

Package ‘BayLum’

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Type Package

Title Chronological Bayesian Models Integrating Optically Stimulated Luminescence and Radiocarbon Age Dating

Description Bayesian analysis of luminescence data and 14C age estimate. Bayesian models are based on the following publications: Combes, B. & Philippe, A. (2017) <doi:10.1016/j.quageo.2017.02.003> and Combes et al (2015) <doi:10.1016/j.jas.2015.02.003>. It includes, amongst others, data import, export, application of age models and palaeodose model.

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BayLum-package	<i>Chronological Bayesian Models Integrating Optically Stimulated Luminescence and 14C Age Dating</i>
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Description

A collection of various R functions for Bayesian analysis of luminescence data and 14C Cal age estimate. This includes, amongst others, data import, export, application of age models and palaeodose model.

Details

This package is based on the functions: [Generate_DataFile](#) and [Generate_DataFile_MG](#) to import luminisecence data. These functions create a list containing all informations to compute age of Single-grain OSL measurements for the first function and Multi-grain OSL measurements for the second.

The functions: [Age_Computation](#) and [AgeS_Computation](#) use Bayesian analysis for OSL age estimation for one or various samples according to differents models (e.g. different growth curves and different equivalent dose distributions around the palaeodose).

It is possible to consider various Bin files per sample, to compute ages of samples in stratigraphic constraints and to integrate systematic errors.

It is possible to calibrate 14C age with the function [AgeC14_Computation](#). We can also estimate chronology containing 14C age and OSL samples with the function [Age_OSLC14](#).

Author(s)

Claire Christophe, Anne Philippe, Sebastian Kreutzer, Guillaume Guerin

AgeC14_Computation	<i>Bayesian analysis for age estimation of 14C age of various samples</i>
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Description

This function calibrate 14C age of samples to get an age (years). The user can choose one of the following radiocarbon calibration curve: Northern or Sourthen Hemisphere or marine atmospheric. It must be the same curve for all samples.

Usage

```
AgeC14_Computation(Data_C14Cal, Data_SigmaC14Cal, SampleNames, Nb_sample,
  PriorAge = rep(c(10000, 50000), Nb_sample), SavePdf = FALSE,
  OutputFileName = c("MCMCplot", "HPD_Cal14CCurve", "summary"),
  OutputFilePath = c(""), SaveEstimates = FALSE,
  OutputTableName = c("DATA"), OutputTablePath = c(""),
  StratiConstraints = c(), sepSC = c(", "), Model = c("full"),
  CalibrationCurve = c("AtmosphericNorth"), Iter = 50000, t = 5,
  Nb_chaines = 3)
```

Arguments

Data_C14Cal	numeric vector: corresponding to 14C age estimate.
Data_SigmaC14Cal	numeric vector: corresponding to the error of 14C age estimates.
SampleNames	character vector: names of sample. The length of this vector is equal to Nb_sample.
Nb_sample	integer: number of samples.
PriorAge	numeric vector (with default): lower and upper bounds for age parameter of each sample in years (not in ka). Note that, <code>length(PriorAge)=2*Nb_sample</code> and <code>PriorAge[2i-1,2i]</code> correponds to the lower and upper bounds of sample whose number ID is equal to i.
SavePdf	boolean (with default): if TRUE save graphs in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE, <code>length(OutputFileName)=3</code> , see PLOT OUTPUT in Value section for more informations.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".
SaveEstimates	boolean (with default): if TRUE save Bayes estimates, credible interval at level 68% and 95% and the result of the gelman en Rudin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".

StratiConstraints

numeric matrix or character (with default): input object for the stratigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints; the user can refer to a matrix (numeric matrix) or to a csv file (character). If there is no stratigraphic relation default value is suitable.

sepSC

character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints csv file.

Model

character (with default): if "full", error on estimate calibration curve is taken account. If "naive" this error is not taken account in the age estimate.

CalibrationCurve

character (with default): calibration curve choosen. Allowed inputs are

- "AtmosphericNorth" for Northern Hemisphere atmospheric radiocarbon calibration curve,
- "Marine" for Marine radiocarbon calibration curve,
- "AtmosphericSouth" for Southern Hemisphere atmospheric radiocarbon calibration curve
- a csv file, with tree columns, the first column is dedicated to "Cal.BP", the second to "X14C.age", the third to "Error". The decimal of this file must be a dot, and the separator must be a comma.

Iter

integer (with default): number of iterations for the MCMC computation (for more information see [jags.model](#)).

t

integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see [jags.model](#)).

Nb_chaines

integer (with default): number of independent chains for the model (for more information see [jags.model](#)).

Details

**** How to fill StratiConstraints ? ****

If there is stratigraphic relations between samples, **14C age in Data_C14Cal must be ordered by order of increasing ages.**

The user can fill the StratiConstraints matrix as follow.

1. **Size of the matrix:** row number of StratiConstraints matrix is equal to Nb_sample+1, and column number is equal to Nb_sample.
2. **First line of the matrix:** for all i in $\{1, \dots, Nb_Sample\}$, $StratiConstraints[1, i] = 1$ that means the lower bound of the sample age (given in $PriorAge[2i-1]$) for the sample whose number ID is equal to i , is taken into account.
3. **Sample relations:** for all j in $\{2, \dots, Nb_Sample+1\}$ and all i in $\{j, \dots, Nb_Sample\}$, $StratiConstraints[j, i] = 1$ if sample age whose number ID is equal to $j-1$ is lower than sample age whose number ID is equal to i . Otherwise, $StratiConstraints[j, i] = 0$.

Note that $StratiConstraints_{\{2:Nb_sample+1, 1:Nb_sample\}}$ is a upper triangular matrix.

The user can also use [SCMatrix](#) or [SC_Ordered](#) (if all samples are ordered) functions to construct the StratiConstraints matrix.

The user can also refer to a .csv file that contains the relation between samples as defined above. The user must take care about the separator used in the csv file using the argument sepSC.

**** More precision on Model ****

We propose two models "full" or "naive". If Model='full' that means measurement error and error on calibration curve are taken account in the Bayesian model; if Model="naive" that means only error on measurement are taken account in the mode.

More precisely, the model considered here, as the one developed by Christen, JA (1994), assume multiplicative effect of errors to address the problem of outliers. In addition, to not penalise variables that are not outliers and damage their estimation, we introduce a structure of mixture, that means only variable that are considered as outlier have in addition a multiplicative error.

Value

NUMERICAL OUTPUT

1. A list containing the following objects:

- **Sampling:** that corresponds to a sample of the posterior distributions of the age parameters;
- **Outlier:** stating the names of samples that are considered as outliers;
- **Model:** stating which model was chosen ("full" or "naive");
- **CalibrationCurve:** stating which radiocarbon calibration curve was chosen;
- **PriorAge:** stating the priors used for the age parameter;
- **StratiConstraints:** stating the stratigraphic relations between samples considered in the model.

2. The Gelman and Rudin test of convergency: print the result of the Gelman and Rudin test of convergency for the age estimate for each sample. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (Iter) or being more precise if it is possible on the PriorAge parameter to reach convergency.

3. Credible intervals and Bayes estimates: prints the Bayes estimates, the credible intervals at 95% and 68% for the age parameters for each sample.

PLOT OUTPUT

1. MCMC trajectories: A graph with the MCMC trajectories and posterior distributions of the age parameter is displayed.

On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

2. Summary of sample age estimates: plot credible intervals and Bayes estimate of each sample age on one graph.

To give the results in a publication, we recommend to give the Bayes estimate of the parameters as well as the credible interval at 95% or 68%.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

References

Christen, JA (1994). Summarizing a set of radiocarbon determinations: a robust approach. *Applied Statistics*, 489-503.

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haffidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. *Radiocarbon* 55(4):1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. *Radiocarbon* 55(4):1889-1903

See Also

[rjags](#), [MCMC_plot](#), [SCMatrix](#)

Examples

```
## Load data
data(DATA_C14,envir = environment())
C14Cal=DATA_C14$C14[,1]
SigmaC14Cal=DATA_C14$C14[,2]
Names=DATA_C14$Names
nb_sample=length(Names)
## Age computation of samples without stratigraphic relations

Age=AgeC14_Computation(Data_C14Cal=C14Cal,Data_SigmaC14Cal=SigmaC14Cal,
  SampleNames=Names,Nb_sample=nb_sample,PriorAge=rep(c(20000,60000),nb_sample),Iter=500)
```

AgeS

Output of [AgeS_Computation](#) function for the samples: "GDB5" and "GDB3"

Description

Output of [AgeS_Computation](#) function for the samples: "GDB5" and "GDB3", there is no stratigraphic relation neither systematic errors.

Usage

```
data("AgeS")
```

Format

A list containing

`Sampling` MCMC.list that corresponds to a sample of the posterior distributions of the ages (in ka), palaeodoses (in Gy) and equivalent dose dispersions (in Gy) parameters of samples "GDB5" and "GDB3";

`Model_GrowthCurve` stating which dose response fitting option was chosen to run the function

Distribution stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;

PriorAge stating the priors used for the age parameter (in ka);

StratiConstraints stating the matrix of stratigraphic relations between samples considered in the model;

CovarianceMatrix stating the covariance matrix of error used in the model, highlighting not common errors between samples in our cases (diagonal matrix).

References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D, Pearson, O , Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

Examples

```
data(AgeS)
str(AgeS)
```

AgeS_Computation

Bayesian analysis for the OSL age estimation of various samples

Description

This function computes the age (in ka) of at least two samples according to the model developed in Combes and Philippe (2017), based on outputs of [Generate_DataFile](#) or [Generate_DataFile_MG](#) or both of them using [Concat_DataFile](#).

