# Package 'specmine'

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absorbance\_to\_transmittance

Convert absorbance to transmittance

# Description

Converts absorbance values to transmittance values.

# Usage

absorbance\_to\_transmittance(dataset)

# Arguments

dataset list representing the dataset from a metabolomics experiment.

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#### Value

Returns the dataset with the data points converted to transmittance values.

## **Examples**

```
## Example of converting transmittance values to absorbance values
data(cassavaPPD)
cassavaPPD = transmittance_to_absorbance(cassavaPPD)
cassavaPPD = absorbance_to_transmittance(cassavaPPD)
```

aggregate\_samples

Aggregate samples

## **Description**

Aggregate samples according to an aggregate function like mean, median, etc.

## Usage

```
aggregate_samples(dataset, indexes, aggreg.fn = "mean",
meta.to.remove = c())
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

indexes index vector with the samples that are going to be aggregated (e.g. c(1,1,2,2),

this index vector will aggregate the first two samples and the last two samples).

aggregation function (e.g. "mean", "median", etc).

meta.to.remove metadata's variables to be removed.

#### Value

Returns the dataset with the samples aggregated.

```
## Example of aggregating samples
data(cassavaPPD)
index.vector = c(1,1,2,2,2,2,2,1,1,3,3,3,3,3,1,4,4,4,4,4,5,5,6,6,6,6,6,5,5,7,7,7,7,8,8,8,8,8,9,9,10,10,10,10,10,9,9,11,11,11,11,11,
9,12,12,12,12,12,13,13,14,14,14,14,14,13,13,15,15,15,15,15,15,15,13,16,16,16,16,16)
dataset = aggregate_samples(cassavaPPD, index.vector, "mean")
```

aov\_all\_vars 7

aov_all_vars Analysis of variance
-----------------------------------

## Description

Perform analysis of variance of all variables in the dataset.

# Usage

```
aov_all_vars(dataset, column.class, doTukey = T, write.file = F,
file.out = "anova-res.csv")
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

column.class string or index indicating what metadata to use.

doTukey boolean value for do or do not TukeyHSD.

write.file boolean value indicating if a file with the results is written or not.

file.out name of the file if write.file is TRUE.

#### Value

Data frame with the results of ANOVA, with p-value, logarithm of p-value, false discovery rate (fdr) and tukey is doTukey is TRUE. The result is ordered by p-value.

## **Examples**

```
## Example of ANOVA with TukeyHSD
data(cassavaPPD)
cassavaPPD = flat_pattern_filter(cassavaPPD, "iqr", by.percent = TRUE,
red.value = 75)
result = aov_all_vars(cassavaPPD, "varieties", doTukey = FALSE)
```

```
apply_by_group Apply by group
```

#### **Description**

Apply a function to samples from a given metadata's group.

#### Usage

```
apply_by_group(dataset, fn.to.apply, metadata.var, var.value)
```

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#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

fn.to.apply function to apply (e.g. mean, max, min).

metadata.var name of the metadata's variable. var.value value of the metadata's variable.

#### Value

Returns a vector with the variables and the value of the applied function.

#### **Examples**

```
## Example of applying a function to a group
  data(cachexia)
  apply.group.result = apply_by_group(cachexia, mean, "Muscle.loss",
"control")
```

apply\_by\_groups

Apply by groups

#### **Description**

Apply a function to samples from a metadata's variable.

#### Usage

```
apply_by_groups(dataset, metadata.var, fn.to.apply = "mean",
variables = NULL, variable.bounds = NULL)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.var name of the metadata's variable.

fn.to.apply function to apply (e.g. mean, max, min).

variables allows to define which variables to calculate the stats (if numbers, indexes are

assumed).

variable.bounds

allow to define an interval of variables (if numeric).

#### Value

Returns a vector with the variables and the value of the applied function on the metadata's groups.

```
## Example of applying a function to groups
data(cachexia)
apply.groups.result = apply_by_groups(cachexia, "Muscle.loss", mean)
```

apply\_by\_sample 9

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annly	/ hv	_sample	

Apply function to samples

# Description

Applies a function to the values of each sample

## Usage

```
apply_by_sample(dataset, fn.to.apply, samples = NULL, ...)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

fn.to.apply function to apply (e.g. mean, max, min).

samples if defined restricts the application to a given set of samples.

. . . additional parameters to apply function.

#### Value

Returns a vector with the samples and the value of the applied function.

## **Examples**

```
## Example of applying a function to variables
data(cachexia)
apply.samples.result = apply_by_sample(cachexia, mean)
```

apply\_by\_variable

Apply function to variables

## **Description**

Applies a function to the values of each variable

## Usage

```
apply_by_variable(dataset, fn.to.apply, variables = NULL,
variable.bounds = NULL, samples = NULL, ...)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

fn. to. apply function to apply (e.g. mean, max, min).

variables allows to define which variables to calculate the stats (if numbers, indexes are

assumed).

variable.bounds

allow to define an interval of variables (if numeric).

samples if defined restricts the application to a given set of samples.

... additional parameters to apply function.

#### Value

Returns a vector with the variables and the value of the applied function.

## **Examples**

```
## Example of applying a function to variables
data(cachexia)
apply.variables.result = apply_by_variable(cachexia, mean)
```

background\_correction Background correction

#### **Description**

Perform background correction on the spectra.

# Usage

```
background_correction(dataset)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

#### Value

Returns the dataset with background correction performed on the data.

```
## Example of background correction
library(hyperSpec)
data(flu)
flu.converted = convert_from_hyperspec(flu)
flu.corrected = background_correction(flu.converted)
```

baseline\_correction 11

baseline\_correction Baseline correction

#### **Description**

Performs baseline correction on the dataset.

#### Usage

```
baseline_correction(dataset, method = "modpolyfit", ...)
```

#### **Arguments**

dataset method list representing the dataset from a metabolomics experiment. string representing the baseline correction method. It can be one of these methods:

- "als" Asymmetric Least Squares, baseline correction by 2nd derivative constrained weighted regression
- "fillPeaks" An iterative algorithm using suppression of baseline by means in local windows
- "irls" Iterative Restricted Least Squares, an algorithm with primary smoothing and repeated baseline suppressions and regressions with 2nd derivative constraint
- "lowpass" Low-pass filter, an algorithm for removing baselines based on Fast Fourier Transform filtering
- "medianWindow" an implementation and extension of Mark S. Friedrichs' model-free algorithm
- "modpolyfit" Modified polynomial fitting, an implementation of Chad A. Lieber and Anita Mahadevan-Jansen's algorithm for polynomial fitting
- "peakDetection" A translation from Kevin R. Coombes et al.'s MATLAB code for detecting peaks and removing baselines
- "rfbaseline" Robust Baseline Estimation, Wrapper for Andreas F. Ruckstuhl, Matthew P. Jacobson, Robert W. Field, James A. Dodd's algorithm based on LOWESS and weighted regression
- "rollingBall" Ideas from Rolling Ball algorithm for X-ray spectra by M. A. Kneen and H. J. Annegarn. Variable window width has been left out

Additional parameters of the baseline correction method.

#### Value

Returns the dataset with the data's baseline corrected.

```
## Example of baseline correction
data(cassavaPPD)
dataset.corrected = baseline_correction(cassavaPPD, method = "modpolyfit")
```

12 boxplot\_vars\_factor

boxplot_variables Boxplot of	of variables
------------------------------	--------------

#### **Description**

Boxplot of each variable of the dataset.

#### Usage

```
boxplot_variables(dataset, variables = NULL, samples = NULL,
horizontal = T, col = "lightblue", nchar.label = 10,
cex.axis = 0.8, ...)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment. variables vector with the variables names or a NULL value indicating all variables. samples vector with the samples names or a NULL value indicating all samples. horizontal boolean value indicating if the boxplots should be horizontal. col string that represents the color of the bodies of the boxplots. nchar.label number of characters to display the variables' names. numeric value that indicates the amount by which the axis is magnified relative cex.axis to the default. additional parameters of boxplot function.

#### **Examples**

```
## Example of showing the boxplot of a few variables
data(cachexia)
boxplot_variables(cachexia, variables = c("Creatine", "Serine", "Lactate"))
```

boxplot\_vars\_factor Boxplot of variables with metadata's variable factors

#### **Description**

Boxplot of variables with metadata's variable factors from the dataset.

## Usage

```
boxplot_vars_factor(dataset, meta.var, variables = NULL,
samples = NULL, horizontal = F, nchar.label = 10, col = NULL,
vec.par = NULL, cex.axis = 0.8, ylabs = NULL, ...)
```

cachexia 13

#### Arguments

dataset list representing the dataset from a metabolomics experiment.

meta.var metadata's variable.

variables vector with the variables names or a NULL value indicating all variables. samples vector with the samples names or a NULL value indicating all samples.

horizontal boolean value indicating if the boxplots should be horizontal.

nchar.label number of characters to display the variables' names.

string that represents the color of the bodies of the boxplots.

vec.par vector with the disposition of the boxplots (rows, columns).

cex.axis numeric value that indicates the amount by which the axis is magnified relative

to the default.

ylabs y-axis labels.

... additional parameters of boxplot function.

## **Examples**

```
## Example of showing the boxplot factors of a few variables
data(cachexia)
boxplot_vars_factor(cachexia, "Muscle.loss", variables = c("Creatine", "Serine",
"Lactate"))
```

cachexia

Human Cachexia data

## **Description**

Cachexia is a complex metabolic syndrome associated with an underlying illness (such as cancer) and characterized by loss of muscle with or without loss of fat mass (Evans et al., 2008). A total of 77 urine samples were collected being 47 of them patients with cachexia, and 30 control patients.

#### Usage

data(cachexia)

#### **Format**

An object of class "list"

#### Source

MetaboAnalyst

#### References

Eisner et al. (2010) Learning to predict cancer-associated skeletal muscle wasting from 1h-nmr profiles of urinary metabolites Metabolomics 7:25-34

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#### **Examples**

```
data(cachexia)
    sum_dataset(cachexia)
```

cassavaPPD

Cassava Postharvest Physiological Deterioration

## **Description**

Cassava is a root well known and widely cultivated in tropical and subtropical regions for its starchy tuberous root, which is a great source of carbohydrates. It also has a great variety of applications, like animal feeding, culinary or alcoholic beverages. In some countries, cassava has also been tested as an ethanol biofuel feedstock.

## Usage

```
data(cassavaPPD)
```

#### **Format**

An object of class "list"

#### References

Uarrota et al. (2014) Metabolomics combined with chemometric tools (pca, hca, pls-da and svm) for screening cassava (manihot esculenta crantz) roots during postharvest physiological deterioration. Food Chemistry 161:67-78

#### **Examples**

```
data(cassavaPPD)
sum_dataset(cassavaPPD)
```

check\_dataset

Check dataset

# Description

Check if the dataset is valid and if not give the proper error message.

## Usage

```
check_dataset(dataset)
```

#### **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

clustering 15

## Value

Message saying if the dataset is valid or invalid, in the last case also gives the reason.

## **Examples**

```
## Example checking a dataset
data(cachexia)
check_dataset(cachexia)
```

clustering

Perform cluster analysis

## **Description**

Perform cluster analysis on the dataset.

## Usage

```
clustering(dataset, method = "hc", distance = "euclidean",
type = "samples", num.clusters = 5, clustMethod = "complete")
```

# Arguments

dataset	list representing the dataset from a metabolomics experiment.
method	a string describing the method of clustering. Possible types are:
	<ul><li> "hc" perform hierarchical clustering.</li><li> "kmeans" perform kmeans clustering.</li></ul>
distance	the distance measure to be used to compute the distances between the rows of a data matrix. Possible types are "euclidean", "manhattan", "pearson" or "spearman". Only for hierarchical clustering.
type	a string indicating if cluster analysis will be performed on samples ("samples") or on variables ("variables").
num.clusters	the number of clusters in k-means cluster analysis.
clustMethod	Cluster method for hierarchical clustering.

#### Value

An object of class kmeans or helust with the clustering results.

## **Description**

Compare two regions of a dataset by samples.

## Usage

```
compare_regions_by_sample(dataset1, dataset2, fn.to.apply,
samples = NULL, ...)
```

# Arguments

dataset1 list representing the dataset from a metabolomics experiment.

dataset2 list representing the dataset from a metabolomics experiment.

fn.to.apply function to apply (e.g. mean, max, min).

samples if defined restricts the application to a given set of samples.

... additional parameters to apply.by.sample function.

#### Value

Returns a data.frame with the results of the function applied to the samples and the ration between the two datasets.

## **Examples**

```
## Example of comparing regions by sample
data(cachexia)
subset1 = subset_x_values(cachexia, 1:31, by.index = TRUE)
subset2 = subset_x_values(cachexia, 32:63, by.index = TRUE)
comp.regions.result = compare_regions_by_sample(subset1, subset2, mean)
```

```
convert_from_chemospec
```

Convert from ChemoSpec

## **Description**

Convert the dataset in the ChemoSpec format to a dataset of this package.

#### Usage

```
convert_from_chemospec(csobj, type = "undefined",
description = "")
```

## **Arguments**

csobj ChemoSpec object representing the dataset.
type string representing the type of the data.

description string representing the description of the dataset.

#### Value

Returns a list representing the dataset converted.

## **Examples**

convert\_from\_hyperspec

Convert from hyperspec

## **Description**

Convert the dataset in the hyperspec format to a dataset of this package.

## Usage

```
convert_from_hyperspec(hsobj, type = "undefined",
description = "")
```

## **Arguments**

hsobj hyperspec object representing the dataset. type string representing the type of the data.

description string representing the description of the dataset.

## Value

Returns a list representing the dataset converted.

## **Examples**

```
## Example of converting a dataset from hyperspec
library(hyperSpec)
data(flu)
dataset = convert_from_hyperspec(flu, type = "undefined",
    description = "some description")
```

convert\_to\_factor

Convert metadata to factor

# Description

Convert a metadata's variable to factor.

#### Usage

```
convert_to_factor(dataset, metadata.var)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

metadata.var name of the metadata's variable.

## Value

Returns the dataset with the metadata's variable converted to factor.

# **Examples**

```
## Example of converting a metadata's variable to factor
data(cassavaPPD)
dataset = convert_to_factor(cassavaPPD, "ppds")
```

convert\_to\_hyperspec Convert to hyperspec

# Description

Convert a dataset to an hyperspec object.

## Usage

```
convert_to_hyperspec(dataset)
```

correlations\_dataset 19

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

#### Value

Returns an hyperspec object representing the dataset converted.

