

# Package ‘BayLum’

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**Type** Package

**Title** Chronological Bayesian Models Integrating Optically Stimulated  
Luminescence Dating

**Description** collection of various R functions for Bayesian analysis of Luminescence data.  
This includes, amongst others, data import, export,  
application of age models and palaeodose model.

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BayLum-package	<i>Chronological Bayesian Models Integrating Optically Stimulated Luminescence Dating</i>
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## Description

A collection of various R functions for Bayesian analysis of Luminescence data. This includes, amongst others, data import, export, application of age models and palaeodose model.

## Details

This package is based on the functions [Generate\\_DataFile](#), [Generate\\_DataFile\\_MG](#), [Age\\_Computation](#) and [AgeS\\_Computation](#). The two first functions create a list containing all informations needed to analyse each Bin file in order to compute its age, of Single-grain OSL measurements for the first function and Multi-grain OSL measurements for the second. The another two functions use Bayesian analysis for OSL age estimation for one or various samples, according differents models that are available in options (different growth curves and different equivalent dose distributions around the palaeodose).

It is possible to consider various Bin files per sample, and to compute ages of samples in stratigraphic constraints and to integrate systematic errors.

## Author(s)

Claire Christophe, Guillaume Guerin, Sebastian Kreutzer, Anne Philippe

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AgeC14_Computation	<i>Bayesian analysis for age estimation of 14C Cal age of various samples</i>
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## Description

This function calibrate 14C Cal age of samples to get an age (years). The user can choose one of the following radiocarbon calibration curve: Northern or Sourthen Hemisphere or marine atmospheric.

**Usage**

```
AgeC14_Computation(Data_C14Cal, Data_SigmaC14Cal, Nb_sample, SampleNames,
  SavePdf = FALSE, OutputFileName = c("MCMCplot", "HPD_Cal14CCurve",
  "summary", "PosteriorDensity", "2per2Laws"), OutputFilePath = c(""),
  SaveEstimates = FALSE, OutputTableName = c("DATA"),
  OutputTablePath = c(""), PriorAge = rep(c(10000, 50000), Nb_sample),
  StratiConstraints = c(), sepSC = c(", "), Model = c("full"),
  CalibrationCurve = c("AtmosphericNorth"), Taille = 50000, t = 5,
  Nb_chaines = 3)
```

**Arguments**

Data_C14Cal	numeric vector: corresponding to 14C Cal age estimate.
Data_SigmaC14Cal	numeric vector: corresponding to the error of 14C Cal age estimates.
Nb_sample	integer: number of samples, Nb_sampe>1.
SampleNames	character vector: names of sample. The length of this vector is equal to Nb_sample.
SavePdf	boolean (with default): if TRUE save graphs in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE.
SaveEstimates	boolean (with default): if TRUE save Bayes estimates, credible interval at level 68% and 95% and , in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE.
PriorAge	numeric vector (with default): lower and upper bounds for age parameter of each sample. length(PriorAge)=2*Nb_sample and PriorAge[2i-1,2i] corresponds to the lower and upper bounds of sample whose number ID is equal to i.
StratiConstraints	numeric matrix or character(with default): input object for the stratigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints. Otherwise, default value is suitable.
sepSC	character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints .csv file.
Model	character (with default): if " <b>full</b> ", error on estimate calibration curve is taken account. If " <b>naive</b> " this error is not taken account in the age estimate.
CalibrationCurve	character (with default): calibration curve choosen. Allowed inputs are <ul style="list-style-type: none"> <li>• "<b>AtmosphericNorth</b>" for Northern Hemisphere atmospheric radiocarbon calibration curve,</li> </ul>

	<ul style="list-style-type: none"> <li>• <b>"Marine"</b> for Marine radiocarbon calibration curve,</li> <li>• <b>"AtmosphericSouth"</b> for Southern Hemisphere atmospheric radiocarbon calibration curve</li> </ul>
Taille	integer (with default): number of iterations for the MCMC computation (for more information see <a href="#">jags.model</a> ).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see <a href="#">jags.model</a> ).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see <a href="#">jags.model</a> ).

## Details

### \*\* How to fill StratiConstraints ? \*\*

If there is stratigraphic relations between samples, **14C Cal estimate age in Data\_C14Cal must be ordered by order of increasing ages.**

The user can fill the StratiConstraints matrix as follow.

1. **Size of the matrix:** row number of StratiConstraints matrix is equal to Nb\_sample+1, and column number is equal to Nb\_sample.
2. **First line of the matrix:** for all  $i$  in  $\{1, \dots, Nb\_Sample\}$ ,  $StratiConstraints[1, i] = 1$  that means the lower bound of the sample age (given in  $PriorAge[2i-1]$ ) for the sample whose number ID is equal to  $i$ , is taken into account.
3. **Sample relations:** for all  $j$  in  $\{2, \dots, Nb\_Sample+1\}$  and all  $i$  in  $\{j, \dots, Nb\_Sample\}$ ,  $StratiConstraints[j, i] = 1$  if sample age whose number ID is equal to  $j-1$  is lower than sample age whose number ID is equal to  $i$ . Otherwise,  $StratiConstraints[j, i] = 0$ .

Note that  $StratiConstraints_{\{2:Nb\_sample+1, 1:Nb\_sample\}}$  is a upper triangular matrix.

The user can also use [SCMatrix](#) function to construc the StratiConstraints matrix.

The user can also refer to a .csv file that contains the relation between samples as defined above.

## Value

### NUMERICAL OUTPUT

1. **A list containing the following objects:**
  - **Sampling:** that corresponds to a sample of the posterior distributions of the age parameters.
  - **Model,** stating which model was chosen ("full" or "naive");
  - **CalibrationCurve,** stating which radiocarbon calibration curve was chosen;
  - **PriorAge,** stating the priors used for the age parameter;
  - **StratiConstraints,** stating the stratigraphic relations between samples considered in the model.
2. **The Gelman and Rudin test of convergency:** prints the result of the Gelman and Rudin test of convergency for the age estimate for each sample. A result close to one is expected. In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (Taille) or be more precise on the PriorAge parameter to reach convergence.

3. **Credible intervals and Bayes estimates:** prints the Bayes estimates, the credible intervals at 95% and 68% for the age parameters for each sample.

## PLOT OUTPUT

1. **MCMC trajectories:** A graph with the MCMC trajectories and posterior distributions of the age parameter is displayed.  
On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.
2. **Age estimate and HPD on calibration curve:** plot age estimate and HPD on calibration plot to check.....
3. **Summary of sample age estimates:** plot credible intervals and Bayes estimate of each sample age on a same graph.
4. **Posterior density:** plot posterior density of each age estimate.
5. **Matrix of scatterplots:** the i,j th scatterplot contains sample of the posterior distribution of the sample ID i plotted against the one of the sample ID j.

## Author(s)

Claire Christophe, Guillaume Guerin

## References

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronk Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Hafflidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4):1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

## See Also

[rjags](#), [MCMC\\_plot](#), [SCMatrix](#)

## Examples

```
## Load data
## Age computation of samples...
# Age=AgeC14_Computation(Data_C14Cal=Data_C14Cal,Data_SigmaC14Cal=Data_SigmaC14Cal,
#   Nb_sample=Nb_sample,SampleNames=SampleNames,PriorAge=PriorAge,Taille=1000)
```

## Description

This function computes the age of at least two samples according to the model developed in Combes and Philippe (2017), based on outputs of [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#) or both of them using [Concat\\_DataFile](#).

Samples, for which data is available in several BIN files, can be analysed.

