function is used to generate, from the BIN file(s), a list of values of: **Multi-grain** OSL intensities and associated uncertainties, regenerative doses, etc., which will be the input of the Bayesian models. To be easy-to-use, this function requires a rigorous organisation - all needed files should be arranged in one folder - of informations concerning each BIN file.

It is possible to process data for various samples simultaneously and to consider more than one BIN file per sample.

### Usage

```
Generate_DataFile_MG(Path, FolderNames, Nb_sample,
  Nb_binfile = length(FolderNames), BinPerSample = rep(1, Nb_sample),
  sepD = c(","), sepDE = c(","), sepDS = c(","), sepR = c("="),
  verbose = TRUE, ...)
```

### **Arguments**

Path

character (**required**): the path to the project folder, containing one or more subfolders in which the BIN files are located. If it is not equal to "", it must be terminated by "/".

FolderNames

character (required) vector: list of names of the sub-folders containing the BIN files

each subfolder must contain a BIN file and associated csv files. See details
for more informations on associated csv files required in the subfolders.
If there is more than one BIN file per sample, see the details section for
instructions regarding how to correctly fill the FolderNames vector.

Nb\_sample integer (**required**): number of samples.

Nb\_binfile integer (with default): number of BIN files. It must be equal to, or greater than

Nb\_sample.

BinPerSample integer vector (with default): vector with the number of BIN files per sample.

The length of this vector must be equal to Nb\_sample and the sum of entries of this vector must be equal to Nb\_binfile. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill BinPerSample vector. Otherwise, this vector must contain a list of 1 values.

sepD character (with default): column separator in the DiscPose.csv files.
sepDE character (with default): column separator in the DoseEnv.csv files.
sepDS character (with default): column separator in the DoseLab.csv files.

sepR character (with default): column separator in the Rule.csv files.

verbose logical (with default): enable/disable verbose mode

... further arguments that can be passed to Luminescence::read BIN2R.

#### Details

With Path and FolderNames, this function goes to the subfolders containing the BIN files and associated information to compute the luminescence data.

### \*\* What are the required files in each subfolder? \*\*

Each subfolder can be named, for example, as the sample name followed by a number; it must contain:

- bin.BIN, the bin file renamed as bin.BIN (note: the name of all files matters);
- **Disc.csv**, a one columns csv file containing the list of disc number of the previously selected grains (typically this list will include the position of grains based on their sensitivity, recycling or other properties);
- **DoseEnv.csv**, a two columns file containing the observation of the natural (or environmental), dose rate, and its non-shared variance (i.e. after removing all shared errors), both in Gy. Note: the user shall provide the squared value of the error associated with the dose rate experienced by the sample grains in nature;
- **DoseSourve.csv**, a two columns file containing the observation of the laboratory dose rate, and its variance (squared error), both in Gy;
- rule.csv, a csv file containing information on
  - beginSignal= the first channel for summing the natural or regenerative OSL signal (typically 1 or 6);
  - endSignal= the last channel for summing the natural or regenerative OSL signal (typically 5 or 10);
  - beginBackground= the first channel for background estimation of the natural or regenerative OSL signal (typically 76 or 81);
  - endBackground= the last channel for background estimation of the natural or regenerative OSL signal (typically 95 or 100);
  - beginTest,
  - endTest,
  - beginTestBackground,
  - endTestBackground= same values as above, for the test dose response (typically the same values should be used);
  - inflatePercent= uncertainty arising from the instrument reproducibility (typically 0.02, i.e. 2%);
  - nbOfLastCycleToRemove= number of cycles at the end of the SAR protocol which should not be included in the dose response curve fitting (typically 1 if only a recycling test is performed, or 2 if both recycling and IR depletion are tested).

### \*\* How to fill the FolderNames vector? \*\*

FolderNames is a vector of length Nb\_binfile. FolderNames[i] is the name (e.g., Sample1-File1, or successive names separated by "/" signs, if BIN files are in subfolders, e.g. Sample1/File1) of the subfolder containing all informations on the BIN file of ID number i. The names in FolderNames are ordered following two rules:

- The names in the FolderNames vector must be ordered following the sample order (the names of subfolders containing BIN files for the same sample should follow each other in the FolderNames vector, e.g. Sample1, Sample2-File1, Sample2-File2, etc.).
- If stratigraphic constraints apply to samples, and so a **Bayesian model with stratigraphic constraints** is implemented, then the names in the FolderNames vector must be ordered by order of increasing ages.

