

Phenotype and genotype input file preparation:

For Phenotype analysis tab

- The phenotype file should contain at least five columns such as Location, Season, Block, Accession, Trait1, Trait2 etc.
Where,
Trait1, Trait2 represents phenotype value of various traits and any number of traits can be added in the phenotype file for phenotype analysis.

For 3k Rice (Default)

Location	Season	BLO CK	Accession	NPT	PL	GYKGPH A
ISARC	20WS	1	POHHERLIMASION::IRGC 62025-1	5	28.9	2011.7
ISARC	21WS	1	POHHERLIMASION::IRGC 62025-1	8	35.33	3929.825
ISARC	21WS	1	NAN TEO 14::IRGC 7304-1	10	25.4	3512.069
ISARC	20WS	1	NAN TEO 14::IRGC 7304-1	6	19.6	
ISARC	21WS	1	URAIBOOL::IRGC 52785-1	7.67	28.9	2601.887
ISARC	20WS	1	URAIBOOL::IRGC 52785-1	5	25.8	3729.09
ISARC	20WS	1	IARI 11387::IRGC 19573-1	6	23.8	2347.06
ISARC	21WS	1	IARI 11387::IRGC 19573-1	7	20.53	3091.667
ISARC	21WS	1	HARRA::GERVEX 501-C1	6.67	18.4	749.1228
ISARC	20WS	1	HARRA::GERVEX 501-C1	7	12.5	478.33
ISARC	20WS	1	PL 3165::IRGC 62827-1	6	13.8	630
ISARC	21WS	1	PL 3165::IRGC 62827-1	8.67	18.13	1878.333
ISARC	20WS	1	ARC 18371::IRGC 42423-2	6	26.1	2444.26
ISARC	21WS	1	ARC 18371::IRGC 42423-2	5.67	21.3	3120.339

For other diploid organism (external)

Trial	Genotype	Replication	TN	RN	ARP	RL	subpopulation
2nd Expt	MG1	3	9	212	33.99	711.243	xx
2nd Expt	MG1	2	9	235	40.06	701.678	xx
2nd Expt	MG1	1	10	283	37.82	524.315	xx
2nd Expt	MG5	1	8	163	34.65	732.077	xx
2nd Expt	MG5	2	9	177	37.64	617.693	xx
2nd Expt	MG5	3	9	196	31.9	892.359	xx
2nd Expt	MG6	1	9	218	24.79	828.369	xx
2nd Expt	MG6	3	9	226	24.7	589.501	xx

2nd Expt	MG6	2	9	239	25.79	507.984	yy
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For other analysis such as Genofile Extract, GWAS, Et-GWAS, Haplopheno tabs

External: Accession/ IDs, should be same

Default

- First column name should be X.Phenotype. of phenotype file(.csv format).
- ID's in X.Phenotype. column should be in the following format e.g; IRIS_313-11586.
- No need to provide a subpopulation column.

External

- The first column name should be X.Phenotype of phenotype file.
- Accession/ IDs in X.Phenotype. column should be the same as the vcf file (genotype file) provided by the user.
- Subpopulation or group information should be provided in the third column of the phenotype file and name of the **column should be “subpopulation”**.

For 3k Rice (Default)

X.Phenotype.	PL
IRIS_313-11586	27.23
IRIS_313-11037	27.83
IRIS_313-8061	19.63
IRIS_313-8060	26.07
IRIS_313-8057	26.83
IRIS_313-9140	24.33
IRIS_313-12185	27.5
IRIS_313-11744	24.73
IRIS_313-12057	29.07

For other diploid organism (external)

X.Phenotype.	ARP(adjusted)	subpopulation
MG1	37.00889	yq
MG10	29.31557	yy
MG100	30.35416	yy
MG101	30.87122	xx
MG106	23.78305	zz
MG109	21.66455	yy
MG11	34.11961	zz
MG110	35.22003	xx
MG114	31.79448	zz

Run HapGUI from docker hub: (follow video manual)

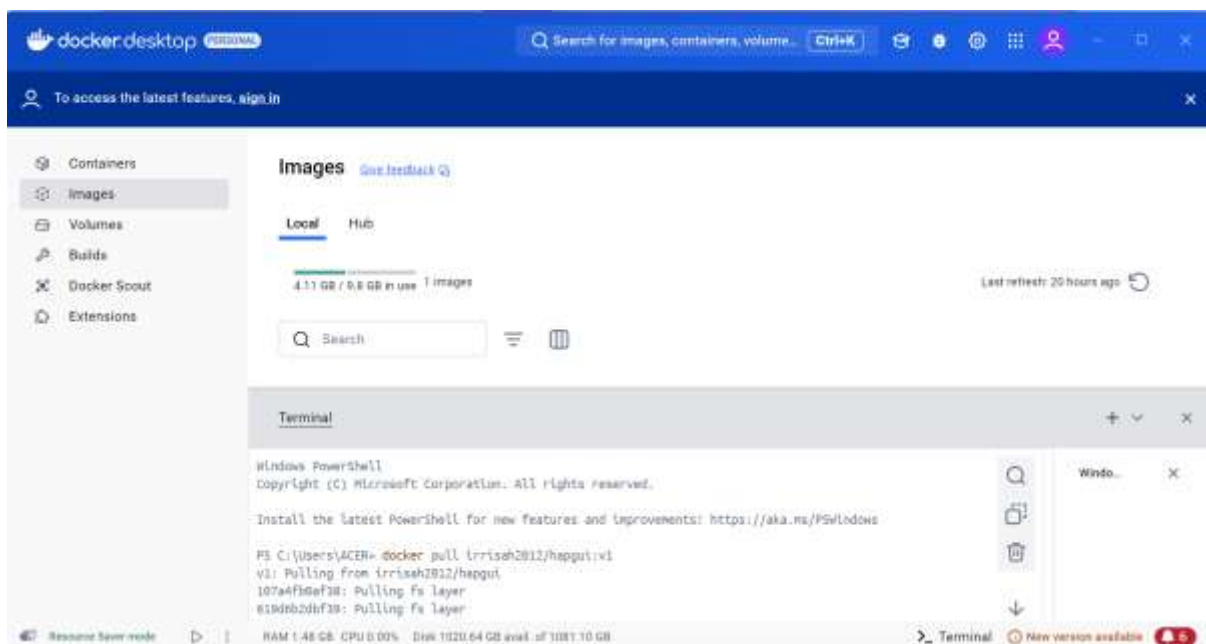
1. Download docker desktop based on operating system from <https://www.docker.com/products/docker-desktop/>
2. Follow the instructions for installing docker by following the provided video manual.
 - While installation of docker, if following window is displayed uncheck above two option and only check third option (Add shortcut to desktop). Everything else will be chosen default as shown in the video manual.

Configuration

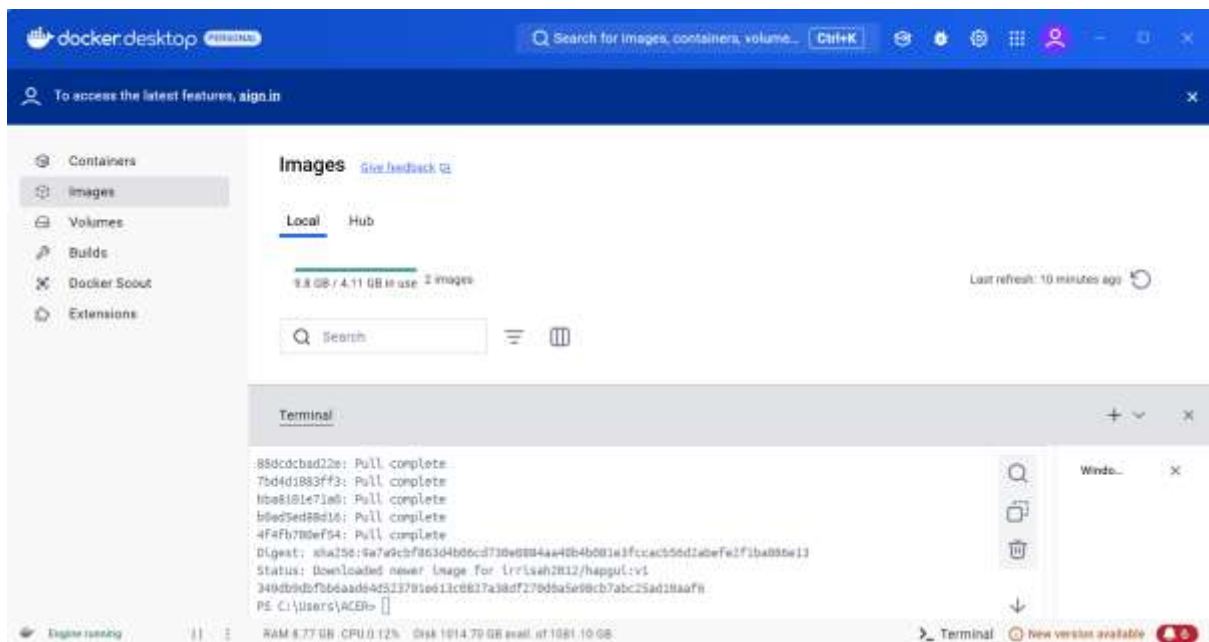
- ☐ Use WSL 2 instead of Hyper-V (recommended)
- ☐ Allow Windows Containers to be used with this installation
- ☒ Add shortcut to desktop