#### **Examples**

```
## Example of converting a dataset to an hyperspec object
data(cassavaPPD)
hyperspec.cassava = convert_to_hyperspec(cassavaPPD)
```

# Description

Calculate the correlations of all variables or samples in the dataset.

## Usage

```
correlations_dataset(dataset, method = "pearson", by.var = T)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

method correlation method, it can be "pearson", "kendall" or "spearman".

by . var if TRUE then the correlations of the variables will be calculated, if not then the

correlations of the samples will be calculated.

#### Value

Returns the correlation matrix

```
## Example of correlations of variables
data(cachexia)
corr.result = correlations_dataset(cachexia,
method = "pearson", by.var = TRUE)
```

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correlations\_test

Correlations test

#### Description

Performs correlations test to the whole dataset.

## Usage

```
correlations_test(dataset, method = "pearson", by.var = T,
alternative = "two.sided")
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

method correlation method, it can be "pearson", "kendall" or "spearman".

by . var if TRUE then the correlations of the variables will be calculated, if not then the

correlations of the samples will be calculated.

alternative alternative argument from cor.test of stats package. Can be "two.sided", "less"

or "greater".

#### Value

Returns a matrix with the correlation values and the p-values

#### **Examples**

```
## Not run:
    ## Example of correlations test of variables (computationally heavy)
    data(cachexia)
    corr.result = correlations_test(cachexia,
method = "pearson", by.var = FALSE)
## End(Not run)
```

correlation\_test

Correlation test of two variables or samples

## **Description**

Performs correlations test of two variables or samples from the dataset.

# Usage

```
correlation_test(dataset, x, y, method = "pearson",
alternative = "two.sided", by.var = T)
```

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## Arguments

dataset list representing the dataset from a metabolomics experiment.

x first variable or sample.y second variable or sample.

method correlation method, it can be "pearson", "kendall" or "spearman".

alternative alternative argument from cor.test of stats package. Can be "two.sided", "less"

or "greater".

by . var if TRUE then the correlations of the variables will be calculated, if not then the

correlations of the samples will be calculated.

#### Value

Returns the correlation result from cor.test function of stats package.

#### **Examples**

```
## Example of correlations test of variables
data(cachexia)
corr.result = correlation_test(cachexia, "Serine", "Creatine", method = "pearson",
by.var = TRUE)
```

count\_missing\_values Count missing values

# Description

Counts the missing values on the dataset.

#### Usage

```
count_missing_values(dataset)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

#### Value

Returns the number of missing values on the dataset.

```
## Example of counting the missing values
data(cachexia)
count_missing_values(cachexia)
```

count\_missing\_values\_per\_sample

Count missing values per sample

## **Description**

Counts the missing values on each sample of the dataset.

#### Usage

```
count_missing_values_per_sample(dataset, remove.zero = T)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

remove.zero boolean value indicating if the results of samples with no missing value are

removed.

#### Value

Returns a vector with the number of missing values on each sample.

## **Examples**

```
## Example of counting the missing values on each sample
data(cachexia)
cachexia$data[10,10] = NA
count_missing_values_per_sample(cachexia)
```

```
count_missing_values_per_variable
```

Count missing values per variable

#### **Description**

Counts the missing values on each variable of the dataset.

#### Usage

```
count_missing_values_per_variable(dataset, remove.zero = T)
```

#### Arguments

dataset list representing the dataset from a metabolomics experiment.

remove.zero boolean value indicating if the results of variables with no missing value are

removed.

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#### Value

Returns a vector with the number of missing values on each sample.

## **Examples**

```
## Example of counting the missing values on each variable
data(cachexia)
cachexia$data[10,10] = NA
count_missing_values_per_variable(cachexia)
```

create\_dataset

Create dataset

#### **Description**

Create a dataset from existing objects

#### Usage

```
create_dataset(datamatrix, type = "undefined", metadata = NULL,
description = "", sample.names = NULL, x.axis.values = NULL,
label.x = NULL, label.values = NULL, xSet = NULL)
```

# Arguments

datamatrix

matrix with numerical data: rows are assumed to be variables and columns assumed to be samples

sumed to be samples.

type

type of data: string that can be one of the following:

- nmr-spectra
- · nmr-peaks
- ir-spectra
- · uvv-spectra
- raman-spectra
- fluor-spectra
- ms-spectra
- lcms-peaks
- · gcms-peaks
- integrated-data
- · concentrations
- · undefined

metadata

data frame with the dataset's metadata: columns represent each metadata variable and rows represent the value of the metadata for the sample.

description

string with a short description of the dataset.

24 cubic\_root\_transform

sample.names vector with sample names, if NULL then the column names of datamatrix or

sequential numbers will be used.

x.axis.values vector with the x axis values, if NULL then the row names of datamatrix or

sequential numbers will be used.

xSet xcmsSet object from xcms package to store the reading and preprocessing re-

sults from MS spectra. Used for metabolite identification purposes.

#### Value

list representing the dataset:

data matrix with the data type type of the data

description short description of the dataset

metadata data frame with the metadata variables labels list with labels of x axis and values

xSet xcmsSet object

# **Examples**

```
## Not run:
    ## Example of creating a dataset
    dataset = create_dataset(datamatrix, type = "nmr-spectra",
metadata = dataset.metadata, description = "shortdescription",
sample.names = NULL, x.axis.values = NULL, label.x = "ppm",
label.values = "intensity")
## End(Not run)
```

 ${\tt cubic\_root\_transform} \quad \textit{Cubic root transformation}$ 

## **Description**

Performs cubic root transformation on the data matrix.

#### Usage

```
cubic_root_transform(datamat)
```

#### **Arguments**

datamat data matrix.

dataset\_from\_peaks 25

## Value

Returns the data matrix with the cubic root transformation applied.

#### **Examples**

```
## Example of cubic root transformation
data(cachexia)
datamat.cubic = cubic_root_transform(cachexia$data)
```

dataset\_from\_peaks

Dataset from peaks

# Description

Converts a peak list to a dataset.

## Usage

```
dataset_from_peaks(sample.list, metadata = NULL,
description = "", type = "nmr-peaks")
```

#### **Arguments**

sample.list list with the peaks from each sample.

metadata data frame with the associated metadata.

description string with the description of the dataset.

type string that represents the type of the data.

#### Value

Returns the dataset from the peak list.

```
## Not run:
    ## Example of converting a peak list to a dataset (computationally heavy)
    data(propolisSampleList)
    dataset = dataset_from_peaks(propolisSampleList, metadata = NULL,
        description = "some text", type = "nmr-peaks")
## End(Not run)
```

26 dendrogram\_plot

data_correction	Data correction
uata_con rection	Daia correction

#### **Description**

Perform spectra corrections with 3 different methods.

## Usage

```
data_correction(dataset, type = "background",
method = "modpolyfit", ...)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

type string that represents the type of correction that will be applied to the spectra.

The three possible types are: "background", to perform background correction; "offset", to perform offset correction; and "baseline", to perform baseline cor-

rection.

method string parameter of baseline correction indicating the correction method.

... additional parameters of baseline correction.

# Value

Returns the dataset with the spectra corrected.

## **Examples**

```
## Example of data correction using background correction
data(cassavaPPD)
dataset.corrected = data_correction(cassavaPPD, type = "background")
```

dendrogram\_plot

Plot dendrogram

## **Description**

Plot dendrogram of hierarchical clustering results.

#### Usage

```
dendrogram_plot(dataset, hc.result, column.metadata = 1,
labels = NULL, ...)
```

dendrogram\_plot\_col 27

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

hc.result object of class helust with the clustering results.

column.metadata

string or index indicating what metadata to use to name the leafs.

labels vector with the leaf names (optional).

... other parameters for plotting.

## **Examples**

```
## Example of a dendrogram
data(cachexia)
hc.result = hierarchical_clustering(cachexia)
dendrogram_plot(cachexia, hc.result)
```

dendrogram\_plot\_col

Plot dendrogram

## **Description**

Plot dendrogram of hierarchical clustering results with different colors

#### Usage

```
dendrogram_plot_col(dataset, hc.result, classes.col, title = "",
lab.cex = 1, leg.pos = "topright",...)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

hc.result object of class helust with the clustering results.

classes.col string or index indicating what metadata to use to name the leafs.

title title of dendrogram.

lab.cex the magnification to be used for x and y labels relative to the current setting of

cex.

leg.pos position of the legend.

... other parameters for plotting.

```
## Example of colored dendrogram
data(cachexia)
hc.result = hierarchical_clustering(cachexia)
dendrogram_plot_col(cachexia, hc.result, "Muscle.loss",
title = "Example")
```

28 feature\_selection

feature\_selection

Perform feature selection

#### **Description**

Perform feature selection on the dataset.

#### Usage

```
feature_selection(dataset, column.class, method = "rfe",
functions, validation = "cv", repeats = 5, number = 10,
subsets = 2^(2:4))
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

column.class string or index indicating what metadata to use.

method method used for feature selection. Possible values are "rfe" (recursive feature

elimination) and "filter" (Selection by filter - sbf) from caret's package.

functions a list of functions for model fitting, prediction and variable importance/filtering. validation the external resampling method: boot, cv, LOOCV or LGOCV (for repeated

training/test splits.

repeats for repeated k-fold cross-validation only: the number of complete sets of folds

to compute.

number either the number of folds or number of resampling iterations.

subsets a numeric vector of integers corresponding to the number of features that should

be retained (rfe only).

#### Value

caret's result from rfe or sbf.

filter\_feature\_selection 29

```
filter_feature_selection
```

Perform selection by filter

# Description

Perform selection by filter using univariate filters, from caret's package.

## Usage

```
filter_feature_selection(datamat, samples.class,
functions = caret::rfSBF, method = "cv", repeats = 5)
```

## **Arguments**

data matrix from dataset.

samples.class string or index indicating what metadata to use.

functions a list of functions for model fitting, prediction and variable filtering.

method the external resampling method: boot, cv, LOOCV or LGOCV (for repeated

training/test splits.

repeats for repeated k-fold cross-validation only: the number of complete sets of folds

to compute.

## Value

A caret's sbf object with the result of selection by filter.

30 first\_derivative

find\_equal\_samples

Find equal samples

## **Description**

Finds samples that have the same peak values - x and y (equal data frames)

## Usage

```
find_equal_samples(sample.list)
```

## **Arguments**

sample.list list of data frames with the samples' peaks.

#### Value

Returns a dataframe with two columns indicating which pair of samples are equal.

## **Examples**

```
## Example of finding equal samples
data(propolisSampleList)
equal.samples = find_equal_samples(propolisSampleList)
```

first\_derivative

First derivative

## **Description**

Calculates the first derivative of the data.

## Usage

```
first_derivative(dataset)
```

#### **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Return the dataset with the first derivative of the data calculated.

```
## Example of calculate the first derivative
data(cassavaPPD)
cassava.derivative = first_derivative(cassavaPPD)
```

flat\_pattern\_filter 31

flat\_pattern\_filter Flat pattern filter

## **Description**

Performs a flat pattern filter over the dataset.

#### Usage

```
flat_pattern_filter(dataset, filter.function = "iqr",
by.percent = T, by.threshold = F, red.value = 0)
```

#### Arguments

dataset

list representing the dataset from a metabolomics experiment.

filter.function

filter function. It can be:

- "iqr" Interquantile Range
- "rsd" Relative Standard Deviation
- "sd" Standard Deviation
- "mad" Median Absolute Deviation
- "mean" Mean
- "median" Median

by.percent

boolean value, if T the number of variables to filter will be a percentage of the number of variables in the dataset; percentage is given by the "red.value" parameter

paramen

by.threshold

boolean value, if T, defines filtering will select variables where values of filtering function are below a given threshold. Threshold is defined by red.value that defines the minimum value of the function needed to keep the variable.

red.value

it can be the percentage or the threshold number. If red.value = "auto", will calculate number of variables to remove automatically

#### Value

Returns the dataset with the data filtered.

```
## Example of flat pattern filter
data(cassavaPPD)
dataset.filtered = flat_pattern_filter(cassavaPPD, "iqr", by.percent = TRUE,
    red.value = 20)
```

32 fold\_change\_var

fold\_change

Fold change analysis

## **Description**

Perform fold change analysis on the dataset.

## Usage

```
fold_change(dataset, metadata.var, ref.value,
threshold.min.fc = NULL, write.file = F,
file.out = "fold_change.csv")
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.var metadata to use to calculate the fold change.

ref.value class name to indicate the initial value.

threshold.min.fc

minimum threshold of the fold change value.

write.file boolean value to write or not a file with the results.

file.out name of the file.

#### Value

Table of results with fold change and log2 of fold change.

## **Examples**

```
## Example of fold change
data(cachexia)
fold.change.results = fold_change(cachexia, "Muscle.loss",
"control", write.file = FALSE)
```

fold\_change\_var

Fold change applied on two variables

## **Description**

Fold change applied on two variables. Instead of having the difference of the variables on two groups, we have the difference of the groups on two variables.

get\_data 33

#### Usage

```
fold_change_var(dataset, metadata.var, variables,
threshold.min.fc = NULL, write.file = F,
file.out = "fold_change_reverse.csv")
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.var metadata to use to calculate the fold change.

variables vector with two positions containing the name of the variables.

threshold.min.fc

minimum threshold of the fold change value.

write.file boolean value to write or not a file with the results.

file.out name of the file.

#### Value

Table of results with fold change and log2 of fold change.

#### **Examples**

```
## Example of fold change reverse
data(cachexia)
fold.change.results = fold_change_var(cachexia, "Muscle.loss",
c("Creatine", "Serine"))
```

get\_data

Get data

# Description

Get the data matrix from dataset

#### Usage

```
get_data(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the data matrix

34 get\_data\_value

#### **Examples**

```
## Example of getting the data matrix
data(cachexia)
cachexia.dm = get_data(cachexia)
```

get\_data\_as\_df

Get data as data frame

## **Description**

Get the data matrix from the dataset as a data frame.

## Usage

```
get_data_as_df(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the data matrix from the dataset as a data.frame object.

#### **Examples**

```
## Example of getting the data matrix as data frame
data(cachexia)
cachexia.dt = get_data_as_df(cachexia)
```

get\_data\_value

Get data value

## **Description**

Get a data value given the x-axis labels and the sample

#### Usage

```
get_data_value(dataset, x.axis.val, sample, by.index = F)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

x.axis.val index or name of the x-axis value. sample index or name of the sample.

by . index boolean value indicating if the x-axis value and sample are represented as index

or not.

get\_data\_values 35

## Value

Returns a numeric with the data point value.

