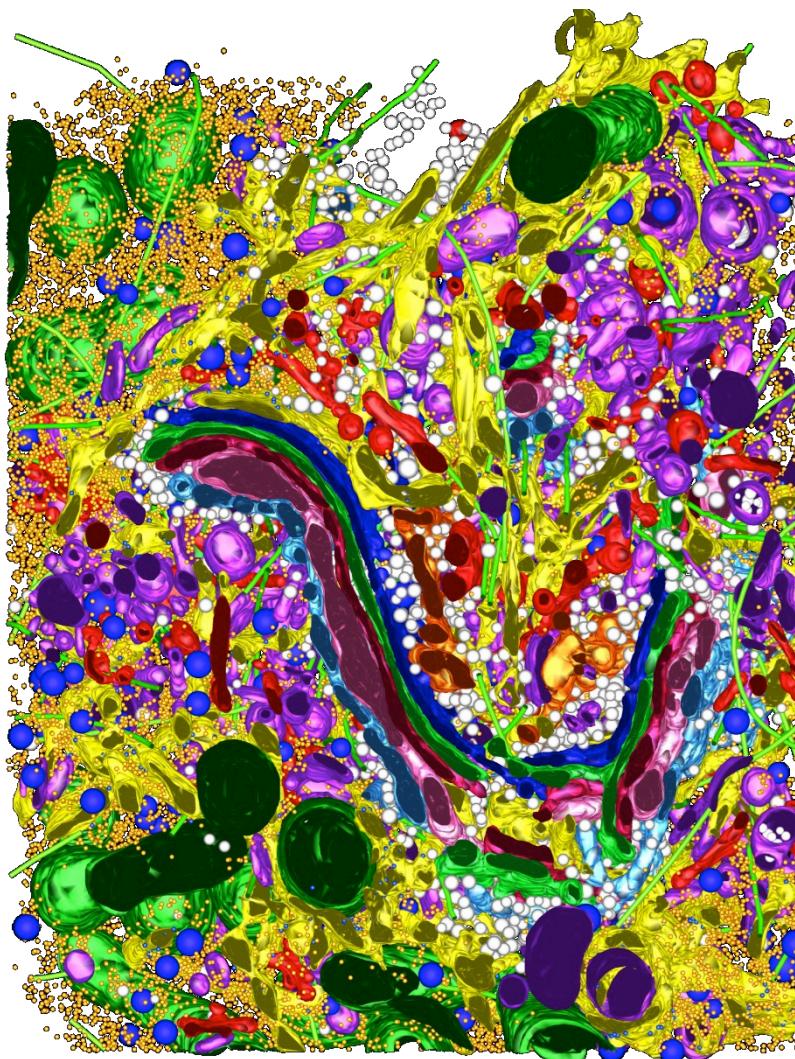


Unravelling the Complexity of Biological Samples using Isotopic Labelling

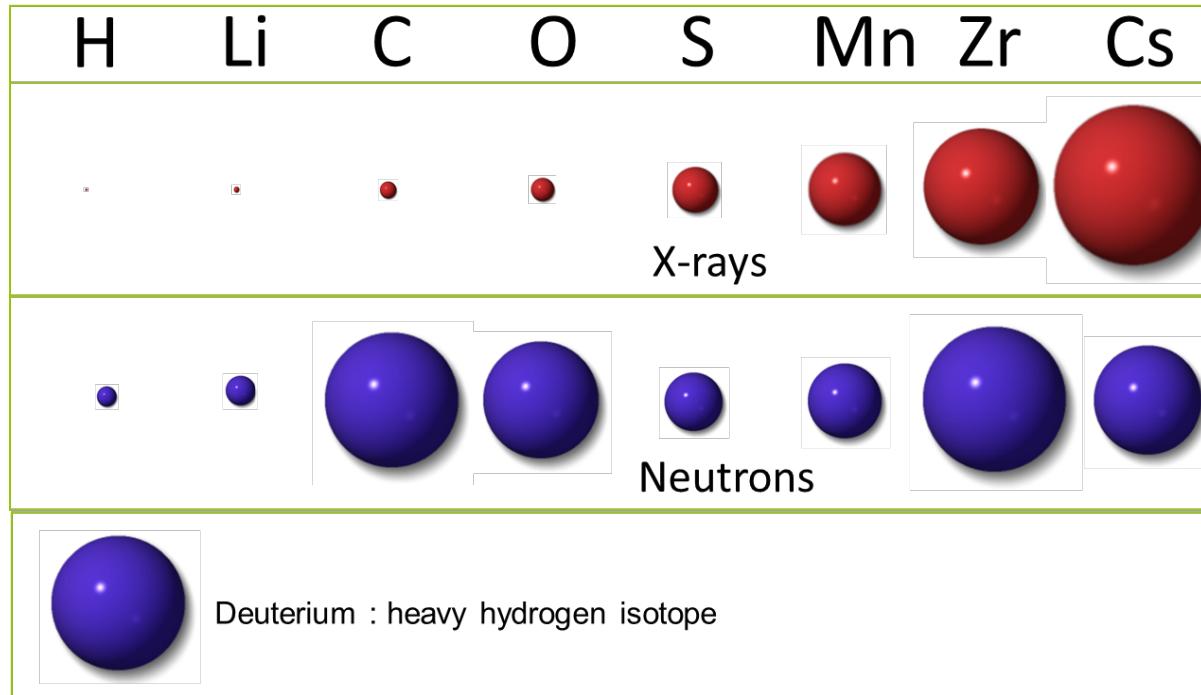
Luke Clifton
Luke.clifton@stfc.ac.uk

Biology is Complex



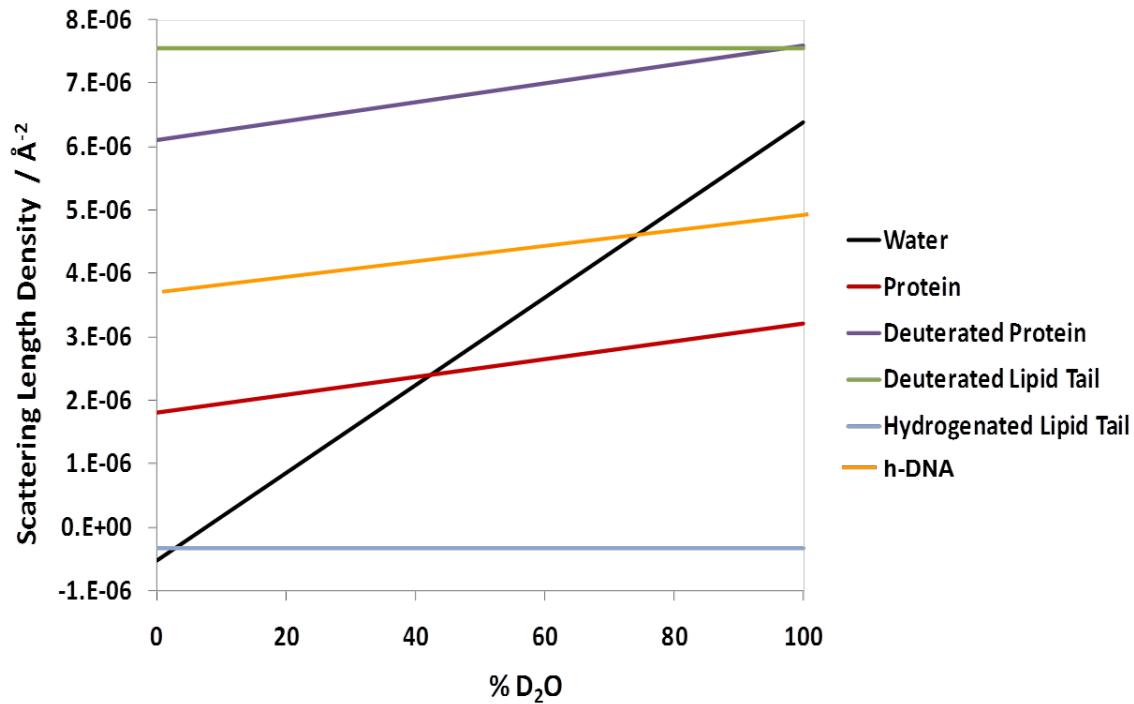
PNAS, 2001, 98, 5, 2399-2406

NS can easily differentiate different hydrogen isotopes

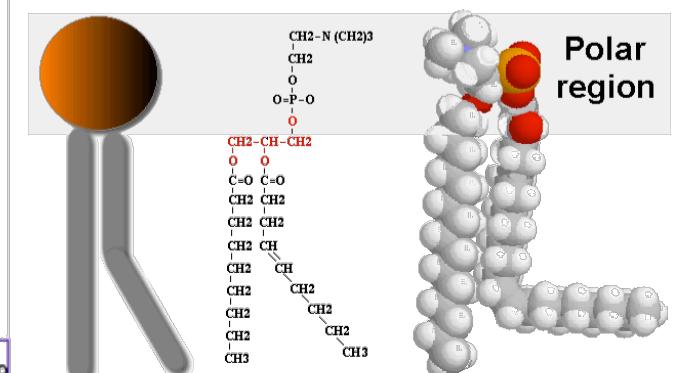
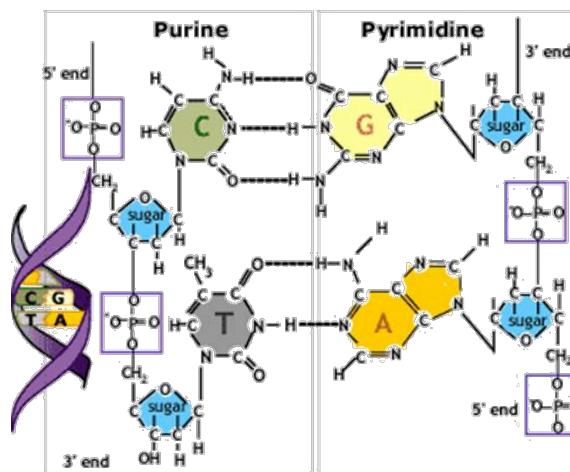