Samples, for which data is available in several BIN files, can be analysed.

Single-grain or Multi-grain OSL measurements can be analysed simultaneously.

Usage

```
AgeS_Computation(DATA, SampleNames, Nb_sample, PriorAge = rep(c(0.01, 100),
  Nb_sample), BinPerSample = rep(1, Nb_sample), SavePdf = FALSE,
  OutputFileName = c("MCMCplot", "summary"), OutputFilePath = c(""),
  SaveEstimates = FALSE, OutputTableName = c("DATA"),
  OutputTablePath = c(""), THETA = c(), sepTHETA = c(", "),
  StratiConstraints = c(), sepSC = c(", "), LIN_fit = TRUE,
  Origin_fit = FALSE, distribution = c("cauchy"), Iter = 50000, t = 5,
  Nb_chaines = 3)
```

Arguments

DATA list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by the function [Generate_DataFile](#) or [Generate_DataFile_MG](#) or [Concat_DataFile](#). DATA contains informations for more than one sample. If there is stratigraphic relations between samples, informations in DATA must be ordered by order of increasing ages. See the details section for more informations.

SampleNames	character vector: names of samples. The length of this vector is equal to Nb_sample.
Nb_sample	integer : number of samples, Nb_sample>1.
PriorAge	numeric vector (with default): lower and upper bounds for age parameter of each sample (in ka). Note that, length(PriorAge)=2*Nb_sample and PriorAge[2i-1, 2i] corresponds to the lower and upper bounds of sample whose number ID is equal to i.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector is equal to Nb_sample. BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in Generate_DataFile or in Generate_DataFile_MG .
SavePdf	boolean (with default): if TRUE save graphs in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE; length(OutputFileName)=2, see PLOT OUTPUT in Value section for more informations.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".
SaveEstimates	boolean (with default): if TRUE save Bayes estimates, credible interval at level 68% and 95% and the result of the gelman en Rudin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".
THETA	numeric matrix or character (with default): input object for systematic and individual error. If systematic errors are considered, see the details section for instructions regarding how to correctly fill THETA; the user can refer to a matrix (numeric matrix) or to a csv file (character). Otherwise, default value is suitable, and only individual errors are considered.
sepTHETA	character (with default): if THETA is character, indicate column separator in THETA csv file.
StratiConstraints	numeric matrix or character (with default): input object for the stratigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints; the user can refer to a matrix (numeric matrix) or to a csv file (character). If there is no stratigraphic relation default value is suitable.
sepSC	character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints.csv file.
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. See details section for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. See details section for more informations on the proposed growth curves.

distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M", see details section for more informations.
Iter	integer (with default): number of iterations for the MCMC computation (for more information see jags.model).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see jags.model).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see jags.model).

Details

** How to fill StratiConstraints ? **

If there is stratigraphic relations between samples, **informations in DATA must be ordered by order of increasing ages**. To do this the user can either fill right Names in [Generate_DataFile](#) or in [Generate_DataFile_MG](#) (as it is indicated in Details section of these function), or ordered by order of increasing ages outputs of [Generate_DataFile](#) or [Generate_DataFile_MG](#) in [Concat_DataFile](#).

The user can fill the StratiConstraints matrix as follow.

1. **Size of the matrix:** row number of StratiConstraints matrix is equal to Nb_sample+1, and column number is equal to Nb_sample.
2. **First line of the matrix:** for all i in $\{1, \dots, Nb_Sample\}$, $StratiConstraints[1, i] = 1$ that means the lower bound of the sample age (given in $PriorAge[2i-1]$) for the sample whose number ID is equal to i , is taken into account.
3. **Sample relations:** for all j in $\{2, \dots, Nb_Sample+1\}$ and all i in $\{j, \dots, Nb_Sample\}$, $StratiConstraints[j, i] = 1$ if sample age whose number ID is equal to $j-1$ is lower than sample age whose number ID is equal to i . Otherwise, $StratiConstraints[j, i] = 0$.

Note that $StratiConstraints_{\{2:Nb_sample+1, 1:Nb_sample\}}$ is a upper triangular matrix.

The user can also use [SCMatrix](#) or [SC_Ordered](#) (if all samples are ordered) functions to construct the StratiConstraints matrix.

The user can also refer to a csv file that contains the relation between samples as defined above. The user must take care about the separator used in the csv file using the argument sepSC.

** How to fill THETA covariance matrix concerning common and individual error? **

If systematic errors are considered, the user can fill the THETA matrix as follow.

- row number of THETA is equal the column number, equal to Nb_sample.
- For all i in $\{1, \dots, Nb_sample\}$, $THETA[i, i]$ contains individual error plus systematic error of the sample whose number ID is equal to i .
- For all i, j in $\{1, \dots, Nb_sample\}$ and i different from j , $THETA[i, j]$ contains common error between samples whose number ID are equal to i and j .

Note that $THETA[i, j]$ is a symetric matrix.

The user can also refer to a .csv file that contains the errors as defined above.

** Option on growth curves **

As for [Age_Computation](#) and [Palaeodose_Computation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** (AgesMultiCS2_EXPLIN):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx+d$; select
 - LIN_fit=TRUE
 - Origin_fit=FALSE
- **Saturating exponential growth** (AgesMultiCS2_EXP):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+d$; select
 - LIN_fit=FALSE
 - Origin_fit=FALSE
- **Saturating exponential plus linear growth and fitting through the origin** (AgesMultiCS2_EXPLINZ0):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx$; select
 - LIN_fit=TRUE
 - Origin_fit=TRUE
- **Saturating exponential growth and fitting through the origin** (AgesMultiCS2_EXPZ0):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))$; select
 - LIN_fit=FALSE
 - Origin_fit=TRUE

** Option on equivalent dose distribution around the palaeodose **

The use can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample;
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample;
- **lognormal_A**: a log-normal distribution with mean or Average equal to the palaeodose of the sample;
- **lognormal_M**: a log-normal distribution with Median equal to the palaeodose of the sample.

Value

NUMERICAL OUTPUT

1. A list containing the following objects:

- **Sampling**: that corresponds to a sample of the posterior distributions of the age (in ka), palaeodose (in Gy) and equivalent dose dispersion (in Gy) parameters for each sample;
- **Model_GrowthCurve**: stating which dose response fitting option was chosen;
- **Distribution**: stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
- **PriorAge**: stating the priors used for the age parameter (in ka);
- **StratiConstraints**: stating the stratigraphic relations between samples considered in the model;
- **CovarianceMatrix**: stating the covariance matrix of error used in the model, highlighting common errors between samples or not.

2. **The Gelman and Rudin test of convergency:** prints the result of the Gelman and Rudin test of convergency for the age, palaeodose and equivalent dose dispersion parameters for each sample. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (*Iter*) or being more precise on the *PriorAge* parameter (for example specify if it is a young sample $c(0.01, 10)$ an old sample $c(10, 100)$), or changing the parameter distribution or the growth curve, to reach convergency.

3. **Credible intervals and Bayes estimates:** prints the Bayes estimates, the credible intervals at 95% and 68% for the age, palaeodose and equivalent dose dispersion parameters for each sample.

PLOT OUTPUT

1. **MCMC trajectories:** A graph with the MCMC trajectories and posterior distributions of the age, palaeodose and equivalent dose dispersion parameters is displayed, there is one page per sample.

The first line of the figure corresponds to the age parameter, the second to the palaeodose parameter and the third to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

2. **Summary of sample age estimates:** plot credible intervals and Bayes estimate of each sample age on a same graph.

To give the results in a publication, we recommend to give the Bayes estimate of the parameters as well as the credible interval at 95% or 68%.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

References

Combes, Benoit and Philippe, Anne, 2017. Bayesian analysis of multiplicative Gaussian error for multiple ages estimation in optically stimulated luminescence dating. *Quaternary Geochronology* (39, 24-34)

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. *Quaternary Geochronology* 28, 62-70. doi:10.1016/j.quageo.2015.04.001

See Also

[Generate_DataFile](#), [Generate_DataFile_MG](#), [rjags](#), [MCMC_plot](#), [SCMatrix Age_Computation](#), [Palaeodose_Computation](#)

Examples

```
## Load data
data(DATA1,envir = environment())
data(DATA2,envir = environment())
Data=Concat_DataFile(DATA2,DATA1)
```

```
## Age computation of samples GDB5 and GDB3,
priorage=c(1,10,20,60) # these samples are not young
## without common error and without stratigraphic constraints
Age=AgeS_Computation(DATA=Data,Nb_sample=2,SampleNames=c("GDB5","GDB3"),
  PriorAge=priorage,Iter=50,Nb_chaines=2)

## Age computation of samples GDB5 and GDB3,
## without common error, assuming GDB5 age younger than GDB3 age
## Not run:
(SC=matrix(data=c(1,1,0,1,0,0),ncol=Nb_sample,nrow = (Nb_sample+1),byrow = T))
Age=AgeS_Computation(DATA=Data,Nb_sample=2,SampleNames=c("GDB5","GDB3"),
  PriorAge=priorage,StratiConstraints=SC,Iter=10000)

## End(Not run)
```

Age_Computation

Bayesian analysis for the OSL age estimation of one sample

Description

This function computes the age (in ka) of a sample according to the model developed in Combes and Philippe (2017), based on an output of [Generate_DataFile](#) or [Generate_DataFile_MG](#). A sample, for which data is available in several BIN files, can be analysed.