# **Examples**

get\_data\_values

Get data values

## Description

Gets the values of all samples in the dataset given a set of x axis names or indexes.

## Usage

```
get_data_values(dataset, x.axis.val, by.index = FALSE)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

x.axis.val vector with the values of the x axis (could be names or indexes).

by.index boolean value indicating if the x.axis.val is a vector of indexes or not.

#### Value

Returns a matrix with the values of all samples in the specified x axis.

```
## Example of getting a metadata value
data(cachexia)
data.values = get_data_values(cachexia, c("Creatine", "Serine", "Lactate"),
by.index = FALSE)
```

36 get\_metadata\_value

get\_metadata

Get metadata

# Description

Get the metadata from the dataset

## Usage

```
get_metadata(dataset)
```

# Arguments

dataset

list representing the dataset from a metabolomics experiment.

#### Value

returns a data frame with the metadata.

## **Examples**

```
## Example of getting the metadata
data(cachexia)
cachexia.mt = get_metadata(cachexia)
```

get\_metadata\_value

Get metadata value

## **Description**

Get the metadata value

## Usage

```
get_metadata_value(dataset, variable, sample)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

variable index or name of the metadata variable.

sample index or name of the sample.

## Value

Return the corresponding metadata value of the sample.

get\_metadata\_var 37

### **Examples**

```
## Example of getting a metadata value
data(cachexia)
metadata.value = get_metadata_value(cachexia, "Muscle.loss", "PIF_178")
```

get\_metadata\_var

Get metadata variable

## Description

Get the values of a metadata variable from the dataset.

# Usage

```
get_metadata_var(dataset, var)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

var index or name of the metadata variable.

#### Value

Returns a vector with the values of the metadata variable.

# **Examples**

```
## Example of getting a metadata variable
data(cachexia)
metadata.variable = get_metadata_var(cachexia, "Muscle.loss")
```

get\_peak\_values

Get peak values

## **Description**

Gets the peak values from a data frame of samples' peaks.

# Usage

```
get_peak_values(samples.df, peak.val)
```

#### **Arguments**

samples.df data frame with the samples' peaks.

peak.val peak name.

## Value

Returns a vector with the peak values.

## **Examples**

```
## Example of getting the peak values
data(propolis)
peak.values = get_peak_values(propolis$data, 2.11)
```

```
get_samples_names_dx Get sample's names from DX files
```

# Description

Function to get the names of the DX files from a folder.

## Usage

```
get_samples_names_dx(foldername)
```

# **Arguments**

foldername string with the path of the data folder.

#### Value

Returns a vector with the sample's names.

```
## Not run:
    ## Example of getting DX sample's names
    dx.sample.names = get_samples_names_dx("data")
## End(Not run)
```

```
get_samples_names_spc Get sample's names from SPC files
```

## **Description**

Function to get the names of the SPC files from a folder.

## Usage

```
get_samples_names_spc(foldername)
```

## **Arguments**

foldername

string with the path of the data folder.

#### Value

Returns a vector with the sample's names.

# **Examples**

```
## Not run:
    ## Example of getting SPC sample's names
    spc.sample.names = get_samples_names_spc("data")
## End(Not run)
```

get\_sample\_names

Get sample names

# Description

Get the sample names from the dataset.

### Usage

```
get_sample_names(dataset)
```

#### **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

## Value

Returns a vector with the sample names.

40 get\_value\_label

### **Examples**

```
## Example of getting the sample names
data(cachexia)
sample.names = get_sample_names(cachexia)
```

get\_type

Get type of data

## **Description**

Get the type of the data from the dataset

## Usage

```
get_type(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns a string with the type of the data.

## **Examples**

```
## Example of getting the type of the data
data(cachexia)
type = get_type(cachexia)
```

get\_value\_label

Get value label

# Description

Get the value label from the dataset

#### Usage

```
get_value_label(dataset)
```

# Arguments

dataset

list representing the dataset from a metabolomics experiment.

get\_x\_label 41

## Value

Returns a string with the value label.

## **Examples**

```
## Example of getting the value label
data(cassavaPPD)
value.label = get_value_label(cassavaPPD)
```

get\_x\_label

Get x-axis label

## **Description**

Get the x-axis label from the dataset.

## Usage

```
get_x_label(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns a string with the x-axis label.

## **Examples**

```
## Example of getting the x-axis label
data(cassavaPPD)
x.label = get_x_label(cassavaPPD)
```

get\_x\_values\_as\_num

Get x-axis values as numbers

## **Description**

Get the x-axis values from the dataset as numbers.

#### Usage

```
get_x_values_as_num(dataset)
```

42 get\_x\_values\_as\_text

### **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns a numeric vector with the x-axis values, if the variable labels are not all numeric then an error message is shown.

# **Examples**

```
## Example of getting the x-axis values as numbers
data(cassavaPPD)
xvalues.numeric = get_x_values_as_num(cassavaPPD)
```

```
get_x_values_as_text Get x-axis values as text
```

# **Description**

Get the x-axis values from the dataset as text.

## Usage

```
get_x_values_as_text(dataset)
```

# Arguments

dataset

list representing the dataset from a metabolomics experiment.

### Value

Returns a character vector with the x-axis values.

```
## Example of getting the x-axis values as text
data(cassavaPPD)
xvalues.text = get_x_values_as_text(cassavaPPD)
```

group\_peaks 43

group_peaks Group peaks
-------------------------

## **Description**

Group peaks with peak alignment.

# Usage

```
group_peaks(sample.list, type, method = "own", metadata = NULL,
samp.classes = 1, description = "", label.x = NULL,
label.values = NULL, step = 0.03)
```

### **Arguments**

sample.list list containing the sample's data. type of the data. type method method of peak alignment. Can be "own" or "metaboanalyst", which the later is for using the peak alignment used in MetaboAnalyst software. data frame containing the metadata. metadata the metadata's variable to be used in the MetaboAnalyst method. samp.classes description short description of the data. label.x the label for the x values. label.values the label for the y values.

#### Value

step

Returns a dataset with the peaks of the data aligned.

## **Examples**

```
## Not run:
    ## Example of grouping peaks (computationally heavy)
    data(propolisSampleList)
    peaks.ds = group_peaks(propolisSampleList, "nmr-peaks", method = "own",
    metadata = NULL, description = "short description",
label.x = "ppm", label.values = "intensity", step = 0.03)
## End(Not run)
```

step value for the peak alignment process.

heatmap\_correlations Correlations heatmap

## **Description**

Plots a heatmap with the correlations.

#### Usage

```
heatmap_correlations(correlations, col = NULL, ...)
```

#### **Arguments**

```
correlations correlation matrix

col colors to be used on heatmap.

... extra parameters to visual purposes.
```

#### **Examples**

```
## Example of correlations heatmap
data(cachexia)
correlations = correlations_dataset(cachexia)
heatmap_correlations(correlations)
```

hierarchical\_clustering

Perform hierarchical clustering analysis

#### **Description**

Perform hierarchical clustering analysis on the dataset.

## Usage

```
hierarchical_clustering(dataset, distance = "euclidean",
clustMethod = "complete", hc.type = "samples")
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

distance the distance measure to be used to compute the distances between the rows of a

data matrix. Possible types are "euclidean", "manhattan", "pearson" or "spear-

man".

clustMethod the agglomeration method to be used. Possible values are "ward", "single",

"complete", "average", "mcquitty", "median" or "centroid".

hc.type a string indicating if hierarchical cluster analysis will be performed on samples

("samples") or on variables ("variables")

impute\_nas\_knn 45

## Value

An object of class helust with the clustering results.

# **Examples**

```
## Example of hierarchical clustering
data(cachexia)
hc.result = hierarchical_clustering(cachexia,
    distance = "euclidean", clustMethod = "complete",
    hc.type = "samples")
```

impute\_nas\_knn

Impute missing values with KNN

# Description

Impute missing values with KNN

## Usage

```
impute_nas_knn(dataset, k = 10, ...)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

k the number of nearest neighbors.

... additional values to impute.knn function.

### Value

Returns the dataset with no missing values.

```
## Example of NA imputation with knn
data(propolis)
dataset = impute_nas_knn(propolis, k=10)
```

impute\_nas\_mean

## **Description**

Impute missing values with linear approximation.

## Usage

```
impute_nas_linapprox(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

## Value

Returns the dataset with no missing values.

# **Examples**

```
## Example of NA imputation with linear approximation
data(propolis)
dataset = impute_nas_linapprox(propolis)
```

impute\_nas\_mean

Impute missing values with mean

## **Description**

Impute missing values with mean

### Usage

```
impute_nas_mean(dataset)
```

#### **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the dataset with no missing values.

```
## Example of NA imputation with mean
data(propolis)
propolis = impute_nas_mean(propolis)
```

impute\_nas\_median 47

impute\_nas\_median

Impute missing values with median

## **Description**

Impute missing values with median

# Usage

```
impute_nas_median(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the dataset with no missing values.

# **Examples**

```
## Example of NA imputation with median
data(propolis)
propolis = impute_nas_median(propolis)
```

impute\_nas\_value

Impute missing values with value replacement

## **Description**

Impute missing values with value replacement.

## Usage

```
impute_nas_value(dataset, value)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

value value to replace the missing values.

## Value

Returns the dataset with no missing values.

is\_spectra

### **Examples**

```
## Example of NA imputation with value replacing
data(propolis)
propolis = impute_nas_value(propolis, 0.0005)
```

```
indexes_to_xvalue_interval
```

Get the x-values of a vector of indexes

# Description

Returns x-values corresponding to a vector of indexes (only to numerical values - spectra)

## Usage

```
indexes_to_xvalue_interval(dataset, indexes)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

indexes numeric vector containing the indexes.

#### Value

Returns a numeric vector with the interval of x-values from the indexes vector

## **Examples**

```
## Example of getting the interval of x-values from indexes
data(cassavaPPD)
xvalue.interval = indexes_to_xvalue_interval(cassavaPPD, c(10,50))
```

is\_spectra

Check type of data

## **Description**

Check if the dataset is from spectral data where x.values are numeric.

# Usage

```
is_spectra(dataset)
```

#### **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

kmeans\_clustering 49

## Value

Returns a boolean indicating if the dataset is from spectral data or not.

# **Examples**

```
## Example of checking if the dataset is from spectral data
data(cassavaPPD)
is_spectra(cassavaPPD)
```

kmeans\_clustering

Perform k-means clustering analysis

# Description

Perform k-means clustering analysis on the dataset.

## Usage

```
kmeans_clustering(dataset, num.clusters, type = "samples")
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

num. clusters the number of clusters.

type a string indicating if k-means will be performed on samples ("samples") or on

variables ("variables")

#### Value

An object of class kmeans with the clustering results.

```
## Example of kmeans clustering
data(cachexia)
kmeans.result = kmeans_clustering(cachexia,
num.clusters = 4, type = "samples")
```

50 kmeans\_result\_df

kmeans\_plot

Plot kmeans clusters

## **Description**

Plot for each formed cluster, in grey the values of all samples of that cluster and in blue the median of that samples.

#### Usage

```
kmeans_plot(dataset, kmeans.result)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

 ${\tt kmeans.result} \quad object \ of \ class \ kmeans \ with \ the \ clustering \ results.$ 

## **Examples**

```
## Example of kmeans plot - dataset filtered for performance purposes
data(cassavaPPD)
cassavaPPD = flat_pattern_filter(cassavaPPD, "iqr", by.percent = TRUE,
red.value = 70)
kmeans.result = kmeans_clustering(cassavaPPD, 3)
kmeans_plot(cassavaPPD, kmeans.result)
```

kmeans\_result\_df

Show cluster's members

## **Description**

Show for each cluster from kmeans analysis the sample names belonging to them.

## Usage

```
kmeans_result_df(kmeans.result)
```

## **Arguments**

kmeans.result object of class kmeans with the clustering results.

## Value

Data frame with the clusters and the samples' names that belong to each one.

kruskalTest\_dataset 51

#### **Examples**

```
## Example of showing kmeans cluster's members
data(cassavaPPD)
kmeans.result = kmeans_clustering(cassavaPPD, 3)
kmeans_result_df(kmeans.result)
```

kruskalTest\_dataset

Kruskal-Wallis tests on dataset

## **Description**

Run Kruskal-Wallis Tests for each row of the data from the dataset.

## Usage

```
kruskalTest_dataset(dataset, metadata.var, threshold = NULL,
write.file = F, file.out = "kruskal.csv")
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

metadata.var metadata variable to use in the t-tests.

threshold threshold value of the p-value.

write.file boolean value to write or not a file with the results.

file.out name of the file.

#### Value

Table with the results of the Kruskal-Wallis tests, with p-value, -log10(p-value) and false discovery rate (fdr).

```
## Example of ks-Tests on dataset
data(cachexia)
kruskaltests.result = kruskalTest_dataset(cachexia, "Muscle.loss",
write.file = FALSE)
```

52 linregression\_onevar

ksTest_dataset	Kolmogorov-Smirnov tests on dataset
----------------	-------------------------------------

# Description

Run Kolmogorov-Smirnov Tests for each row of the data from the dataset.

#### Usage

```
ksTest_dataset(dataset, metadata.var, threshold = NULL,
write.file = F, file.out = "ks.csv")
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

metadata.var metadata variable to use in the t-tests.

threshold threshold value of the p-value.

write.file boolean value to write or not a file with the results.

file.out name of the file.

### Value

Table with the results of the Kolmogorov-Smirnov tests, with p-value, -log10(p-value) and false discovery rate (fdr).

#### **Examples**

```
## Example of ks-Tests on dataset
data(cachexia)
kstests.result = ksTest_dataset(cachexia, "Muscle.loss",
write.file = FALSE)
```

linregression\_onevar Linear regression on one variable

# Description

Performs linear regression on one variable of the dataset.

## Usage

```
linregression_onevar(dataset, x.val, metadata.vars, combination)
```

linreg\_all\_vars 53

## Arguments

dataset list representing the dataset from a metabolomics experiment.

x.val the x-value to be tested.

metadata.vars metadata variables to use in linear regression.

combination a formula specifying the model.

#### Value

Returns a summary of the result from the lm function from stats package.

#### **Examples**

linreg\_all\_vars

Linear Regression

### **Description**

Performs linear regression analysis over the dataset with the selected metadata's variables.

#### Usage

```
linreg_all_vars(dataset, metadata.vars, combination)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.vars metadata variables to use in linear regression.

combination a formula specifying the model.

#### Value

Returns a list where each element is the linear regression result of a variable on the dataset.