Single-grain or Multi-grain OSL measurements can be analysed simultaneously.

## Usage

```
AgeS_Computation(DATA, SampleNames, Nb_sample, BinPerSample = rep(1,
  Nb_sample), SavePdf = FALSE, OutputFileName = c("MCMCplot", "summary",
  "2per2Laws"), OutputFilePath = c(""), SaveEstimates = FALSE,
  OutputTableName = c("DATA"), OutputTablePath = c(""), THETA = c(),
  sepTHETA = c(", "), PriorAge = rep(c(0.01, 100), Nb_sample),
  StratiConstraints = c(), sepSC = c(", "), LIN_fit = TRUE,
  Origin_fit = FALSE, distribution = c("cauchy"), Taille = 50000, t = 5,
  Nb_chaines = 3)
```

## Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by the function <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> or <a href="#">Concat_DataFile</a> . DATA contains information for more than one sample.
SampleNames	character vector: names of sample. The length of this vector is equal to Nb_sample.
Nb_sample	integer : number of samples, Nb_sampe>1.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector is equal to Nb_sample. BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in <a href="#">Generate_DataFile</a> or in <a href="#">Generate_DataFile_MG</a> .
SavePdf	boolean (with default): if TRUE save graphs in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE.
SaveEstimates	boolean (with default): if TRUE save Bayes estimates, credible interval at level 68% and 95% and , in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE.

THETA	numeric matrix or character (with default): input object for systematic and individual error. If systematic errors are considered, see the details section for instructions regarding how to correctly fill THETA. Otherwise, default value is suitable, and only individual error is considered.
sepTHETA	character (with default): if THETA is character, indicate column separator in THETA .csv file.
PriorAge	numeric vector (with default): lower and upper bounds for age parameter of each sample. $\text{length}(\text{PriorAge})=2 \times \text{Nb\_sample}$ and $\text{PriorAge}[2i-1, 2i]$ corresponds to the lower and upper bounds of sample whose number ID is equal to $i$ .
StratiConstraints	numeric matrix or character (with default): input object for the stratigraphic relation between samples. If there is stratigraphic relation between samples see the details section for instructions regarding how to correctly fill StratiConstraints. Otherwise, default value is suitable.
sepSC	character (with default): if StratiConstraints is character, indicate column separator in StratiConstraints .csv file.
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. Please see details for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. Please see details for more informations on the proposed growth curves.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M".
Taille	integer (with default): number of iterations for the MCMC computation (for more information see <a href="#">jags.model</a> ).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see <a href="#">jags.model</a> ).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see <a href="#">jags.model</a> ).

## Details

### \*\* How to fill StratiConstraints ? \*\*

If there is stratigraphic relations between samples, **information in DATA must be ordered by order of increasing ages**. To do this the user can either fill right Names in [Generate\\_DataFile](#) or in [Generate\\_DataFile\\_MG](#) (as it is indicated in Details section of these function), or ordered by order of increasing ages outputs of [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#) in [Concat\\_DataFile](#). The user can fill the StratiConstraints matrix as follow.

1. **Size of the matrix:** row number of StratiConstraints matrix is equal to  $\text{Nb\_sample}+1$ , and column number is equal to  $\text{Nb\_sample}$ .
2. **First line of the matrix:** for all  $i$  in  $\{1, \dots, \text{Nb\_Sample}\}$ ,  $\text{StratiConstraints}[1, i]=1$  that means the lower bound of the sample age (given in  $\text{PriorAge}[2i-1]$ ) for the sample whose number ID is equal to  $i$ , is taken into account.

3. **Sample relations:** for all  $j$  in  $\{2, \dots, \text{Nb\_Sample}+1\}$  and all  $i$  in  $\{j, \dots, \text{Nb\_Sample}\}$ ,  $\text{StratiConstraints}[j, i] = 1$  if sample age whose number ID is equal to  $j-1$  is lower than sample age whose number ID is equal to  $i$ . Otherwise,  $\text{StratiConstraints}[j, i] = 0$ .

Note that  $\text{StratiConstraints}_{\{2:\text{Nb\_sample}+A, 1:\text{Nb\_sample}\}}$  is a upper triangular matrix.

The user can also use [SCMatrix](#) function to construc the StratiConstraints matrix.

The user can also refer to a .csv file that contains the relation between samples as defined above.

**\*\* How to fill THETA covariance matrix concerning common and individual error? \*\***

If systematic errors are considered, the user can fill the THETA matrix as follow.

- row number of THETA is equal the column number, equal to  $\text{Nb\_sample}$ .
- For all  $i$  in  $\{1, \dots, \text{Nb\_sample}\}$ ,  $\text{THETA}[i, i]$  contains individual error plus systematic error of the sample whose number ID is equal to  $i$ .
- For all  $i, j$  in  $\{1, \dots, \text{Nb\_sample}\}$  and  $i$  different from  $j$ ,  $\text{THETA}[i, j]$  contains common error between samples whose number ID are equal to  $i$  and  $j$ .

Note that  $\text{THETA}[i, j]$  is a symetric matrix.

The user can also refer to a .csv file that contains the errors as defined above.

**\*\* Option on growth curves \*\***

As for [AgeComputation](#) and [PalaeodoseComputation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** ( $\text{AgesMultiCS2\_EXPLIN}$ ):  
for all  $x$  in  $\text{IR}^+$ ,  $f(x) = a(1 - \exp(-x/b)) + cx + d$ ; select
  - $\text{LIN\_fit} = \text{TRUE}$
  - $\text{Origin\_fit} = \text{FALSE}$
- **Saturating exponential growth** ( $\text{AgesMultiCS2\_EXP}$ ):  
for all  $x$  in  $\text{IR}^+$ ,  $f(x) = a(1 - \exp(-x/b)) + d$ ; select
  - $\text{LIN\_fit} = \text{FALSE}$
  - $\text{Origin\_fit} = \text{FALSE}$
- **Saturating exponential plus linear growth and fitting through the origin** ( $\text{AgesMultiCS2\_EXPLINZ0}$ ):  
for all  $x$  in  $\text{IR}^+$ ,  $f(x) = a(1 - \exp(-x/b)) + cx$ ; select
  - $\text{LIN\_fit} = \text{TRUE}$
  - $\text{Origin\_fit} = \text{TRUE}$
- **Saturating exponential growth and fitting through the origin** ( $\text{AgesMultiCS2\_EXPZ0}$ ):  
for all  $x$  in  $\text{IR}^+$ ,  $f(x) = a(1 - \exp(-x/b))$ ; select
  - $\text{LIN\_fit} = \text{FALSE}$
  - $\text{Origin\_fit} = \text{TRUE}$

**\*\* Option on equivalent dose distribution around the palaeodose \*\***

The use can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample
- **lognormal\_A**: a log-normal distribution with mean or Average equal to the palaeodose of the sample
- **lognormal\_M**: a log-normal distribution with Median equal to the palaeodose of the sample



**Value****NUMERICAL OUTPUT****1. A list containing the following objects:**

- **Sampling:** that corresponds to a sample of the posterior distributions of the Age, palaeodose and equivalent dose dispersion parameters.
- **Model\_GrowthCurve**, stating which dose response fitting option was chosen;
- **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
- **PriorAge**, stating the priors used for the age parameter;
- **StratiConstraints**, stating the stratigraphic relations between samples considered in the model;
- **CovarianceMatrix**, stating the covariance matrix of error used in the model, highlighting common errors between samples or not.