Usage

```
Age_Computation(DATA, SampleName, PriorAge = c(0.01, 100),
  BinPerSample = c(1), SavePdf = FALSE, OutputFileName = c("MCMCplot"),
  OutputFilePath = c(""), SaveEstimates = FALSE,
  OutputTableName = c("DATA"), OutputTablePath = c(""), LIN_fit = TRUE,
  Origin_fit = FALSE, distribution = c("cauchy"), I = 1, Iter = 50000,
  t = 5, Nb_chaines = 3, quiet = FALSE)
```

Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by the function Generate_DataFile or Generate_DataFile_MG . DATA can contain informations for more than one sample.
SampleName	character: name of the sample.
PriorAge	numeric vector (with default): lower and upper bounds for the sample age parameter (in ka). Note that, length(PriorAge)=2.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. If in DATA there is more than one sample, the BinPerSample vector must be the same as that used to run the function Generate_DataFile or in Generate_DataFile_MG for generating the DATA object.
SavePdf	boolean (with default): if TRUE save graph in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE; length(OutputFileName)=2, see PLOT OUTPUT in Value section for more informations.

OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".
SaveEstimates	boolean (with default): if TRUE save Bayes estimates and credible interval at level 68% and 95% and the result of the gelman en Rudin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. See details section for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. See details section for more informations on the proposed growth curves.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M", see details section for more informations.
I	integer (with default): if DATA contains data from more than one sample, I indicates the ID number of the sample to be analysed.
Iter	integer (with default): number of iterations for the MCMC computation (for more information see jags.model).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see jags.model).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see jags.model).
quiet	logical (with default): enables/disables rjags messages

Details

** Option on growth curves **

As for [AgeS_Computation](#) and [Palaeodose_Computation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** (AgeMultiBF_EXPLIN):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx+d$; select
 - LIN_fit=TRUE
 - Origin_fit=FALSE
- **Saturating exponential growth** (AgeMultiBF_EXP):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+d$; select
 - LIN_fit=FALSE
 - Origin_fit=FALSE
- **Saturating exponential plus linear growth and fitting through the origin** (AgeMultiBF_EXPLINZO):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx$; select

- LIN_fit=TRUE
- Origin_fit=TRUE
- **Saturating exponential growth and fitting through the origin** (AgeMultiBF_EXPZO):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))$; select
 - LIN_fit=FALSE
 - Origin_fit=TRUE

**** Option on equivalent dose distribution around the palaeodose ****

The user can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample
- **lognormal_A**: a log-normal distribution with mean or **Average** equal to the palaeodose of the sample
- **lognormal_M**: a log-normal distribution with **Median** equal to the palaeodose of the sample

Value

NUMERICAL OUTPUT

1. **A list containing the following objects:**

- **Sampling** that corresponds to a sample of the posterior distributions of the age (in ka), palaeodose (in Gy) and equivalent dose dispersion (in Gy) parameters.
- **Model_GrowthCurve**, stating which dose response fitting option was chosen;
- **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
- **PriorAge**, stating the priors used for the age parameter (in ka).

2. **The Gelman and Rudin test of convergency**: prints the result of the Gelman and Rudin test of convergency for the age, palaeodose and equivalent dose dispersion parameters. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the printed estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (*Iter*), or being more precise on the *PriorAge* parameter (for example specify if it is a young sample $c(0.01, 10)$ an old sample $c(10, 100)$), or changing the parameter distribution or the growth curve, to reach convergency.

3. **Credible intervals and Bayes estimates**: prints the Bayes estimates, the credible intervals at 95% and 68% for the age, palaeodose and equivalent dose dispersion parameters of the sample.

PLOT OUTPUT

A graph with the MCMC trajectories and posterior distributions of the age, palaeodose and equivalent dose dispersion parameters is displayed.

The first line of the figure corresponds to the age parameter, the second to the palaeodose parameter and the third to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

To give the results in a publication, we recommend to give the Bayes estimate of the parameter as well as the credible interval at 95% or 68%.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

References

Combes, Benoit and Philippe, Anne, 2017. Bayesian analysis of multiplicative Gaussian error for multiple ages estimation in optically stimulated luminescence dating. *Quaternary Geochronology* (39, 24-34)

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. *Quaternary Geochronology* 28, 62-70. doi:10.1016/j.quageo.2015.04.001

See Also

[Generate_DataFile](#), [Generate_DataFile_MG](#), [rjags](#), [MCMC_plot](#) [AgeS_Computation](#), [Palaeodose_Computation](#)

Examples

```
## load data file generated by the function Generate_DataFile
data(DATA1,envir = environment())
priorage <- c(10,60) # GDB3 is an old sample
Age <- Age_Computation(
  DATA = DATA1,
  SampleName = "GDB3",
  PriorAge = priorage,
  Iter = 100,
  quiet = TRUE)
```

Age_OSLC14

Bayesian analysis for age estimation of OSL measerments and 14C ages of various samples

Description

This function compute an age of OSL data of at least two samples and calibrate 14C ages of samples to get an age (in ka).

Age of OSL data are computed according to the model given in Combes and Philippe (2017). Single-grain or Multi-grain OSL measurements can be analysed simultaneously (with output of [Generate_DataFile](#) or [Generate_DataFile_MG](#) or both of them using [Concat_DataFile](#)). Samples, for which data is available in several BIN files, can be analysed.

For C14 data, the user can choose one of the following radiocarbon calibration curve: Northern or Southern Hemisphere or marine atmospheric.

Usage

```
Age_OSLC14(DATA, Data_C14Cal, Data_SigmaC14Cal, Nb_sample, SampleNames,
  SampleNature, PriorAge = rep(c(10, 60), Nb_sample), SavePdf = FALSE,
  OutputFileName = c("MCMCplot", "HPD_Cal14CCurve", "summary"),
  OutputFilePath = c(""), SaveEstimates = FALSE,
  OutputTableName = c("DATA"), OutputTablePath = c("")),
```

```
StratiConstraints = c(), sepSC = c(", "), BinPerSample = rep(1,
sum(SampleNature[1, ])), THETA = c(), sepTHETA = c(", "), LIN_fit = TRUE,
Origin_fit = FALSE, distribution = c("cauchy"), Model_C14 = c("full"),
CalibrationCurve = c("AtmosphericNorth"), Iter = 50000, t = 5,
Nb_chaines = 3)
```

Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by the function Generate_DataFile or Generate_DataFile_MG or Concat_DataFile . DATA contains information for more than one sample. If there is stratigraphic relations between samples, informations in DATA must be ordered by order of increasing ages. See the details section to for more informations.
Data_C14Cal	numeric vector: corresponding to 14C age estimate (in years, conversion in ka is automatically done in the function). If there is stratigraphic relations between samples, Data_C14Cal must be ordered by order of increasing ages.
Data_SigmaC14Cal	numeric vector: corresponding to the error of 14C age estimates.
Nb_sample	integer: number of samples (OSL data and 14C age), (Nb_sample>3, at least to sample of OSL data and one sample of 14C age).
SampleNames	character vector: sample names for both OSL data and C14 data. The length of this vector is equal to Nb_sample. If there is stratigraphic relation, this vector must be ordered by increasing order (to mix OSL samples and 14C ages if it is needed).
SampleNature	numeric matrix: states the nature of the sample. Row number of SampleNature matrix is equal to 2 and column number is equal to Nb_sample. First line of the matrix: SampleNature[1,i] states if sample whose number ID is equal to i, is an OSL sample 1 or not 0 . Second line of the matrix: SampleNature[2,i] states if sample whose number ID is equal to i, is an 14C sample 1 or not 0 .
PriorAge	numeric vector (with default): lower and upper bounds for age parameter of each sample (in ka). Note that, length(PriorAge)=2*Nb_sample and PriorAge[2i-1,2i] corresponds to the lower and upper bounds of sample whose number ID is equal to i.
SavePdf	boolean (with default): if TRUE save graphs in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE, length(OutputFileName)=3, see PLOT OUTPUT in Value section for more informations.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE. If it is not equal to "", it must be terminated by "/".
SaveEstimates	boolean (with default): if TRUE save Bayes estimates, credible interval at level 68% and 95% and the result of the gelman en Rudin test of convergency, in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE. If it is not equal to "", it must be terminated by "/".

StratiConstraints

	numeric matrix or character (with default): input object for the stratigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints, the user can refer to a matrix (numeric matrix) or to a csv file (character). Otherwise, default value is suitable.
sepSC	character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints csv file.
BinPerSample	integer vector (with default): vector with the number of BIN files per OSL sample. The length of this vector is equal to the number of OSL samples. BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in Generate_DataFile or in Generate_DataFile_MG .
THETA	numeric matrix or character (with default): input object for systematic and individual error for OSL samples. If systematic errors are considered, see the details section for instructions regarding how to correctly fill THETA; the user can refer to a matrix (numeric matrix) or to a csv file (character). Otherwise, default value is suitable, and only individual error is considered.
sepTHETA	character (with default): if THETA is character, indicate column separator in THETA csv file.
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves, for OSL samples. See details for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. See details for more informations on the proposed growth curves, for OSL samples.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose, for OSL samples. Allowed inputs are "cauchy" , "gaussian" , "lognormal_A" and "lognormal_M" , see details for more informations.
Model_C14	character (with default): if "full" , error on estimate calibration curve is taken account, for 14C samples. If "naive" this error is not taken account in the age estimate.
CalibrationCurve	character (with default): calibration curve chosen, for 14C samples. Allowed inputs are <ul style="list-style-type: none"> • "AtmosphericNorth" for Northern Hemisphere atmospheric radiocarbon calibration curve, • "Marine" for Marine radiocarbon calibration curve, • "AtmosphericSouth" for Southern Hemisphere atmospheric radiocarbon calibration curve, • a csv file, with tree columns, the first column is dedicated to "Cal.BP", the second to "X14C.age", the third to "Error". The decimal of this file must be a dot, and the separator must be a comma.
Iter	integer (with default): number of iterations for the MCMC computation (for more information see jags.model).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see jags.model).