54 linreg\_pvalue\_table

linreg\_coef\_table

Linear regression coefficient table

#### **Description**

Gets a data.frame with the coefficient values.

### Usage

```
linreg_coef_table(linreg.results, write.file = F,
file.out = "linreg-coefs.csv")
```

### **Arguments**

linreg.results Linear regression results from linreg.all.vars function.

write.file boolean value to indicate if a file should be written with the results.

file.out name of the file.

#### Value

Returns a data.frame with the coefficient values.

#### **Examples**

```
## Not run:
    ## Example of linear regression coefficient table
    linreg.coef.tab = linreg_coef_table(linreg.results)
## End(Not run)
```

linreg\_pvalue\_table

Linear regression p-values table

# Description

Gets the p-values table from the linear regression analysis.

## Usage

```
linreg_pvalue_table(linreg.results, write.file = F,
file.out = "linreg-pvalues.csv")
```

#### **Arguments**

 ${\tt linreg.results} \ \ Linear\ regression\ results\ from\ linreg. all. vars\ function.$ 

write.file boolean value to indicate if a file should be written with the results.

file.out name of the file.

linreg\_rsquared 55

## Value

Returns a data.frame with the p-values.

## **Examples**

```
## Not run:
    ## Example of linear regression p-values table
    linreg.pvalue.tab = linreg_pvalue_table(linreg.results)
## End(Not run)
```

linreg\_rsquared

Linear regression r-squared

# Description

Gets the linear regression r-squared values.

#### Usage

```
linreg_rsquared(linreg.results, write.file = F,
file.out = "linreg-rsquared.csv")
```

# Arguments

linreg.results Linear regression results from linreg.all.vars function.

write.file boolean value to indicate if a file should be written with the results.

file.out name of the file.

#### Value

Returns a data.frame with the r-squared values.

```
## Not run:
    ## Example of linear regression rsquared values and the adjusted
    ## rsquared values
    linreg_rsquared(linreg.results)
## End(Not run)
```

low\_level\_fusion

 $log\_transform$ 

Logarithmic transformation.

## Description

Performs logarithmic transformation on the data matrix.

## Usage

```
log_transform(datamat)
```

## Arguments

datamat

data matrix.

## Value

Returns the data matrix with the logarithmic transformation applied.

# **Examples**

```
## Example of logarithmic transformation
data(cassavaPPD)
datamat.log = log_transform(cassavaPPD$data)
```

low\_level\_fusion

Low level fusion

# Description

Low level fusion method for integrate different datasets (only samples with the same name on all datasets will be merged)

# Usage

```
low_level_fusion(datasets)
```

## **Arguments**

datasets

list containing the datasets to be fused (each dataset is a list from the pre defined format for the dataset).

## Value

Return a single dataset with all the datasets merged.

#### **Examples**

```
## Not run:
    ## Example of low level fusion
    datasets = list(dataset1, dataset2)
    dataset.fusion = low_level_fusion(datasets)
## End(Not run)
```

MAIT\_identify\_metabolites

MAIT metabolite identification

# **Description**

Performs metabolite identification using MAIT.

## Usage

```
MAIT_identify_metabolites(dataset, metadata.variable,
xSet = NULL, data.folder = NULL, features = NULL,
mass.tolerance = 0.5)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.variable

metadata's variable.

xSet xcmsSet object that can be passed.

data.folder string indicating the data folder.

features features that can be used to help to identify the metabolites.

mass.tolerance mass tolerance.

## Value

Returns an object resulted from identifyMetabolites function from MAIT package.

## References

http://www.bioconductor.org/packages/release/bioc/html/MAIT.html

58 mean\_centering

## **Examples**

```
## Not run:
    ## Example of MAIT metabolite identification
    data(spinalCord)
    library(MAIT)
    mait.metabolites = MAIT_identify_metabolites(spinalCord, "type",
features = "all", data.folder = "data",
xSet = spinalCord$xSet)
    mait.metab.table = mait.metabolites@FeatureInfo@metaboliteTable
    mait.metab.table[which(mait.metab.table$Name != "Unknown"),
    c(1,3,6)]
## End(Not run)
```

mean\_centering

Mean centering

# Description

Performs mean centering on the dataset.

## Usage

```
mean_centering(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the dataset with mean centering applied.

```
## Example of mean centering
data(cassavaPPD)
cassavaPPD = mean_centering(cassavaPPD)
```

merge\_datasets 59

#### **Description**

Merges two datasets with the same variables and metadata's variables.

#### Usage

```
merge_datasets(dataset1, dataset2)
```

### **Arguments**

dataset1 list representing the first dataset from a metabolomics experiment.

dataset2 list representing the second dataset from a metabolomics experiment.

#### Value

Returns one dataset with the data from the two datasets merged.

#### **Examples**

```
## Not run:
    ## Example of merging two datasets
    dataset = merge_datasets(dataset1, dataset2)
## End(Not run)
```

merge\_data\_metadata

Merge data and metadata

# Description

Merges the data and metadata from the dataset into a single data.frame.

#### Usage

```
merge_data_metadata(dataset, samples = NULL,
metadata.vars = NULL, x.values = NULL, by.index = F)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

samples vector with indexes or names of the samples to select

metadata.vars metadata's variables.

x.values vector with the desired x-values to get from the dataset. by.index if TRUE, the values of the x.values argument are indexes. 60 metadata\_as\_variables

## Value

Returns a data.frame with the data and metadata from the dataset merged.

# **Examples**

```
## Example of merging data and metadata
data(cachexia)
dt.merged = merge_data_metadata(cachexia)
```

# Description

Use one or more metadata variables as variables.

## Usage

```
metadata_as_variables(dataset, metadata.vars, by.index = F)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

metadata.vars name or index of the metadata variables that are going to be used as data vari-

ables.

by . index boolean value indicating if the metadata variables are indexes or names

#### Value

Returns the dataset with the metadata variables removed from the metadata and added on the data matrix.

```
## Example of using a metadata variable as data variable
data(propolis)
propolis = metadata_as_variables(propolis, "seasons", by.index = FALSE)
```

```
missingvalues_imputation
```

Missing values imputation

# Description

Treats the missing values of a dataset according to a specific method.

#### Usage

```
missingvalues_imputation(dataset, method = "value",
value = 5e-04, k = 5)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

method imputation method. It can be:

• "value" - replaces the missing values with a specific value

• "mean" - replaces the missing values with the mean of the variables' values

• "median" - replaces the missing values with the median of the variables' values

• "knn" - replaces the missing values with k nearest neighbor averaging

• "linapprox" - replaces the missing values with linear approximation

value the value to replace the missing values if the method is "value".

k the number of neighbors if the method is "knn".

## Value

Returns the dataset with no missing values.

```
## Example of impute missing values
data(propolis)
dataset = missingvalues_imputation(propolis, method = "value",
   value = 0.0005)
```

62 multiClassSummary

msc\_correction

Multiplicative scatter correction

## **Description**

Perform multiplicative scatter correction on the spectra.

### Usage

```
msc_correction(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Return the dataset with the multiplicative scatter correction employed on the data.

## **Examples**

```
## Example of multiplicative scatter correction
data(cassavaPPD)
propolis.msc = msc_correction(cassavaPPD)
```

multiClassSummary

Multi Class Summary

## **Description**

Summary function for caret to compute AUC.

# Usage

```
multiClassSummary(data, lev = NULL, model = NULL)
```

## **Arguments**

data data parameter.

lev lev parameter.

model model parameter.

### References

www.r-bloggers.com/error-metrics-for-multi-class-problems-in-r-beyond-accuracy-and-kappa/

# Description

Perform multi-factor ANOVA on all variables with the selected metadata variables.

## Usage

```
multifactor_aov_all_vars(dataset, metadata.vars, combination)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.vars metadata variables to use in ANOVA. combination a formula specifying the model.

#### Value

List where each element is the multifactor anova result of a variable on the dataset.

#### **Examples**

```
## Example of multifactor ANOVA on all variables
data(propolis)
propolis = missingvalues_imputation(propolis, "value", value = 0.00005)
m.aov.results = multifactor_aov_all_vars(propolis,
c("seasons", "agroregions"), "seasons*agroregions")
```

## **Description**

Gets the p-values table from the multifactor ANOVA results.

#### Usage

```
multifactor_aov_pvalues_table(multifactor.aov.results,
write.file = F, file.out = "multi-anova-pvalues.csv")
```

## Arguments

```
multifactor.aov.results
multifactor anova results.

write.file boolean value to indicate if a file is written.

file.out name of the file.
```

#### Value

Returns a data.frame with the p-values.

## **Examples**

```
## Example of multifactor ANOVA p-values table
data(propolis)
propolis = missingvalues_imputation(propolis, "value", value = 0.00005)
m.aov.results = multifactor_aov_all_vars(propolis,
c("seasons", "agroregions"), "seasons*agroregions")
m.aov.pvalues = multifactor_aov_pvalues_table(m.aov.results)
```

# Description

Gets the variability explained table from the multifactor ANOVA results.

#### Usage

```
multifactor_aov_varexp_table(multifactor.aov.results,
write.file = F, file.out = "multi-anova-varexp.csv")
```

#### Arguments

```
multifactor.aov.results
multifactor anova results.

write.file boolean value to indicate if a file is written.

file.out name of the file.
```

# Value

Returns a data.frame with the variability explained.

multiplot 65

#### **Examples**

```
## Example of multifactor ANOVA variability explained table
data(propolis)
propolis = missingvalues_imputation(propolis, "value", value = 0.00005)
m.aov.results = multifactor_aov_all_vars(propolis,
c("seasons", "agroregions"), "seasons*agroregions")
m.aov.varepx = multifactor_aov_varexp_table(m.aov.results)
```

multiplot

Multiplot

## **Description**

Multiplot from ggplot2 - function taken from (see references).

#### Usage

```
multiplot(plots, plotlist = NULL, file, cols = 1, layout = NULL)
```

## Arguments

```
plots list with the plots to display.
plotlist plot list.
file file.
cols number of columns.
layout layout of the plot.
```

#### References

http://www.cookbook-r.com/Graphs/Multiple\_graphs\_on\_one\_page\_

```
## Example of multiplot
   data(cachexia)
   pca.result = pca_analysis_dataset(cachexia)
   plot1 = pca_scoresplot2D(cachexia, pca.result, "Muscle.loss",
ellipses = TRUE)
   plot2 = pca_scoresplot2D(cachexia, pca.result, "Muscle.loss",
ellipses = FALSE, labels = TRUE)
   plts = list(plot1,plot2)
   multiplot(plts, cols = 2)
```

66 normalize\_samples

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norma	- 1	i	70

Normalize data

## Description

Normalize the data from the dataset with a specific method.

#### Usage

```
normalize(dataset, method, ref = NULL, constant = 1000)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

method string specifying the normalization method. The possible values are:

• "sum" normalization by sum.

• "median" normalization by median.

• "ref.sample" normalization by reference sample.

• "ref.feature" normalization by reference feature.

ref the reference if method is "ref.sample" or "ref.feature".

constant the constant value if method is "sum".

#### Value

Returns the dataset with the data normalized.

# **Examples**

```
## Example of normalization by median
data(cassavaPPD)
cassava.norm = normalize(cassavaPPD, "median")
```

normalize\_samples

Normalize samples

## **Description**

Normalize the data from a datamatrix with a specific method.

## Usage

```
normalize_samples(datamat, method, ref = NULL, constant = 1000)
```

num\_samples 67

## Arguments

datamat data matrix.

method string specifying the normalization method. The possible values are:

• "sum" normalization by sum.

• "median" normalization by median.

• "ref.sample" normalization by reference sample.

• "ref.feature" normalization by reference feature.

ref the reference if method is "ref.sample" or "ref.feature".

constant the constant value if method is "sum".

## Value

Returns the data matrix normalized.

# **Examples**

```
## Example of normalization by median
data(cassavaPPD)
datamat.norm = normalize_samples(cassavaPPD$data, "median")
```

num\_samples

Get number of samples

## **Description**

Get the number of samples from a dataset.

## Usage

```
num_samples(dataset)
```

# Arguments

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns an integer with the number of samples in the dataset.

```
## Example of getting the number of samples
data(cachexia)
number.of.samples = num_samples(cachexia)
```

68 offset\_correction

num\_x\_values

Get number of x values

## **Description**

Get the number of x-axis values.

## Usage

```
num_x_values(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

## Value

Returns an integer with the number of x-axis values.

# **Examples**

```
## Example of getting the number of x-axis values
data(cassavaPPD)
number.x.values = num_x_values(cassavaPPD)
```

 $offset\_correction$ 

Offset correction

## **Description**

Perform offset correction on the data.

### Usage

```
offset_correction(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the dataset

```
## Example of offset correction
data(cassavaPPD)
cassava.corrected = offset_correction(cassavaPPD)
```

pca\_analysis\_dataset 69

# Description

Performs a classical PCA analysis over the dataset.

# Usage

```
pca_analysis_dataset(dataset, scale = T, center = T,
write.file = F, file.out = "pca", ...)
```

#### **Arguments**

dataset	list representing the dataset from a metabolomics experiment.
scale	boolean value indicating if the variables are going to be scaled or not.
center	booleam value indicating if the variables are going to be centered or not.
write.file	boolean value that indicates if the results from PCA analysis are going to be written on a file.
file.out	name of the file that will store the results.
	additional parameters to ggplot function.

## Value

object of class 'prcomp' with the results from the PCA analysis.

## **Examples**

```
## Example of performing a classical PCA analysis
data(cachexia)
pca.results = pca_analysis_dataset(cachexia)
```

pca\_biplot PCA biplot

# Description

Shows a PCA biplot.

## Usage

```
pca_biplot(dataset, pca.result, cex = 0.8, legend.cex = 0.8, x.colors = 1, inset = c(0, 0), legend.place = "topright", ...)
```

70 pca\_biplot3D

## Arguments

dataset list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

cex cex value.

legend.cex cex value of the legend.

x.colors colors of a metadata's variable.
inset inset parameter of legend function.

legend.place legend place.

... additional parameters passed to biplot function.

## **Examples**

```
## Example of a PCA biplot
data(cachexia)
pca.result = pca_analysis_dataset(cachexia)
pca_biplot(cachexia, pca.result, cex = 0.8)
```

pca\_biplot3D

3D PCA biplot (interactive)

#### **Description**

Shows a interactive 3D PCA biplot.

#### Usage

```
pca_biplot3D(dataset, pca.result, column.class = NULL,
pcas = c(1, 2, 3))
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

column.class metadata's variable.

pcas the three principal components.