3. To run HapGUI, users need to follow the following instructions just once.
 - Then open the terminal of docker, type the following commands and run one by one.

```
docker pull irrisah2012/hapgui:v1
```

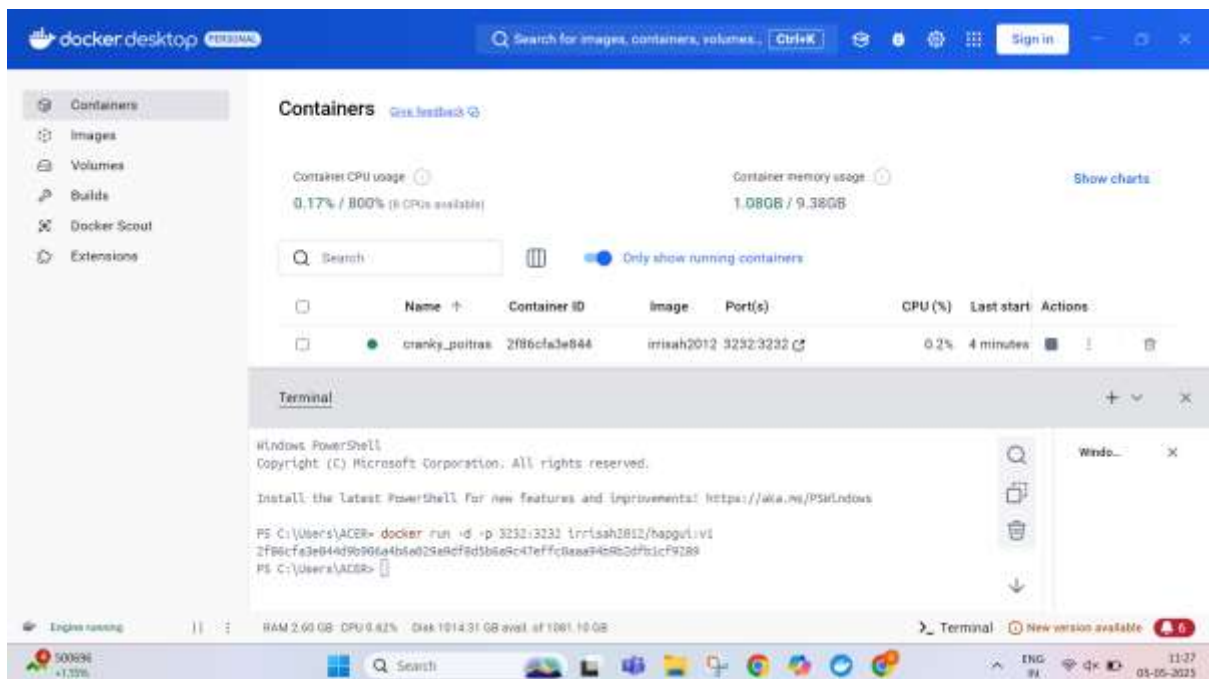


- After the pull completes the docker terminal will look like the following.



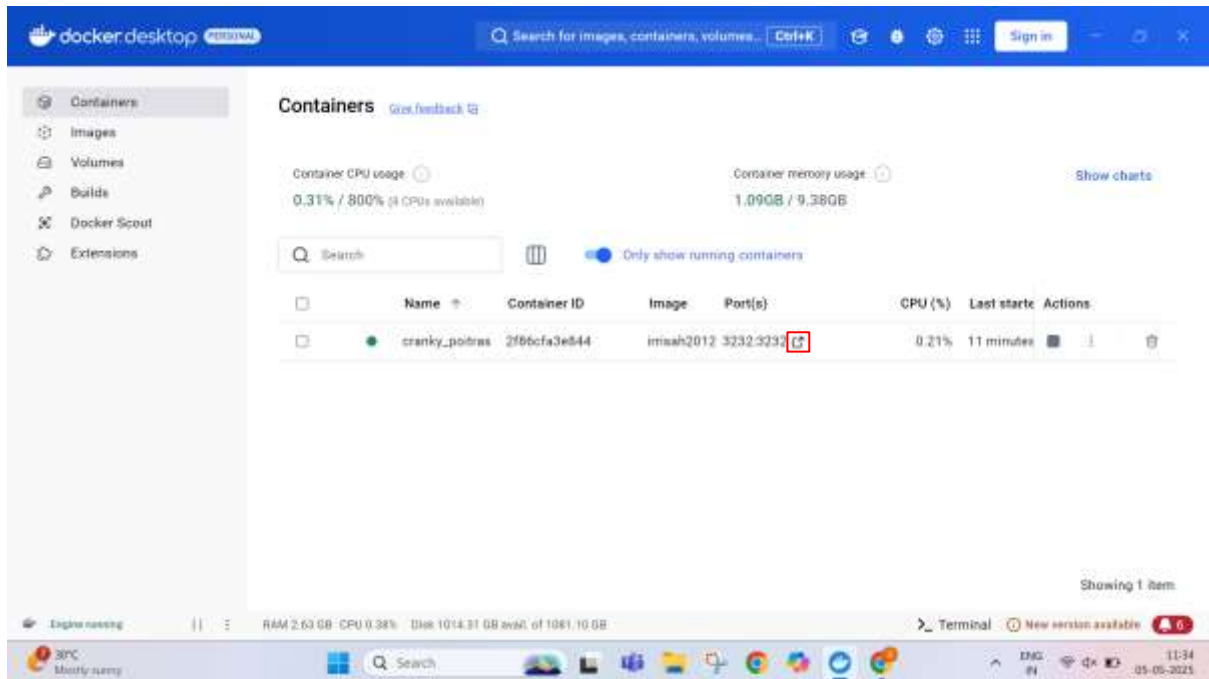
- Then type following command and run.

`docker run -d -p 3232:3232 irrisah2012/hapgui:v1`

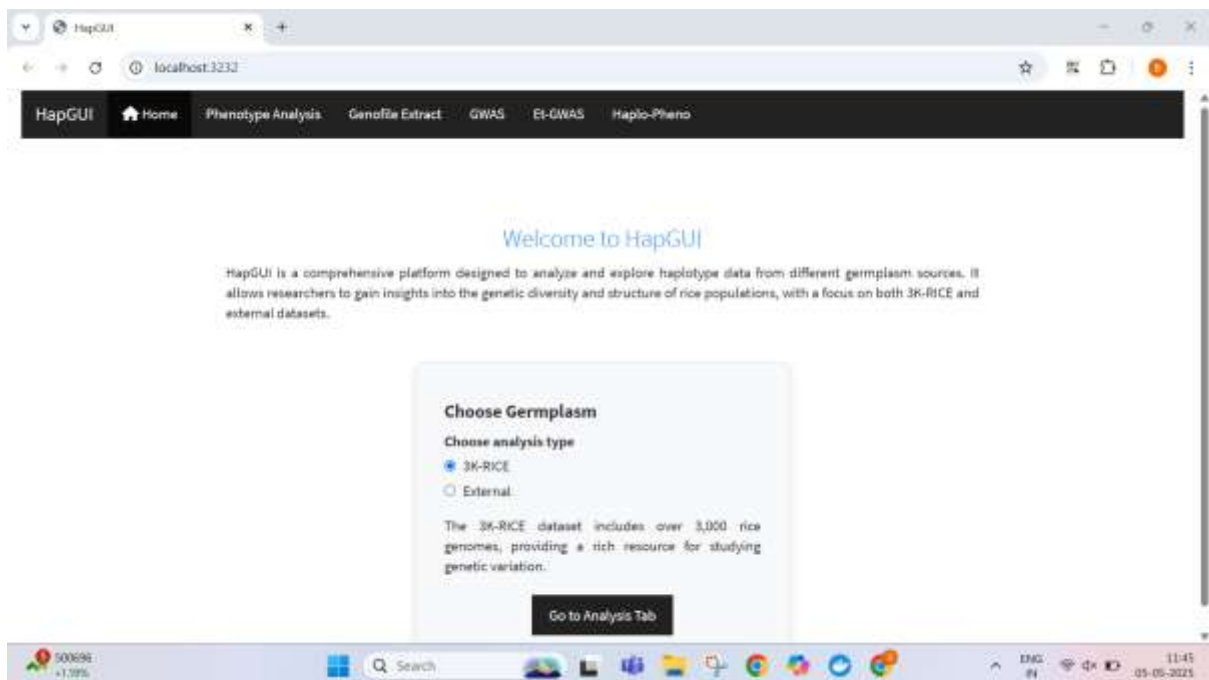


4. Then click on the highlighted symbol, after that on the browser (chrome, firefox etc.) of the user's PC HapGUI will open as provided in the screenshot below.