Nb_chaines integer (with default): number of independent chains for the model (for more information see [jags.model](#)).

Details

Note that there is tree type of arguments in the previous list. There are arguments for informations concerning only OSL samples: DATA, BinPerSample, THETA, sepTHETA, LIN_fit, Origin_fit, distribution.

There are arguments for informations concerning only C14 samples: Data_C14Cal, Data_SigmaC14Cal, Model_C14, CalibrationCurve.

There are arguments for informations concerning all the samples: Nb_sample, SampleNames, SampleNature, PriorAge, SavePdf, OutputFileName, OutputFilePath, SaveEstimates, OutputTableName, OutputTablePath, StratiConstraints, sepSC.

**** How to fill StratiConstraints ? ****

If there is stratigraphic relations between samples, **14C estimate age in Data_C14Cal must be ordered by order of increasing ages, as informations in DATA**. Names in SampleNames must be ordered and corresponds to the order in Data_C14Cal and in DATA, also if it is needed to mix names of OSL samples and 14C samples.

The user can fill the StratiConstraints matrix as follow.

1. **Size of the matrix:** row number of StratiConstraints matrix is equal to Nb_sample+1, and column number is equal to Nb_sample.
2. **First line of the matrix:** for all i in $\{1, \dots, Nb_Sample\}$, $StratiConstraints[1, i] = 1$ that means the lower bound of the sample age (given in $PriorAge[2i-1]$) for the sample whose number ID is equal to i , is taken into account.
3. **Sample relations:** for all j in $\{2, \dots, Nb_Sample+1\}$ and all i in $\{j, \dots, Nb_Sample\}$, $StratiConstraints[j, i] = 1$ if sample age whose number ID is equal to $j-1$ is lower than sample age whose number ID is equal to i . Otherwise, $StratiConstraints[j, i] = 0$.

Note that $StratiConstraints_{\{2:Nb_sample+1, 1:Nb_sample\}}$ is a upper triangular matrix.

The user can also use [SCMatrix](#) or [SC_Ordered](#) (if all samples are ordered) function to construc the StratiConstraints matrix.

The user can also refer to a csv file that contains the relation between samples as defined above. The user must take care about the separator used in the csv file using the argument sepSC.

**** How to fill THETA covariance matrix concerning common and individual error? ****

If systematic errors are considered, the user can fill the THETA matrix as follow.

- row number of THETA is equal the column number, equal to Nb_sample.
- For all i in $\{1, \dots, Nb_sample\}$, $THETA[i, i]$ contains individual error plus systematic error of the sample whose number ID is equal to i .
- For all i, j in $\{1, \dots, Nb_sample\}$ and i different from j , $THETA[i, j]$ contains common error between samples whose number ID are equal to i and j .

Note that $THETA[i, j]$ is a symetric matrix.

The user can also refer to a .csv file that contains the errors as defined above.

**** Option on growth curves ****

As for [Age_Computation](#) and [Palaeodose_Computation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** (AgesMultiCS2_EXPLIN):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx+d$; select
 - LIN_fit=TRUE
 - Origin_fit=FALSE
- **Saturating exponential growth** (AgesMultiCS2_EXP):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+d$; select
 - LIN_fit=FALSE
 - Origin_fit=FALSE
- **Saturating exponential plus linear growth and fitting through the origin** (AgesMultiCS2_EXPLINZ0):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx$; select
 - LIN_fit=TRUE
 - Origin_fit=TRUE
- **Saturating exponential growth and fitting through the origin** (AgesMultiCS2_EXPZ0):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))$; select
 - LIN_fit=FALSE
 - Origin_fit=TRUE

**** Option on equivalent dose distribution around the palaeodose ****

The use can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample
- **lognormal_A**: a log-normal distribution with mean or **Average** equal to the palaeodose of the sample
- **lognormal_M**: a log-normal distribution with **Median** equal to the palaeodose of the sample

**** More precision on Model ****

We propose two models "full" or "naive". If Model='full' that means measurement error and error on calibration curve are taken account in the Bayesian model; if Model="naive" that means only error on measurement are taken account in the mode.

More precisely, the model considered here, as the one developped by Christen, JA (1994), assume multiplicative effect of errors to address the problem of outliers. In addition, to not penalise variables that are not outliers and damage theirs estimation, we introduce a structure of mixture, that means only variable that are considered as outlier have in addition a multiplicative error.

Value

NUMERICAL OUTPUT

1. **A list containing the following objects:**

- **Sampling:** that corresponds to a sample of the posterior distributions of the age parameters (in ka for both C14 samples and OSL samples);
 - **PriorAge:** stating the priors used for the age parameter;
 - **StratiConstraints:** stating the stratigraphic relations between samples considered in the model;
 - **Model_OSL_GrowthCurve:** stating which dose response fitting option was chosen;
 - **Model_OSL_Distribution:** stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
 - **Model_C14:** stating which model was chosen ("full" or "naive");
 - **CalibrationCurve:** stating which radiocarbon calibration curve was chosen;
 - **Outlier:** stating the names of samples that must be outliers.
2. **The Gelman and Rudin test of convergency:** prints the result of the Gelman and Rudin test of convergency for the age estimate for each sample. A result close to one is expected. In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations). If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (Iter) or be more precise on the PriorAge parameter to reach convergency.
 3. **Credible intervals and Bayes estimates:** prints the Bayes estimates, the credible intervals at 95% and 68% for the age parameters for each sample.

PLOT OUTPUT

1. **MCMC trajectories:** A graph with the MCMC trajectories and posterior distributions of the age parameter is displayed. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.
2. **Age estimate and HPD at 95% of 14C samples on calibration curve:** plot age estimate and HPD on calibration plot.
3. **Summary of sample age estimates:** plot credible intervals and Bayes estimate of each sample age on a same graph.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronk Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4):1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

See Also

[rjags](#), [MCMC_plot](#), [SCMatrix](#)

Examples

```
## Load data
# OSL data
data(DATA1,envir = environment())
data(DATA2,envir = environment())
Data <- Concat_DataFile(DATA2,DATA1)

# 14C data
C14Cal <- DATA_C14$C14[1,1]
SigmaC14Cal <- DATA_C14$C14[1,2]
Names <- DATA_C14$Names[1]

# Prior Age
prior=rep(c(1,60),3)
samplenature=matrix(data=c(1,0,1,0,1,0),ncol=3,nrow=2,byrow=TRUE)
SC <- matrix(data=c(1,1,1,0,1,1,0,0,1,0,0,0),ncol=3,nrow=4,byrow=TRUE)

## Age computation of samples
Age <- Age_OSLC14(DATA=Data,Data_C14Cal=C14Cal,Data_SigmaC14Cal=SigmaC14Cal,
  SampleNames=c("GDB5",Names,"GDB3"),Nb_sample=3,SampleNature=samplenature,
  PriorAge=prior,StratiConstraints=SC,Iter=50,Nb_chaines=2)
```

AtmosphericNorth_CalC14

Atmospheric North data for calibration of 14C age

Description

As 14C years is not equal to calendar years because atmospheric 14C concentration varies through time. Hence, data in AtmosphericNorth_CalC14 allows a calibration for mid-latitude Northern Hemisphere atmosphere reservoir.

Usage

```
data("AtmosphericNorth_CalC14")
```

Format

A data frame with 5 variables.

CAL.BP a numeric vector corresponding to calendar years before present

X14C.age a numeric vector corresponding to 14C age

Error a numeric vector corresponding to error around 14C age measurement

References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronk Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafliðason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Examples

```
data(AtmosphericNorth_CalC14)
## maybe str(AtmosphericNorth_CalC14) ; head(AtmosphericNorth_CalC14) ...
```

AtmosphericSouth_CalC14

Atmospheric South data for calibration of 14C age

Description

As 14C years is not equal to calendar years because atmospheric 14C concentration varies through time. Hence, data in AtmosphericSouth_CalC14 allows a calibration for mid-latitude Southern Hemisphere atmosphere reservoir.

Usage

```
data("AtmosphericSouth_CalC14")
```

Format

A data frame with 3 variables.

CAL.BP a numeric vector corresponding to calendar years before present

X14C.age a numeric vector corresponding to 14C age

Error a numeric vector corresponding to error around 14C age measurement

References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronk Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Examples

```
data(AtmosphericSouth_CalC14)
## maybe str(AtmosphericSouth_CalC14) ; head(AtmosphericSouth_CalC14) ...
```

Concat_DataFile	<i>Concatenates two outputs of Generate_DataFile or Generate_DataFile_MG</i>
-----------------	--

Description

This function allows concatenating two lists provided by [Generate_DataFile](#) or [Generate_DataFile_MG](#) functions.

Only concatenation of 2 files is possible.

Usage

```
Concat_DataFile(L1, L2)
```

Arguments

L1	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement.
L2	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement.

Details

For more information on the structure of the input list, we refer to **Value** section of [Generate_DataFile](#) or [Generate_DataFile_MG](#).

This function is especially usefull in two cases:

- if informations concerning samples are yet saved in RData file (that allow to not run again [Generate_DataFile](#) or [Generate_DataFile_MG](#) that can take time);
- the user want to analyse simultaneously Single-grain and Multi-grain OSL measurements, because sample are in under stratigraphic constraints or they share systematic errors.