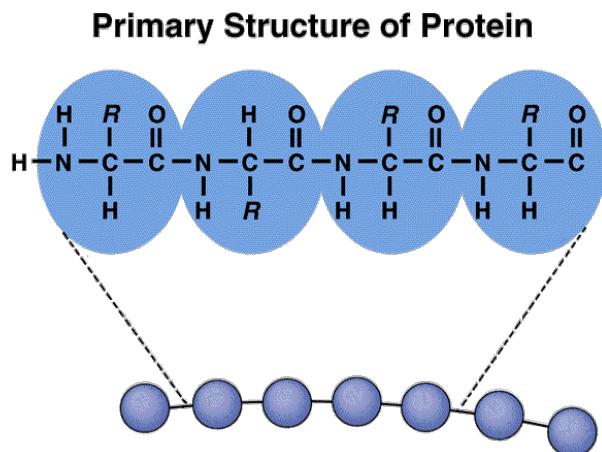


| Element | Coherent Scattering Length (b)/ 10^{-5} Å |
|-------------|---|
| Hydrogen | -3.74 |
| Deuterium | 6.671 |
| Carbon | 6.646 |
| Nitrogen | 9.36 |
| Oxygen | 5.803 |
| Sulphur | 2.847 |
| Phosphorous | 5.13 |

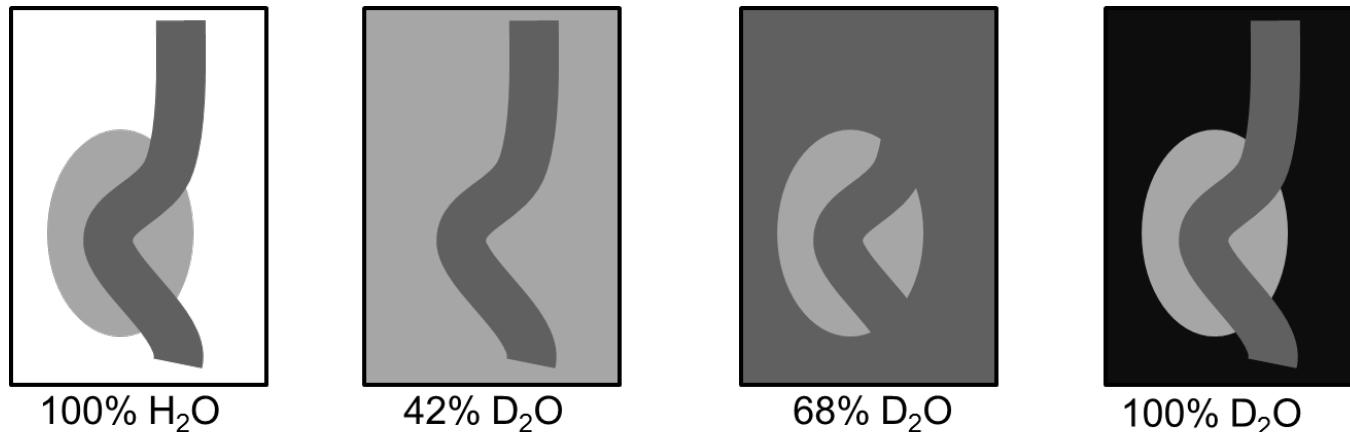
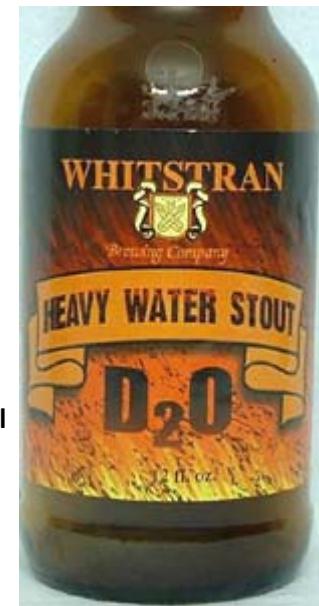
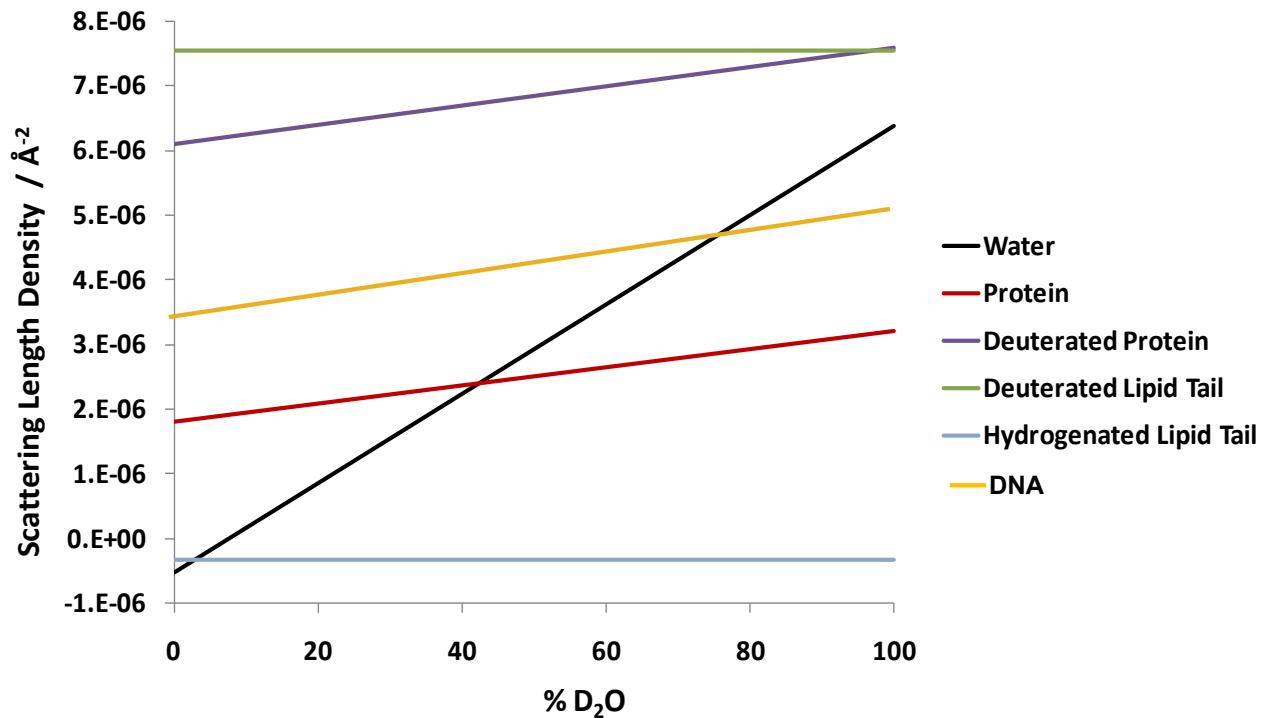
Natural contrast between biomolecules is present due to ^{14}N