Else, copy the link (<http://localhost:3232/>) and paste on any browser and press enter.



Then HapGUI interface will open as shown below.



Before running other analysis for external organism Please download the gff3 file as demonstrated below.

Before performing Genofile Extract (Genotype file extraction) for GWAS, Et-GWAS and Haplopheno analysis for other panels of Rice as well as other diploid organism gff3 file (General Feature Format) need to be downloaded from ensemble database (<https://ensemblgenomes.org/>) as explained in the video manual.

The screenshot shows the Ensembl Plants homepage. At the top, there's a navigation bar with links like 'HOME', 'BLAST', 'Biomart', 'Tools', 'Downloads', 'Help & Docs', and 'Blog'. A search bar is prominently displayed with the text 'Search Ensembl Plants'. Below the search bar, the 'All genomes' section shows a list of species, including Arabidopsis thaliana, Oryza sativa, and Zea mays. The 'Favorite genomes' section allows users to mark their preferred species. The 'What's new in release 61' section provides a summary of the latest updates, including new genomes and improvements to the genome assembly. The 'Archive sites' section offers links to previous releases of the database. The 'Funding' section acknowledges the support from the European Union and the Max Planck Society.

This screenshot shows the 'Zea mays' page on the Ensembl Plants website. The page is dedicated to the Zea mays genome assembly. It features a search bar at the top. The 'About Zea mays' section provides a brief overview of the species and its genome. The 'Genome assembly' section details the 3x-R10-REFERENCE-ASMv3.1 assembly, including its size and the number of contigs. The 'Gene annotation' section describes the gene models and the Ensembl Gene annotation pipeline. The 'Comparative genomics' section offers tools for comparing the Zea mays genome with other species. The 'Regulation' section provides information about the regulatory elements of the genome. The 'Links' section offers additional resources and links to related databases.

- Be careful while downloading the gff3 file. Because the gff3 file will be available separately for each chromosome and one gff3 file will be with all chromosomes. So, we need the gff3 file with all chromosomes as mentioned below (e.g., for Arabidopsis_thaliana).
- After downloading extract the gff3 file and use in the analysis.

Index of /pub/plants/release-61/gff3/arabidopsis_thaliana

Name	Last modified	Size	Description
Parent Directory			
Arabidopsis_thaliana.TAIR10.61.chromosome.1.gff3.gz	2025-04-15 12:49	2.4M	
Arabidopsis_thaliana.TAIR10.61.chromosome.2.gff3.gz	2025-04-15 12:49	1.4M	
Arabidopsis_thaliana.TAIR10.61.chromosome.3.gff3.gz	2025-04-15 12:49	1.8M	
Arabidopsis_thaliana.TAIR10.61.chromosome.4.gff3.gz	2025-04-15 12:49	1.4M	
Arabidopsis_thaliana.TAIR10.61.chromosome.5.gff3.gz	2025-04-15 12:49	2.1M	
Arabidopsis_thaliana.TAIR10.61.chromosome.Mt.gff3.gz	2025-04-15 12:49	9.7K	
Arabidopsis_thaliana.TAIR10.61.chromosome.Pt.gff3.gz	2025-04-15 12:49	9.3K	
Arabidopsis_thaliana.TAIR10.61.gff3.gz	2025-04-15 12:49	9.1M	
README	2025-04-15 12:45	6.2K	

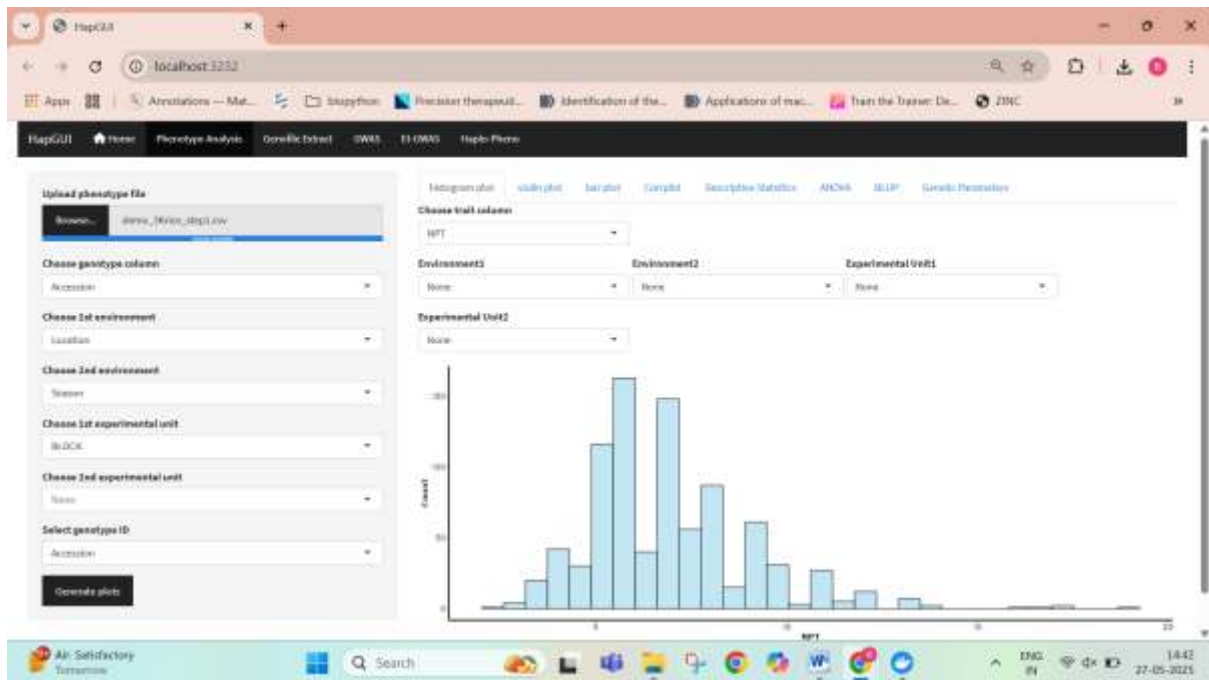
← Download this file

Phenotype analysis:

- Choose phenotype file.
- Choose genotype column.
- Choose 1st environment. If user has 2nd environment, then choose (if available in your phenotype data).
- Next choose 1st and 2nd experimental unit (if available in your phenotype data).
- Select genotype ID.
- Click on generate plots.