Value

A List of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

See Also

[Generate_DataFile](#), [Generate_DataFile_MG](#)

Examples

```
# load data files
data(DATA1,envir = environment())
data(DATA2,envir = environment())
# concatenate two data files
DATA3=Concat_DataFile(L1=DATA1,L2=DATA2)
str(DATA3)
```

DATA1

*DATA of sample named GDB3***Description**

list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J,K,Nb_measurement obtained using [Generate_DataFile](#) function with single-grain OSL measurements of the sample GDB3.

Usage

```
data("DATA1")
```

Format

A list containing:

LT: (one list per sample): each list contains all L/T values for the corresponding sample;

sLT: (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes: (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab= a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot_env: a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose: (one list per sample): each list contains all regenerated doses;

J: a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K: a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb_measurement: a vector giving, for each BIN file, the number of measurements;

References

For more informations on this sample we refer to the following publication:

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D, Pearson, O , Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

Examples

```
data(DATA1)
str(DATA1)
```


DATA2

*DATA on sample named GDB5***Description**

list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J,K,Nb_measurement obtained using [Generate_DataFile](#) function with single-grain OSL measurements of the sample GDB5.

Usage

```
data("DATA2")
```

Format

A data frame containing:

LT: (one list per sample): each list contains all L/T values for the corresponding sample;

sLT: (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes: (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab: a matrix containing in line *i*, the laboratory dose rate and its variance for sample *i*;

ddot_env: a matrix containing in line *i*, the environmental dose rate and its variance (excluding the common error terms) for sample *i*;

regDose: (one list per sample): each list contains all regenerated doses;

J: a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K: a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb_measurement: , a vector giving, for each BIN file, the number of measurements;

References

For more informations on this sample we refer to the following publication:

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D, Pearson, O , Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

Examples

```
data(DATA2)
str(DATA2)
```

DATA3

*DATA of sample named FER1***Description**

list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J,K,Nb_measurement obtained using [Generate_DataFile](#) function with multi-grain OSL measurements of the sample FER1.

Usage

```
data("DATA3")
```

Format

A list containing:

LT: (one list per sample): each list contains all L/T values for the corresponding sample;

sLT: (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes: (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab= a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot_env: a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose: (one list per sample): each list contains all regenerated doses;

J: a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K: a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb_measurement: a vector giving, for each BIN file, the number of measurements;

References

For more informations on this sample we refer to the following publication:

Guerin, G., Frouin, M., Talamo, S., Aldeias, V., Bruxelles, L., Chiotti, L., Goldberg, P., Hublin, J.J., Jain, M., Lahaye, C., Madelaine, S., Maureille, B., McPherron, S., Mercier, N., Murray, A., Sandgathe, D., Steele, T., Thomsen, K., Turq, A. (2015). A multi-method luminescence dating of the Palaeolithic sequence of La Ferrassie based on new excavations adjacent to the La Ferrassie 1 and 2 skeletons. *Journal of Archaeological Science*, 58, 147-166.

Examples

```
data(DATA3)
str(DATA3)
```

DATA_C14

*C14 cal age estiamte and its error***Description**

C14 cal age estiamtes and theirs error of samples S-EVA-26510, S-EVA-26506, S-EVA-26507, S-EVA-26508.

Usage

```
data("DATA_C14")
```

Format

A list containing:

Names: character vector of the sample names;

C14: numeric matrix, in the first column the 14C Cal age of the samples, and in the second column theirs errors.

References

For more informations on this sample we refer to the following publication:

Guerin, G., Frouin, M., Talamo, S., Aldeias, V., Bruxelles, L., Chiotti, L., Goldberg, P., Hublin, J.J., Jain, M., Lahaye, C., Madelaine, S., Maureille, B., McPherron, S., Mercier, N., Murray, A., Sandgathe, D., Steele, T., Thomsen, K., Turq, A. (2015). A multi-method luminescence dating of the Palaeolithic sequence of La Ferrassie based on new excavations adjacent to the La Ferrassie 1 and 2 skeletons. *Journal of Archaeological Science*, 58, 147-166.

Examples

```
data(DATA_C14)
(DATA_C14)
```

Generate_DataFile

Generates, from one (or several) BIN-file(s) of Single-grain OSL measurements, a list of luminescence data and information before statistical analysis

Description

This function is used to generate, from the BIN file(s), a list of values of: **Single-grain** OSL intensities and associated uncertainties, regenerative doses, etc., which will be the input of the Bayesian models. To be easy-to-use, this function requires a rigorous organisation - all needed files should be arranged in one folder - of informations concerning each BIN file.

It is possible to process data for various samples simultaneously and to consider more than one BIN file per sample.

Usage

```
Generate_DataFile(Path, FolderNames, Nb_sample,
  Nb_binfile = length(FolderNames), BinPerSample = rep(1, Nb_sample),
  sepDP = c(", "), sepDE = c(", "), sepDS = c(", "), sepR = c("="),
  verbose = TRUE, ...)
```

Arguments

Path	character (required) : the path to the project folder, containing one or more subfolders in which the BIN files are located. If it is not equal to "", it must be terminated by "/".
FolderNames	character (required) : list of names of the sub-folders containing the BIN files <ul style="list-style-type: none"> • each subfolder must contain a BIN file and associated csv files. See details for more informations on associated csv files required in the subfolders. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill the FolderNames vector.
Nb_sample	integer (required) : number of samples.
Nb_binfile	integer (with default): number of BIN files. It must be equal to, or greater than Nb_sample.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector must be equal to Nb_sample and the sum of entries of this vector must be equal to Nb_binfile. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill BinPerSample vector. Otherwise, this vector must contain a list of 1 values.
sepDP	character (with default): column separator in the DiscPose.csv files.
sepDE	character (with default): column separator in the DoseEnv.csv files.
sepDS	character (with default): column separator in the DoseLab.csv files.
sepR	character (with default): column separator in the Rule.csv files.
verbose	logical (with default): enable/disable verbose mode
...	further arguments that can be passed to Luminescence::read_BIN2R .

Details

With Path and FolderNames, this function goes to the subfolders containing the BIN files and associated information to compute the luminescence data.

**** What are the required files in each subfolder? ****

Each subfolder can be named, for example, as the sample name followed by a number; it must contain:

- **bin.BIN**: the bin file renamed as bin.BIN (note: the name of all files matters);
- **DiscPos.csv**: a two columns csv file containing the list of disc and grain position number of the previously selected grains (typically this list will include the position of grains based on their sensitivity, recycling or other properties);
- **DoseEnv.csv**: a two columns file containing the observation of the natural (or environmental), dose rate, and its non-shared variance (i.e. after removing all shared errors), both in Gy. Note: the user shall provide the squared value of the error associated with the dose rate experienced by the sample grains in nature;

- **DoseSource.csv**: a two columns file containing the observation of the laboratory dose rate, and its variance (squared error) both in Gy;
- **rule.csv**: a csv file containing information on
 - beginSignal= the first channel for summing the natural or regenerative OSL signal (typically 1 or 6);
 - endSignal= the last channel for summing the natural or regenerative OSL signal (typically 5 or 10);
 - beginBackground= the first channel for background estimation of the natural or regenerative OSL signal (typically 76 or 81);
 - endBackground= the last channel for background estimation of the natural or regenerative OSL signal (typically 95 or 100);
 - beginTest=,
 - endTest=,
 - beginTestBackground=,
 - endTestBackground= same values as above, for the test dose response (typically the same values should be used);
 - inflatePercent= uncertainty arising from the instrument reproducibility (typically 0.02, i.e. 2%);
 - nbOfLastCycleToRemove= number of cycles at the end of the SAR protocol which should not be included in the dose response curve fitting (typically 1 if only a recycling test is performed, or 2 if both recycling and IR depletion are tested).

**** How to fill the FolderNames vector? ****

FolderNames is a vector of length Nb_binfile. FolderNames[i] is the name (e.g., Sample1-File1, or successive names separated by "/" signs, if BIN files are in subfolders, e.g. Sample1/File1) of the subfolder containing all informations on the BIN file of ID number i. The names in FolderNames are ordered following two rules:

- The names in the FolderNames vector must be ordered following the sample order (the names of subfolders containing BIN files for the same sample should follow each other in the FolderNames vector, e.g. Sample1, Sample2-File1, Sample2-File2, etc.).
- If stratigraphic constraints apply to samples, and so a **Bayesian model with stratigraphic constraints** is implemented, then the names in the FolderNames vector must be ordered by order of increasing ages.

For example, FolderNames=c(noun1, noun2), in which case noun1 (respectively, noun2) corresponds to the subfolder name containing the BIN file of sample 1 (respectively of sample 2). In addition, if we know that sample 1 is younger than sample 2, then FolderNames vector is correctly filled.

If conversely, FolderNames=c(noun2, noun1), the analysis performed by [AgeS_Computation](#) would not be consistent.

**** How to fill the BinPerSample vector? ****

BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i.

For example, let us consider a case with two samples (Sample1 and Sample2), with 2 BIN files for Sample1 and 1 for Sample2. In this case, Nb_binfile=3 and Nb_sample=2. The user may then set FolderNames=c("Sample1-File1", "Sample1-File2", "Sample2-File1"), in which

case "Sample1-File1" is the name of the subfolder containing the first BIN file for Sample1, "Sample1-File2" the name of the subfolder for the second BIN file of Sample1; eventually, "Sample2-File1" is the name of the subfolder containing the BIN file for the second sample. In this case, BinPerSample=c(2,1).