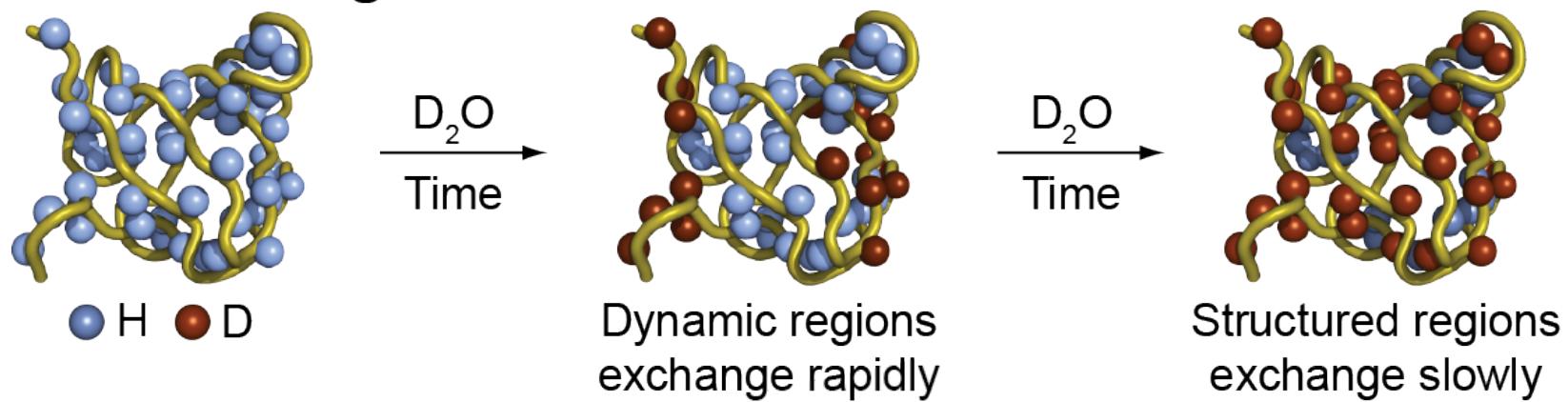
| Element | Coherent Scattering Length (b) / 10^{-5} Å |
|-------------|--|
| Hydrogen | -3.74 |
| Deuterium | 6.671 |
| Carbon | 6.646 |
| Nitrogen | 9.36 |
| Oxygen | 5.803 |
| Sulphur | 2.847 |
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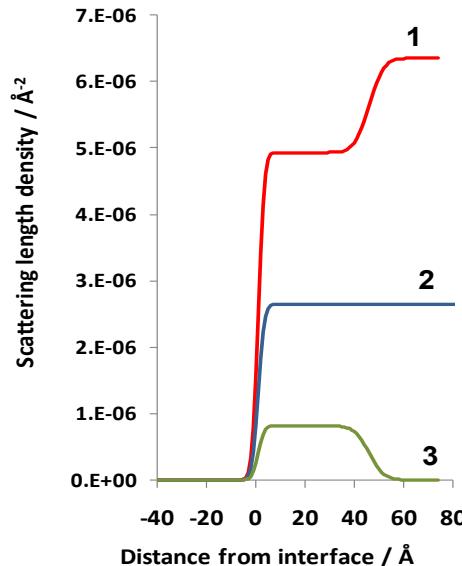
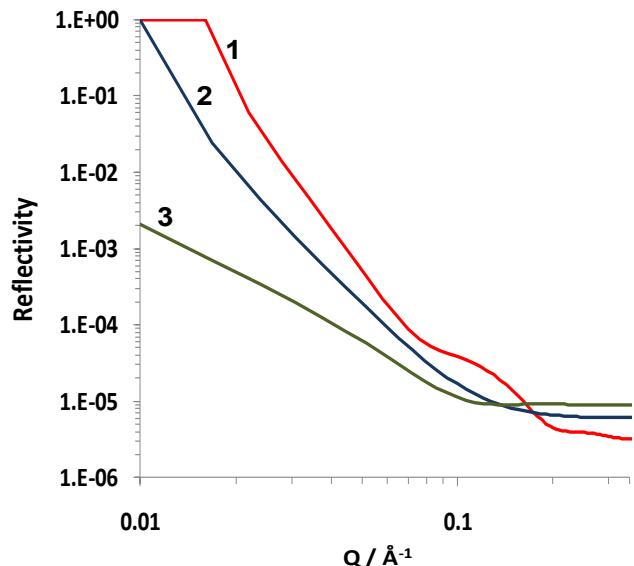
The easiest Labelling technique is to change the Labelling of the solution