For example, FolderNames=c(noun1, noun2), in which case noun1 (respectively, noun2) corresponds to the subfolder name containing the BIN file of sample 1 (respectively of sample 2). In addition, if we know that sample 1 is younger than sample 2, then FolderNames vector is correctly filled.

If conversely, FolderNames=c(noun2, noun1), the analysis performed by AgeS\_Computation would not be consistent.

### \*\* How to fill the BinPerSample vector? \*\*

BinPerSample[i] correponds to the number of BIN files for the sample whose number ID is equal to i.

For example, let us consider a case with two samples (Sample1 and Sample2), with 2 BIN files for Sample1 and 1 for Sample2. In this case, Nb\_binfile=3 and Nb\_sample=2. The user may then set FolderNames=c("Sample1-File1", "Sample1-File2", "Sample2-File1"), in which case "Sample1-1" is the name of the subfolder containing the first BIN file for Sample1, "Sample1-File2" the name of the subfolder for the second BIN file of Sample1; eventually, "Sample2-1" is the name of the subfolder containing the BIN file for the second sample. In this case, BinPerSample=c(2,1).

For the general BIN-file structure, the reader is referred to the following website: http://www.nutech.dtu.dk/ The function read\_BIN2R developed in Luminescence package is used to read the BIN files.

# Value

A list containing the following objects:

- LT (one list per sample); each list contains all L/T values for the corresponding sample;
- sLT (one list per sample); each list contains all uncertainties on L/T values for the corresponding sample;
- **ITimes** (one list per sample); each list contains irradiation time values for the corresponding sample;
- dLab, a matrix containing in line i, the laboratory dose rate and its variance for sample i;
- **ddot\_env**, a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;
- regDose (one list per sample); each list contains all regenerated doses;
- J, a vector giving, for each BIN file, the number of aliquots selected for the analysis;
- **K**, a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;
- **Nb\_measurement**, a vector giving, for each BIN file, the number of measurements;

# \*\* How to save this list \*\*

You can save this list in a .RData object. To do this, you can use the fonction save. Then, to load this list you can use the function load (see example section fore more details).

### Note

The function imports only BIN-file records which have been previously selected.

LT\_RegenDose 35

#### Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guérin

#### See Also

read\_BIN2R, combine\_DataFiles, LT\_RegenDose Age\_Computation, AgeS\_Computation, Palaeodose\_Computation

# **Examples**

```
path<- system.file("extdata/FER1", "", package="BayLum")
folder=""
# give the number of sample
nbsample=1
DATA=Generate_DataFile_MG(Path=path,FolderNames=folder,Nb_sample=nbsample)
str(DATA)
# to save information in RData object in folder containing bin file
#save(DATA,file=c(paste(path,folder,'DATA.RData',sep="")))
# to load information containing DATA.RData object
#load(file=c(paste(path,folder,"DATA.RData",sep="")))</pre>
```

LT\_RegenDose

Plots Lx/Tx as a function of the regenerative dose

### **Description**

This function plots Lx/Tx values as a function of regenerative dose, for every selected aliquot and for each sample.

### Usage

```
LT_RegenDose(DATA, Path, FolderNames, SampleNames = FolderNames, Nb_sample,
BinPerSample = rep(1, Nb_sample), SG = rep(TRUE, Nb_sample),
sepDP = c(","), nrow = 3L, ncol = nrow)
```

### **Arguments**

DATA	list ( <b>required</b> ): list of objects LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by Generate_DataFile or Generate_DataFile_MG or combine_DataFiles. DATA can contain information from more than one sample.
Path	<pre>character (required): path to the project folder (the same as the one used in Generate_DataFile or Generate_DataFile_MG to provide DATA)</pre>
FolderNames	character ( <b>required</b> ): vector of names of the sub-folders containing the BIN files, which were used by Generate_DataFile or Generate_DataFile_MG to generate the DATA object.
SampleNames	character (with default): Names of samples. To use if there is more than one bin

file per sample.