For the general BIN-file structure, the reader is referred to the following website: <http://www.nutech.dtu.dk/>

The function `read_BIN2R` developed in `Luminescence` package is used to read the BIN files.

Value

A list containing the following objects:

- **LT** (one list per sample); each list contains all L/T values for the corresponding sample;
- **sLT** (one list per sample); each list contains all uncertainties on L/T values for the corresponding sample;
- **ITimes** (one list per sample); each list contains irradiation time values for the corresponding sample;
- **dLab**, a matrix containing in line *i*, the laboratory dose rate and its variance for sample *i*;
- **ddot_env**, a matrix containing in line *i*, the environmental dose rate and its variance (excluding the common error terms) for sample *i*;
- **regDose** (one list per sample); each list contains all regenerated doses;
- **J**, a vector giving, for each BIN file, the number of aliquots selected for the analysis;
- **K**, a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;
- **Nb_measurement**, a vector giving, for each BIN file, the number of measurements.

** How to save this list **

You can save this list in a `.RData` object. To do this, you can use the fonction `save`. Then, to load this list you can use the function `load` (see example section fore more details).

Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guerin

See Also

[read_BIN2R](#), [Concat_DataFile](#), [Generate_DataFile_MG](#), [LT_RegenDose Age_Computation](#), [AgeS_Computation](#), [Palaeodose_Computation](#)

Examples

```
## Example for one sample with one Bin File
path<- system.file("extdata/samp1", "", package="BayLum")
folder=""
nbsample=1 # give the number of sample
Data <- Generate_DataFile(
  Path = path,
  FolderNames = folder,
  Nb_sample = nbsample,
  verbose = FALSE)
str(Data)
```

```
## to save information in RData object in folder containing bin file
# save(Data,file=c(paste(path,folder,'Data.RData',sep="")))
## to load information containing Data.RData object
# load(file=c(paste(path,folder,"Data.RData",sep="")))
```

Generate_DataFile_MG	<i>Generates, from one (or several) BIN file(s) of Multi-grain OSL measurements, a list of luminescence data and information before statistical analysis</i>
----------------------	--

Description

This function is used to generate, from the BIN file(s), a list of values of: **Multi-grain** OSL intensities and associated uncertainties, regenerative doses, etc., which will be the input of the Bayesian models. To be easy-to-use, this function requires a rigorous organisation - all needed files should be arranged in one folder - of informations concerning each BIN file.

It is possible to process data for various samples simultaneously and to consider more than one BIN file per sample.

Usage

```
Generate_DataFile_MG(Path, FolderNames, Nb_sample,
  Nb_binfile = length(FolderNames), BinPerSample = rep(1, Nb_sample),
  sepD = c(","), sepDE = c(","), sepDS = c(","), sepR = c("="),
  verbose = TRUE, ...)
```

Arguments

Path	character (required) : the path to the project folder, containing one or more subfolders in which the BIN files are located. If it is not equal to "", it must be terminated by "/".
FolderNames	character (required) vector: list of names of the sub-folders containing the BIN files <ul style="list-style-type: none"> each subfolder must contain a BIN file and associated csv files. See details for more informations on associated csv files required in the subfolders. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill the FolderNames vector.
Nb_sample	integer (required) : number of samples.
Nb_binfile	integer (with default): number of BIN files. It must be equal to, or greater than Nb_sample.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector must be equal to Nb_sample and the sum of entries of this vector must be equal to Nb_binfile. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill BinPerSample vector. Otherwise, this vector must contain a list of 1 values.
sepD	character (with default): column separator in the DiscPose.csv files.
sepDE	character (with default): column separator in the DoseEnv.csv files.
sepDS	character (with default): column separator in the DoseLab.csv files.

sepR [character](#) (with default): column separator in the Rule.csv files.
 verbose [logical](#) (with default): enable/disable verbose mode
 ... further arguments that can be passed to [Luminescence::read_BIN2R](#).

Details

With Path and FolderNames, this function goes to the subfolders containing the BIN files and associated information to compute the luminescence data.

**** What are the required files in each subfolder? ****

Each subfolder can be named, for example, as the sample name followed by a number; it must contain:

- **bin.BIN**, the bin file renamed as bin.BIN (note: the name of all files matters);
- **Disc.csv**, a one columns csv file containing the list of disc number of the previously selected grains (typically this list will include the position of grains based on their sensitivity, recycling or other properties);
- **DoseEnv.csv**, a two columns file containing the observation of the natural (or environmental), dose rate, and its non-shared variance (i.e. after removing all shared errors), both in Gy. Note: the user shall provide the squared value of the error associated with the dose rate experienced by the sample grains in nature;
- **DoseSource.csv**, a two columns file containing the observation of the laboratory dose rate, and its variance (squared error), both in Gy;
- **rule.csv**, a csv file containing information on
 - beginSignal= the first channel for summing the natural or regenerative OSL signal (typically 1 or 6);
 - endSignal= the last channel for summing the natural or regenerative OSL signal (typically 5 or 10);
 - beginBackground= the first channel for background estimation of the natural or regenerative OSL signal (typically 76 or 81);
 - endBackground= the last channel for background estimation of the natural or regenerative OSL signal (typically 95 or 100);
 - beginTest,
 - endTest,
 - beginTestBackground,
 - endTestBackground= same values as above, for the test dose response (typically the same values should be used);
 - inflatePercent= uncertainty arising from the instrument reproducibility (typically 0.02, i.e. 2%);
 - nbOfLastCycleToRemove= number of cycles at the end of the SAR protocol which should not be included in the dose response curve fitting (typically 1 if only a recycling test is performed, or 2 if both recycling and IR depletion are tested).

**** How to fill the FolderNames vector? ****

FolderNames is a vector of length Nb_binfile. FolderNames[i] is the name (e.g., Sample1-File1, or successive names separated by "/" signs, if BIN files are in subfolders, e.g. Sample1/File1) of the subfolder containing all informations on the BIN file of ID number i. The names in FolderNames are ordered following two rules:

- The names in the FolderNames vector must be ordered following the sample order (the names of subfolders containing BIN files for the same sample should follow each other in the FolderNames vector, e.g. Sample1, Sample2-File1, Sample2-File2, etc.).
- If stratigraphic constraints apply to samples, and so a **Bayesian model with stratigraphic constraints** is implemented, then the names in the FolderNames vector must be ordered by order of increasing ages.
For example, FolderNames=c(noun1, noun2), in which case noun1 (respectively, noun2) corresponds to the subfolder name containing the BIN file of sample 1 (respectively of sample 2). In addition, if we know that sample 1 is younger than sample 2, then FolderNames vector is correctly filled.
If conversely, FolderNames=c(noun2, noun1), the analysis performed by [AgeS_Computation](#) would not be consistent.

**** How to fill the BinPerSample vector? ****

BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i.

For example, let us consider a case with two samples (Sample1 and Sample2), with 2 BIN files for Sample1 and 1 for Sample2. In this case, Nb_binfile=3 and Nb_sample=2. The user may then set FolderNames=c("Sample1-File1", "Sample1-File2", "Sample2-File1"), in which case "Sample1-1" is the name of the subfolder containing the first BIN file for Sample1, "Sample1-File2" the name of the subfolder for the second BIN file of Sample1; eventually, "Sample2-1" is the name of the subfolder containing the BIN file for the second sample. In this case, BinPerSample=c(2,1).

For the general BIN-file structure, the reader is referred to the following website: <http://www.nutech.dtu.dk/>

The function [read_BIN2R](#) developed in [Luminescence](#) package is used to read the BIN files.

Value

A list containing the following objects:

- **LT** (one list per sample); each list contains all L/T values for the corresponding sample;
- **sLT** (one list per sample); each list contains all uncertainties on L/T values for the corresponding sample;
- **ITimes** (one list per sample); each list contains irradiation time values for the corresponding sample;
- **dLab**, a matrix containing in line i, the laboratory dose rate and its variance for sample i;
- **ddot_env**, a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;
- **regDose** (one list per sample); each list contains all regenerated doses;
- **J**, a vector giving, for each BIN file, the number of aliquots selected for the analysis;
- **K**, a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;
- **Nb_measurement**, a vector giving, for each BIN file, the number of measurements;

**** How to save this list ****

You can save this list in a .RData object. To do this, you can use the function [save](#). Then, to load this list you can use the function [load](#) (see example section for more details).

Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guerin

See Also

[read_BIN2R](#), [Concat_DataFile](#), [LT_RegenDose](#) [Age_Computation](#), [AgeS_Computation](#), [Palaeodose_Computation](#)

Examples

```
path<- system.file("extdata/FER1", "", package="BayLum")
folder=""
# give the number of sample
nbsample=1
DATA=Generate_DataFile_MG(Path=path,FolderNames=folder,Nb_sample=nbsample)
str(DATA)

# to save information in RData object in folder containing bin file
#save(DATA,file=c(paste(path,folder,'DATA.RData',sep="")))
# to load information containing DATA.RData object
#load(file=c(paste(path,folder,"DATA.RData",sep="")))
```

LT_RegenDose

Plots Lx/Tx as a function of the regenerative dose

Description

This function plots Lx/Tx values as a function of regenerative dose, for every selected aliquot and for each sample.