Histogram plot

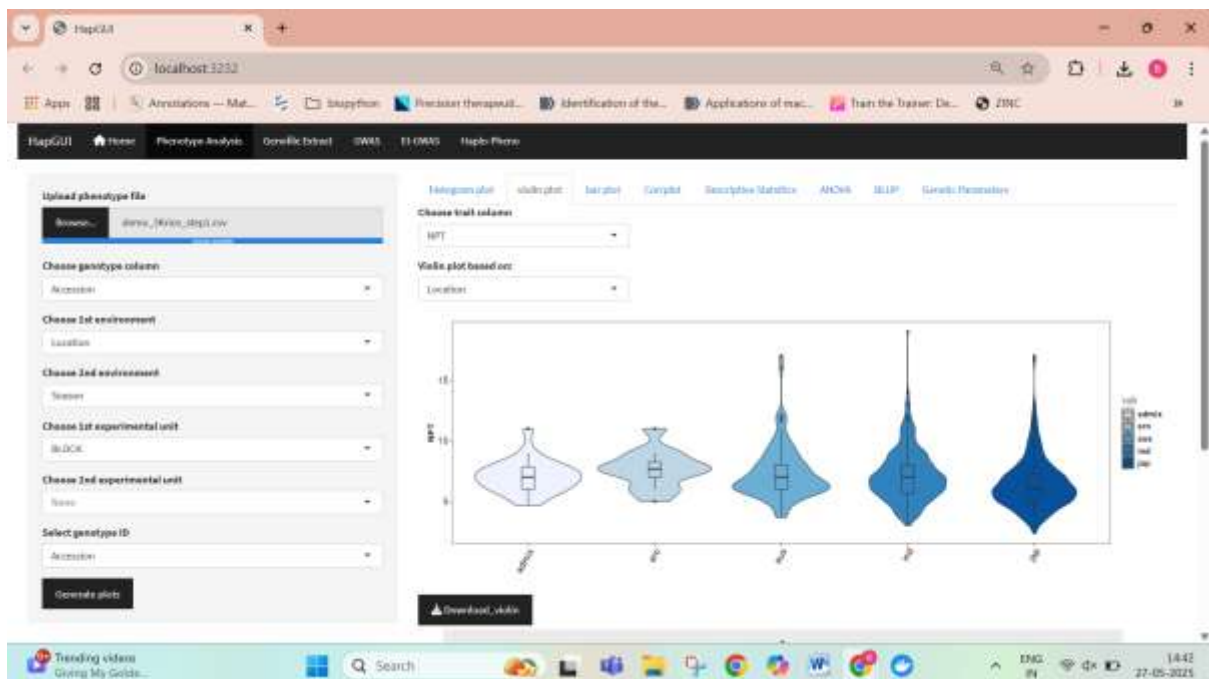
- Choose trait column that needs to be visualized.
- The histogram plot can be downloaded by clicking on “Download_histogram”.



Violin plot

Select appropriate option of interest from the steps mentioned below:

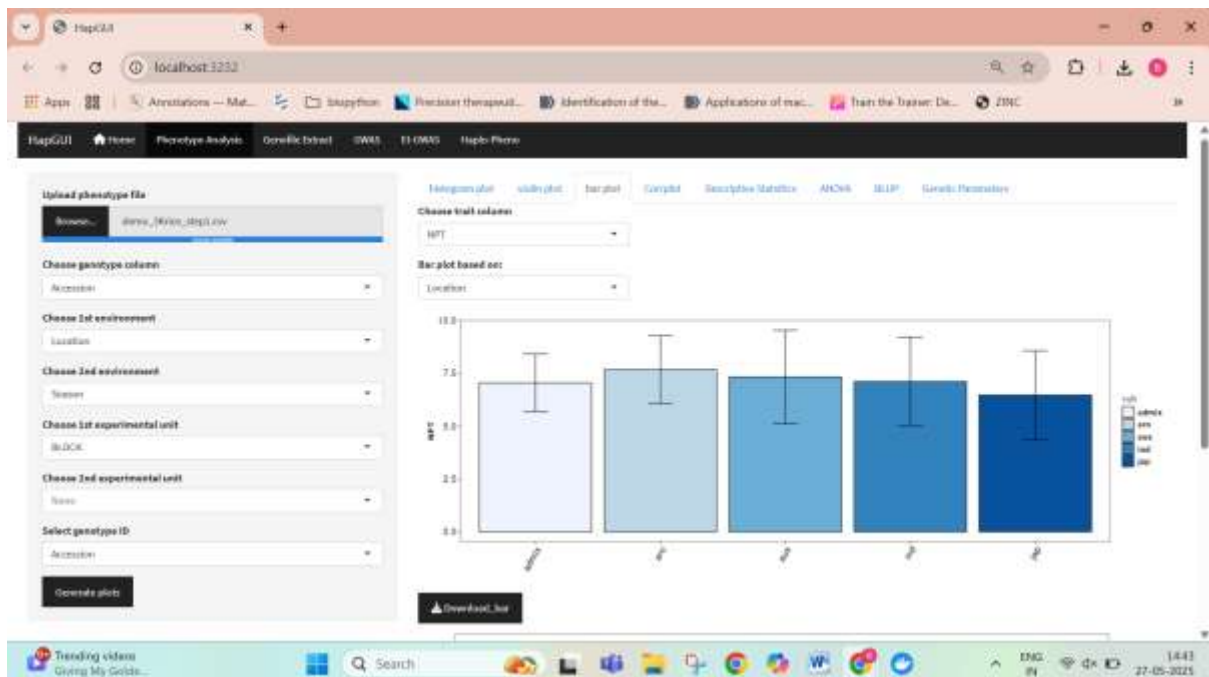
- i. Choose trait column that needs to be visualised.
- ii. Select “Violin plot based on” option that needs to be visualized.
- iii. The histogram plot can be downloaded by clicking on “Download_violin”.



Box plot

Select appropriate option of interest from the steps mentioned below:

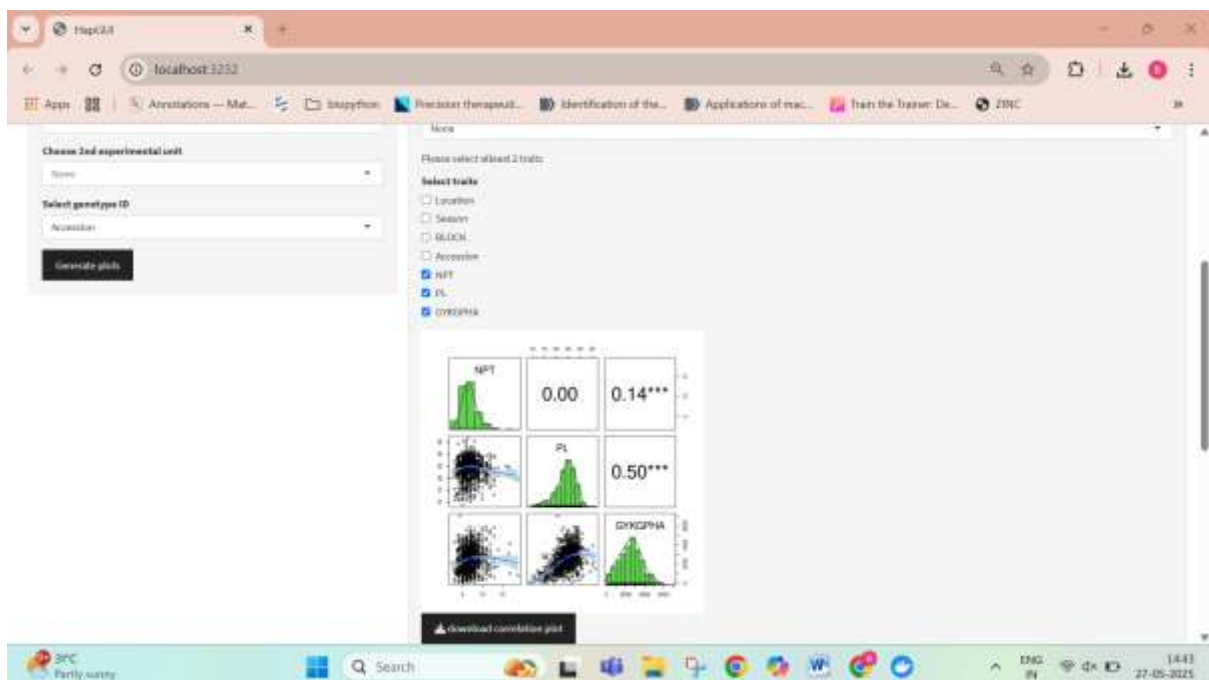
- i. “Choose trait column”.
- ii. “Bar plot based on”.
- iii. The bar plot can be downloaded by clicking on “Download_bar”.