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Nb_sample	integer ( <b>required</b> ): ID number (in [1,Nb_sample]) of the sample selected for plotting L/T as a function of regenerative doses. Required if the DATA object contains information for more than one sample.
BinPerSample	<pre>integer (with default): integer vector (with default): vector with the number of BIN files per sample, which was used in Generate_DataFile or Generate_DataFile_MG to generate the DATA object.</pre>
SG	logical (with default): vector to set the type of measurement for each sample (length(SG)=Nb_sample). If the sample of number ID equal to i, SG[i]=TRUE if it is a Single-grain OSL measurements, SG[i]=FALSE if it is a Multi-grain OSL measurements.
sepDP	character (with default): column separator in the DiscPose.csv file or in Disc.csv file. It must be the same separator for all samples, for Single-grain OSL measurements or Multi-grain OSL measurements.
nrow	integer (with default): controls the arangment of the plots, here the number of rows. Can be set to NULL.
ncol	integer (with default): controls the arangment of the plots, here the number of columns. Can be set to NULL.

# **Details**

To fill FolderNames and BinPerSample, we refer to the **Detail** section from the Generate\_DataFile or Generate\_DataFile function. As well for a precise description of input DATA.

### Value

Lx/Tx plots; there are as many plots as selected aliquots in the DiscPos.csv file. There are 9 plots per page. There is not interpolation.

# Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guérin

# See Also

```
Generate_DataFile, Generate_DataFile_MG
```

# **Examples**

```
## load data file generated by the function Generate_DataFile
data(DATA3,envir = environment())
path<- system.file("extdata/FER1", "", package="BayLum")
folder=""
samplename="FER1"
LT_RegenDose(DATA=DATA3,Path=path,FolderNames=folder,SampleNames=samplename,Nb_sample=1,SG=FALSE)</pre>
```

Marine\_CalC14 37

Marine\_CalC14

Marine data for calibration of 14C age

### **Description**

As 14C years is not equal to calendar years because atmospheric 14C concentration varies through time. Hence, data in marine\_CalC14 allows a calibration for hypothetical "global" marine reservoir.

### Usage

```
data("Marine_CalC14")
```

#### **Format**

A data frame with 3 variables.

CAL.BP a numeric vector correpondig to calendar years befor present

X14C. age a numeric vector correponding to 14C age

Error a numeric vector correponding to error arround 14C age measurement

#### References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine 13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

# **Examples**

```
data(Marine_CalC14)
## maybe str(Marine_CalC14); head(Marine_CalC14) ...
```

MCMCsample

MCMC sample from the posterior distribution of the dataset GDB5

### **Description**

MCMC samples from the posterior distribution of "A" for age, "D" for palaeodose and "sD" for dispersion of equivalent doses around "D", of the data set GDB5.

### Usage

```
data("MCMCsample")
```

### **Format**

It is a matric with 6000 row and tree column.

- A The first column of the matrice are sampled from the posterior distribution of the paramete A
- D The first column of the matrice are sampled from the posterior distribution of the paramete D
- sD The first column of the matrice are sampled from the posterior distribution of the paramete sD

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#### References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L, Pleurdeau, D, Pearson, O, Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

# **Examples**

```
data(MCMCsample)
## maybe str(MCMCsample); plot(MCMCsample[,1],type="1") ...
```

ModelC14

Likelihood of C14 samples for JAGS models use in Age\_OSLC14

## **Description**

A list of models for C14 data to define likelyhood in JAGS models.

### Usage

```
data("ModelC14")
```

### **Format**

This list contains:

full a model considering error on calibration curve.

naive a model not considering error on calibration curve.

### References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine 13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

```
data(Model_AgeC14)
## ModelC14[[full]]
## ModelC14$full
```

ModelOSL 39

Mode10SI

Likelihood of OSL samples for JAGS models use in Age\_OSLC14

### **Description**

A list of models for OSL data to define likelyhood in JAGS models.

### Usage

```
data("ModelOSL")
```

#### **Format**

This list contains:

AgesMultiCS2\_EXPLIN a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgesMultiCS2\_EXP a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgesMultiCS2\_EXPZO a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgesMultiCS2\_EXPLINZO a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

### **Details**

The different distibutions to describe equivalent dose values around the palaeodose are:

cauchy a Cauchy distribution with postition parameter equal to the palaeodose of the sample gaussian a Gaussian distribution with mean equal to the palaeodose of the sample

 ${\tt lognormal\_A} \ \ a \ log-normal \ distribution \ with \ mean \ or \ {\bf A} verage \ equal \ to \ the \ palaeodose \ of \ the \ sample$ 

lognormal\_M a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function AgeS\_Computation, section Details.