**2. The Gelman and Rudin test of convergency:** prints the result of the Gelman and Rudin test of convergency for the age, palaeodose and equivalent dose dispersion parameters for each sample. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (*Taille*) or be more precise on the *PriorAge* parameter (for example specify if it is a young sample  $c(0.01, 10)$  or an old sample  $c(10, 100)$ ) to reach convergency.

**3. Credible intervals and Bayes estimates:** prints the Bayes estimates, the credible intervals at 95% and 68% for the age, palaeodose and equivalent dose dispersion parameters for each sample.**PLOT OUTPUT****1. MCMC trajectories:** A graph with the MCMC trajectories and posterior distributions of the age, palaeodose and equivalent dose dispersion parameters is displayed, there is one page per sample.

The first line of the figure corresponds to the age parameter, the second to the palaeodose parameter and the third to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

**2. Summary of sample age estimates:** plot credible intervals and Bayes estimate of each sample age on a same graph.**3. Matrix of scatterplots:** the  $i, j$  th scatterplot contains sample of the posterior distribution of the sample ID  $i$  plotted against the one of the sample ID  $j$ .**Author(s)**

Claire Christophe, Guillaume Guerin

**References**

Combes, Benoit and Philippe, Anne, 2017. Bayesian analysis of multiplicative Gaussian error for multiple ages estimation in optically stimulated luminescence dating. *Quaternary Geochronology* (39, 24-34)

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. Quaternary Geochronology 28, 62-70. doi:10.1016/j.quageo.2015.04.001

### See Also

[Generate\\_DataFile](#), [Generate\\_DataFile\\_MG](#), [rjags](#), [MCMC\\_plot](#), [SCMatrix](#) [Age\\_Computation](#), [Palaeodose\\_Computation](#)

### Examples

```
## Load data
# data(DATA1,envir = environment())
# data(DATA2,envir = environment())
# DATA=Concat_DataFile(DATA2,DATA1)

## Age computation of samples GDB5 and GDB3,
## without common error and without stratigraphic constraints
# Age=AgeS_Computation(DATA,Nb_sample=2,SampleNames=c("GDB5","GDB3"),Taille=10000)

## Age computation of samples GDB5 and GDB3,
## without common error, assuming GDB5 age younger than GDB3 age
# (SC=matrix(data=c(1,1,0,1,0,0),ncol=Nb_sample,nrow = (Nb_sample+1),byrow = T))
# Age=AgeS_Computation(DATA,Nb_sample=2,SampleNames=c("GDB5","GDB3"),
#   StratiConstraints=SC,Taille=10000)
```

---

Age\_Computation

*Bayesian analysis for the OSL age estimation of one sample*

---

### Description

This function computes the age of a sample according to the model developed in Combes and Philippe (2017), based on an output of [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#). A sample, for which data is available in several BIN files, can be analysed.

### Usage

```
Age_Computation(DATA, samplename, SavePdf = FALSE,
  OutputFileName = c("MCMCplot"), OutputFilePath = c(""),
  SaveEstimates = FALSE, OutputTableName = c("DATA"),
  OutputTablePath = c(""), BinPerSample = c(1), PriorAge = c(0.01, 100),
  LIN_fit = TRUE, Origin_fit = FALSE, distribution = c("cauchy"), I = 1,
  Taille = 50000, t = 5, Nb_chaines = 3)
```

### Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by the function <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> . DATA can contain information for more than one sample.
samplename	character: name of the sample.
SavePdf	boolean (with default): if TRUE save graph in pdf file named OutputFileName in folder OutputFilePath.

OutputFileName	character (with default): name of the pdf file that will be generated by the function if SavePdf=TRUE.
OutputFilePath	character (with default): path to the pdf file that will be generated by the function if SavePdf=TRUE.
SaveEstimates	boolean (with default): if TRUE save Bayes estimates and credible interval at level 68% and 95%, in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. If in DATA there is more than one sample, the BinPerSample vector must be the same as that used to run the function <a href="#">Generate_DataFile</a> or in <a href="#">Generate_DataFile_MG</a> for generating the DATA object.
PriorAge	numeric vector (with default): lower and upper bounds for the sample age parameter. $\text{length}(\text{PriorAge})=2$ .
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. Please see details for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. Please see details for more informations on the proposed growth curves.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M".
I	integer (with default): if DATA contains data from more than one sample, I indicates the ID number of the sample to be analysed.
Taille	integer (with default): number of iterations for the MCMC computation (for more information see <a href="#">jags.model</a> ).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see <a href="#">jags.model</a> ).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see <a href="#">jags.model</a> ).

## Details

### \*\* Option on growth curves \*\*

As for [AgeS\\_Computation](#) and [Palaeodose\\_Computation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** (AgeMultiBF\_EXPLIN):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))+cx+d$ ; select
  - LIN\_fit=TRUE
  - Origin\_fit=FALSE
- **Saturating exponential growth** (AgeMultiBF\_EXP):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))+d$ ; select

- LIN\_fit=FALSE
- Origin\_fit=FALSE
- **Saturating exponential plus linear growth and fitting through the origin** (AgeMultiBF\_EXPLINZO):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))+cx$ ; select
  - LIN\_fit=TRUE
  - Origin\_fit=TRUE
- **Saturating exponential growth and fitting through the origin** (AgeMultiBF\_EXPZO):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))$ ; select
  - LIN\_fit=FALSE
  - Origin\_fit=TRUE

**\*\* Option on equivalent dose distribution around the palaeodose \*\***

The use can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample
- **lognormal\_A**: a log-normal distribution with mean or **A**verage equal to the palaeodose of the sample
- **lognormal\_M**: a log-normal distribution with **M**edian equal to the palaeodose of the sample

**Value**

**NUMERICAL OUTPUT**

1. **A list containing the following objects:**

- **Sampling** that corresponds to a sample of the posterior distributions of the Age, palaeodose and equivalent dose dispersion parameters.
- **Model\_GrowthCurve**, stating which dose response fitting option was chosen;
- **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
- **PriorAge**, stating the priors used for the age parameter.

2. **The Gelman and Rudin test of convergency**: prints the result of the Gelman and Rudin test of convergency for the age, palaeodose and equivalent dose dispersion parameters. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the graph generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the printed estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (*Taille*) to reach convergency.

3. **Credible intervals and Bayes estimates**: prints the Bayes estimates, the credible intervals at 95% and 68% for the age, palaeodose and equivalent dose dispersion parameters of the sample.

**PLOT OUTPUT**

A graph with the MCMC trajectories and posterior distributions of the age, palaeodose and equivalent dose dispersion parameters is displayed.

The first line of the figure corresponds to the age parameter, the second to the palaeodose parameter and the third to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

**Author(s)**

Claire Christophe, Guillaume Guerin

**References**

Combes, Benoit and Philippe, Anne, 2017. Bayesian analysis of multiplicative Gaussian error for multiple ages estimation in optically stimulated luminescence dating. *Quaternary Geochronology* (39, 24-34)

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. *Quaternary Geochronology* 28, 62-70. doi:10.1016/j.quageo.2015.04.001

**See Also**

[Generate\\_DataFile](#), [Generate\\_DataFile\\_MG](#), [rjags](#), [MCMC\\_plot AgeS\\_Computation](#), [Palaeodose\\_Computation](#)

**Examples**

```
## load data file generated by the function Generate_DataFile
# data(DATA1,envir = environment())
# Age=Age_Computation(DATA1,samplename="GDB3",Taille=10000)
```

---

Concat_DataFile	Concatenates two outputs of <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a>
-----------------	---

---

**Description**

This function allows concatenating two lists provided as output of the [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#) function.

Only concatenation of 2 files is possible.