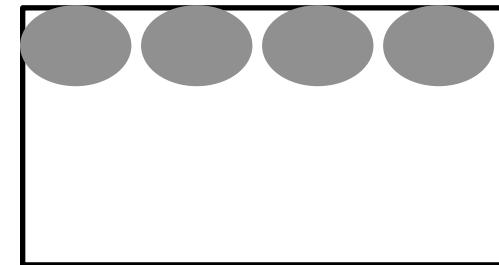
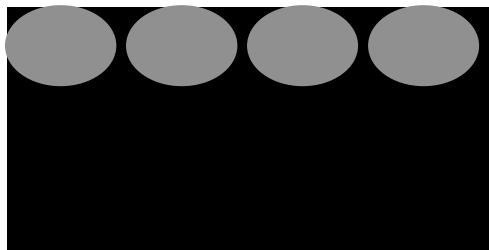
H/D Exchange



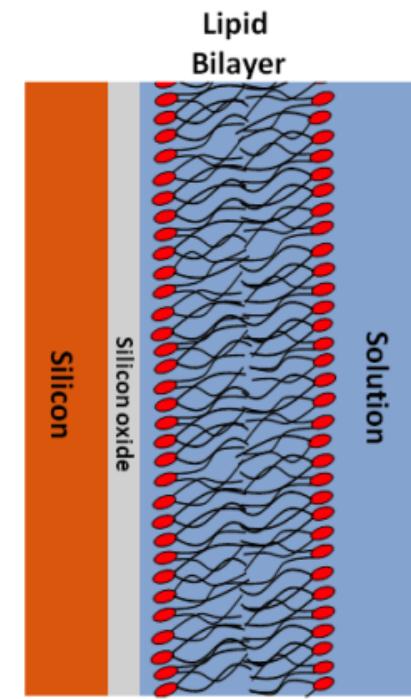
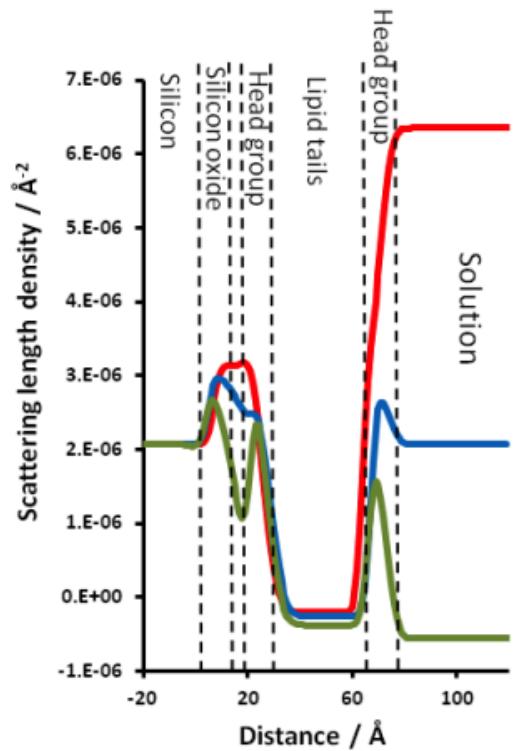
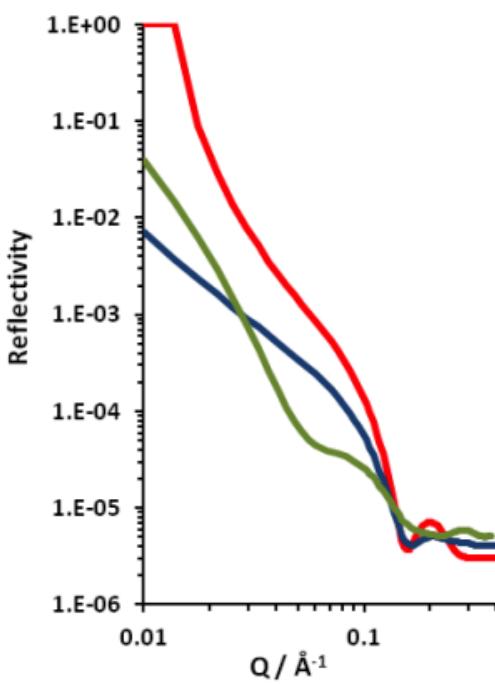
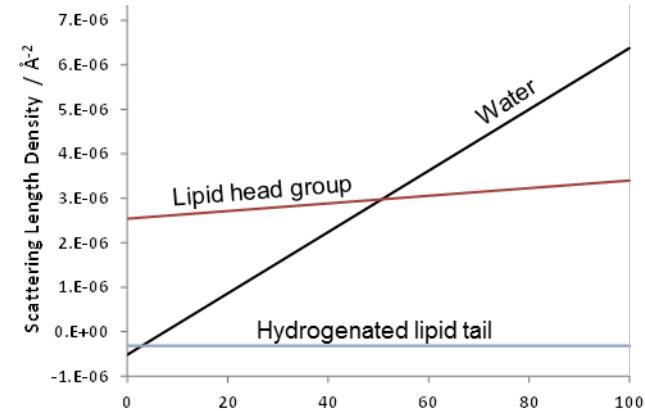
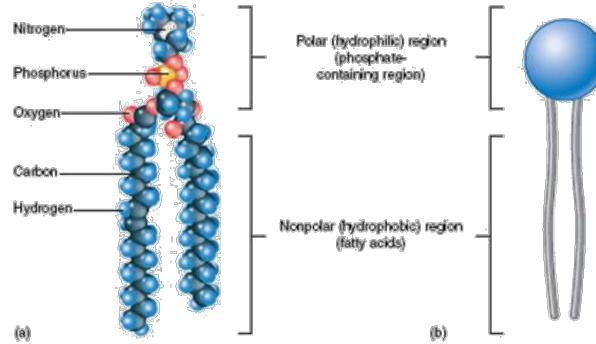
Matching sample and solution SLD