```
## Example of a 3D PCA biplot
data(cachexia)
pca.result = pca_analysis_dataset(cachexia)
pca_biplot3D(cachexia, pca.result, "Muscle.loss", pcas = c(1,2,3))
```

pca\_importance 71

|--|

# Description

Gets the importance from the PCs.

## Usage

```
pca_importance(pca.res, pcs = 1:length(pca.res$sdev), sd = T,
prop = T, cumul = T, min.cum = NULL)
```

# Arguments

pca.res	prcomp object with the PCA results.
pcs	vector with the PCs to get.
sd	boolean value indicating if standard deviation will be returned or not.
prop	boolean value that indicates if the proportion of variance is returned or not.
cumul	boolean value that indicates if the cumulative variance is returned or not.
min.cum	allows to define minimum cumulative % of variance

### Value

Returns the information about the importance of the PCs.

## **Examples**

```
## Example of performing a classical PCA analysis
data(cachexia)
pca.result = pca_analysis_dataset(cachexia)
pca_importance(pca.result, pcs = 1:5)
```

```
pca_kmeans_plot2D 2D PCA k-means plot
```

## **Description**

Groups the points with the clusters given by k-means in a 2D PCA scores plot.

# Usage

```
pca_kmeans_plot2D(dataset, pca.result, num.clusters = 3,
pcas = c(1, 2), kmeans.result = NULL, labels = FALSE,
ellipses = FALSE, leg.pos = "right", xlim = NULL, ylim = NULL)
```

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### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

promp object with the PCA results. num.clusters number of clusters of k-means.

pcas vector with the principal components to be plotted.

kmeans.result result from k-means. If null k-means is performed in the function. boolean value indicating if the samples' labels will be shown.

ellipses boolean value that indicates if an ellipse will be drawn on each group of the

metadata's variable.

leg.pos legend position.

xlim vector with two positions with the x-axis limits.
ylim vector with two positions with the y-axis limits.

## **Examples**

```
## Not run:
    ## Example of a 2D PCA k-means plot
    data(cachexia)
    pca.result = pca_analysis_dataset(cachexia)
    pca_kmeans_plot2D(cachexia, pca.result, num.clusters = 3, pcas = c(1,2))
## End(Not run)
```

pca\_kmeans\_plot3D

3D PCA k-means plot (interactive)

# **Description**

Groups the points with the clusters given by k-means in a interactive 3D PCA scores plot.

## Usage

```
pca_kmeans_plot3D(dataset, pca.result, num.clusters = 3, pcas = c(1, 2, 3), kmeans.result = NULL, labels = FALSE, size = 1, ...)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

promp object with the PCA results.

num.clusters

number of clusters of k-means.

pcas vector with the principal components to be plotted.

kmeans.result result from k-means. If null k-means is performed in the function. boolean value indicating if the samples' labels will be shown.

size parameter of plot3d from rgl package.

. . . additional parameters of plot3d function from rgl package.

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### **Examples**

```
pca_pairs_kmeans_plot PCA k-means pairs plot
```

# **Description**

Groups the points with the clusters from k-means in a PCA pairs plot.

### Usage

```
pca_pairs_kmeans_plot(dataset, pca.result, num.clusters = 3,
kmeans.result = NULL, pcas = c(1, 2, 3, 4, 5))
```

### **Arguments**

list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

num.clusters number of clusters of k-means.

kmeans.result result from k-means. If null k-means is performed in the function.

pcas vector with the principal components to be plotted.

```
## Not run:
    ## Example of a PCA k-means pairs plot (computationally heavy)
    data(cachexia)
    pca.result = pca_analysis_dataset(cachexia)
    kmeans.res = kmeans_clustering(cachexia, 3)
    pca_pairs_kmeans_plot(cachexia, pca.result, num.clusters = 3,
    kmeans.result = kmeans.res, pcas = c(1,2,3,4,5))
## End(Not run)
```

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pca\_pairs\_plot

PCA pairs plot

## **Description**

Shows a PCA pairs plot.

## Usage

```
pca_pairs_plot(dataset, pca.result, column.class = NULL,
pcas = c(1, 2, 3, 4, 5), ...)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

column.class metadata's variable.

pcas the principal components to be shown.

... additional parameters to ggpairs function from GGally package.

### **Examples**

```
## Example of a PCA pairs plot
data(cachexia)
pca.result = pca_analysis_dataset(cachexia)
pca_pairs_plot(cachexia, pca.result, "Muscle.loss", pcas = c(1,2,3))
```

pca\_plot\_3d

3D pca plot

## Description

3D plot from 3 components

## Usage

```
pca_plot_3d(dataset, model, var.class, pcas = 1:3, colors = NULL,
legend.place = "topright", ...)
```

pca\_robust 75

# **Arguments**

dataset list representing the dataset from a metabolomics experiment.

model model with pca scores (pls model).

var.class metadata column class.

pcas the components to be plotted.

colors colors of the groups.

legend.place legend place.

... additional parameters to legend function.

## **Examples**

```
## Not run:
    ## Example of a 3d pca plot
    data(cachexia)
    train.result = train_models_performance(cachexia, "pls",
    "Muscle.loss", "cv")
    pca_plot_3d(cachexia, train.result$final.models$pls, "Muscle.loss")
## End(Not run)
```

pca\_robust

PCA analysis (robust)

### **Description**

Performs a robust PCA analysis.

### Usage

```
pca_robust(dataset, center = "median", scale = "mad", k = 10,
write.file = F, file.out = "robpca", ...)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

center indicates how the data is to be centered. Can be a function or a vector with the

center values of each column.

scale indicates how the data is to be rescaled. Can be a function or a vector with the

scale value of each column.

k the desired number of components to compute

write.file boolean value that indicates if the results from PCA analysis are going to be

written on a file.

file.out name of the file that will store the results.

... additional parameters pass to or from other functions.

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#### Value

Returns an object of class 'princomp' with the PCA results.

### **Examples**

```
## Example of performing a robust PCA analysis
data(cachexia)
pca.results = pca_robust(cachexia, center = "mean", scale = "mad",
k = 10)
```

pca\_scoresplot2D

2D PCA scores plot

#### **Description**

Shows a 2D PCA scores plot of two principal componets.

#### **Usage**

```
pca_scoresplot2D(dataset, pca.result, column.class,
pcas = c(1, 2), labels = FALSE, ellipses = FALSE,
pallette = 2, leg.pos = "right", xlim = NULL, ylim = NULL)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

column.class metadata's variable.

pcas vector of two elements with the PCs that will be plotted.

labels boolean value indicating if the sample's labels will be displayed.

ellipses boolean value that indicates if an ellipse will be drawn on each group of the

metadata's variable.

pallette parameter of scale\_colour\_brewer from ggplot2.

leg.pos position of the legend.

xlim vector with two numeric values indicating the limits of the x axis.
ylim vector with two numeric values indicating the limits of the y axis.

pca\_scoresplot3D 77

pca_scoresplot3D	3D PCA scores plot
------------------	--------------------

### **Description**

Shows a 3D PCA scores plot of three principal componets.

#### Usage

```
pca_scoresplot3D(dataset, pca.result, column.class,
pcas = c(1, 2, 3))
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

column.class metadata's variable.

pcas vector with the principal components to be plotted.

### **Examples**

```
## Example of a 3D PCA scores plot
data(cachexia)
pca.result = pca_analysis_dataset(cachexia)
pca_scoresplot3D(cachexia, pca.result, "Muscle.loss", pcas = c(1,2,3))
```

### **Description**

Shows a interactive 3D PCA scores plot of three principal components.

## Usage

```
pca_scoresplot3D_rgl(dataset, pca.result, column.class,
pcas = c(1, 2, 3), size = 1, labels = FALSE)
```

# **Arguments**

dataset list representing the dataset from a metabolomics experiment.

pca.result prcomp object with the PCA results.

column.class metadata's variable.

pcas vector with the principal components to be plotted.

size parameter of plot3d from rgl package.

labels boolean value indicating if the samples' labels will be shown.

78 pca\_screeplot

## **Examples**

pca\_screeplot

PCA scree plot

# Description

PCA scree plot with the proportion and cumulative variance of the PCs.

# Usage

```
pca_screeplot(pca.result, num.pcs = NULL, cex.leg = 0.8,
leg.pos = "right", lab.text = c("individual percent",
"cumulative percent"), fill.col = c("blue", "red"),
ylab = "Percentage", xlab = "Principal components", ...)
```

## **Arguments**

pca.result	prcomp object with the PCA results.
num.pcs	number of principal components.
cex.leg	cex value of legend.
leg.pos	legend position.
lab.text	legend's labels.
fill.col	color of the legend's boxes.
ylab	y-axis label.
xlab	x-axis label
	additional parameters to matplot.

```
## Example of a scree plot
data(cachexia)
pca.result = pca_analysis_dataset(cachexia)
pca_screeplot(pca.result)
```

peaks\_per\_sample 79

peaks\_per\_sample

Peaks per sample

## **Description**

Counts number of peaks in a sample (given its index).

# Usage

```
peaks_per_sample(sample.list, sample.index)
```

## **Arguments**

```
sample.list list of data frames with the samples' peaks. sample.index sample index.
```

#### Value

Return a integer value with the number of peaks in the sample.

# **Examples**

```
## Example of counting the peaks in a sample
data(propolisSampleList)
num.peaks.sample = peaks_per_sample(propolisSampleList, 4)
```

peaks\_per\_samples

Peaks per samples

# Description

Calculates the number of peaks on each sample.

# Usage

```
peaks_per_samples(sample.list)
```

### **Arguments**

```
sample.list list of data frames with the samples' peaks.
```

# Value

Returns a numeric vector with the number of peaks on each sample.

80 plotvar\_twofactor

### **Examples**

```
## Example of counting the peaks in each sample
data(propolisSampleList)
num.peaks.samples = peaks_per_samples(propolisSampleList)
```

plotvar\_twofactor

Plot variable distribution on two factors

## **Description**

Plot variable distribution on two factors from the dataset.

## Usage

```
plotvar_twofactor(dataset, variable, meta.var1, meta.var2,
colour = "darkblue", title = "", xlabel = NULL, ylabel = NULL)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

variable variable's name.

meta.var1 first metadata's variable.

meta.var2 second metadata's variable.

colour colours of the bodies of the boxplots.

title title of the plot.

xlabel x-axis label.

ylabel y-axis label.

#### Value

Returns the plot object from ggplot function.

```
## Example of plotting a variable's distribution with 2 factors
data(cassavaPPD)
plotvar_twofactor(cassavaPPD, "399.3", "varieties", "ppds")
```

plot\_anova 81

plot\_anova

Plot ANOVA results

### **Description**

Function for plotting the results from ANOVA.

## Usage

```
plot_anova(dataset, anova.results, anova.threshold = 0.01,
reverse.x = F)
```

## Arguments

```
dataset list representing the dataset from a metabolomics experiment.

anova.results ANOVA results.

anova.threshold ANOVA threshold for the p-value.

reverse.x boolean value to indicate if the x-axis is plotted in reverse.
```

## **Examples**

```
## Example of plotting the ANOVA results - first filter the
## dataset to reduce computation time
data(cassavaPPD)
  cassavaPPD = flat_pattern_filter(cassavaPPD, "iqr", by.percent = TRUE,
red.value = 75)
  anova.results = aov_all_vars(cassavaPPD, "varieties", doTukey = FALSE)
  plot_anova(cassavaPPD, anova.results, 0.05e-8)
```

plot\_fold\_change

Plot fold change results

# Description

Function for plotting the results from fold change.

## Usage

```
plot_fold_change(dataset, fc.results, fc.threshold, plot.log = T,
var = F, xlab = "")
```

82 plot\_kruskaltest

## Arguments

dataset list representing the dataset from a metabolomics experiment.

fc.results fold change results.

fc.threshold fold change threshold for the p-value.

plot.log boolean value to determine if the fold change values are transformed logarith-

mically or not.

var boolean value, if TRUE it uses the xlab argument to represent the xlabel of the

plot.

xlab string with the x axis description.

## **Examples**

```
## Example of plotting the fold change results
  data(cachexia)
  fc.results = fold_change(cachexia, "Muscle.loss",
"control")
  plot_fold_change(cachexia, fc.results, 2)
```

plot\_kruskaltest

Plot Kruskal-Wallis tests results

## Description

Function for plotting the results from Kruskal-Wallis tests.

### Usage

```
plot_kruskaltest(dataset, kr.results, kr.threshold = 0.01)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

kr.results Kruskal-Wallis tests results.

kr. threshold Kruskal-Wallis test threshold for the p-value.

```
## Example of plotting the Kolmogorov-Smirnov tests results
data(cachexia)
kr.results = kruskalTest_dataset(cachexia, "Muscle.loss",
write.file = FALSE)
plot_kruskaltest(cachexia, kr.results, 0.05)
```

plot\_kstest 83

plot_kstest Pa	ot Kolmogorov-Smirnov tests results
----------------	-------------------------------------

## **Description**

Function for plotting the results from Kolmogorov-Smirnov tests.

### Usage

```
plot_kstest(dataset, ks.results, ks.threshold = 0.01)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

ks.results Kolmogorov-Smirnov tests results.

ks. threshold Kolmogorov-Smirnov test threshold for the p-value.

# **Examples**

```
## Example of plotting the Kolmogorov-Smirnov tests results
data(cachexia)
ks.results = ksTest_dataset(cachexia, "Muscle.loss",
write.file = FALSE)
plot_kstest(cachexia, ks.results, 0.05)
```

```
plot_regression_coefs_pvalues
```

Plot regression coefficient and p-values

## **Description**

Plots the linear regression coefficient and the p-values.

## Usage

```
plot_regression_coefs_pvalues(linreg.results, bar.col = NULL,
coef.size = 5, ...)
```

## **Arguments**

```
linreg.results linear regression results.
```

```
bar.col color of the bars.

coef.size coefficient font size.
```

. . . additional parameters to geom\_text and geom\_bar from ggplot.

84 plot\_spectra

### **Examples**

```
## Not run:
    ## Example of multiplot
    plot_regression_coefs_pvalues(linreg.results)
## End(Not run)
```

plot\_spectra

Plot spectra

### **Description**

Plot spectra from dataset.

### Usage

```
plot_spectra(dataset, column.class, func = NULL, samples = NULL,
variable.bounds = NULL, xlab = NULL, ylab = NULL, lty = 1,
legend.place = "topright", cex = 0.8, reverse.x = F, ...)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

column.class string indicating the metadata's variable.

func function to compute the summary statistics to apply to the data.

samples vector with samples' names, if NULL all the samples will be considered.

variable.bounds

numeric vector with two elements indicating the interval of x-values to plot.

xlab x-axis label. ylab y-axis label.