**Usage**

```
Concat_DataFile(u1, u2)
```

**Arguments**

u1	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement.
u2	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement.

**Details**

For more information on the structure of the input list, we refer to **Value** section of [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#).

This function is especially usefull in two cases:

- information concerning samples are yet saved in .RData file (that allow to not run again [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#) that can take time);
- the user want to analyse simultaneously Single-grain and Multi-grain OSL measurements, because sample are in under stratigraphic constraints or they share systematic errors.

**Value**

A List of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J, K, Nb\_measurement.

**Author(s)**

Claire Christophe, Guillaume Guerin

**See Also**

[Generate\\_DataFile](#), [Generate\\_DataFile\\_MG](#)

**Examples**

```
# load data files
data(DATA1,envir = environment())
data(DATA2,envir = environment())
# concatenate two data files
DATA3=Concat_DataFile(DATA1,DATA2)
str(DATA3)
```

---

DATA1

---

*DATA of sample named GDB3*


---

**Description**

list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J,K,Nb\_measurement obtained using Generate\_DataFile function from luminescence data.

**Usage**

```
data("DATA1")
```

**Format**

A list containing:

LT (one list per sample): each list contains all L/T values for the corresponding sample;

sLT (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot\_env a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose (one list per sample): each list contains all regenerated doses;

J a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb\_measurement a vector giving, for each BIN file, the number of measurements;

## References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D., Pearson, O., Puaud, S., Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

## Examples

```
data(DATA1)
str(DATA1)
```

---

DATA2

*DATA on sample named GDB5*

---

## Description

list of objects: LT, sLT, ITimes, dLab, ddot\_env, regDose, J,K,Nb\_measurement obtained using Generate\_DataFile function from luminescence data.

## Usage

```
data("DATA2")
```

## Format

A list containing:

LT (one list per sample): each list contains all L/T values for the corresponding sample;

sLT (one list per sample): each list contains all uncertainties on L/T values for the corresponding sample;

ITimes (one list per sample): each list contains irradiation time values for the corresponding sample;

dLab a matrix containing in line i, the laboratory dose rate and its variance for sample i;

ddot\_env a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

regDose (one list per sample): each list contains all regenerated doses;

J a vector giving, for each BIN file, the number of aliquots selected for the analysis;

K a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;

Nb\_measurement , a vector giving, for each BIN file, the number of measurements;

## References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D., Pearson, O., Puaud, S., Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. PloS one, 12(1), e0169418.

## Examples

```
data(DATA2)
str(DATA2)
```

---

Generate_DataFile	<i>Generates, from one (or several) BIN file(s) of Single-grain OSL measurements, a list of luminescence data and information before statistical analysis</i>
-------------------	---

---

## Description

This function is used to generate, from the BIN file(s), a list of values of: **Single-grain** OSL intensities and associated uncertainties, regenerative doses, etc., which will be the input of the Bayesian models. To be easy-to-use, this function requires a rigorous organisation - all needed files should be arranged in one folder - of informations concerning each BIN file.

It is possible to process data for various samples simultaneously and to consider more than one BIN file per sample.

## Usage

```
Generate_DataFile(Path, Names, Nb_sample, Nb_binfile = length(Names),
  BinPerSample = rep(1, Nb_sample), sepDP = c(","), sepDE = c(","),
  sepDS = c(","), sepR = c("="))
```

## Arguments

Path	character: the path to the project folder, containing one or more subfolders in which the BIN files are located.
Names	character vector: list of names of the sub-folders containing the BIN files - each subfolder must contain a BIN file and associated .csv files. See details for more informations on associated .csv files required in the subfolders. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill the Names vector.
Nb_sample	integer: number of samples.
Nb_binfile	integer (with default): number of BIN files. It must be equal to, or greater than Nb_sample.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector must be equal to Nb_sample and the sum of entries of this vector must be equal to Nb_binfile. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill BinPerSample vector. Otherwise, this vector must contain a list of 1 values.
sepDP	character (with default): column separator in the DiscPose.csv files.
sepDE	character (with default): column separator in the DoseEnv.csv files.
sepDS	character (with default): column separator in the DoseLab.csv files.
sepR	character (with default): column separator in the Rule.csv files.

## Details

With Path and Names, this function goes to the subfolders containing the BIN files and associated information to compute the luminescence data.

**\*\* What are the required files in each subfolder? \*\***

Each subfolder can be named, for example, as the sample name followed by a number; it must contain:



- **bin.BIN**, the bin file renamed as bin.BIN (note: the name of all files matters);
- **DiscPos.csv**, a two columns .csv file containing the list of disc and grain position number of the previously selected grains (typically this list will include the position of grains based on their sensitivity, recycling or other properties);
- **DoseEnv.csv**, a two columns file containing the observation of the natural (or environmental), dose rate, and its non-shared variance (i.e. after removing all shared errors). Note: the user shall provide the squared value of the error associated with the dose rate experienced by the sample grains in nature;
- **DoseSource.csv**, a two columns file containing the observation of the laboratory dose rate, and its variance (squared error);
- **rule.csv**, a .csv file containing information on
  - beginSignal= the first channel for summing the natural or regenerative OSL signal (typically 1 or 6);
  - endSignal= the last channel for summing the natural or regenerative OSL signal (typically 5 or 10);
  - beginBackground= the first channel for background estimation of the natural or regenerative OSL signal (typically 76 or 81);
  - endBackground= the last channel for background estimation of the natural or regenerative OSL signal (typically 95 or 100);
  - beginTest,
  - endTest,
  - beginTestBackground,
  - endTestBackground= same values as above, for the test dose response (typically the same values should be used);
  - inflatePercent= uncertainty arising from the instrument reproducibility (typically 0.02, i.e. 2%);
  - nbOfLastCycleToRemove= number of cycles at the end of the SAR protocol which should not be included in the dose response curve fitting (typically 1 if only a recycling test is performed, or 2 if both recycling and IR depletion are tested).

#### **\*\* How to fill the Names vector? \*\***

Names is a vector of length Nb\_binfile. Names[i] is the name (e.g., Sample1-File1, or successive names separated by "/" signs, if BIN files are in subfolders, e.g. Sample1/File1) of the subfolder containing all informations on the BIN file of ID number i. The names in Names are ordered following two rules:

- The names in the Names vector must be ordered following the sample order (the names of subfolders containing BIN files for the same sample should follow each other in the Names vector, e.g. Sample1, Sample2-File1, Sample2-File2, etc.).
- If stratigraphic constraints apply to samples, and so a **Bayesian model with stratigraphic constraints** is implemented, then the names in the Names vector must be ordered by order of increasing ages.  
For example, Names=c(noun1 , noun2), in which case noun1 (respectively, noun2) corresponds to the subfolder name containing the BIN file of sample 1 (respectively of sample 2). In addition, if we know that sample 1 is younger than sample 2, then Names vector is correctly filled. If conversely, Names=c(noun2, noun1), the analysis performed by [AgeS\\_Computation](#) would not be consistent.

**\*\* How to fill the BinPerSample vector? \*\***

BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i.

For example, let us consider a case with two samples (Sample1 and Sample2), with 2 BIN files for Sample1 and 1 for Sample2. In this case, Nb\_binfile=3 and Nb\_sample=2. The user may then set Names=c("Sample1-File1", "Sample1-File2", "Sample2-File1"), in which case "Sample1-1" is the name of the subfolder containing the first BIN file for Sample1, "Sample1-File2" the name of the subfolder for the second BIN file of Sample1; eventually, "Sample2-1" is the name of the subfolder containing the BIN file for the second sample. In this case, BinPerSample=c(2, 1).