Usage

```
LT_RegenDose(DATA, Path, FolderNames, SampleNames = FolderNames, Nb_sample,
  BinPerSample = rep(1, Nb_sample), SG = rep(TRUE, Nb_sample),
  sepDP = c(", "), nrow = 3L, ncol = nrow)
```

Arguments

DATA	list (required) : list of objects LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by Generate_DataFile or Generate_DataFile_MG or Concat_DataFile . DATA can contain information from more than one sample.
Path	character (required) : path to the project folder (the same as the one used in Generate_DataFile or Generate_DataFile_MG to provide DATA)
FolderNames	character (required) : vector of names of the sub-folders containing the BIN files, which were used by Generate_DataFile or Generate_DataFile_MG to generate the DATA object.
SampleNames	character (with default): Names of samples. To use if there is more than one bin file per sample.
Nb_sample	integer (required) : ID number (in [1,Nb_sample]) of the sample selected for plotting L/T as a function of regenerative doses. Required if the DATA object contains information for more than one sample.
BinPerSample	integer (with default): integer vector (with default): vector with the number of BIN files per sample, which was used in Generate_DataFile or Generate_DataFile_MG to generate the DATA object.

SG	logical (with default): vector to set the type of measurement for each sample (length(SG)=Nb_sample). If the sample of number ID equal to i, SG[i]=TRUE if it is a Single-grain OSL measurements, SG[i]=FALSE if it is a Multi-grain OSL measurements.
sepDP	character (with default): column separator in the DiscPose.csv file or in Disc.csv file. It must be the same separator for all samples, for Single-grain OSL measurements or Multi-grain OSL measurements.
nrow	integer (with default): controls the arangement of the plots, here the number of rows. Can be set to NULL.
ncol	integer (with default): controls the arangement of the plots, here the number of columns. Can be set to NULL.

Details

To fill FolderNames and BinPerSample, we refer to the **Detail** section from the [Generate_DataFile](#) or [Generate_DataFile](#) function. As well for a precise description of input DATA.

Value

Lx/Tx plots; there are as many plots as selected aliquots in the DiscPos.csv file. There are 9 plots per page. There is not interpolation.

Author(s)

Claire Christophe, Sebastian Kreutzer, Anne Philippe, Guillaume Guérin

See Also

[Generate_DataFile](#), [Generate_DataFile_MG](#)

Examples

```
## load data file generated by the function Generate_DataFile
data(DATA3,envir = environment())
path<- system.file("extdata/FER1", "", package="BayLum")
folder=""
samplename="FER1"
LT_RegenDose(DATA=DATA3,Path=path,FolderNames=folder,SampleNames=samplename,Nb_sample=1,SG=FALSE)
```

Marine_CalC14

Marine data for calibration of 14C age

Description

As 14C years is not equal to calendar years because atmospheric 14C concentration varies through time. Hence, data in marine_CalC14 allows a calibration for hypothetical "global" marine reservoir.

Usage

```
data("Marine_CalC14")
```

Format

A data frame with 3 variables.

CAL.BP a numeric vector correpondig to calendar years befor present

X14C.age a numeric vector corresponding to 14C age

Error a numeric vector corresponding to error arround 14C age measurement

References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 ans Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4)=1869-1887.

Examples

```
data(Marine_CalC14)
## maybe str(Marine_CalC14) ; head(Marine_CalC14) ...
```

MCMCsample

MCMC sample from the posterior distribution of the dataset GDB5

Description

MCMC samples from the posterior distribution of "A" for age, "D" for palaeodose and "sD" for dispersion of equivalent doses around "D", of the data set GDB5.

Usage

```
data("MCMCsample")
```

Format

It is a matric with 6000 row and tree column.

A The first column of the matrice are sampled from the posterior distribution of the paramete A

D The first column of the matrice are sampled from the posterior distribution of the paramete D

sD The first column of the matrice are sampled from the posterior distribution of the paramete sD

References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D, Pearson, O , Puaud, S, Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

Examples

```
data(MCMCsample)
## maybe str(MCMCsample) ; plot(MCMCsample[,1],type="l") ...
```

Description

This function uses the output of `coda.samples` to plot the trajectories of MCMC and densities of the posterior distributions of the age - if it is calculated, palaeodose and equivalent dose dispersion parameters of the sample. This function is used in the function `Age_Computation`, `AgeS_Computation` and `Palaeodose_Computation`.

Usage

```
MCMC_plot(sample, size, SampleNames, Nb_sample = 1, Nb_chaines = 3,
  value = c(0, Nb_sample, 2 * Nb_sample), param = c("A", "D", "sD"))
```

Arguments

<code>sample</code>	MCMC.list: that is generated by <code>jags.model</code> in <code>Age_Computation</code> .
<code>size</code>	integer: length of each chain.
<code>SampleNames</code>	character vector: names of the samples, used in the figure titles.
<code>Nb_sample</code>	integer (with default): number of analysed samples.
<code>Nb_chaines</code>	integer (with default): number of independent chains for the model (for more informations, see <code>jags.model</code>).
<code>value</code>	integer vector (with default): position index used to select age (if available), palaeodose and equivalent dose dispersion parameters for the first sample.
<code>param</code>	character vector (with default): names of the selected parameters with value A for age (if available), D for palaeodose and sD for equivalent dose dispersion.

Value

A pdf file with the MCMC trajectories and posterior distributions for each parameter defined in `param`. There is one page per sample, which is divided by `length(param)` vertically and by 2 horizontally.

The first line of the figure corresponds to the first parameter defined in `param`, and so on. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distributions of the parameter.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

See Also

`Age_Computation`, `AgeS_Computation`, `Palaeodose_Computation`, `coda.samples` and `rjags` packages.

Examples

```
data(MCMCsample, envir = environment())
MCMC_plot(sample=MCMCsample, size=2000, SampleNames="GDB3")
```

ModelC14

Likelihood of C14 samples for JAGS models use in Age_OSLC14

Description

A list of models for C14 data to define likelihood in JAGS models.

Usage

```
data("ModelC14")
```

Format

This list contains:

full a model considering error on calibration curve.

naive a model not considering error on calibration curve.

References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafliðason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4):1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

Examples

```
data(Model_AgeC14)
## ModelC14[[full]]
## ModelC14$full
```

ModelOSL

Likelihood of OSL samples for JAGS models use in Age_OSLC14

Description

A list of models for OSL data to define likelihood in JAGS models.

Usage

```
data("ModelOSL")
```

Format

This list contains:

`AgesMultiCS2_EXPLIN` a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgesMultiCS2_EXP` a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgesMultiCS2_EXPZO` a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgesMultiCS2_EXPLINZO` a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

Details

The different distributions to describe equivalent dose values around the palaeodose are:

`cauchy` a Cauchy distribution with position parameter equal to the palaeodose of the sample

`gaussian` a Gaussian distribution with mean equal to the palaeodose of the sample

`lognormal_A` a log-normal distribution with mean or Average equal to the palaeodose of the sample

`lognormal_M` a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [AgeS_Computation](#), section Details.

References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

Examples

```
data(ModelOSL)
## The JAGS model of the likelihood for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
## ModelOSL[[AgesMultiOSL_EXPLIN]][[gaussian]]
```

ModelPrior

Prior for JAGS models use in Age_OSLC14

Description

A list to define prior in JAGS models, taking account OSL data and C14 data in stratigraphic constraint. The difficulty is in the fact that each cases is different. The youngest sample can be a C14 as well as a OSL sample. To resolve this problem we consider diferent cases thanks to this list.

Usage

```
data("ModelPrior")
```

Format

This list contains:

Sample1_C14 model considering that the youngest sample is a C14 sample

Sample1_OSL model considering that the youngest sample is a OSL sample

C14_OSL model considering that the second sample is a C14 sample

OSL_C14 model considering that the second sample is a OSL sample

C14 model considering that the last sample is a C14 sample

OSL model considering that the last sample is a OSL sample

References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

Examples

```
data(ModelPrior)
## ModelPrior[[OSL]]
## ModelPrior$OSL
```

Model_Age

JAGS models use in Age_Computation

Description

A list of JAGS models use to a Bayesian analysis of OSL age of one sample. There are models for various growth curves and various distrubution to describe equivalent dose distribution around the palaeodose.

Usage

```
data("Model_Age")
```


Format

This list contains:

`AgeMultiBF_EXPLIN` a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgeMultiBF_EXP` a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgeMultiBF_EXPZO` a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgeMultiBF_EXPLINZO` a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

Details

The different distributions to describe equivalent dose values around the palaeodose are:

`cauchy` a Cauchy distribution with position parameter equal to the palaeodose of the sample

`gaussian` a Gaussian distribution with mean equal to the palaeodose of the sample

`lognormal_A` a log-normal distribution with mean or Average equal to the palaeodose of the sample

`lognormal_M` a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [Age_Computation](#), section Details.

References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

See Also

[rjags](#)

Examples

```
data(Model_Age)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[AgeMultiBF_EXPLIN]][[gaussian]]
```

Model_AgeC14

*JAGS models use in AgeC14_Computation***Description**

A list of JAGS models use to a Bayesian analysis of C14 calibration age of various sample. Stratigraphic relations can be taken in count to calibrate C14 ages. This ages take into account that some data can be an outlier.

Usage

```
data("Model_AgeC14")
```

Format

This list contains:

full a model considering error on calibration curve.

naive a model not considering error on calibration curve.

References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafliadason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4):1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

See Also

[rjags](#)

Examples

```
data(Model_AgeC14)
## Model_AgeC14[[full]]
## Model_AgeC14$full
```

Model_AgeS

*JAGS models use in AgeS_Computation***Description**

A list of JAGS models use to a Bayesian analysis of OSL age of various samples. There are models for various growth curves and various distrubution to describe equivalent dose distribution around the palaeodose.