1ty parameter of matplot.

legend.place string indicating the place that the legend's box will be placed.

cex numeric value that indicates the amount by which the legend is magnified rela-

tive to the default.

reverse.x boolean value indicating if the x-axis will be shown reversed or not.

... additional parameters to matplot.

```
## Example of plotting spectra (simple)
data(cassavaPPD)
plot_spectra(cassavaPPD, "varieties", func = NULL,
  samples = c("BRA_1", "IAC5_4"), variable.bounds = c(1000,2000),
  legend.place = "topright")
```

plot\_spectra\_simple 85

# Description

Plot spectra from dataset (simple version).

# Usage

```
plot_spectra_simple(dataset, samples = NULL,
variable.bounds = NULL, xlab = NULL, ylab = NULL,
lty = 1, lwd = 1, col = 1, reverse.x = F, ...)
```

# Arguments

dataset	list representing the dataset from a metabolomics experiment.			
samples	vector with samples' names, if NULL all the samples will be considered.			
variable.bounds				
	numeric vector with two elements indicating the interval of x-values to plot.			
xlab	x-axis label.			
ylab	y-axis label			
lty	parameter of matplot.			
lwd	parameter of matplot.			
col	parameter of matplot.			
reverse.x	boolean value indicating if the x-axis will be shown reversed or not.			
• • •	additional parameters to pass to matplot.			

```
## Example of plotting spectra (simple)
data(cassavaPPD)
plot_spectra_simple(cassavaPPD, samples = c("IAC5_4", "BRA_1"),
variable.bounds = c(1000,2000))
```

86 predict\_samples

plot\_ttests

Plot t-tests results

## **Description**

Function for plotting the results from t-tests.

## Usage

```
plot_ttests(dataset, tt.results, tt.threshold = 0.01)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

tt.results t-tests results.

tt.threshold t-test threshold for the p-value.

## **Examples**

```
## Example of plotting the t-tests results
data(cachexia)
ttests.results = tTests_dataset(cachexia, "Muscle.loss")
plot_ttests(cachexia, ttests.results, 0.05)
```

predict\_samples

Predict samples

# **Description**

Predict new samples.

# Usage

```
predict_samples(train.result, new.samples)
```

## **Arguments**

```
train.result result from training a classifier.
new.samples dataframe with new samples.
```

## Value

Returns a data frame with the samples and the predicted class.

propolis 87

### **Examples**

```
## Example of predicting samples
data(cachexia)
training.result = train_models_performance(cachexia, "pls",
"Muscle.loss", "cv")
result = predict_samples(training.result$final.models$pls, cachexia$data)
```

propolis

Brazilian Propolis from different Harvest Seasons and different Agroecological Regions (dataset)

### **Description**

Propolis or bee glue is a sticky dark-colored substance produced from the collected buds or exudates of plants (resin) by bees (Apis mellifera L.). The resin is masticated, salivary enzymes are added, and the partially digested material is mixed with beewax and used in the hive to seal the walls, strengthen the borders of combs, and embalm dead invaders (Wollenweber et al., 1990). The propolis samples are from NMR data and were collected in the autumn (AU), winter (WI), spring (SP), and summer (SM) of 2010 from Apis mellifera hives located in southern Brazil (Santa Catarina State). A total of 59 samples were collected, and the distribution of samples by seasons being: SM - 16 samples, AU and SP - 15 samples, WI - 13 samples. Also, three agroecological regions were defined for the different apiaries, and one distributed as follows: Highlands - 12 samples, Plain - 11 samples, Plateau - 36 samples.

## Usage

data(propolis)

#### Format

An object of class "list"

#### References

E. Wollenweber, B. M. Hausen, and W. Greenaway. Phenolic constituents and sensitizing properties of propolis, poplar balsam and balsam of peru. Bulletin de Groupe Polyphenol, 15:112-120, 1990. M. Maraschin, A. Somensi-Zeggio, S. K. Oliveira, S. Kuhnen, M. M. Tomazzoli, A. C. M. Zeri, R. Carreira, and M. Rocha. A machine learning and chemometrics assisted interpretation of spectroscopic data - a nmr-based metabolomics platform for the assessment of brazilian propolis. 2012

```
data(propolis)
sum_dataset(propolis)
```

88 propolisSampleList

propolisSampleList Brazilian Propolis from different Harvest Seasons and different Agroecological Regions (sample list)

### Description

Propolis or bee glue is a sticky dark-colored substance produced from the collected buds or exudates of plants (resin) by bees (Apis mellifera L.). The resin is masticated, salivary enzymes are added, and the partially digested material is mixed with beewax and used in the hive to seal the walls, strengthen the borders of combs, and embalm dead invaders (Wollenweber et al., 1990). The propolis samples are from NMR data and were collected in the autumn (AU), winter (WI), spring (SP), and summer (SM) of 2010 from Apis mellifera hives located in southern Brazil (Santa Catarina State). A total of 59 samples were collected, and the distribution of samples by seasons being: SM - 16 samples, AU and SP - 15 samples, WI - 13 samples. Also, three agroecological regions were defined for the different apiaries, and one distributed as follows: Highlands - 12 samples, Plain - 11 samples, Plateau - 36 samples.

### Usage

data(propolisSampleList)

### **Format**

An object of class "list"

### References

E. Wollenweber, B. M. Hausen, and W. Greenaway. Phenolic constituents and sensitizing properties of propolis, poplar balsam and balsam of peru. Bulletin de Groupe Polyphenol, 15:112-120, 1990. M. Maraschin, A. Somensi-Zeggio, S. K. Oliveira, S. Kuhnen, M. M. Tomazzoli, A. C. M. Zeri, R. Carreira, and M. Rocha. A machine learning and chemometrics assisted interpretation of spectroscopic data - a nmr-based metabolomics platform for the assessment of brazilian propolis. 2012

### **Examples**

data(propolisSampleList)
propolisSampleList[[1]]

read\_csvs\_folder 89

read\_csvs\_folder

Read CSVs from folder

# **Description**

Reads multiple CSV files in a given folder.

### Usage

```
read_csvs_folder(foldername, ...)
```

## **Arguments**

```
foldername string with the name of the folder.
... additional parameters to read.csv function.
```

#### Value

Returns a list of data frames.

## **Examples**

```
## Not run:
    ## Example of reading multiple csvs
    sample.list = read_csvs_folder("foldername")
## End(Not run)
```

read\_dataset\_csv

Read dataset from CSV

## Description

Reads the data from a CSV file and creates the dataset.

# Usage

```
read_dataset_csv(filename.data, filename.meta = NULL,
type = "undefined", description = "", label.x = NULL,
label.values = NULL, sample.names = NULL, format = "row",
header.col = TRUE, header.row = TRUE, sep = ",",
header.col.meta = TRUE, header.row.meta = TRUE, sep.meta = ",")
```

90 read\_dataset\_csv

### **Arguments**

filename.data name of the data file.

filename.meta name of the metadata file.

type type of the data.

description a short text describing the dataset.

label.x the label for the x values.

label.values the label for the y values.

sample.names the names of the samples.

format format which the data are in the CSV file. It can be "row" if the samples are in

the rows or "col" if the samples are in the columns.

header.col boolean value indicating if the CSV contains a header column with the names

of the samples or variables.

header.row boolean value indicating if the CSV contains a header row with the names of the

samples or variables.

sep the separator character.

header.col.meta

boolean value indicating if the metadata CSV file contains a header column with

the name of the metadata variables.

header.row.meta

boolean value indicating if the metadata CSV file contains a header row with the

name of the samples.

sep.meta the separator character of the metadata file.

### Value

Returns the dataset from the CSV file.

```
## Not run:
    ## Example of reading a dataset from a CSV file
    dataset = read_dataset_csv("data.csv", "metadata.csv",
        type = "nmr-spectra",
        description ="description of the dataset",
        label.x = "ppm", label.values = "intensity")
## End(Not run)
```

read\_dataset\_dx 91

## **Description**

Reads the data from the (J)DX files and creates the dataset.

## Usage

```
read_dataset_dx(folder.data, filename.meta = NULL,
type = "undefined", description = "", label.x = NULL,
label.values = NULL, header.col.meta = TRUE,
header.row.meta = TRUE, sep.meta = ",")
```

# Arguments

folder.data string containing the path of the data folder.

filename.meta name of the metadata file.

type type of the data.

description a short text describing the dataset.

label.x the label for the x values. label.values the label for the y values.

header.col.meta

boolean value indicating if the metadata CSV file contains a header column with

the name of the metadata variables.

header.row.meta

boolean value indicating if the metadata CSV file contains a header row with the

name of the samples.

sep.meta the separator character of the metadata file.

#### Value

Returns the dataset from the (J)DX files.

```
## Not run:
    ## Example of reading a dataset from (J)DX files
    dataset = read_dataset_dx("data", "metadata.csv", type = "nmr-spectra",
        description = "description of the dataset", label.x = "ppm",
        label.values = "intensity")
## End(Not run)
```

92 read\_dataset\_spc

read\_dataset\_spc Read a

Read dataset from SPC files

## **Description**

Reads the data from the SPC files and creates the dataset.

### Usage

```
read_dataset_spc(folder.data, filename.meta = NULL,
type = "undefined", description = "", nosubhdr = F,
label.x = NULL, label.values = NULL, header.col.meta = TRUE,
header.row.meta = TRUE, sep.meta = ",")
```

### **Arguments**

folder.data string containing the path of the data folder.

filename.meta name of the metadata file.

type type of the data.

description a short text describing the dataset.

nosubhdr if TRUE, it won't read the subheader.

label.x the label for the x values. label.values the label for the y values.

header.col.meta

boolean value indicating if the metadata CSV file contains a header column with

the name of the metadata variables.

header.row.meta

boolean value indicating if the metadata CSV file contains a header row with the

name of the samples.

sep.meta the separator character of the metadata file.

### Value

Returns the dataset from the SPC files.

```
## Not run:
    ## Example of reading a dataset from SPC files
    dataset = read_dataset_spc("data", "metadata.csv", type = "ir-spectra",
        description = "description of the dataset", label.x = "wavelength",
        label.values = "transmittance")
## End(Not run)
```

read\_data\_csv 93

read_data_csv I	Read	<b>CSV</b>	data
-----------------	------	------------	------

# Description

Reads the data from the CSV file.

# Usage

```
read_data_csv(filename, format = "row", header.col = TRUE,
header.row = TRUE, sep = ",")
```

## Arguments

filename name of the file with the data.

format which the data are in the CSV file. It can be "row" if the samples are in

the rows or "col" if the samples are in the columns.

header.col boolean value indicating if the CSV contains a header column with the names

of the samples or variables.

header.row boolean value indicating if the CSV contains a header row with the names of the

samples or variables.

sep the separator character.

## Value

Returns a numeric matrix with the data.

# **Examples**

```
## Not run:
    ## Example of reading a dataset from a CSV file
    data.matrix = read_data_csv("data.csv", format = "row")
## End(Not run)
```

read\_data\_dx

Read data from (J)DX files

## **Description**

Reads the data from the (J)DX files.

# Usage

```
read_data_dx(foldername, debug = F)
```

94 read\_data\_spc

## **Arguments**

foldername string with the path of the data folder.

debug option for ChemoSpec's readJDX function.

## Value

Returns a list with the samples and the respective data points.

## **Examples**

```
## Not run:
    ## Example of reading a dataset from (J)DX file
s sample.list = read_data_dx("data")
## End(Not run)
```

read\_data\_spc

Read data from SPC files

# Description

Reads the data from the SPC files.

## Usage

```
read_data_spc(foldername, nosubhdr = F)
```

## Arguments

foldername string with the path of the data folder.
nosubhdr if TRUE, it won't read the subheader.

### Value

Returns a list with the samples and the respective data points.

```
## Not run:
    ## Example of reading a dataset from SPC files
    sample.list = read_data_spc("data")
## End(Not run)
```

read\_metadata 95

read_metadata	
---------------	--

Read metadata

### **Description**

Read the metadata from a file.

### Usage

```
read_metadata(filename, header.col = T, header.row = T,
sep = ",")
```

### **Arguments**

filename name of the file with the metadata.

header.col boolean value indicating if the metadata CSV file contains a header column with

the name of the metadata variables.

header.row boolean value indicating if the metadata CSV file contains a header row with the

name of the samples.

sep the separator character.

### Value

Returns a data frame with the metadata.

#### **Examples**

```
## Not run:
    ## Example of reading metadata from a file
    metadata = read_metadata("metadata.csv")
## End(Not run)
```

read\_ms\_spectra

Read MS spectra

#### **Description**

Read the data from the MS files and creates the dataset.

# Usage

```
read_ms_spectra(folder.name, type = "undefined",
filename.meta = NULL, description = "", prof.method = "bin",
fwhm = 30, bw = 30, intvalue = "into", header.col.meta = TRUE,
header.row.meta = TRUE, sep.meta = ",")
```

96 read\_multiple\_csvs

### **Arguments**

folder.name string containing the path of the data folder.

type type of the data.

filename.meta name of the metadata file.

description a short text describing the dataset.

prof.method profmethod parameter from xcmsSet function from xcms package.

fwhm fwhm parameter from xcmsSet function from xcms package. A commonly used

value is 30 (seconds) for LC-MS and 4 (seconds) for GC-MS spectra.

bw bw parameter from group function from xcms package.

intvalue value parameter from groupval function from xcms package. It can be:

• "into" - integrated area of original (raw) peak

• "intf" - integrated area of filtered peak.

• "maxo" - maximum intensity of original (raw) peak.

• "maxf" - maximum intensity of filtered peak.

header.col.meta

boolean value indicating if the metadata CSV file contains a header column with the name of the metadata variables.

header.row.meta

boolean value indicating if the metadata CSV file contains a header row with the

name of the samples.

sep.meta the separator character of the metadata file.

### Value

Returns a dataset from the MS files.

### **Examples**

```
## Not run:
    ## Example of reading a dataset from MS files
    dataset = read_ms_spectra("data", type = "nmr-spectra", "metadata.csv",
        description = "description of the dataset")
## End(Not run)
```

read\_multiple\_csvs

Read multiple CSVs

### **Description**

Reads multiple CSVs, each one with a sample.

#### Usage

```
read_multiple_csvs(filenames, ext = ".csv", ...)
```

### **Arguments**

filenames list of file names of the files to read.

ext extension name.

... additional parameters to read.csv function.

#### Value

returns a list of dataframes.

## **Examples**

```
## Not run:
    ## Example of reading multiple csvs
    filenames = c("sample1", "sample2", "sample3", "sample4")
    sample.list = read_multiple_csvs(filenames, ext = ".csv")
## End(Not run)
```

recursive\_feature\_elimination

Perform recursive feature elimination

## **Description**

Perform recursive feature elimination on the dataset using caret's package.

### Usage

```
recursive_feature_elimination(datamat, samples.class,
functions = caret::rfFuncs, method = "cv", repeats = 5,
number = 10, subsets = 2^(2:4))
```

## **Arguments**

data matrix from dataset.

samples.class string or index indicating what metadata to use.

functions a list of functions for model fitting, prediction and variable importance.

method the external resampling method: boot, cv, LOOCV or LGOCV (for repeated

training/test splits.

repeats for repeated k-fold cross-validation only: the number of complete sets of folds

to compute.

number either the number of folds or number of resampling iterations.

subsets a numeric vector of integers corresponding to the number of features that should

be retained.

98 remove\_data

### Value

A caret's rfe object with the result of recursive feature selection.

## **Examples**

remove\_data

Remove data

## **Description**

Remove data from the dataset.

## Usage

```
remove_data(dataset, data.to.remove, type = "sample",
by.index = F, rebuild.factors = T)
```

# **Arguments**

### Value

Returns the dataset with the specified data removed.

remove\_data\_variables 99

## **Examples**

```
## Example of removing data
data(cachexia)
dataset = remove_data(cachexia, c("Creatine", "Serine"), type = "data",
  by.index = FALSE)
```

remove\_data\_variables Remove data variables

# Description

Remove data variables from the dataset.

# Usage

```
remove_data_variables(dataset, variables.to.remove,
by.index = FALSE)
```

# Arguments

```
dataset list representing the dataset from a metabolomics experiment.

variables.to.remove

vector with the variables' names to remove.

by.index if TRUE, the values of the variables.to.remove argument are indexes.
```

### Value

Returns the dataset with the specified data variables removed.