For the general BIN-file structure, the reader is referred to the following website: <http://www.nutech.dtu.dk/>

The function `read_BIN2R` developed in `Luminescence` package is used to read the BIN files.

## Value

A list containing the following objects:

- **LT** (one list per sample); each list contains all L/T values for the corresponding sample;
- **sLT** (one list per sample); each list contains all uncertainties on L/T values for the corresponding sample;
- **ITimes** (one list per sample); each list contains irradiation time values for the corresponding sample;
- **dLab**, a matrix containing in line i, the laboratory dose rate and its variance for sample i;
- **ddot\_env**, a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;
- **regDose** (one list per sample); each list contains all regenerated doses;
- **J**, a vector giving, for each BIN file, the number of aliquots selected for the analysis;
- **K**, a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;
- **Nb\_measurement**, a vector giving, for each BIN file, the number of measurements;

**\*\* How to save this list \*\***

You can save this list in a .RData object. To do this, you can use the fonction `save`. Then, to load this list you can use the function `load` (see example section fore more details).

## Author(s)

Claire Christophe, IRAMAT-CRP2A, Universite Bordeaux Montaigne (France), Guillaume Guerin, IRAMAT-CRP2A, Universite Bordeaux Montaigne (France), Sebastian Kreutzer, IRAMAT-CRP2A, Universite Bordeaux Montaigne (France)

## See Also

[read\\_BIN2R](#), [Concat\\_DataFile](#), [Generate\\_DataFile\\_MG](#), [LT\\_RegenDose](#), [Age\\_Computation](#), [AgeS\\_Computation](#), [Palaeodose\\_Computation](#)

## Examples

```
## 1) Example for one sample with one Bin File
## Put path in quotes to go to the folder containing bin.BIN file and associated .csv files
## Path must be determinated by "/"
# Path=""
## Enter in quotes the name of the folder containings bin.BIN file
```

```

# Names=""
## give the number of sample
# Nb_sample=1
# DATA=Generate_DataFile(Path,Names,Nb_sample)
# str(DATA)

## to save information in .RData object
# save(DATA,file=c(paste(Path,'DATA.RData',sep="")))
## to load information containing DATA.RData object
# load(file=c(paste(Path,"DATA.RData",sep="")))

## 2) Example for 2 samples and one Bin file each
## Put path in quotes to go to the folders containing bin.BIN file for each sample.
## Path must be determined by "/"
# Path=""
## Enter between the "" the names of each folders containsing bin.BIN file
# Names=c("Nom1","Nom2")
## give the number of sample
# Nb_sample=2
# DATA=Generate_DataFile(Path,Names,Nb_sample)
# str(DATA)

## to save information in .RData object
# save(DATA,file=c(paste(Path,'DATA.RData',sep="")))
## to load information containing DATA.RData object
# load(file=c(paste(Path,"DATA.RData",sep="")))

```

---

Generate_DataFile_MG	<i>Generates, from one (or several) BIN file(s) of Multi-grain OSL measurements, a list of luminescence data and information before statistical analysis</i>
----------------------	--

---

## Description

This function is used to generate, from the BIN file(s), a list of values of: **Multi-grain** OSL intensities and associated uncertainties, regenerative doses, etc., which will be the input of the Bayesian models. To be easy-to-use, this function requires a rigorous organisation - all needed files should be arranged in one folder - of informations concerning each BIN file.

It is possible to process data for various samples simultaneously and to consider more than one BIN file per sample.

## Usage

```

Generate_DataFile_MG(Path, Names, Nb_sample, Nb_binfile = length(Names),
  BinPerSample = rep(1, Nb_sample), sepD = c(", "), sepDE = c(", "),
  sepDS = c(", "), sepR = c("="))

```

## Arguments

Path	character: the path to the project folder, containing one or more subfolders in which the BIN files are located.
------	--

Names	character vector: list of names of the sub-folders containing the BIN files - each subfolder must contain a BIN file and associated .csv files. See details for more informations on associated .csv files required in the subfolders. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill the Names vector.
Nb_sample	integer: number of samples.
Nb_binfile	integer (with default): number of BIN files. It must be equal to, or greater than Nb_sample.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector must be equal to Nb_sample and the sum of entries of this vector must be equal to Nb_binfile. If there is more than one BIN file per sample, see the details section for instructions regarding how to correctly fill BinPerSample vector. Otherwise, this vector must contain a list of 1 values.
sepD	character (with default): column separator in the DiscPose.csv files.
sepDE	character (with default): column separator in the DoseEnv.csv files.
sepDS	character (with default): column separator in the DoseLab.csv files.
sepR	character (with default): column separator in the Rule.csv files.

## Details

With Path and Names, this function goes to the subfolders containing the BIN files and associated information to compute the luminescence data.

### **\*\* What are the required files in each subfolder? \*\***

Each subfolder can be named, for example, as the sample name followed by a number; it must contain:

- **bin.BIN**, the bin file renamed as bin.BIN (note: the name of all files matters);
- **Disc.csv**, a one columns .csv file containing the list of disc number of the previously selected grains (typically this list will include the position of grains based on their sensitivity, recycling or other properties);
- **DoseEnv.csv**, a two columns file containing the observation of the natural (or environmental), dose rate, and its non-shared variance (i.e. after removing all shared errors). Note: the user shall provide the squared value of the error associated with the dose rate experienced by the sample grains in nature;
- **DoseSource.csv**, a two columns file containing the observation of the laboratory dose rate, and its variance (squared error);
- **rule.csv**, a .csv file containing information on
  - beginSignal= the first channel for summing the natural or regenerative OSL signal (typically 1 or 6);
  - endSignal= the last channel for summing the natural or regenerative OSL signal (typically 5 or 10);
  - beginBackground= the first channel for background estimation of the natural or regenerative OSL signal (typically 76 or 81);
  - endBackground= the last channel for background estimation of the natural or regenerative OSL signal (typically 95 or 100);
  - beginTest,
  - endTest,
  - beginTestBackground,

- endTestBackground= same values as above, for the test dose response (typically the same values should be used);
- inflatePercent= uncertainty arising from the instrument reproducibility (typically 0.02, i.e. 2%);
- nbOfLastCycleToRemove= number of cycles at the end of the SAR protocol which should not be included in the dose response curve fitting (typically 1 if only a recycling test is performed, or 2 if both recycling and IR depletion are tested).

**\*\* How to fill the Names vector? \*\***

Names is a vector of length Nb\_binfile. Names[i] is the name (e.g., Sample1-File1, or successive names separated by "/" signs, if BIN files are in subfolders, e.g. Sample1/File1) of the subfolder containing all informations on the BIN file of ID number i. The names in Names are ordered following two rules:

- The names in the Names vector must be ordered following the sample order (the names of subfolders containing BIN files for the same sample should follow each other in the Names vector, e.g. Sample1, Sample2-File1, Sample2-File2, etc.).
- If stratigraphic constraints apply to samples, and so a **Bayesian model with stratigraphic constraints** is implemented, then the names in the Names vector must be ordered by order of increasing ages.

For example, Names=c(noun1, noun2), in which case noun1 (respectively, noun2) corresponds to the subfolder name containing the BIN file of sample 1 (respectively of sample 2). In addition, if we know that sample 1 is younger than sample 2, then Names vector is correctly filled. If conversely, Names=c(noun2, noun1), the analysis performed by [AgeS\\_Computation](#) would not be consistent.

**\*\* How to fill the BinPerSample vector? \*\***

BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i.