# References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

```
data(ModelOSL) ## The JAGS model of the likelyhood for a saturating exponential plus linear growth ## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d}) ## and a gaussian distribution of equivalent doses around the palaeodose: ## ModelOSL[[AgesMultiOSL_EXPLIN]][[gaussian]]
```

40 Model\_Age

ModelPrior

Prior for JAGS models use in Age\_OSLC14

### **Description**

A list to define prior in JAGS models, taking acount OSL data and C14 data in stratigraphic constraint. The difficulty is in the fact that each cases is different. The youngest sample can be a C14 as well as a OSL sample. To resolve this problem we consider different cases thanks to this list.

#### **Usage**

```
data("ModelPrior")
```

#### **Format**

This list contains:

Sample1\_C14 model considering that the youngest sample is a C14 sample

Sample1\_OSL model considering that the youngest sample is a OSL sample

C14\_OSL model considering that the second sample is a C14 sample

OSL\_C14 model considering that the second sample is a OSL sample

C14 model considering that the last sample is a C14 sample

OSL model considering that the last sample is a OSL sample

#### References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

### **Examples**

```
data(ModelPrior)
## ModelPrior[[OSL]]
## ModelPrior$OSL
```

Model\_Age

JAGS models use in Age\_Computation

# **Description**

A list of JAGS models use to a Bayesian analysis of OSL age of one sample. There are models for various growth curves and various distribution to describe equivalent dose distribution around the palaeodose.

### Usage

```
data("Model_Age")
```

Model\_Age 41

#### **Format**

This list contains:

AgeMultiBF\_EXPLIN a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgeMultiBF\_EXP a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgeMultiBF\_EXPZO a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgeMultiBF\_EXPLINZO a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

#### **Details**

The different distibutions to describe equivalent dose values around the palaeodose are:

cauchy a Cauchy distribution with postition parameter equal to the palaeodose of the sample gaussian a Gaussian distribution with mean equal to the palaeodose of the sample

 $lognormal\_A$  a log-normal distribution with mean or Average equal to the palaeodose of the sample

lognormal\_M a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function Age\_Computation, section Details.

# References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

# See Also

rjags

```
data(Model_Age)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[AgeMultiBF_EXPLIN]][[gaussian]]
```

42 Model\_AgeC14

Model\_AgeC14

JAGS models use in AgeC14\_Computation

# **Description**

A list of JAGS models use to a Bayesian analysis of C14 calibration age of various sample. Stratigraphic relations can be taken in count to calibrate C14 ages. This ages take into account that some data can be an outlier.

# Usage

```
data("Model_AgeC14")
```

# **Format**

This list contains:

full a model considering error on calibration curve.

naive a model not considering error on calibration curve.

#### References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine 13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

#### See Also

rjags

```
data(Model_AgeC14)
## Model_AgeC14[[full]]
## Model_AgeC14$full
```

Model\_AgeS 43

Model\_AgeS

JAGS models use in AgeS\_Computation

### **Description**

A list of JAGS models use to a Bayesian analysis of OSL age of various samples. There are models for various growth curves and various distribution to describe equivalent dose distribution around the palaeodose.

### Usage

```
data("Model_AgeS")
```

#### **Format**

This list contains:

AgesMultiCS2\_EXPLIN a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgesMultiCS2\_EXP a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgesMultiCS2\_EXPZO a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

AgesMultiCS2\_EXPLINZO a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

# Details

The different distibutions to describe equivalent dose values around the palaeodose are:

cauchy a Cauchy distribution with postition parameter equal to the palaeodose of the sample gaussian a Gaussian distribution with mean equal to the palaeodose of the sample

lognormal\_A a log-normal distribution with mean or Average equal to the palaeodose of the sample

 $lognormal\_M$  a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function AgeS\_Computation, section Details.

#### References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

### See Also

rjags

44 Model\_Palaeodose

### **Examples**

```
data(Model_AgeS)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[AgesMultiCS2_EXPLIN]][[gaussian]]
```

Model\_Palaeodose

JAGS models use in Palaeodose\_Computation

#### **Description**

A list of JAGS models use to a Bayesian analysis of OSL palaeodose of one or various samples. There are models for various growth curves and various distribution to describe equivalent dose distribution around the palaeodose.