Correlation plot

Select appropriate option of interest from the steps mentioned below:

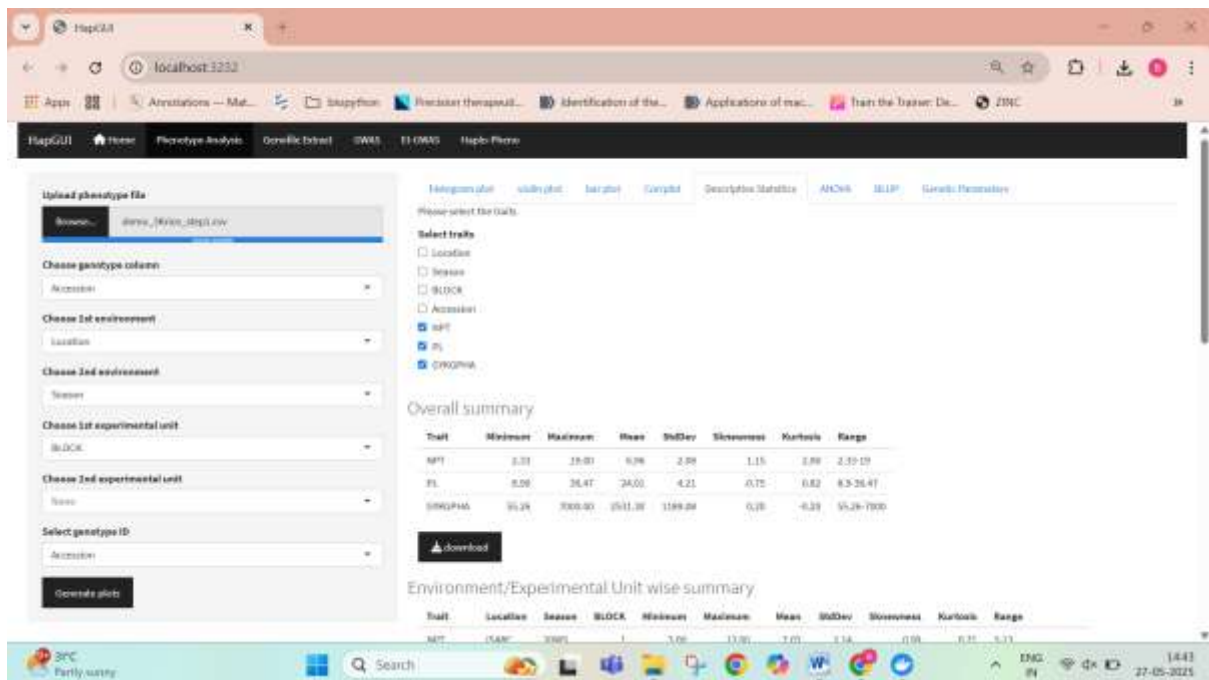
- i. Please select more than 2 traits for getting correlation plot.
- ii. The correlation plot can be downloaded by clicking on “Download_correlation plot”.



Descriptive statistics

Select appropriate option of interest from the steps mentioned below:

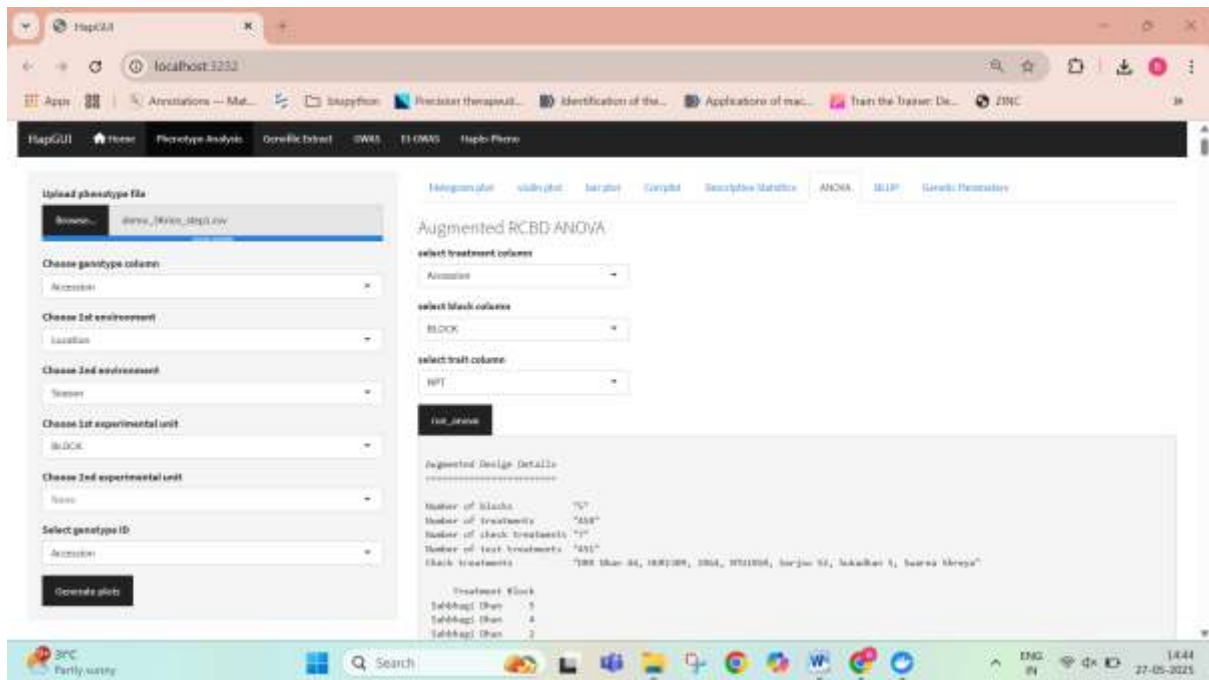
- i. Select traits of interest
- ii. The descriptive statistics can be downloaded by clicking on “download” button.



ANOVA

Select appropriate option of interest from the steps mentioned below:

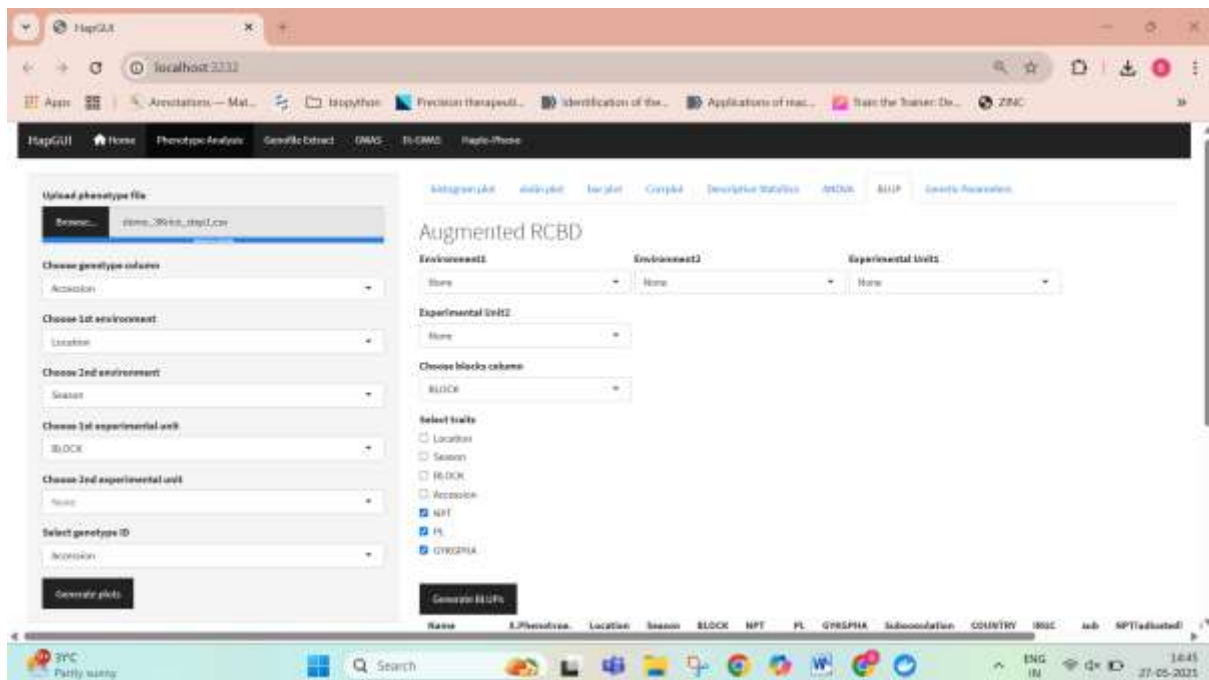
- i. Select treatment column (e.g, Accession).
- ii. Select block column.
- iii. Select trait column.
- iv. Click “Run_anova”.
- v. The results will be displayed as given below.



BLUP

Select appropriate option of interest from the steps mentioned below:

- “Select traits” (check on traits of interest).
- Click “Generate BLUPs”.
- Blup results



- Displaying the result of BLUP.