For example, let us consider a case with two samples (Sample1 and Sample2), with 2 BIN files for Sample1 and 1 for Sample2. In this case, Nb\_binfile=3 and Nb\_sample=2. The user may then set Names=c("Sample1-File1", "Sample1-File2", "Sample2-File1"), in which case "Sample1-1" is the name of the subfolder containing the first BIN file for Sample1, "Sample1-File2" the name of the subfolder for the second BIN file of Sample1; eventually, "Sample2-1" is the name of the subfolder containing the BIN file for the second sample. In this case, BinPerSample=c(2, 1).

For the general BIN-file structure, the reader is referred to the following website: <http://www.nutech.dtu.dk/>

The function [read\\_BIN2R](#) developed in [Luminescence](#) package is used to read the BIN files.

## Value

A list containing the following objects:

- **LT** (one list per sample); each list contains all L/T values for the corresponding sample;
- **sLT** (one list per sample); each list contains all uncertainties on L/T values for the corresponding sample;
- **ITimes** (one list per sample); each list contains irradiation time values for the corresponding sample;
- **dLab**, a matrix containing in line i, the laboratory dose rate and its variance for sample i;
- **ddot\_env**, a matrix containing in line i, the environmental dose rate and its variance (excluding the common error terms) for sample i;

- **regDose** (one list per sample); each list contains all regenerated doses;
- **J**, a vector giving, for each BIN file, the number of aliquots selected for the analysis;
- **K**, a vector giving, for each BIN file, the number of regenerative doses in the SAR protocol;
- **Nb\_measurement**, a vector giving, for each BIN file, the number of measurements;

**\*\* How to save this list \*\***

You can save this list in a .RData object. To do this, you can use the fonction [save](#). Then, to load this list you can use the function [load](#) (see example section fore more details).

### Author(s)

Claire Christophe, Guillaume Guerin

### See Also

[read\\_BIN2R](#), [Concat\\_DataFile](#), [LT\\_RegenDose](#) [Age\\_Computation](#), [AgeS\\_Computation](#), [Palaeodose\\_Computation](#)

### Examples

```
## 1) Example for one sample with one Bin File
## Put path in quotes to go to the folder containing bin.BIN file and associated .csv files
## Path must be determinated by "/"
# Path=""
## Enter between the "" the name of the folder containsing bin.BIN file
# Names=""
## give the number of sample
# Nb_sample=1
# DATA=Generate_DataFile_MG(Path,Names,Nb_sample)
# str(DATA)

## to save information in .RData object
# save(DATA,file=c(paste(Path,'DATA.RData',sep="")))
## to load information containing DATA.RData object
# load(file=c(paste(Path,"DATA.RData",sep="")))
```

---

LT\_RegenDose

*Plots Lx/Tx as a function of regenerative dose*

---

### Description

This function plots Lx/Tx values as a function of Regenerative Dose, for every selected aliquot and for each sample.

### Usage

```
LT_RegenDose(DATA, Path, Names, Nb_sample, BinPerSample = rep(1, Nb_sample),
  SampleNames = Names, SG = TRUE, sepDP = c(","))
```

## Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> or <a href="#">Concat_DataFile</a> . DATA can contain information from more than one sample.
Path	character: path to the project folder (the same as the one used in <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> to provide DATA)
Names	character vector: list of names of the sub-folders containing the BIN files, which were used by <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> to generate the DATA object.
Nb_sample	integer: ID number (in [1,Nb_sample]) of the sample selected for plotting L/T as a function of regenerative doses. Required if the DATA object contains information for more than one sample.
BinPerSample	(with default): integer vector (with default): vector with the number of BIN files per sample, which was used in <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> to generate the DATA object.
SampleNames	character vector (with default): Names of samples. To use if there is more than one bin file per sample.
SG	boolean (with default): TRUE if it a Single-grain OSL measurements, else it is Multi-grain OSL measurements.
sepDP	character (with default): column separator in the DiscPose.csv file.

## Details

To fill Names and BinPerSample, we refer to the **Detail** section from the [Generate\\_DataFile](#) or [Generate\\_DataFile](#) function. As well for a precise description of input DATA.

## Value

Lx/Tx plots; there are as many plots as selected aliquots in the DiscPos.csv file. There are 9 plots per page. There is not interpolation.

## Author(s)

Claire Christophe, Guillaume Guerin

## See Also

[Generate\\_DataFile](#), [Generate\\_DataFile\\_MG](#)

## Examples

```
## load data file generated by the function Generate_DataFile
# data(DATA1,envir = environment())
# Path="" #<- Put in quote the path to reach \bf{DiscPos.csv} file of the corresponding sample.
# Names="" #<- Put in quote the Names of the folder containing \bf{DiscPos.csv} file sample.
# LT_RegenDose(DATA1,Path=Path,Names=Names,Nb_sample=1)
```

---

MCMCsample

*MCMC sample from the posterior distribution of the dataset GDB5*


---

### Description

MCMC samples from the posterior distribution of "A" for age, "D" for palaeodose and "sD" for dispersion of equivalent doses around "D", of the data set GDB5.

### Usage

```
data("MCMCsample")
```

### Format

It is a matrix with 6000 rows and three columns.

A The first column of the matrix is sampled from the posterior distribution of the parameter A

D The first column of the matrix is sampled from the posterior distribution of the parameter D

sD The first column of the matrix is sampled from the posterior distribution of the parameter sD

### References

Tribolo, C., Asrat, A., Bahain, J. J., Chapon, C., Douville, E., Fragnol, C., Hernandez, M., Hovers, E., Leplongeon, A., Martin, L., Pleurdeau, D., Pearson, O., Puaud, S., Assefa, Z. (2017). Across the Gap: Geochronological and Sedimentological Analyses from the Late Pleistocene-Holocene Sequence of Goda Buticha, Southeastern Ethiopia. *PloS one*, 12(1), e0169418.

### Examples

```
data(MCMCsample)
## maybe str(MCMCsample) ; plot(MCMCsample[,1],type="l") ...
```

---

MCMC\_plot

*MCMC trajectories and posterior distributions plot*


---

### Description

This function uses the output of [coda.samples](#) to plot the trajectories of MCMC and densities of the posterior distributions of the age - if it is calculated, palaeodose and equivalent dose dispersion parameters of the sample. This function is used in the function [Age\\_Computation](#), [AgeS\\_Computation](#) and [Palaeodose\\_Computation](#).

### Usage

```
MCMC_plot(sample, size, SampleNames, Nb_sample = 1, Nb_chains = 3,
  value = c(0, Nb_sample, 2 * Nb_sample), nom = c("A", "D", "sD"))
```



**Arguments**

sample	MCMC.list: this is generated by <code>jags.model</code> in <code>Age_Computation</code> .
size	integer: length of each chain.
SampleNames	character vector: names of the samples, used in the figure titles.
Nb_sample	integer (with default): number of analysed samples.
Nb_chaines	integer (with default): number of independent chains for the model (for more informations, see <code>jags.model</code> ).
value	integer vector (with default): position index used to select age (if available), palaeodose and equivalent dose dispersion parameters for the first sample.
nom	character vector (with default): names of the selected parameters with value A for age (if available), D for palaeodose and sD for equivalent dose dispersion.

**Value**

A pdf file with the MCMC trajectories and posterior distributions for each parameter defined in `nom`. There is one page per sample, which is divided by `length(nom)` vertically and by 2 horizontally. The first line of the figure corresponds to the first parameter defined in `nom`, and so on. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distributions of the parameter.