```
## Example of removing data variables
data(cachexia)
dataset = remove_data_variables(cachexia, c("Creatine", "Serine"),
   by.index = FALSE)
```

```
remove_metadata_variables
```

Remove metadata's variables

### **Description**

Remove metadata's variables from the dataset

# Usage

```
remove_metadata_variables(dataset, variables.to.remove)
```

## **Arguments**

```
dataset list representing the dataset from a metabolomics experiment. variables.to.remove vector with the metadata's variables to remove.
```

#### Value

Returns the dataset with the selected metadata's variables removed.

# **Examples**

```
## Example of removing metadata's variables
data(cassavaPPD)
dataset = remove_metadata_variables(cassavaPPD, c("varieties"))
```

remove\_peaks\_interval Remove interval of peaks

### **Description**

Removes peaks from a given interval.

# Usage

```
remove_peaks_interval(sample.df, peak.val.min, peak.val.max)
```

# **Arguments**

```
sample.df data frame with the samples' peaks.
```

peak.val.min minimum peak value.
peak.val.max maximum peak value.

## Value

Returns a data frame with the filtered samples' peaks.

# **Examples**

```
## Example of removing a interval of peaks
data(propolisSampleList)
samples.df = remove_peaks_interval(propolisSampleList[[1]], 2, 8.05)
```

```
remove_peaks_interval_sample_list

*Remove interval of peaks (sample list)
```

# Description

Removes peaks on a sample list given a peak interval.

## Usage

```
remove_peaks_interval_sample_list(sample.list, peak.val.min,
peak.val.max)
```

# **Arguments**

```
sample.list list of data frames with the samples' peaks.

peak.val.min minimum peak value.

peak.val.max maximum peak value.
```

### Value

Returns the sample list with the filtered peaks.

```
## Example of removing a interval of peaks in a sample list
data(propolisSampleList)
samples.list = remove_peaks_interval_sample_list(propolisSampleList, 2, 8.05)
```

remove\_samples

Remove samples

### **Description**

Remove samples from the dataset.

# Usage

```
remove_samples(dataset, samples.to.remove, rebuild.factors = T)
```

## **Arguments**

```
dataset list representing the dataset from a metabolomics experiment. samples.to.remove vector with the sample's names to remove. rebuild.factors

if TRUE, rebuilds the factors from metadata.
```

#### Value

Returns the dataset with the specified samples removed.

# **Examples**

```
## Example of removing samples
data(cachexia)
cachexia = remove_samples(cachexia, c("PIF_178","PIF_090"))
```

## Description

Remove samples from the dataset by the number of NAs

### Usage

```
remove_samples_by_nas(dataset, max.nas = 0, by.percent = F)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

max.nas number of NAs (or percentage) to which samples with more NAs will be re-

moved.

by percent if TRUE the max.nas argument is a percentage.

## Value

Returns the dataset with the samples with more NAs than the limit removed.

## **Examples**

```
## Example of removing samples by NAs
data(propolis)
propolis = remove_samples_by_nas(propolis, 40, by.percent = TRUE)
```

```
remove_samples_by_na_metadata
```

Remove samples by NA on metadata

# Description

Remove samples from the dataset with the metadata's variable value with NAs.

## Usage

```
remove_samples_by_na_metadata(dataset, metadata.var)
```

# **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.var metadata's variable.

## Value

Returns the dataset with the samples with NA on metadata's variable removed.

```
## Example of removing samples that have NAs on metadata's variable
data(cachexia)
cachexia$metadata$Muscle.loss[10] = NA
cachexia = remove_samples_by_na_metadata(cachexia, "Muscle.loss")
```

```
remove_variables_by_nas
```

Remove variables by NAs

# Description

Remove variables from the dataset by the number of NAs

# Usage

```
remove_variables_by_nas(dataset, max.nas = 0, by.percent = F)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

max.nas number of NAs (or percentage) to which samples with more NAs will be re-

moved.

by percent if TRUE the max.nas argument is a percentage.

### Value

Returns the dataset with the variables with more NAs than the limit removed.

# Examples

```
## Example of removing variables by NAs
data(propolis)
propolis = remove_variables_by_nas(propolis, 40, by.percent = TRUE)
```

```
remove_x_values_by_interval
```

Remove x-values by interval

## **Description**

Remove an interval of x-values from the dataset.

# Usage

```
remove_x_values_by_interval(dataset, min.value, max.value)
```

### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

min.value minimum value of the interval.
max.value maximum value of the interval.

replace\_data\_value 105

## Value

Returns the dataset with the specified interval of x-values removed.

## **Examples**

```
## Example of removing x-values by interval
data(cassavaPPD)
cassavaPPD = remove_x_values_by_interval(cassavaPPD, 200, 300)
```

replace\_data\_value

Replace data value

## **Description**

Replace a data value for a new value on the dataset.

## Usage

```
replace_data_value(dataset, x.axis.val, sample, new.value,
by.index = F)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

x.axis.val variable index or name.

sample sample name.

new.value new value to replace the old value.

by . index boolean value to indicate if the x.axis.val is an index or not.

#### Value

Returns the dataset with the data value replaced.

```
## Example of replacing a data value from the dataset
data(cachexia)
dataset = replace_data_value(cachexia, "Creatine", "PIF_178", 10.3,
   by.index = FALSE)
```

106 savitzky\_golay

```
replace_metadata_value
```

Replace metadata's value

# Description

Replace a metadata's variable value of a sample.

### Usage

```
replace_metadata_value(dataset, variable, sample, new.value)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

variable metadata's variable. sample name of the sample.

new.value new value of the metadata's variable.

### Value

Returns the dataset with the metadata updated.

# **Examples**

```
## Example of replacing metadata's variable value of a sample
data(cachexia)
dataset = replace_metadata_value(cachexia, "Muscle.loss", "PIF_178",
    "control")
```

savitzky\_golay

Savitzky-golay transformation

# Description

Smoothing and derivative of the data using Savitzky-Golay.

## Usage

```
savitzky_golay(dataset, p.order, window, deriv = 0)
```

scaling 107

# Arguments

dataset list representing the dataset from a metabolomics experiment.

p. order integer value representing the polynomial order.

window odd number indicating the window size.

deriv integer value indicating the differentiation order.

#### Value

Returns the dataset with the spectra smoothed using Savitzky-Golay.

## **Examples**

```
## Example of smoothing the spectra from dataset with Savitzky-Golay
data(cassavaPPD)
dataset.smoothed = savitzky_golay(cassavaPPD, p.order = 3, window = 11,
    deriv = 0)
```

scaling

Scale dataset

## **Description**

Performs scaling according to a method.

## Usage

```
scaling(dataset, method = "auto")
```

## Arguments

dataset

method

list representing the dataset from a metabolomics experiment. string specifying the scaling method. The possible values are:

- "auto" auto scaling.
- "range" range scaling.
- "pareto" pareto scaling.
- "tointerval" scaling to an interval.

#### Value

Returns the dataset scaled.

```
## Example of auto scaling
data(cassavaPPD)
cassava.scal = scaling(cassavaPPD, "auto")
```

108 set\_metadata

scaling\_samples

Scale data matrix

## **Description**

Performs scaling according to a method.

# Usage

```
scaling_samples(datamat, method = "auto")
```

# Arguments

datamat data matrix.

method string specifying the scaling method. The possible values are:

"auto" auto scaling. "range" range scaling. "pareto" pareto scaling.

• "tointerval" scaling to an interval.

### Value

Returns the data matrix scaled.

## **Examples**

```
## Example of auto scaling
data(cassavaPPD)
cassava.scal = scaling_samples(cassavaPPD$data, "auto")
```

set\_metadata

Set new metadata

# Description

Updates the dataset's metadata with a new one.

## Usage

```
set_metadata(dataset, new.metadata)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

new.metadata matrix or dataframe with the new metadata.

set\_sample\_names 109

## Value

Returns the dataset with the updated metadata.

## **Examples**

```
## Example of setting a new metadata to the dataset
data(cachexia)
new.metadata = c(rep("meta1", 39), rep("meta2", 38))
new.metadata = data.frame(var_meta = new.metadata)
rownames(new.metadata) = get_sample_names(cachexia)
cachexia = set_metadata(cachexia, new.metadata)
```

set\_sample\_names

Set samples names

## **Description**

Set new samples names to the dataset.

## Usage

```
set_sample_names(dataset, new.sample.names)
```

## **Arguments**

```
dataset list representing the dataset from a metabolomics experiment.

new.sample.names

vector with the new samples names.
```

## Value

Returns the dataset with the updated samples names.

```
## Example of setting a new value label to the dataset
data(cachexia)
new.samples.names = as.character(1:77)
cachexia = set_sample_names(cachexia, new.samples.names)
```

set\_x\_label

set\_value\_label

Set value label

## Description

Set a new value label for the dataset.

## Usage

```
set_value_label(dataset, new.val.label)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

new.val.label string with the new value label.

## Value

Returns the dataset with the updated value label.

## **Examples**

```
## Example of setting a new value label to the dataset
data(cachexia)
cachexia = set_value_label(cachexia, "new value label")
```

set\_x\_label

Set x-label

## **Description**

Set a new x-label to the dataset.

## Usage

```
set_x_label(dataset, new.x.label)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

new.x.label string with the x-label.

## Value

Returns the dataset with the updated x-label.

set\_x\_values 111

## **Examples**

```
## Example of setting a new x-label to the dataset
data(cachexia)
cachexia = set_x_label(cachexia, "new x-label")
```

set\_x\_values

Set new x-values

## Description

Set new x-values to the dataset

## Usage

```
set_x_values(dataset, new.x.values, new.x.label = NULL)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

new.x.values vector with the new x-values.

new.x.label string with the new x-label (can be NULL).

# Value

Returns the dataset with the updated x-values.

#### **Examples**

```
## Example of setting new x-values to the dataset
data(cachexia)
new.xvalues = 1:63
cachexia = set_x_values(cachexia, new.xvalues, new.x.label = NULL)
```

 $shift\_correction$ 

Shift correction

## **Description**

Shifts the spectra according to a specific method.

#### Usage

```
shift_correction(dataset, method = "constant", shift.val = 0,
interp.function = "linear", ref.limits = NULL)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

method string that indicates the shifting method. It can be:

• "constant" uses a constant shift that is added to the x-values

• "interpolation" uses interpolation according to "interp.function"

shift.val value of the shift (for constant and interpolation methods); can be a single value

for all spectra, can be the string "auto", the shifts are automatically determined or a vector with the size of the number of samples with the shifts for each spectra.

interp.function

string that represents the interpolation function, can be "linear" or "spline".

ref.limits vector with 2 elements that represents the reference limits to calculate the shifts.

## Value

Returns the dataset with the spectras shifted.

## **Examples**

smoothing\_interpolation

Smoothing interpolation

## Description

Performs smoothing interpolation according to a specific method.

#### Usage

```
smoothing_interpolation(dataset, method = "bin",
reducing.factor = 2, x.axis = NULL, p.order = 3,
window = 11, deriv = 0)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

method string specifying the smoothing method. The three possible methods are: "bin",

"loess" and "savitzky.golay".

reducing.factor

numeric value indicating the reducing factor for bin method.

snv\_dataset 113

x.axis	numeric vector representing the new x-axis for loess method.
p.order	numeric value representing the polynomial order for savitzky-golay method.
window	numeric value indicating the size of the window for savitzky-golay method. (must be an odd number)
deriv	numeric value indicating the differentiation order for savitzky-golay method.

## Value

Returns the dataset with the spectra smoothed.

## **Examples**

```
## Example of smoothing the spectra from dataset
data(cassavaPPD)
dataset.smoothed = smoothing_interpolation(cassavaPPD, method = "bin",
    reducing.factor = 2)
```

snv\_dataset

Standard Normal Variate

## Description

Performs Standard Normal Variate on the dataset.

## Usage

```
snv_dataset(dataset)
```

## **Arguments**

dataset

list representing the dataset from a metabolomics experiment.

#### Value

Returns the dataset with the data normalized by SNV.