Genetic parameters

Select appropriate option of interest from the steps mentioned below:

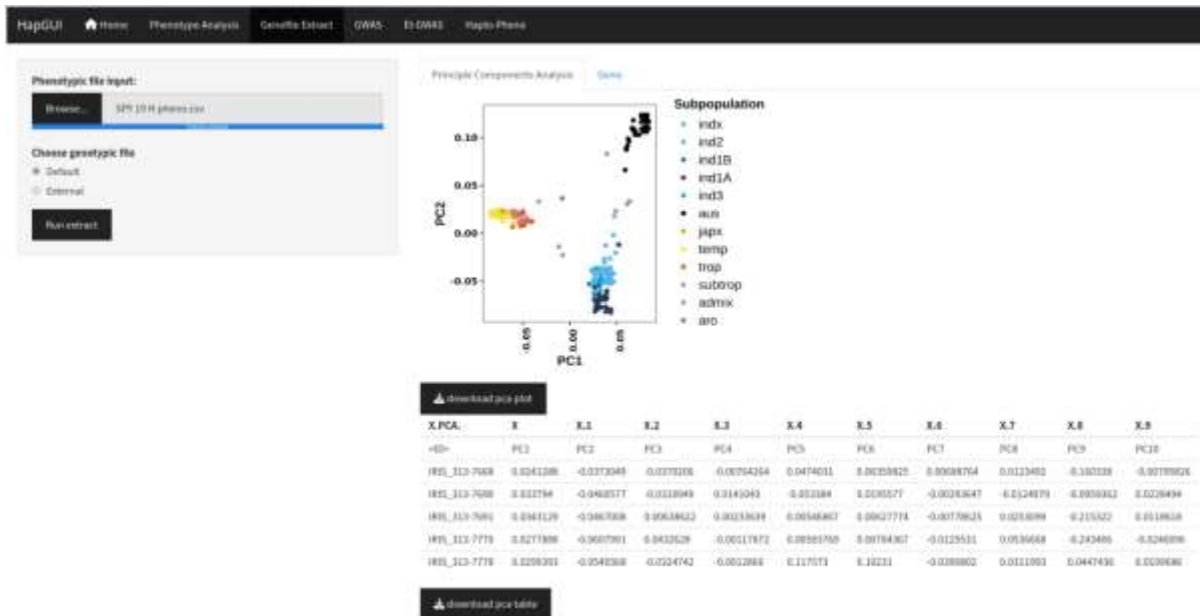
- Choose blocks column
- Select treatment column
- “Select traits” Check on traits for those BLUP values needs to be generated.

Genofile Extract

For 3k Rice panel

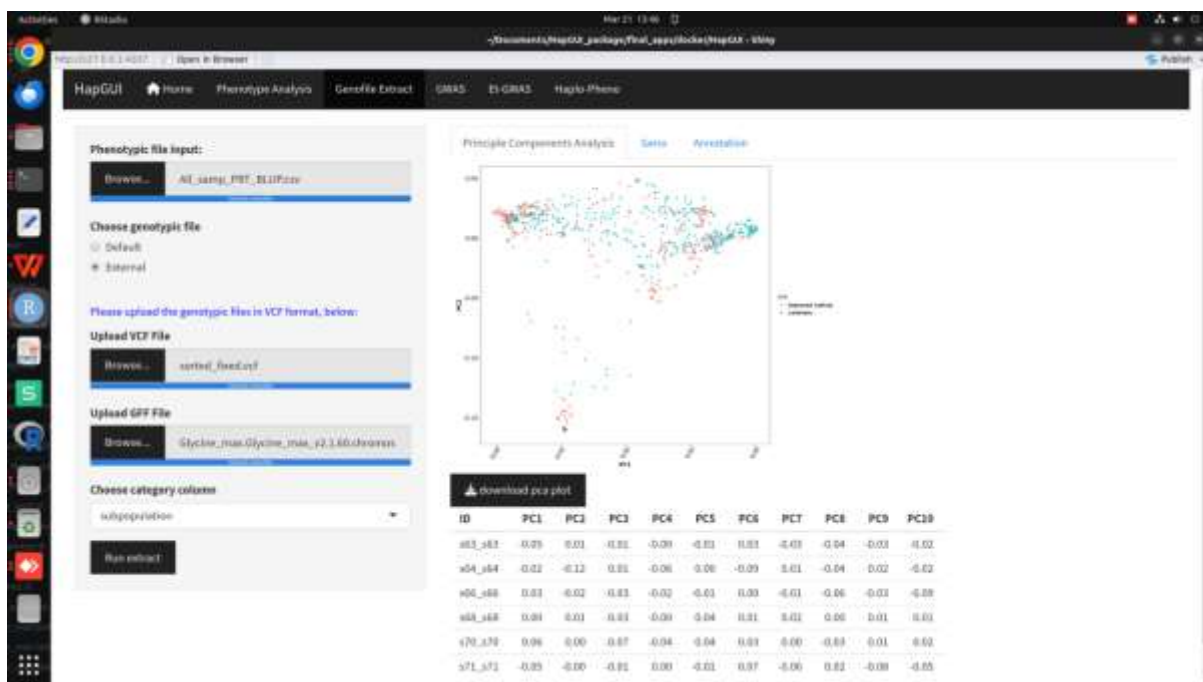
- Upload your phenotype file in appropriate format as mentioned in the page no. 2

- ii. Select “Default” in “Choose genotypic file” section.



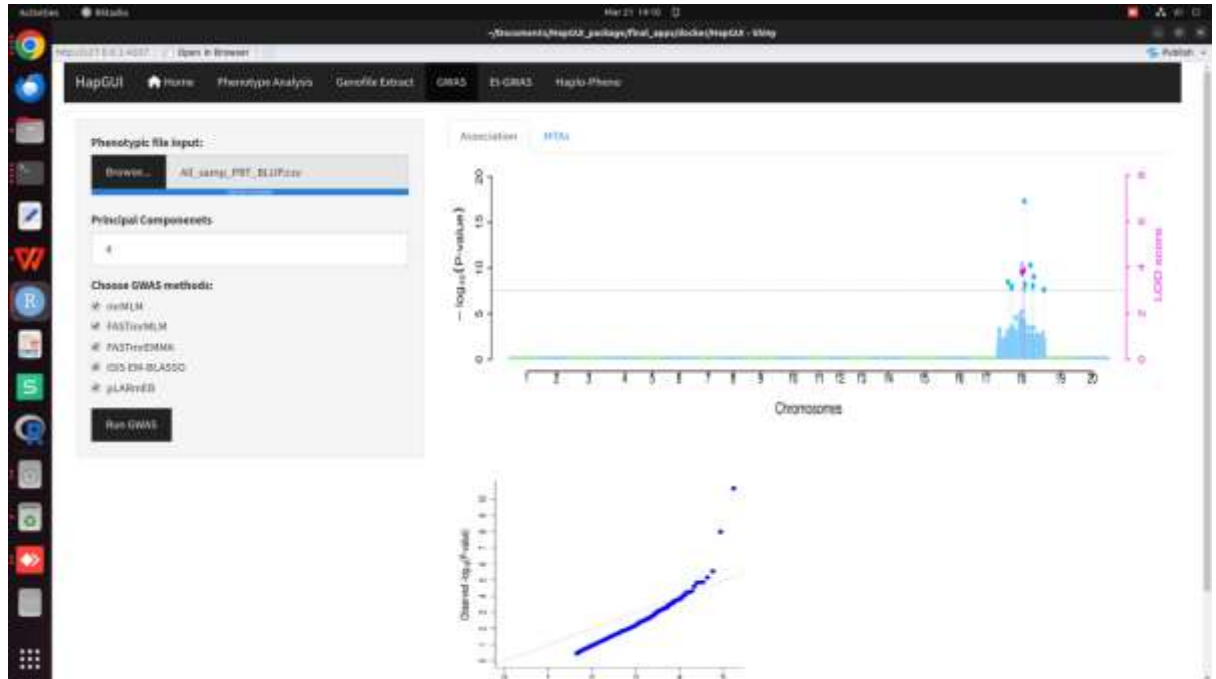
For external diploid organism

- i. Upload your phenotype file in appropriate format as mentioned in the page no. 2.
- ii. Select “External” in “Choose genotypic file” section.
- iii. Upload genotype file in vcf format in “Upload VCF File” section.
- iv. Upload General Feature Format file in gff3 format in “Upload GFF File” section.
- v. In “Choose category column” choose “subpopulation”.