**Author(s)**

Claire Christophe, Guillaume Guerin

**See Also**

`Age_Computation`, `AgeS_Computation`, `Palaeodose_Computation`, `coda.samples` and `rjags` packages.

**Examples**

```
data(MCMCsample,envir = environment())
MCMC_plot(sample=MCMCsample,size=2000,SampleNames="GDB3")
```

---

Model\_Age

*JAGS models use in Age\_Computation*


---

**Description**

A list of JAGS models use to a Bayesian analysis of OSL age of one sample. There are models for various growth curves and various distribution to describe equivalent dose distribution around the palaeodose.

**Usage**

```
data("Model_Age")
```

## Format

This list contains:

`AgeMultiBF_EXPLIN` a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgeMultiBF_EXP` a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgeMultiBF_EXPZO` a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

`AgeMultiBF_EXPLINZO` a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

## Details

The different distributions to describe equivalent dose values around the palaeodose are:

`cauchy` a Cauchy distribution with position parameter equal to the palaeodose of the sample

`gaussian` a Gaussian distribution with mean equal to the palaeodose of the sample

`lognormal_A` a log-normal distribution with mean or Average equal to the palaeodose of the sample

`lognormal_M` a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [Age\\_Computation](#), section Details.

## References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

## See Also

[rjags](#)

## Examples

```
data(Model_Age)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[AgeMultiBF_EXPLIN]][[gaussian]]
```

---

Model\_AgeC14*JAGS models use in AgeC14\_Computation*

---

**Description**

A list of JAGS models use to a Bayesian analysis of C14 calibration age of various sample. Stratigraphic relations can be taken in count to calibrate C14 ages.

**Usage**

```
data("Model_AgeC14")
```

**Format**

This list contains:

full a model considering error on calibration curve.

naive a model not considering error on calibration curve.

**References**

Reimer PJ, Bard E, Bayliss A, Beck JW, Blackwell PC, Bronl Ramsey C, Buck CE, Cheng H, Edwards RL, Friedrich M, Grootes PM, Guilderson TP, Haffidason H, Hajdas I, Hatte C, Heaton TJ, Hoffmann DL, Hogg AG, Hughen KA, Kaiser KF, Kromer B, Manning SW, Niu M, Reimer RW, Richards DA, Scott EM, Southon JR, Staff RA, Turney CSM, van der Plicht J. 2013. IntCal13 and Marine13 radiocarbon age calibration curves 0-50000 years cal BP. Radiocarbon 55(4):1869-1887.

Hogg AG, Hua Q, Blackwell PG, Niu M, Buck CE, Guilderson TP, Heaton TJ, Palmer JG, Reimer PJ, Reimer RW, Turney CSM, Zimmerman SRH. 2013. SHCal13 Southern Hemisphere calibration, 0-50000 years cal BP. Radiocarbon 55(4):1889-1903

**See Also**

[rjags](#)

**Examples**

```
data(Model_AgeC14)
## Model_AgeC14[[full]]
## Model_AgeC14$full
```

---

Model\_AgeS

---

*JAGS models use in AgeS\_Computation*


---

## Description

A list of JAGS models use to a Bayesian analysis of OSL age of various samples. There are models for various growth curves and various distrubution to describe equivalent dose distribution around the palaeodose.

## Usage

```
data("Model_AgeS")
```

## Format

This list contains:

**AgesMultiCS2\_EXPLIN** a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

**AgesMultiCS2\_EXP** a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

**AgesMultiCS2\_EXPZ0** a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

**AgesMultiCS2\_EXPLINZ0** a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

## Details

The different distributions to describe equivalent dose values around the palaeodose are:

**cauchy** a Cauchy distribution with postition parameter equal to the palaeodose of the sample

**gaussian** a Gaussian distribution with mean equal to the palaeodose of the sample

**lognormal\_A** a log-normal distribution with mean or Average equal to the palaeodose of the sample

**lognormal\_M** a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [AgeS\\_Computation](#), section Details.

## References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

## See Also

[rjags](#)

## Examples

```
data(Model_AgeS)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type  $f(x)=a(1-\exp(-x/b))+cx+d$ )
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[AgesMultiCS2_EXPLIN]][[gaussian]]
```

---

Model_Palaeodose	<i>JAGS models use in Palaeodose_Computation</i>
------------------	--

---

## Description

A list of JAGS models use to a Bayesian analysis of OSL palaeodose of one or various samples. There are models for various growth curves and various distribution to describe equivalent dose distribution around the palaeodose.

## Usage

```
data("Model_Palaeodose")
```

## Format

This list contains:

**PalaeodosesMultiBF\_EXPLIN** a list of 4 models that all consider a saturating exponential plus linear growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

**PalaeodosesMultiBF\_EXP** a list of 4 models that all consider a saturating exponential growth. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

**PalaeodosesMultiBF\_EXPZO** a list of 4 models that all consider a saturating exponential plus linear growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

**PalaeodosesMultiBF\_EXPLINZO** a list of 4 models that all consider a saturating exponential growth and fitting through the origin. These 4 models have different distribution to describe equivalent dose values around the palaeodose.

## Details

The different distributions to describe equivalent dose values around the palaeodose are:

**cauchy** a Cauchy distribution with position parameter equal to the palaeodose of the sample

**gaussian** a Gaussian distribution with mean equal to the palaeodose of the sample

**lognormal\_A** a log-normal distribution with mean or Average equal to the palaeodose of the sample

**lognormal\_M** a log-normal distribution with Median equal to the palaeodose of the sample

For more information we refer to the function [Palaeodose\\_Computation](#), section Details.

## References

Plummer, M. (2003). JAGS: A program for analysis of Bayesian graphical models using Gibbs sampling. In Proceedings of the 3rd international workshop on distributed statistical computing, volume 124, page 125. Technische Universit at Wien, Austria.

Plummer, M. (2015). JAGS Version 4.0. 0 user manual.

## See Also

[rjags](#)

## Examples

```
data(Model_Palaeodose)
## The JAGS model for a saturating exponential plus linear growth
## (a function of the type \code{f(x)=a(1-exp(-x/b))+cx+d})
## and a gaussian distribution of equivalent doses around the palaeodose:
# Model_Age[[PalaeodosesMultiBF_EXPLIN]][[gaussian]]
```

---

Palaeodose\_Computation

*Bayesian analysis for the palaeodose estimation of various samples*

---

## Description

This function computes the palaeodose of one or various samples according to the model developed in Combes et al (2015), based on an output of [Generate\\_DataFile](#) or [Generate\\_DataFile\\_MG](#) or both of them using [Concat\\_DataFile](#).

Samples, for which data is available in several BIN files, can be analysed.

Single-grain or Multi-grain OSL measurements can be analysed simultaneously.

## Usage

```
Palaeodose_Computation(DATA, SampleNames, Nb_sample, BinPerSample = rep(1,
  Nb_sample), SavePdf = FALSE, OutputFileName = c("MCMCplot"),
  OutputFilePath = c(""), SaveEstimates = FALSE,
  OutputTableName = c("DATA"), OutputTablePath = c(""), LIN_fit = TRUE,
  Origin_fit = FALSE, distribution = c("cauchy"), Taille = 50000, t = 5,
  Nb_chaines = 3)
```

## Arguments

DATA	list of objects: LT, sLT, ITimes, dLab, ddot_env, regDose, J, K, Nb_measurement, provided by <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> . DATA contains information for more than one sample.
SampleNames	character vector: names of sample. The length of this vector is equal to Nb_sample.
Nb_sample	integer: number of samples, Nb_sample>1.
BinPerSample	integer vector (with default): vector with the number of BIN files per sample. The length of this vector is equal to Nb_sample. BinPerSample[i] corresponds to the number of BIN files for the sample whose number ID is equal to i. For more information to fill this vector, we refer to details in <a href="#">Generate_DataFile</a> or <a href="#">Generate_DataFile_MG</a> .