```
## Example of SNV on a dataset
data(cassavaPPD)
dataset = snv_dataset(cassavaPPD)
```

stats\_by\_sample

spinalCord	Brazilian Propolis from different Harvest Seasons and different Agroe- cological Regions

## **Description**

This dataset consists of 12 LC-MS samples of spectra in the netCDF format, from mice spinal cord divided into 2 groups: the wild type and the knockout group. The data was obtained from the MetaboAnalyst site, originating from a study which describes a general strategy for identifying endogenous substrates of enzymes by untargeted LC-MS analysis of tissue metabolomes from wild-type and enzyme-inactivated organisms

## Usage

```
data(spinalCord)
```

#### **Format**

An object of class "list"

#### **Source**

MetaboAnalyst

#### References

A. Saghatelian, S.A. Trauger, E.J. Want, E.G. Hawkins, G. Siuzdak, B.F. Cravatt Assignment of Endogenous Substrates to Enzymes by Global Metabolite Profiling Biochemistry, 43:14332-14339, 2004.

## **Examples**

```
data(spinalCord)
sum_dataset(spinalCord)
```

stats\_by\_sample

Statistics of samples

## Description

Get a summary of statistics of the samples.

#### Usage

```
stats_by_sample(dataset, samples = NULL)
```

stats\_by\_variable 115

## Arguments

dataset list representing the dataset from a metabolomics experiment. samples if defined restricts the application to a given set of samples.

#### Value

Returns a vector with the a summary of statistics of the samples.

## **Examples**

```
## Example of getting stats of samples
data(cachexia)
samples.stats.result = stats_by_sample(cachexia)
```

stats\_by\_variable

Statistics of variables

## **Description**

Get a summary of statistics of the variables.

## Usage

```
stats_by_variable(dataset, variables = NULL,
variable.bounds = NULL)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

variables allows to define which variables to calculate the stats (if numbers, indexes are

assumed).

variable.bounds

allow to define an interval of variables (if numeric).

#### Value

Returns a vector with the a summary of statistics of the variables.

```
## Example of getting stats of variables
data(cachexia)
variable.stats.result = stats_by_variable(cachexia)
```

116 subset\_metadata

## Description

Gets a subset of specific samples and x-values.

## Usage

```
subset_by_samples_and_xvalues(dataset, samples, variables = NULL,
by.index = F, variable.bounds = NULL, rebuild.factors = T)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

samples vector with indexes or names of the samples to select

variables vector with the desired variables to get from the dataset.

by.index if TRUE, the values of the variables argument are indexes.

variable.bounds

variable bounds used if by.index is FALSE and variables are NULL.

rebuild.factors

if TRUE the metadata factors are rebuilded.

#### Value

Returns the dataset with the selected samples and x-values.

#### **Examples**

```
## Example of subsetting samples and x-values
data(cachexia)
subset = subset_by_samples_and_xvalues(cachexia, c("PIF_178","NETL_022_V1"),
    variables = c("Creatine", "Serine"))
```

subset\_metadata

Subset metadata

## **Description**

Subsets the metadata according to the specified metadata's variables.

# Usage

```
subset_metadata(dataset, variables)
```

subset\_random\_samples 117

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

variables metadata's variables.

#### Value

Returns the dataset with the metadata subsetted.

## **Examples**

```
## Example of subsetting samples
data(propolis)
subset = subset_metadata(propolis, c("seasons"))
```

subset\_random\_samples Subset random samples

## Description

Gets a subset of random samples from the dataset.

## Usage

```
subset_random_samples(dataset, nsamples)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

nsamples integer representing the number of samples that we want to get.

## Value

Returns the dataset with a number of random samples.

```
## Example of subsetting random samples
data(cachexia)
subset = subset_random_samples(cachexia, 15)
```

subset\_samples

Subset samples

## **Description**

Gets a subset of specific samples from the dataset.

## Usage

```
subset_samples(dataset, samples, rebuild.factors = T)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment. samples vector with indexes or names of the samples to select rebuild.factors

if TRUE the metadata factors are rebuilded.

#### Value

Returns the dataset with the selected set of samples.

## **Examples**

```
## Example of subsetting samples
data(cachexia)
subset = subset_samples(cachexia, c("PIF_178","PIF_132"))
```

```
subset_samples_by_metadata_values
```

Subset samples by metadata values

## **Description**

Gets a subset of specific samples according to metadata's values from the dataset.

# Usage

```
subset_samples_by_metadata_values(dataset, metadata.varname,
values)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment. metadata.varname

name of the metadata's variable.

values of the metadata's variable.

subset\_x\_values 119

## Value

Returns the dataset with the set of samples according to the metadata's values.

## **Examples**

```
## Example of subsetting samples by metadata's values
data(cassavaPPD)
subset = subset_samples_by_metadata_values(cassavaPPD, "varieties",
c("BRA","IAC"))
```

subset\_x\_values

Subset x-values

## **Description**

Gets a subset of specific x-values from the dataset.

## Usage

```
subset_x_values(dataset, variables, by.index = FALSE)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

variables vector with the desired variables to get from the dataset.

by index if TRUE, the values of the variables argument are indexes.

## Value

Returns the dataset with the selected set of x-values.

```
## Example of subsetting x-values
data(cachexia)
subset = subset_x_values(cachexia, c(1,2,10,20), by.index = TRUE)
```

## **Description**

Gets a subset of a specific interval of x-values.

## Usage

```
subset_x_values_by_interval(dataset, min.value, max.value)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

min.value the minimum value of the interval.

max.value the maximum value of the interval.

#### Value

Returns the dataset with the selected interval of x-values.

## **Examples**

```
## Example of subsetting x-values by an interval
data(cassavaPPD)
subset = subset_x_values_by_interval(cassavaPPD, 200, 800)
```

```
summary_var_importance
```

Summary of variables importance

## **Description**

Summary of variables importance of the models

## Usage

```
summary_var_importance(performances, number.rows)
```

## **Arguments**

performances the result from training the models.
number.rows number of variables to display.

sum\_dataset 121

## Value

Returns list with the variables importance of each model.

#### **Examples**

```
## Example of getting a summary of variables importance
data(cachexia)
training.result = train_models_performance(cachexia, "pls",
"Muscle.loss", "cv")
result = summary_var_importance(training.result, 10)
```

sum\_dataset

Dataset summary

## **Description**

Returns a summary with its main features.

## Usage

```
sum_dataset(dataset, stats = T)
```

# **Arguments**

dataset list representing the dataset from a metabolomics experiment. stats if TRUE prints some global statistics of the data values.

## Value

Returns the summary of the dataset containing:

- Description
- Type of data
- Number of samples
- Number of datapoints
- Number of metadata variables if metadata not null
- Labels of x axis values and data points if labels not null

If the parameter 'stats' is TRUE then returns also:

- Number of missing values in data
- Mean of data values
- Median of data values
- · Standard deviation
- · Range of values
- Quantiles

122 train\_and\_predict

## **Examples**

ain_and_predict
1

# Description

Train a model and predict new unlabeled samples with that model.

## Usage

```
train_and_predict(dataset, new.samples, column.class, model,
validation, num.folds = 10, num.repeats = 10, tunelength = 10,
tunegrid = NULL, metric = NULL, summary.function =
defaultSummary)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment. new.samples dataframe with new samples to predict the class label. column.class metadata column class. model model to be used in training. validation validation method. number of folds in cross validation. num.folds num.repeats number of repeats. tunelength number of levels for each tuning parameters. tunegrid dataframe with possible tuning values. metric metric used to evaluate the model's performance. Can be "Accuracy" or "ROC". summary.function summary function. For "ROC" the multiClassSummary function must be used.

## Value

Returns a list with the training result and the predictions result.

```
## Example of training and predicting
data(cachexia)
result = train_and_predict(cachexia, new.samples = cachexia$data,
"Muscle.loss", "pls", "cv")
```

train\_classifier 123

train_classifier	Train classifier
------------------	------------------

#### **Description**

Train a specific classifier.

## Usage

```
train_classifier(dataset, column.class, model, validation,
num.folds = 10, num.repeats = 10, tunelength = 10,
tunegrid = NULL, metric = NULL,
summary.function = defaultSummary, class.in.metadata = T)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

column.class metadata column class.

model model to be used in training.

validation validation method.

num. folds number of folds in cross validation.

num.repeats number of repeats.

tunelength number of levels for each tuning parameters.

tunegrid dataframe with possible tuning values.

metric metric used to evaluate the model's performance. Can be "Accuracy" or "ROC".

summary.function

summary function. For "ROC" the multiClassSummary function must be used.

class.in.metadata

boolean value to indicate if the class is in metadata.

## Value

Returns the train result object from caret.

```
## Example of training a classifier
data(cachexia)
result = train_classifier(cachexia, "Muscle.loss", "pls", "cv")
```

train\_models\_performance

Train models

## Description

Train various models.

## Usage

```
train_models_performance(dataset, models, column.class,
validation, num.folds = 10, num.repeats = 10, tunelength = 10,
tunegrid = NULL, metric = NULL, summary.function = "default",
class.in.metadata = T, compute.varimp = T)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

models models to be used in training.
column.class metadata column class.

validation validation method.

num. folds number of folds in cross validation.

num.repeats number of repeats.

tunelength number of levels for each tuning parameters.

tunegrid dataframe with possible tuning values.

metric metric used to evaluate the model's performance. Can be "Accuracy" or "ROC".

summary.function

summary function. For "ROC" the multiClassSummary function must be used.

class.in.metadata

boolean value to indicate if the class is in metadata.

compute.varimp boolean value to indicate if the var importance is calculated.

## Value

Returns a list with the results from training

performance The results from the best tunes of the models vips The variable importance from the models

full.results The full results from the tuning parameters of each model

best.tunes The best tune of each model

confusion.matrices

The confusion matrices of the models (only in classification)

final.models The final models

transform\_data 125

## **Examples**

```
## Example of training models
data(cachexia)
result = train_models_performance(cachexia, "pls",
   "Muscle.loss", "cv")
```

transform\_data

Transform data

## Description

Performs data transformation according to a method.

## Usage

```
transform_data(dataset, method = "log")
```

## Arguments

dataset list representing

list representing the dataset from a metabolomics experiment.

method

string specifying the transformation method. The possible values are:

- "log" logarithmic transformation.
- "cubicroot" cubic root transformation.

#### Value

Returns the dataset with the data transformation applied.

```
## Example of logarithmic transformation
data(cachexia)
dataset.log = transform_data(cachexia, "log")
```

126 tTests\_dataset

```
transmittance_to_absorbance
```

Convert transmittance to absorbance

#### **Description**

Converts transmittance values to absorbance values.

## Usage

```
transmittance_to_absorbance(dataset, percent = T)
```

## Arguments

dataset list representing the dataset from a metabolomics experiment.

percent boolean value to indicate if the absorbance values are in a percentage or not.

#### Value

Returns the dataset with the data points converted to absorbance values.

#### **Examples**

```
## Example of converting transmittance values to absorbance values
data(cassavaPPD)
dataset = transmittance_to_absorbance(cassavaPPD, percent = TRUE)
```

tTests\_dataset

t-Tests on dataset

## **Description**

Run t-Tests for each row of the data from the dataset.

## Usage

```
tTests_dataset(dataset, metadata.var, threshold = NULL,
write.file = F, file.out = "ttests.csv")
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

metadata.var metadata variable to use in the t-tests.

threshold threshold value of the p-value.

write.file boolean value to write or not a file with the results.

file.out name of the file.

values\_per\_peak 127

## Value

Table with the results of the t-tests, with p-value, -log10(p-value) and false discovery rate (fdr).

## **Examples**

```
## Example of t-Tests on dataset
data(cachexia)
ttests.result = tTests_dataset(cachexia, "Muscle.loss",
write.file = FALSE)
```

values\_per\_peak

Values per peak

## **Description**

Gets the number of values on each peak.

## Usage

```
values_per_peak(samples.df)
```

## **Arguments**

samples.df data frame

data frame with the samples' peaks.

# Value

Returns a vector with the number of values for each peak.

```
## Example of getting the number of values for each peak
data(propolis)
num.peaks = values_per_peak(propolis$data)
```

128 variables\_as\_metadata

values\_per\_sample

Values per peak

#### **Description**

Gets the number of values on each sample.

## Usage

```
values_per_sample(samples.df)
```

## **Arguments**

samples.df data frame with the samples' peaks.

#### Value

Returns a vector with the number of values for each sample.

## **Examples**

```
## Example of getting the number of values for each sample
data(propolis)
num.values.samples = values_per_sample(propolis$data)
```

variables\_as\_metadata Variables as metadata

## **Description**

Use one or more data variables as metadata variables.

# Usage

```
variables_as_metadata(dataset, variables, by.index = F)
```

# Arguments

dataset list representing the dataset from a metabolomics experiment.

variables name or index of the variables that are going to be used as metadata variables.

by . index boolean value indicating if the variables are indexes or names

#### Value

Returns the dataset with the variables removed from the data and added on the metadata.

volcano\_plot\_fc\_tt 129

## **Examples**

```
## Example of using a variable as metadata variable
data(cachexia)
dataset = variables_as_metadata(cachexia, "Creatine", by.index = FALSE)
```

## **Description**

Volcano plot to intersect the results from t-tests and fold change.

#### Usage

```
volcano_plot_fc_tt(dataset, fc.results, tt.results,
fc.threshold = 2, tt.threshold = 0.01)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

fc.results fold change results.

tt.results t-tests results.

fc.threshold fold change threshold value.

tt.threshold t-test p-value threshold.

## Value

Returns the name of the samples which intersects both fold change and t-tests results above the given thresholds.

```
## Example of a volcano plot
data(cachexia)
foldchange.results = fold_change(cachexia, "Muscle.loss", "control")
ttests.results = tTests_dataset(cachexia, "Muscle.loss")
volcano_plot_fc_tt(cachexia, foldchange.results, ttests.results,
    fc.threshold = 2, tt.threshold = 0.01)
```

130 x\_values\_to\_indexes

```
xvalue_interval_to_indexes
```

Get indexes of an interval of x-values

#### **Description**

Returns indexes corresponding to an interval of x-values (only to numerical values - spectra)

## Usage

```
xvalue_interval_to_indexes(dataset, min.value, max.value)
```

## **Arguments**

dataset list representing the dataset from a metabolomics experiment.

min.value mininum x-value of the interval.

max.value maximum x-value of the interval.

## Value

Returns a numeric vector with the indexes of the x-values interval

#### **Examples**

```
## Example of getting the indexes of an interval of x-values
data(propolis)
indexes.interval = xvalue_interval_to_indexes(propolis, 2.0, 5.5)
```

## **Description**

Returns the indexes corresponding to a vector of x-values (only to numerical values - spectra)

## Usage

```
x_values_to_indexes(dataset, x.values)
```

#### **Arguments**

dataset list representing the dataset from a metabolomics experiment.

x.values vector of x-values.

x\_values\_to\_indexes 131

# Value

Returns a numeric vector with the indexes of the x-values.

```
## Example of getting the indexes of the x-values
data(propolis)
indexes = x_values_to_indexes(propolis, c(1.3, 3.51, 6.03))
```

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