## Usage

```
data("Model_Palaeodose")
```

### **Format**

This list contains:

- PalaeodosesMultiBF\_EXPLIN a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.
- PalaeodosesMultiBF\_EXP a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.
- PalaeodosesMultiBF\_EXPZO a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.
- PalaeodosesMultiBF\_EXPLINZO a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

# Details

The different distibutions to describe equivalent dose values around the palaeodose are:

cauchy a Cauchy distribution with postition parameter equal to the palaeodose of the sample gaussian a Gaussian distribution with mean equal to the palaeodose of the sample

lognormal\_A a log-normal distribution with mean or Average equal to the palaeodose of the sample

lognormal\_M a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function Palaeodose\_Computation, section Details.

#### References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

### See Also

rjags

### **Examples**

```
data(Model_Palaeodose)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[PalaeodosesMultiBF_EXPLIN]][[gaussian]]
```

Palaeodose\_Computation

Bayesian analysis for the palaeodose estimation of various samples

# **Description**

This function computes the palaeodose (in Gy) of one or various samples according to the model developed in Combes et al (2015), based on an output of Generate\_DataFile or Generate\_DataFile\_MG or both of them using combine\_DataFiles.

Samples, for which data is avalilable in several BIN files, can be analysed.

Single-grain or Multi-grain OSL measurements can be analysed simultaneouly.

# Usage

```
Palaeodose_Computation(DATA, SampleNames, Nb_sample, BinPerSample = rep(1,
Nb_sample), SavePdf = FALSE, OutputFileName = c("MCMCplot"),
OutputFilePath = c(""), SaveEstimates = FALSE,
OutputTableName = c("DATA"), OutputTablePath = c(""), LIN_fit = TRUE,
Origin_fit = FALSE, distribution = c("cauchy"), Iter = 50000, t = 5,
Nb_chaines = 3)
```

### **Arguments**

DATA list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J, K, Nb\_measurement,

provided by  $Generate\_DataFile$  or  $Generate\_DataFile\_MG$ . DATA contains

information for more than one sample.

SampleNames character vector: names of sample. The length of this vector is equal to Nb\_sample.

Nb\_sample integer: number of samples.

BinPerSample integer vector (with default): vector with the number of BIN files per sample.

The length of this vector is equal to Nb\_sample. BinPerSample[i] correponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in Generate\_DataFile

or Generate\_DataFile\_MG.

SavePdf	boolean (with default): if TRUE save graph in pdf file named $OutputFileName$ in folder $OutputFilePath$ .	
OutputFileName	character (with default): name of the pdf files that will be generated by the function.	
OutputFilePath	character (with default): path to the pdf files that will be generated by the function.	
SaveEstimates	boolean (with default): if TRUE save Bayes estimates and credible interval at level 68 in a csv table named OutputFileName in folder OutputFilePath.	
OutputTableName		
	character (with default): name of the table that will be generated by the function if $SaveEstimates=TRUE$ .	
OutputTablePath		
	character (with default): path to the table that will be generated by the function if $SaveEstimates=TRUE$ .	
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. Please see details for more informations on the proposed dose response curves.	
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. Please see details for more informations on the proposed growth curves.	
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M".	
Iter	integer (with default): number of iterations for the MCMC computation (for more information see $jags.model$ ).	
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see jags.model).	
Nb_chaines	integer (with default): number of independent chains for the model (for more information see jags.model).	

### **Details**

# \*\* Option on growth curves \*\*

As for Age\_Computation and AgeS\_Computation, the user can choose from 4 dose response curves:

• Saturating exponential plus linear growth (PalaeodosesMultiBF\_EXPLIN):

for all x in IR+, f(x)=a(1-exp(-x/b))+cx+d; select

- LIN\_fit=TRUE
- Origin\_fit=FALSE
- **Saturating exponential growth** (PalaeodosesMultiBF\_EXP):

for all x in IR+, f(x)=a(1-exp(-x/b))+d; select

- LIN\_fit=FALSE
- Origin\_fit=FALSE
- Saturating exponential plus linear growth and fitting through the origin (PalaeodosesMultiBF\_EXPLINZO): for all x in IR+, f(x)=a(1-exp(-x/b))+cx; select
  - LIN\_fit=TRUE

- Origin\_fit=TRUE
- Saturating exponential growth and fitting through the origin (PalaeodosesMultiBF\_EXPZ0): for all x in IR+, f(x)=a(1-exp(-x/b)); select
  - LIN\_fit=FALSE
  - Origin\_fit=TRUE

# \*\* Option on equivalent dose distribution around the palaeodose \*\*

The use can choose between:

- cauchy: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- gaussian: a Gaussian distribution with mean equal to the palaeodose of the sample
- lognormal\_A: a log-normal distribution with mean or Average equal to the palaeodose of the sample
- lognormal\_M: a log-normal distribution with Median equal to the palaeodose of the sample

#### Value

### **NUMERICAL OUTPUT**

- 1. A list containing the following objects:
  - **Sampling** that corresponds to a sample of the posterior distributions of palaeodose and equivalent dose dispersion parameters (both in Gy).
  - Model\_GrowthCurve, stating which dose response fitting option was chosen;
  - **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample.
- 2. **The Gelman and Rubin test of convergency**: prints the result of the Gelman and Rubin test of convergency for palaeodose and equivalent dose dispersion parameters for each sample. A result close to one is expected.
  - In addition, the user must visually assess the convergency of the trajectories by looking at the pdf file generated by the function (see **PLOT OUTPUT** for more informations).
  - If both convergencies (Gelman and Rubin test and plot checking) are satisfactory, the user can consider the printed estimates as valid. Otherwise, the user may try increasing the number of MCMC interations (Iter) to reach convergency.
- 3. **Credible intervals and Bayes estimates**: prints the Bayes esitmates, the credible intervals at 95% and 68% for the palaeodose and equivalent dose dispersion parameters for each sample.

# PLOT OUTPUT

- MCMC trajectories A graph with the MCMC trajectories and posterior distributions of the palaeodose and equivalent dose dispersion parameters is displayed, there is one page per sample.
  - The first line of the figure correponds to the palaeodose parameter and the second to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.
- 2. **Summary of palaeodose estimates**: plot credible intervals and Bayes estimate of each sample palaeodose on a same graph.

To give result in a publication, we recommend to give the Bayes estimate of the parameters as well as the credible interval at 95% or 68%.

48 plot\_MCMC

#### Note

Please note that the initial values for all MCMC are currently all the same for all chains since we rely on the automatic initial value generation of JAGS. This is not optimal and will be changed in future. However, it does not affect the quality of the age estimates if the chains have converged.

#### Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guérin

#### References

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. Quaternary Geochronology 28, 62-70. doi:10.1016/j.quageo.2015.04.001

#### See Also

Generate\_DataFile, Generate\_DataFile\_MG, combine\_DataFiles, rjags, plot\_MCMC, Age\_Computation, AgeS\_Computation

### **Examples**

```
## Load data
data(DATA1,envir = environment())
## Palaeodose computation of samples GDB3
P=Palaeodose_Computation(DATA=DATA1,Nb_sample=1,SampleNames=c("GDB5"),Iter=100)
```

plot\_MCMC

Plot MCMC trajectories and posterior distributions

# Description

This function uses the output of rjags::jags.model to visualise the traces of the MCMC and the corresponding densities. In particular it displays the posterior distributions of the age, if it is calculated, palaeodose and the equivalent dose dispersion parameters of the sample. The function output is very similar to plot output produced with the 'coda' package, but tailored to meet the needs in the context of the 'BayLum' package.

### Usage

```
plot_MCMC(object, sample_names = NULL, variables = c("A", "D", "sD"),
   axes_labels = c(A = "Age (ka)", D = "D (Gy)", sD = "sD (Gy)"),
   n.chains = NULL, n.iter = 1000L, smooth = FALSE, rug = TRUE,
   plot_single = FALSE, ...)
```

plot\_MCMC 49

# **Arguments**

object	coda::mcmc.list or coda::mcmc ( <b>required</b> ): Output generated by rjags::jags.model, e.g., in Age_Computation
sample_names	character (optional): Names of the used samples. This argument overrides the optional argument mtext.
variables	character (with default): Variables in your coda::mcmc object to be plotted.
axes_labels	character (with default): Axes labels used for the trace and density plots. The labels should be provided as named character vector with the parameter names as the names used to asign the axes labelling. The labelling for the x-axis (trace plots) and y-axis (density plot) cannot be modified.
n.chains	numeric (optional): Set the number of chains to visualise, if nothing is provided the number of chains is determined from the input object
n.iter	integer (with default): Set the number of iterations to be visualised in the trace plots, regardless of the size of the input dataset as long as the real number of iterations is > n.iter. Please note that large numbers impact the plot performance.
smooth	logical (with default): Enable/disables smooth of trace plots using stats::smooth
rug	logical (with default): Enable/disables rug under density plots
plot_single	logical (with default): Enables/disables the single plot mode of the function, i.e. if set to TRUE every plot is returned in a single plot and own par settings can be applied.
•••	further arguments that can be passed to modify the plot output. Supported arguments are lwd, lty, col, type, cex,mtext, cf. mtext for mtext and plot.default for the other arguments.