Usage

```
data("Model_AgeS")
```

Format

This list contains:

`AgesMultiCS2_EXPLAIN` a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgesMultiCS2_EXP` a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgesMultiCS2_EXPZ0` a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgesMultiCS2_EXPLAINZ0` a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

Details

The different distributions to describe equivalent dose values around the palaeodose are:

`cauchy` a Cauchy distribution with postition parameter equal to the palaeodose of the sample

`gaussian` a Gaussian distribution with mean equal to the palaeodose of the sample

`lognormal_A` a log-normal distribution with mean or Average equal to the palaeodose of the sample

`lognormal_M` a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [AgeS_Computation](#), section Details.

References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

See Also

[rjags](#)

Examples

```
data(Model_AgeS)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type  $f(x)=a(1-\exp(-x/b))+cx+d$ )
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[AgesMultiCS2_EXPLIN]][[gaussian]]
```

Model_Palaeodose	<i>JAGS models use in Palaeodose_Computation</i>
------------------	--

Description

A list of JAGS models use to a Bayesian analysis of OSL palaeodose of one or various samples. There are models for various growth curves and various distribution to describe equivalent dose distribution around the palaeodose.

Usage

```
data("Model_Palaeodose")
```

Format

This list contains:

PalaeodosesMultiBF_EXPLIN a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

PalaeodosesMultiBF_EXP a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

PalaeodosesMultiBF_EXPZO a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

PalaeodosesMultiBF_EXPLINZO a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

Details

The different distributions to describe equivalent dose values around the palaeodose are:

cauchy a Cauchy distribution with position parameter equal to the palaeodose of the sample

gaussian a Gaussian distribution with mean equal to the palaeodose of the sample

lognormal_A a log-normal distribution with mean or Average equal to the palaeodose of the sample

lognormal_M a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [Palaeodose_Computation](#), section Details.

References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

See Also

[rjags](#)

Examples

```
data(Model_Palaeodose)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[PalaeodosesMultiBF_EXPLIN]][[gaussian]]
```

Palaeodose_Computation

Bayesian analysis for the palaeodose estimation of various samples

Description

This function computes the palaeodose (in Gy) of one or various samples according to the model developed in Combes et al (2015), based on an output of [Generate_DataFile](#) or [Generate_DataFile_MG](#) or both of them using [Concat_DataFile](#).

Samples, for which data is available in several BIN files, can be analysed.

Single-grain or Multi-grain OSL measurements can be analysed simultaneously.

Usage

```
Palaeodose_Computation(DATA, SampleNames, Nb_sample, BinPerSample = rep(1,
  Nb_sample), SavePdf = FALSE, OutputFileName = c("MCMCplot"),
  OutputFilePath = c(""), SaveEstimates = FALSE,
  OutputTableName = c("DATA"), OutputTablePath = c(""), LIN_fit = TRUE,
  Origin_fit = FALSE, distribution = c("cauchy"), Iter = 50000, t = 5,
  Nb_chaines = 3)
```

Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by Generate_DataFile or Generate_DataFile_MG . DATA contains information for more than one sample.
SampleNames	character vector: names of sample. The length of this vector is equal to Nb_sample.
Nb_sample	integer: number of samples.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector is equal to Nb_sample. BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in Generate_DataFile or Generate_DataFile_MG .

SavePdf	boolean (with default): if TRUE save graph in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf files that will be generated by the function.
OutputFilePath	character (with default): path to the pdf files that will be generated by the function.
SaveEstimates	boolean (with default): if TRUE save Bayes estimates and credible interval at level 68 in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE.
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. Please see details for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. Please see details for more informations on the proposed growth curves.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M".
Iter	integer (with default): number of iterations for the MCMC computation (for more information see jags.model).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see jags.model).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see jags.model).

Details

** Option on growth curves **

As for [Age_Computation](#) and [AgeS_Computation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** (PalaeodosesMultiBF_EXPLIN):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx+d$; select
 - LIN_fit=TRUE
 - Origin_fit=FALSE
- **Saturating exponential growth** (PalaeodosesMultiBF_EXP):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+d$; select
 - LIN_fit=FALSE
 - Origin_fit=FALSE
- **Saturating exponential plus linear growth and fitting through the origin** (PalaeodosesMultiBF_EXPLINZ0):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))+cx$; select
 - LIN_fit=TRUE

- Origin_fit=TRUE
- **Saturating exponential growth and fitting through the origin** (PalaeodosesMultiBF_EXPZO):
for all x in \mathbb{R}^+ , $f(x)=a(1-\exp(-x/b))$; select
 - LIN_fit=FALSE
 - Origin_fit=TRUE

** Option on equivalent dose distribution around the palaeodose **

The use can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample
- **lognormal_A**: a log-normal distribution with mean or Average equal to the palaeodose of the sample
- **lognormal_M**: a log-normal distribution with Median equal to the palaeodose of the sample

Value

NUMERICAL OUTPUT

1. A list containing the following objects:

- **Sampling** that corresponds to a sample of the posterior distributions of palaeodose and equivalent dose dispersion parameters (both in Gy).
- **Model_GrowthCurve**, stating which dose response fitting option was chosen;
- **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample.

2. The Gelman and Rudin test of convergency:

prints the result of the Gelman and Rudin test of convergency for palaeodose and equivalent dose dispersion parameters for each sample. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the pdf file generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the printed estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (*Iter*) to reach convergency.

3. Credible intervals and Bayes estimates:

prints the Bayes estimates, the credible intervals at 95% and 68% for the palaeodose and equivalent dose dispersion parameters for each sample.

PLOT OUTPUT

1. MCMC trajectories

A graph with the MCMC trajectories and posterior distributions of the palaeodose and equivalent dose dispersion parameters is displayed, there is one page per sample.

The first line of the figure corresponds to the palaeodose parameter and the second to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

2. Summary of palaeodose estimates:

plot credible intervals and Bayes estimate of each sample palaeodose on a same graph.

To give result in a publication, we recommend to give the Bayes estimate of the parameters as well as the credible interval at 95% or 68%.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

References

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. Quaternary Geochronology 28, 62-70. doi:10.1016/j.quageo.2015.04.001

See Also

[Generate_DataFile](#), [Generate_DataFile_MG](#), [Concat_DataFile](#), [rjags](#), [MCMC_plot](#), [Age_Computation](#), [AgeS_Computation](#)

Examples

```
## Load data
data(DATA1,envir = environment())
## Palaeodose computation of samples GDB3
P=Palaeodose_Computation(DATA=DATA1,Nb_sample=1,SampleNames=c("GDB5"),Iter=100)
```

ScatterSamples

Scatterplots of samples

Description

This fonction compute a matrix of scatterplots of at most 8 samples.

Usage

```
ScatterSamples(Sampling, SampleNames, SelectedSamples, sc = 3,
  Nb_chaines = 3)
```

Arguments

Sampling	MCMC.list: that is generated by jags.model in AgeS_Computation , AgeC14_Computation or Age_OSLC14 .
SampleNames	character vector: names of samples.
SelectedSamples	numeric vector: indicate if the sample is selected to be plot with the other samples. If the sample with names <code>SampleNames[i]</code> is selecteted, then <code>SelectedSamples[i]=1</code> ; if not <code>SelectedSamples=0</code> . <code>length(SelectedSamples)=length(SampleNames)</code> . At least two samples must be selected.
sc	numeric (with default): to define the number of values on the axis.
Nb_chaines	integer (with default): number of independent chains used in AgeS_Computation to generate the sampling (for more information see jags.model).

Value

A matrix that contains at row *i* and column *j* the scatterplot of the posterior distribution of the sample ID *i* plotted against the one of the sample ID *j*

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

See Also

[Age_Computation](#), [AgeS_Computation](#), [AgeC14_Computation](#), and [rjags](#) packages.

Examples

```
data(AgeS,envir = environment())
ScatterSamples(Sampling=AgeS$Sampling,SampleNames=c("GDB5","GDB3"),SelectedSamples=c(1,1))
```

SCMatrix

Definition of the stratigraphic constraint matrix

Description

This function helps to define the stratigraphic relation between samples, with questions. The output of this function can be used in function `AgeS_Computation`.

Usage

```
SCMatrix(Nb_sample, SampleNames)
```

Arguments

`Nb_sample` interger: the sample number.
`SampleNames` charcater vector: sample names.

Details

Ask if sample *i* is younger than sample *j* to construc the stratigraphic constrain matrix.

Value

A Matrix that summarise the ordered relation between samples. This matrix can be intergrate in `AgeS_Computation` function. We refer to detail on `AgeS_Computation` for more information concerning this matrix.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

See Also

`AgeS_Computation`

Examples

```
## Assume that "sample1" is younger than "sample2"
## That means the expected value is 1.
## It is an interactive function.
## Not run:
SCMatrix(Nb_sample=2,SampleNames=c("sample1","sample2"))
## Enter the value 1

## End(Not run)
```

SC_Ordered

*Stratigraphic matrix for samples ordered***Description**

Construc the stratigraphic matrix used in fonction AgeS_Computation and AgeC14_Computation for samples that are all ordered by increasing oder.

Usage

```
SC_Ordered(Nb_sample)
```

Arguments

Nb_sample interger: the sample number.

Value

Stratigraphic matrix where each sample are ordered by incresing order This matrix can be intergrate in AgeS_Computation function. We refer to detail on AgeS_Computation for more information concerning this matrix.

Author(s)

Claire Christophe, Anne Philippe, Guillaume Guerin

See Also

AgeS_Computation, SC_Matrix

Examples

```
# compute the stratigraphic matrix for 3 samples such that the first sample is younger
# than the second, and the second is younger than the third
(SC=SC_Ordered(Nb_sample=3))
```

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