GWAS

- i. Upload your phenotype file in appropriate format as mentioned in the page no. 2.
- ii. Insert a number, in Principal Components. Insert more than three till 10.
- iii. In “Choose GWAS methods”, choose your model of interest.
- iv. Then Click on “Run GWAS”.



Et-GWAS

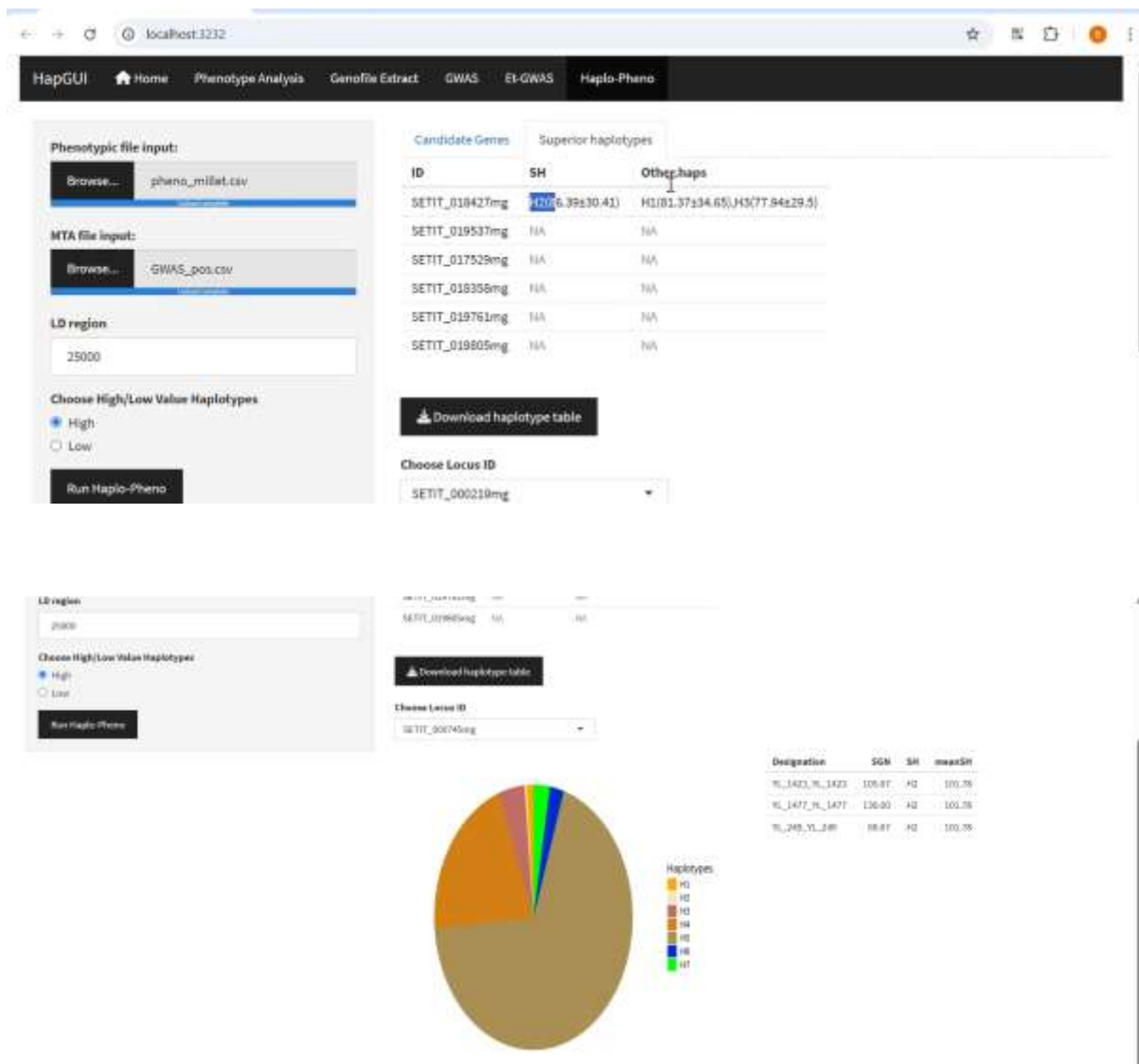
- i. Select “Bulk size:”
- ii. Select “Trait”
- iii. Upload your phenotype file in appropriate format as mentioned in the page no. 2.
- iv. Click “Run”



Haplo-pheno

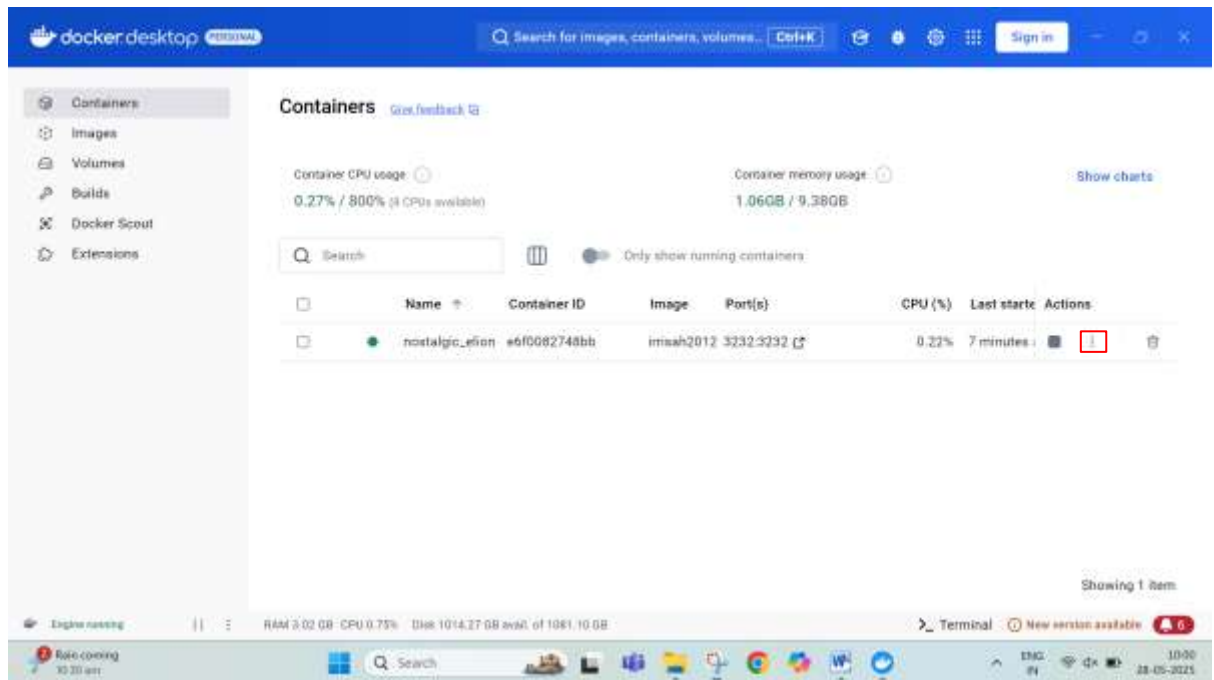
- i. Upload your phenotype file in appropriate format as mentioned in the page no. 2.
- ii. Upload the MTA file with extension “GWAS_pos” or “Et-GWAS_pos” from GWAS results, Et-GWAS_results folder.
- iii. Write LD region (only $0 \leq 250000$) for identification of candidate genes.
- iv. Choose “High” or “Low” based on the traits behaviour for identification of superior haplotypes e.g., for yield related traits “High” and disease scoring related traits “Low”.
- v. Candidate genes and Superior haplotypes table will be displayed as follows.
- vi. By clicking on “download” button, the results of candidate genes and superior haplotypes can be downloaded.