# **Details**

The function is used in the function Age\_Computation, AgeS\_Computation and Palaeodose\_Computation, but can be used also as standalone plot function.

# Value

Two plots: Traces of the MCMC chains and the corresponding density plots. This plots are similar to coda::traceplot and coda::densplot.

# **Function version**

0.1.1

# Author(s)

Sebastian Kreutzer, IRAMAT-CRP2A, Université Bordeaux Montaigne (France). This function is a re-written version of the function MCMC\_plot() by Claire Christophe

# See Also

Age\_Computation, AgeS\_Computation, Palaeodose\_Computation, rjags::coda.samples and rjags packages.

50 ScatterSamples

#### **Examples**

```
data(MCMCsample,envir = environment())
object <- coda::as.mcmc(MCMCsample)
plot_MCMC(object)</pre>
```

ScatterSamples

Scatterplots of samples

### **Description**

This fonction compute a matrix of scatterplots of at most 8 samples.

# Usage

```
ScatterSamples(Sampling, SampleNames, SelectedSamples, sc = 3,
  Nb_chaines = 3)
```

# **Arguments**

Sampling MCMC.list: that is generated by jags.model in AgeS\_Computation, AgeC14\_Computation

or Age\_OSLC14.

SampleNames character vector: names of samples.

SelectedSamples

numeric vector: indicate if the sample is selected to be plot with the other sam-

 $ples. \ If the sample with names \verb|SampleNames[i]| is selected, then \verb|SelectedSamples[i]=1|;$ 

if not SelectedSamples=0. length(SelectedSamples)=length(SampleNames).

At least two samples must be selected.

sc numeric (with default): to define the number of values on the axis.

Nb\_chaines integer (with default): number of independent chains used in AgeS\_Computation

to generate the sampling (for more information see jags.model).

### Value

A matrix that contains at row i and column j the scatterplot of the posterior distribution of the sample ID i plotted against the one of the sample ID j

### Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

# See Also

```
Age_Computation, AgeS_Computation, AgeC14_Computation, and rjags packages.
```

```
data(AgeS,envir = environment())
ScatterSamples(Sampling=AgeS$Sampling,SampleNames=c("GDB5","GDB3"),SelectedSamples=c(1,1))
```

SCMatrix 51

SCMatrix

Definition of the stratigraphic constraint matrix

### **Description**

This function helps to define the stratigraphic relation between samples, with questions. The output of this function can be used in function AgeS\_Computation.

# Usage

```
SCMatrix(Nb_sample, SampleNames)
```

### **Arguments**

Nb\_sample interger: the sample number.

SampleNames charcater vector: sample names.

### **Details**

Ask if sample i is younger than sample j to construc the stratigraphic constrain matrix.

### Value

A Matrix that summarise the ordered relation between samples. This matrix can be intergrate in AgeS\_Computation function. We refer to detail on AgeS\_Computation for more information concerning this matrix.

# Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

### See Also

```
AgeS_Computation
```

```
## Assume that "sample1" is younger than "sample2"
## That means the expected value is 1.
## It is an interactive function.
## Not run:
SCMatrix(Nb_sample=2,SampleNames=c("sample1","sample2"))
## Enter the value 1
## End(Not run)
```

52 SC\_Ordered

SC\_Ordered

Create stratigraphically ordered sample matrix

# **Description**

Construct the stratigraphic matrix used in the functions AgeS\_Computation and AgeC14\_Computation for samples that are all ordered by increasing age.

# Usage

```
SC_Ordered(Nb_sample)
```

# **Arguments**

Nb\_sample integer (required): the number of samples

#### Value

Stratigraphic matrix where each sample are ordered by increasing order. This matrix can be intergrated in the function AgeS\_Computation. Please see AgeS\_Computation for more information on this matrix.

### Author(s)

Claire Christophe, Anne Philippe, Sebastian Kreutzer, Guillaume Guérin

### See Also

AgeS\_Computation, SCMatrix

```
# compute the stratigraphic matrix for 3 samples such that the first sample is younger
# than the second, and the second is younger than the third
SC <- SC_Ordered(Nb_sample = 3)</pre>
```

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