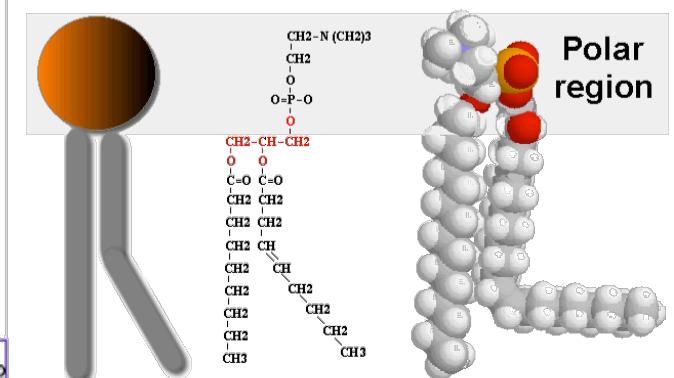
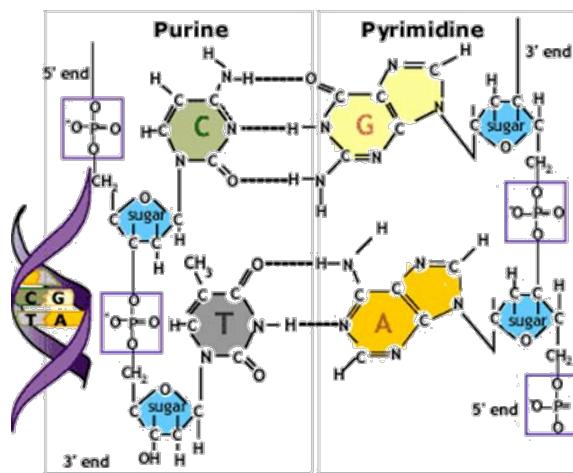
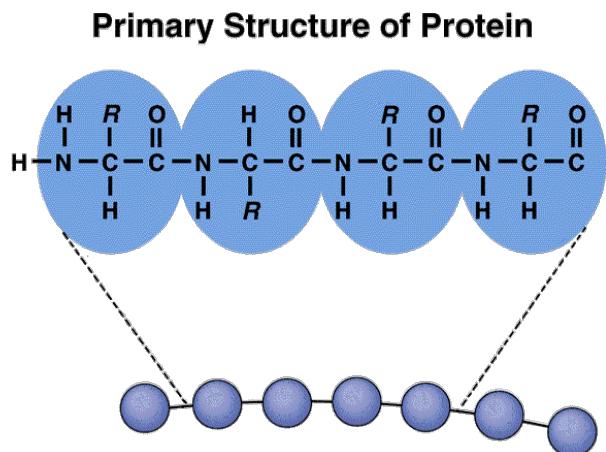
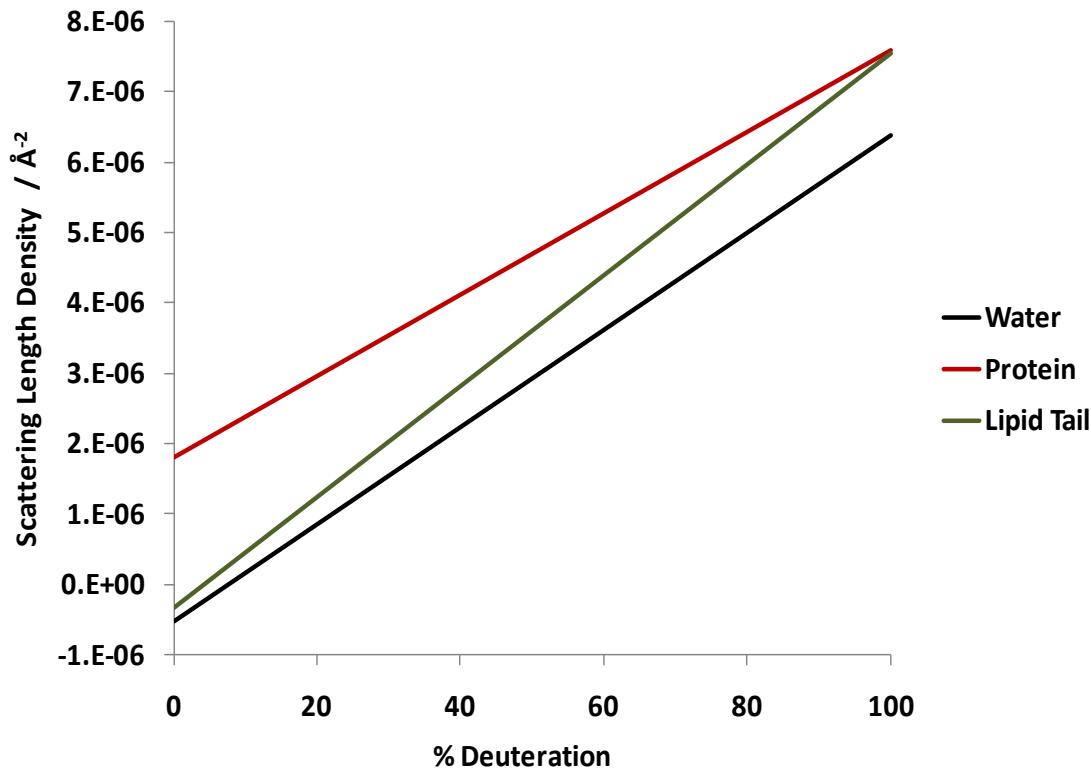
- 1. D_2O
- 2. 42% D_2O
- 3. H_2O



The easiest Labelling technique is to change the Labelling of the solution



Isotopic Labelling Changes the SLD of Biomolecules



Where do you get your labelled samples?

Oxford Isotope Facility

The ISIS Isotope Facility (formerly the Oxford Isotope facility) is able to produce deuterated small molecules for ISIS and ILL experiments.

Details of what is possible are below. If you think you will need to use the services of the isotope facility for your experiment please tick the box on the ISIS proposal form when you submit a proposal, and also contact John Webster (john.webster@stfc.ac.uk) to discuss your requirements.

Materials categories:

Category A

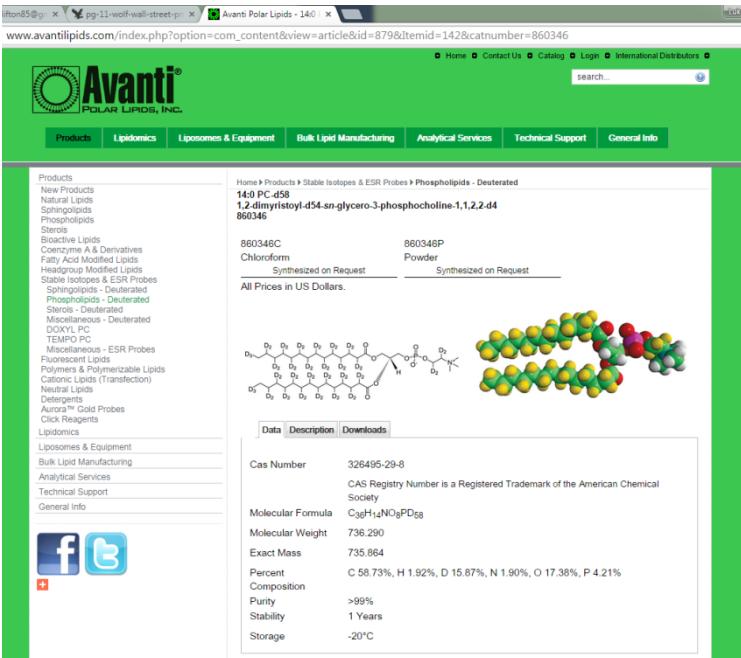
Perdeuterated fatty acids of most chain lengths from C6 to C20, including odd numbered chains, and the alcohol and bromoalkane versions of these. It is envisaged that up to 10 g of any compound could be made available instantly (a case will have to be made for quantities larger than 1g).

Sodium dodecyl sulphate. Some chain deuterated alkyl trimethylammonium bromides (C12, C14, C16, enquire about others). Some chain deuterated oligoethyleneglycol monoether non-ionic surfactants (enquire).