Chromosome	start	stop	strand	gene_id	QTN
1	729723	735220	+	SETIT_018427mg	1_755915
1	738793	738834	+	SETIT_018537mg	1_755915
1	747341	748929	-	SETIT_017529mg	1_755915
1	760715	763460	+	SETIT_018358mg	1_755915
1	770121	771041	+	SETIT_019761mg	1_755915
1	778137	779297	+	SETIT_019805mg	1_755915

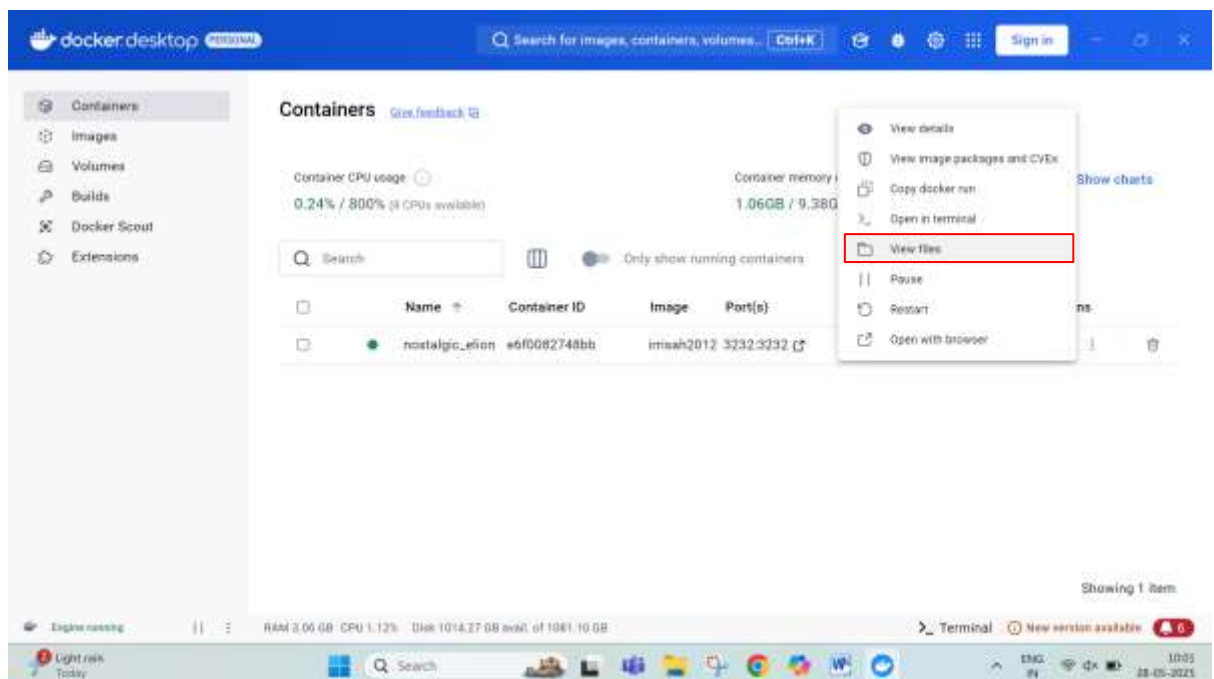


Accessing/ downloading the results of GWAS, Et-GWAS and Haplopheno

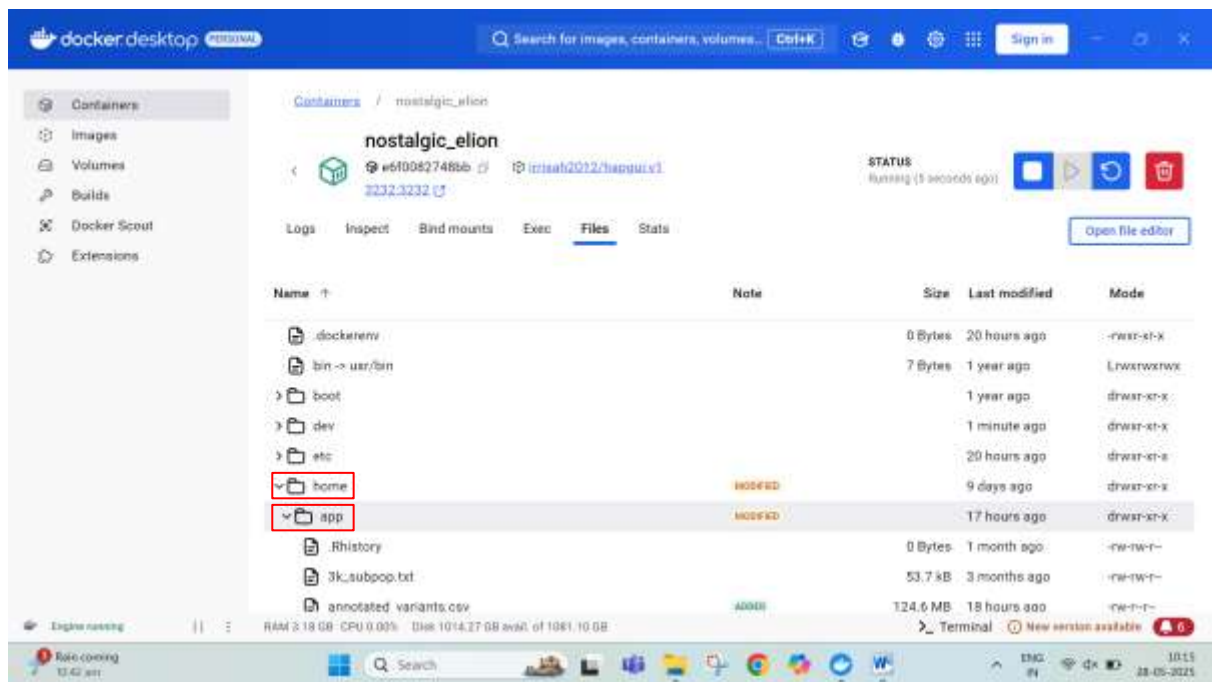
- After performing the analysis of GWAS, Et-GWAS and Haplopheno, the results can be downloaded by following the provided screenshots.
 - Click on the symbol “ ” as shown below.



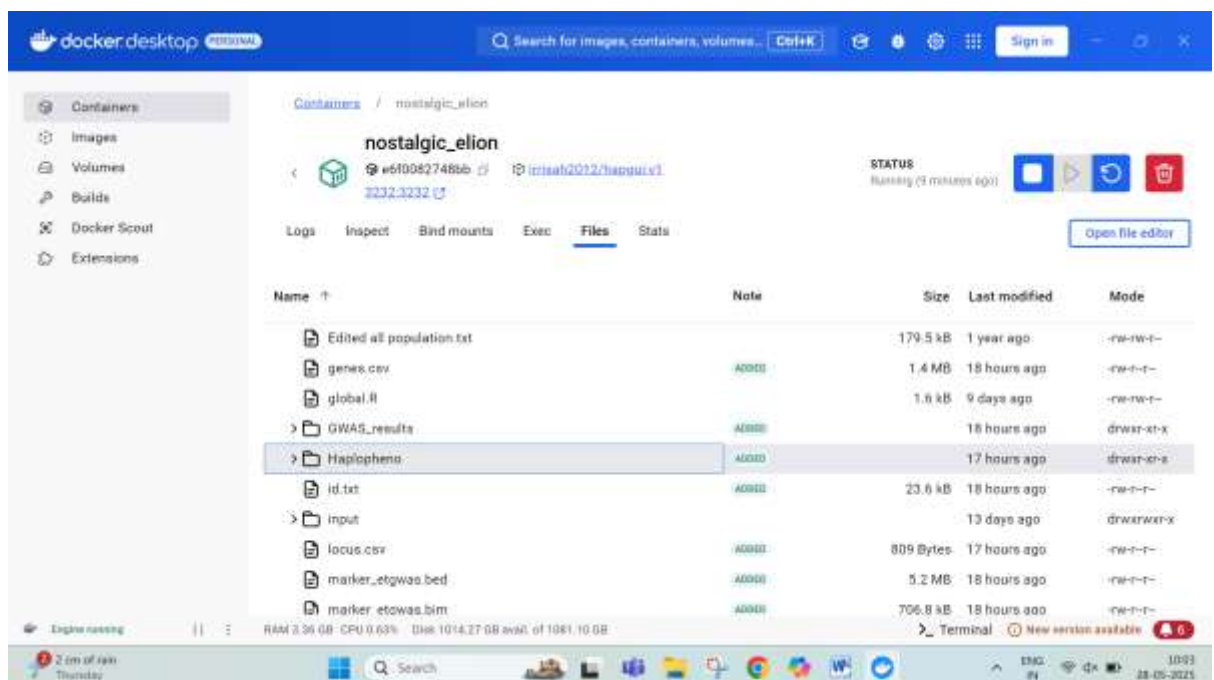
- Click on “view files”.



- Then Click on “home” and later on “app”.



- Now you'll find the results namely "GWAS_results", "EtGWAS_results" and "Haplopheno" folders where you can find respective results in respective folders.



- All these results can be downloaded by "right clicking". Then the results can be downloaded by clicking on "save" button, Delete by clicking on the "Delete" button.
- **Important Note:** In the similar way like save, please delete after saving the results, before running another analysis.