SavePdf	boolean (with default): if TRUE save graph in pdf file named OutputFileName in folder OutputFilePath.
OutputFileName	character (with default): name of the pdf files that will be generated by the function.
OutputFilePath	character (with default): path to the pdf files that will be generated by the function.
SaveEstimates	boolean (with default): if TRUE save Bayes estimates and credible interval at level 68 in a csv table named OutputFileName in folder OutputFilePath.
OutputTableName	character (with default): name of the table that will be generated by the function if SaveEstimates=TRUE.
OutputTablePath	character (with default): path to the table that will be generated by the function if SaveEstimates=TRUE.
LIN_fit	logical (with default): if TRUE (default) allows a linear component, on top of the (default) saturating exponential curve, for the fitting of dose response curves. Please see details for more informations on the proposed dose response curves.
Origin_fit	logical (with default): if TRUE, forces the dose response curves to pass through the origin. Please see details for more informations on the proposed growth curves.
distribution	character (with default): type of distribution that defines how individual equivalent dose values are distributed around the palaeodose. Allowed inputs are "cauchy", "gaussian", "lognormal_A" and "lognormal_M".
Taille	integer (with default): number of iterations for the MCMC computation (for more information see <a href="#">jags.model</a> ).
t	integer (with default): 1 every t iterations of the MCMC is considered for sampling the posterior distribution (for more information see <a href="#">jags.model</a> ).
Nb_chaines	integer (with default): number of independent chains for the model (for more information see <a href="#">jags.model</a> ).

## Details

### \*\* Option on growth curves \*\*

As for [Age\\_Computation](#) and [AgeS\\_Computation](#), the user can choose from 4 dose response curves:

- **Saturating exponential plus linear growth** (PalaeodosesMultiBF\_EXPLIN):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))+cx+d$ ; select
  - LIN\_fit=TRUE
  - Origin\_fit=FALSE
- **Saturating exponential growth** (PalaeodosesMultiBF\_EXP):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))+d$ ; select
  - LIN\_fit=FALSE
  - Origin\_fit=FALSE
- **Saturating exponential plus linear growth and fitting through the origin** (PalaeodosesMultiBF\_EXPLINZ0):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))+cx$ ; select
  - LIN\_fit=TRUE

- Origin\_fit=TRUE
- **Saturating exponential growth and fitting through the origin** (PalaeodosesMultiBF\_EXPZ0):  
for all  $x$  in  $\mathbb{R}^+$ ,  $f(x)=a(1-\exp(-x/b))$ ; select
  - LIN\_fit=FALSE
  - Origin\_fit=TRUE

**\*\* Option on equivalent dose distribution around the palaeodose \*\***

The user can choose between :

- **cauchy**: a Cauchy distribution with location parameter equal to the palaeodose of the sample
- **gaussian**: a Gaussian distribution with mean equal to the palaeodose of the sample
- **lognormal\_A**: a log-normal distribution with mean or Average equal to the palaeodose of the sample
- **lognormal\_M**: a log-normal distribution with Median equal to the palaeodose of the sample

## Value

### NUMERICAL OUTPUT

1. **A list containing the following objects:**

- **Sampling** that corresponds to a sample of the posterior distributions of palaeodose and equivalent dose dispersion parameters.
- **Model\_GrowthCurve**, stating which dose response fitting option was chosen;
- **Distribution**, stating which distribution was chosen to model the dispersion of individual equivalent dose values around the palaeodose of the sample;
- **PriorAge**, stating the priors used for the age parameter.

2. **The Gelman and Rudin test of convergency**: prints the result of the Gelman and Rudin test of convergency for palaeodose and equivalent dose dispersion parameters for each sample. A result close to one is expected.

In addition, the user must visually assess the convergency of the trajectories by looking at the pdf file generated by the function (see **PLOT OUTPUT** for more informations).

If both convergencies (Gelman and Rudin test and plot checking) are satisfactory, the user can consider the printed estimates as valid. Otherwise, the user may try increasing the number of MCMC iterations (Taille) to reach convergency.

3. **Credible intervals and Bayes estimates**: prints the Bayes estimates, the credible intervals at 95% and 68% for the palaeodose and equivalent dose dispersion parameters for each sample.

### PLOT OUTPUT

1. **MCMC trajectories** A graph with the MCMC trajectories and posterior distributions of the palaeodose and equivalent dose dispersion parameters is displayed, there is one page per sample.

The first line of the figure corresponds to the palaeodose parameter and the second to the equivalent dose dispersion parameter. On each line, the plot on the left represents the MCMC trajectories, and the one on the right the posterior distribution of the parameter.

2. **Summary of palaeodose estimates**: plot credible intervals and Bayes estimate of each sample palaeodose on a same graph.



**Author(s)**

Claire Christophe, Guillaume Guerin

**References**

Combes, B., Philippe, A., Lanos, P., Mercier, N., Tribolo, C., Guerin, G., Guibert, P., Lahaye, C., 2015. A Bayesian central equivalent dose model for optically stimulated luminescence dating. *Quaternary Geochronology* 28, 62-70. doi:10.1016/j.quageo.2015.04.001

**See Also**

[Generate\\_DataFile](#), [Generate\\_DataFile\\_MG](#), [Concat\\_DataFile](#), [rjags](#), [MCMC\\_plot](#), [Age\\_Computation](#), [AgeS\\_Computation](#)

**Examples**

```
## Load data
# data(DATA1,envir = environment())
## Palaeodose computation of samples GDB3
# P=Palaeodose_Computation(DATA1,Nb_sample=1,SampleNames=c("GDB5"),Taille=10000)
```

---

SCMatrix

---

*Definition of the stratigraphic constraint matrix*


---

**Description**

This function helps to define the stratigraphic relation between samples, with questions. The output of this function can be used in function `AgeS_Computation`.

**Usage**

```
SCMatrix(Nb_sample, Names)
```

**Arguments**

Nb_sample	interger: the sample number.
Names	charcater vector: sample names.

**Details**

Ask if sample *i* is younger than sample *j* to construc the stratigraphic constrain matrix.

**Value**

A Matrix that summarise the ordered relation between samples. This matrix can be intergrate in `AgeS_Computation` function. We refer to detail on `AgeS_Computation` for more information concerning this matrix.

**Author(s)**

Claire Christophe, Guillaume Guerin

**See Also**

AgeS\_Computation

**Examples**

```
## Assume that "sample1" is younger than "sample2"
# SCMatrix(2,c("sample1","sample2"))
# 1
```

---

SC\_Ordered

*Stratigraphic matrix for samples ordered*

---

**Description**

Construc the stratigraphic matrix used in fonction AgeS\_Computation and AgeC14\_Computation for samples that are all ordered by increasing oder.

**Usage**

```
SC_Ordered(Nb_sample)
```

**Arguments**

Nb\_sample            interger: the sample number.

**Value**

Stratigraphic matrix where each sample are ordered by incresing order This matrix can be intergrate in AgeS\_Computation function. We refer to detail on AgeS\_Computation for more information concerning this matrix.

**Author(s)**

Claire Christophe, Guillaume Guerin

**See Also**

AgeS\_Computation, SC\_Matrix

**Examples**

```
SC=SC_Ordered(Nb_sample=3)
```

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