Category B

The compounds in this category include most compounds with a straight chain alkyl group, e.g. alkyl trichlorosilanes, alkane thiols, alkane sulphonates, dialkyl dimethyl ammonium halides.

Some dicarboxylic acids (C12, enquire about others) and their corresponding dialcohols and dibromides. Ethylhexanoic acid and corresponding alcohol and bromide.





Silantes

Stable Isotope Labeled Biomolecules

The Deuteration Laboratory

DLab

- Home
- The Lab
 - Introduction
 - Equipment Available
 - Protocols
 - Collaborations
 - Staff
 - Positions Available
 - Selected Publications
- Users/Access
 - Application Form
 - Safety
- Contracts and funding
- EPSRC project description
- Photo Galleries
- Presentations & Videos
- Useful Links
 - Workshops/Meetings
 - ILL Forms
 - Useful Links
 - Facilities for analysis
- PSB

News

- Monday meetings schedule
- D-Lab Forum
- The new PSB-Get together meetings
- D-Lab's artwork

Quick links:

- The HTX Lab
- CISB Stores
- Site plan
- Phone - How to
- EPSRC Medical services



The Deuteration Laboratory
Institut Laue Langevin, CSF
6 Rue Jules Horowitz
38042 Grenoble cedex 9, France.

Send suggestions/comments concerning this website to Silantes@ill.fr or Maxine.Cupper@ill.fr
Last updated on May 24, 2012
Visitors (since 12/04/2007): www.freecounter.net
network: 130.246.0.0/16 Rutherford Appleton Laboratory

Many thanks to R. Leal and C. Bages for help & suggestions with this website.



Starting an experiment : Calculating ρ

$$\rho = \frac{\sum b}{V}$$

| Lipid / Solvent | Neutron scattering length density (ρ) (10^{-6} \AA^{-2}) |
|-----------------------------------|---|
| D ₂ O | 6.35 |
| H ₂ O | -0.56 |
| Silicon | 2.07 |
| Silicon oxide (SiO ₂) | 3.41 |
| Deuterated-tails (gel phase) | 7.45 |
| Hydrogenous-tails (gel phase) | -0.37 |
| h-Protein in D ₂ O | ~3.4 |
| h-Protein in H ₂ O | ~2.0 |
| h-DNA in D ₂ O | ~3.2 |
| h-DNA in H ₂ O | ~3.8 |

Web tools can help : NIST

<https://www.ncnr.nist.gov/resources/activation/>

NIST Center for Neutron Research

Home Instruments Science

Material: Si

Neutron Activation: For rabbit system Calculate

| | | |
|---------------------|--------------------|-------------------------|
| Thermal flux 1e8 | Cd ratio 0 | Thermal/fast ratio 0 |
| Mass 1 | Time on beam 10 | Time off beam 1 y |

Absorption and Scattering: Calculate

| | |
|--------------------------|------------------------|
| Density 2.32 | Thickness 1 |
| Source neutrons 1 Ang | Source X-rays Cu Ka |

Si at 2.32 g/cm³

Source neutrons: 1.000 Å = 81.80 meV = 3956 m/s
Source X-rays: 1.542 Å = 8.042 keV

| 1/e penetration depth (cm) | | Scattering length density (10 ⁻⁶ /Å ²) | | Scattering cross section (1/cm) | | X-ray SLD (10 ⁻⁶ /Å ²) | |
|-------------------------------|---------|--|--------|------------------------------------|-------|--|--------|
| abs | 211.367 | real | 2.065 | coh | 0.108 | real | 19.984 |
| abs+incoh | 202.836 | imag | -0.000 | abs | 0.005 | imag | -0.456 |
| abs+incoh+coh | 8.886 | incoh | 0.089 | incoh | 0.000 | | |

Neutron transmission is 99.51% for 1 cm of sample (after absorption and incoherent scattering).
Transmitted flux is 9.951e+7 n/cm²/s for a 1e8 n/cm²/s beam.

Biological Scattering Tool Website : Scattering Length Density Calculator

<http://psldc.isis.rl.ac.uk/>

Biomolecular Scattering Length Density Calculator

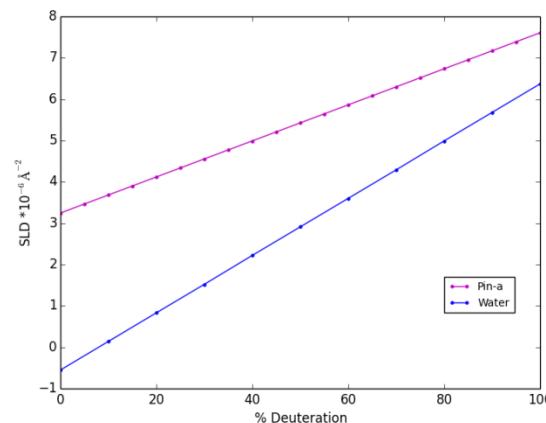
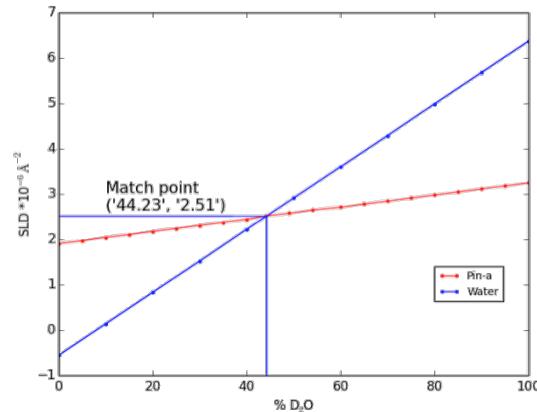
TITLE:

Type of biomolecule :
 Protein / Peptide
 RNA (A,U,G,C)
 DNA(A,T,G,C)

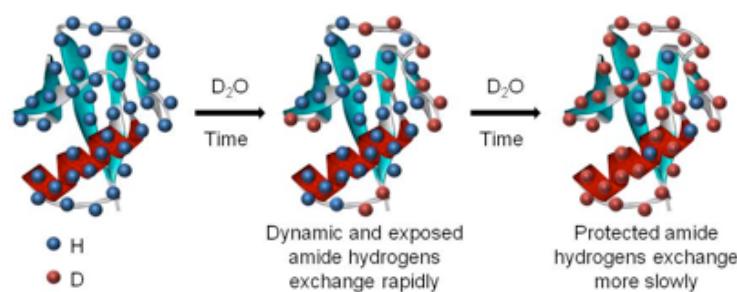
Primary structure:
DV AGGGGAQQCP VETKLNSCRN YLLDRCSTMK DFPPVTWRWIK WWKGCCQELL
GECSSRLGQM PPQCRCNIIQ GSIQGDLLGI FGFQRDRASK VIQEAKNLPP
RCNQGPPCNI PGTI|

OPTIONS:

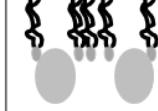
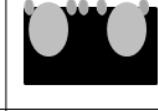
1. Solution D₂O% *¹
2. Deuteration % *²
3. Exchange % *³
4. Concentration of sample (mg/mL)

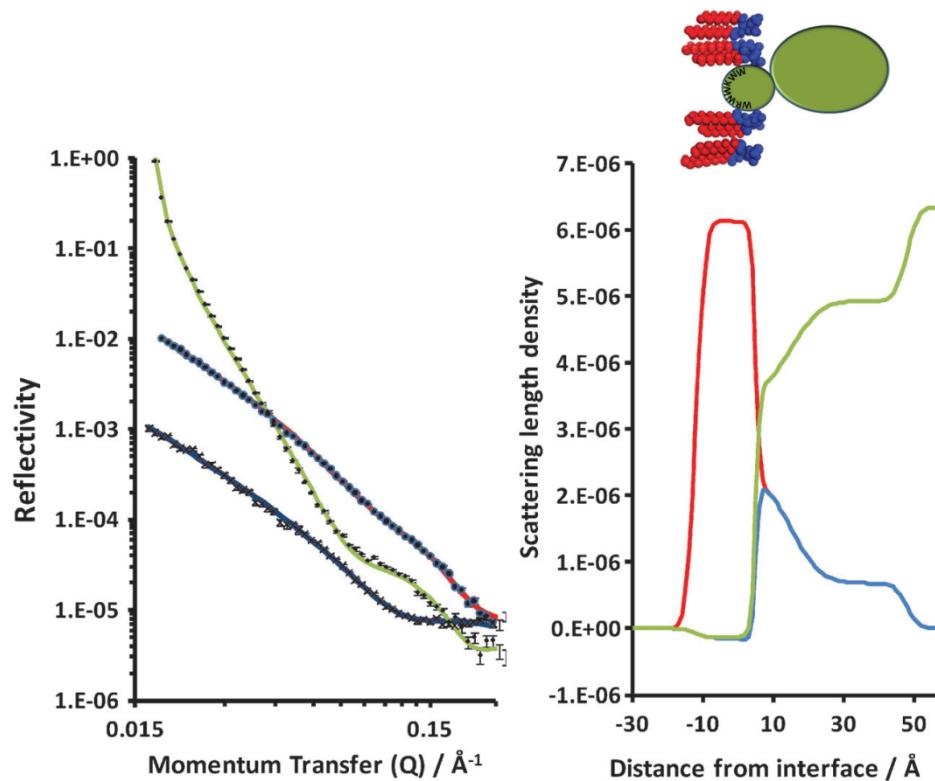


H/D Exchange



Choose your Contrasts

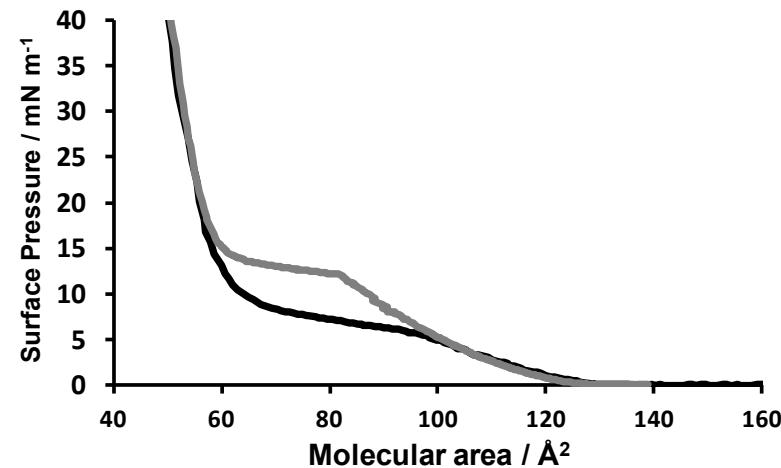
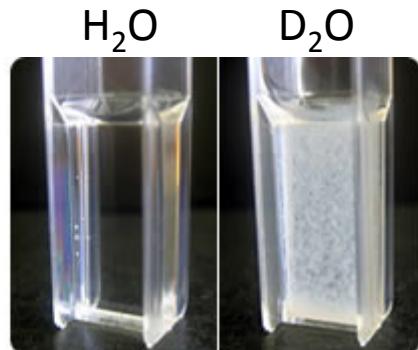
| # | Isotopic Contrast | Main Information Content of Reflectivity Data | Contrast Cartoon |
|---|--|---|---|
| 1 | Deuterated-lipid : Hydrogenated-lipid : Protein on Null Reflective Water | Lipid component |  |
| 2 | Hydrogenated-lipid : Hydrogenated-lipid : Protein on Null Reflective Water | Protein component |  |
| 3 | Hydrogenated-lipid : Hydrogenated-lipid : Protein on D ₂ O | Solution and Hydrogenous material - both lipid and protein components |  |
| 4 | Deuterated-lipid : Hydrogenated-lipid : Protein on D ₂ O | Solution and Protein component |  |
| 5 | Hydrogenated-lipid : Deuterated-lipid : Protein on Null Reflective Water | Protein component |  |
| 6 | Deuterated-lipid : Deuterated-lipid : Protein on Null Reflective Water | Deuterated material - both lipid and protein components |  |



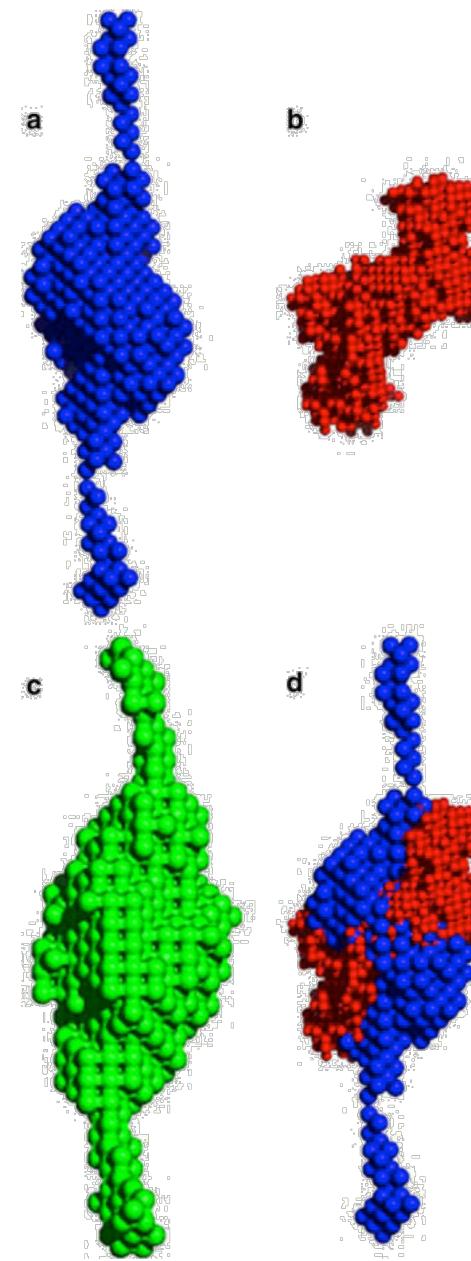
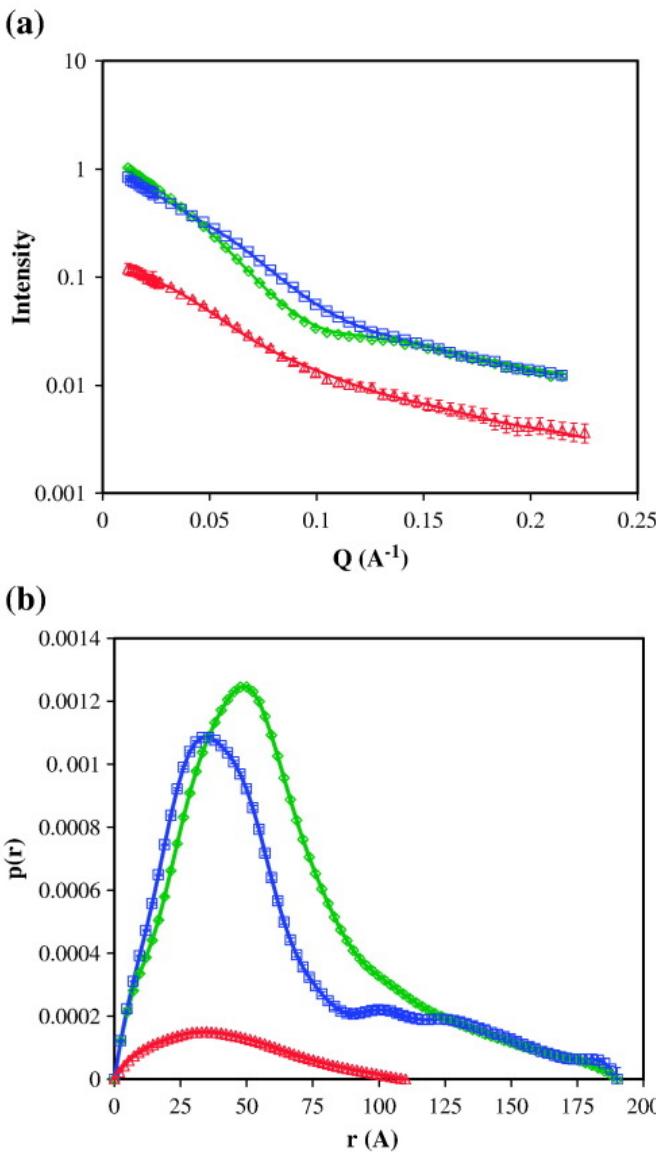
$$\rho_{fitted} = (\varphi_{lipid} \cdot \rho_{lipid}) + (\varphi_{protein} \cdot \rho_{protein}) + (\varphi_{water} \cdot \rho_{water})$$

BEWARE

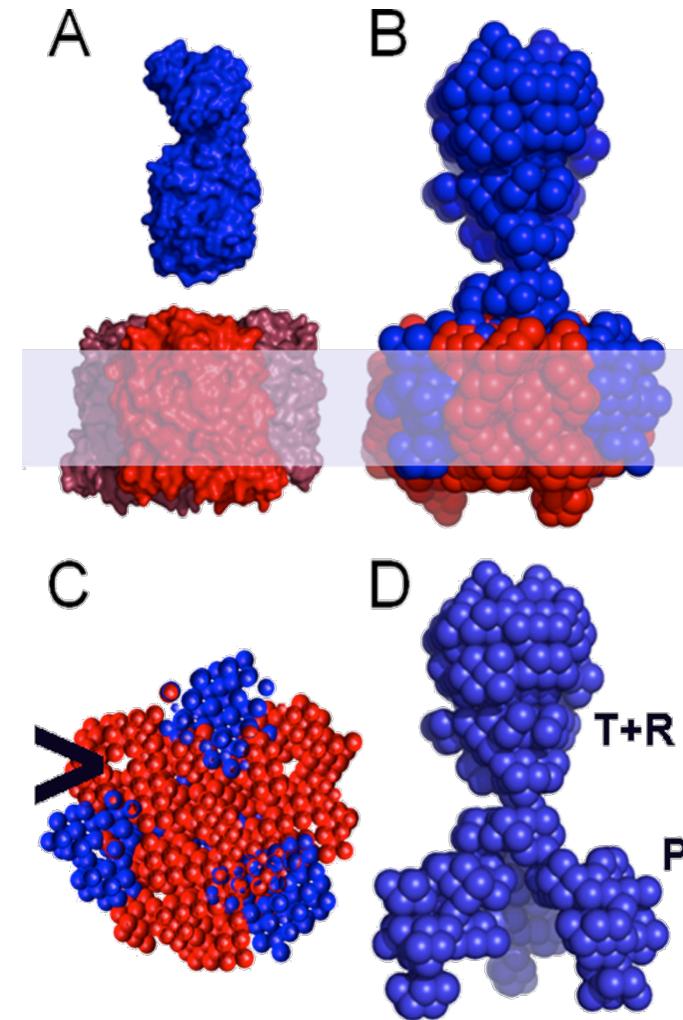
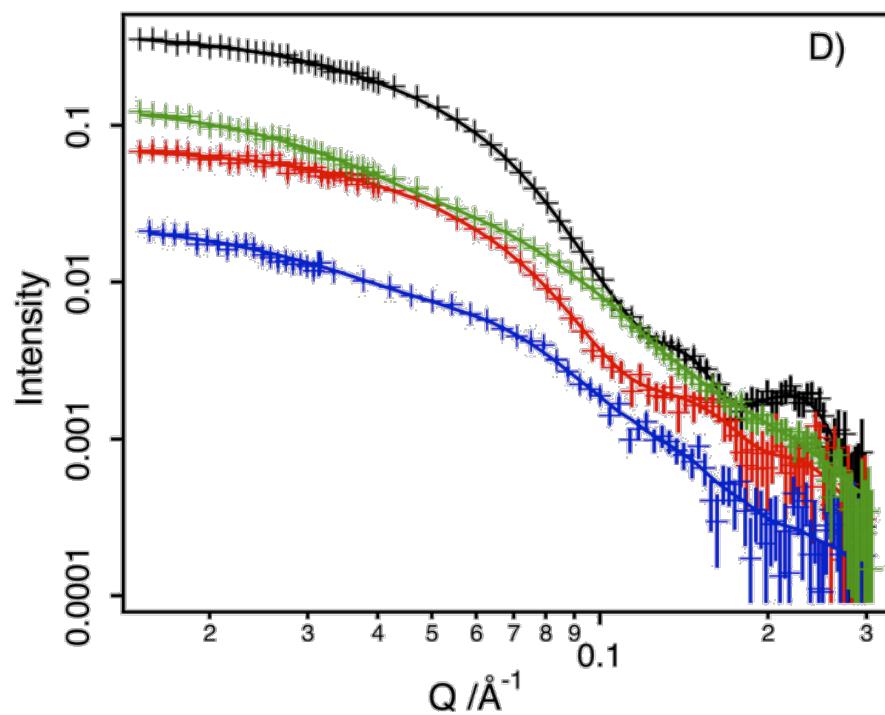
- D₂O and H₂O although chemically very similar are not the same!
 - Slight Differences in Nature of Hydrogen and Deuterium Bonding!
 - Due to more restricted O-D bond vibration vs. O-H, D₂O forms stronger dipole-dipole bind.
 - D₂O Melts at 3.7°C vs. 0°C.



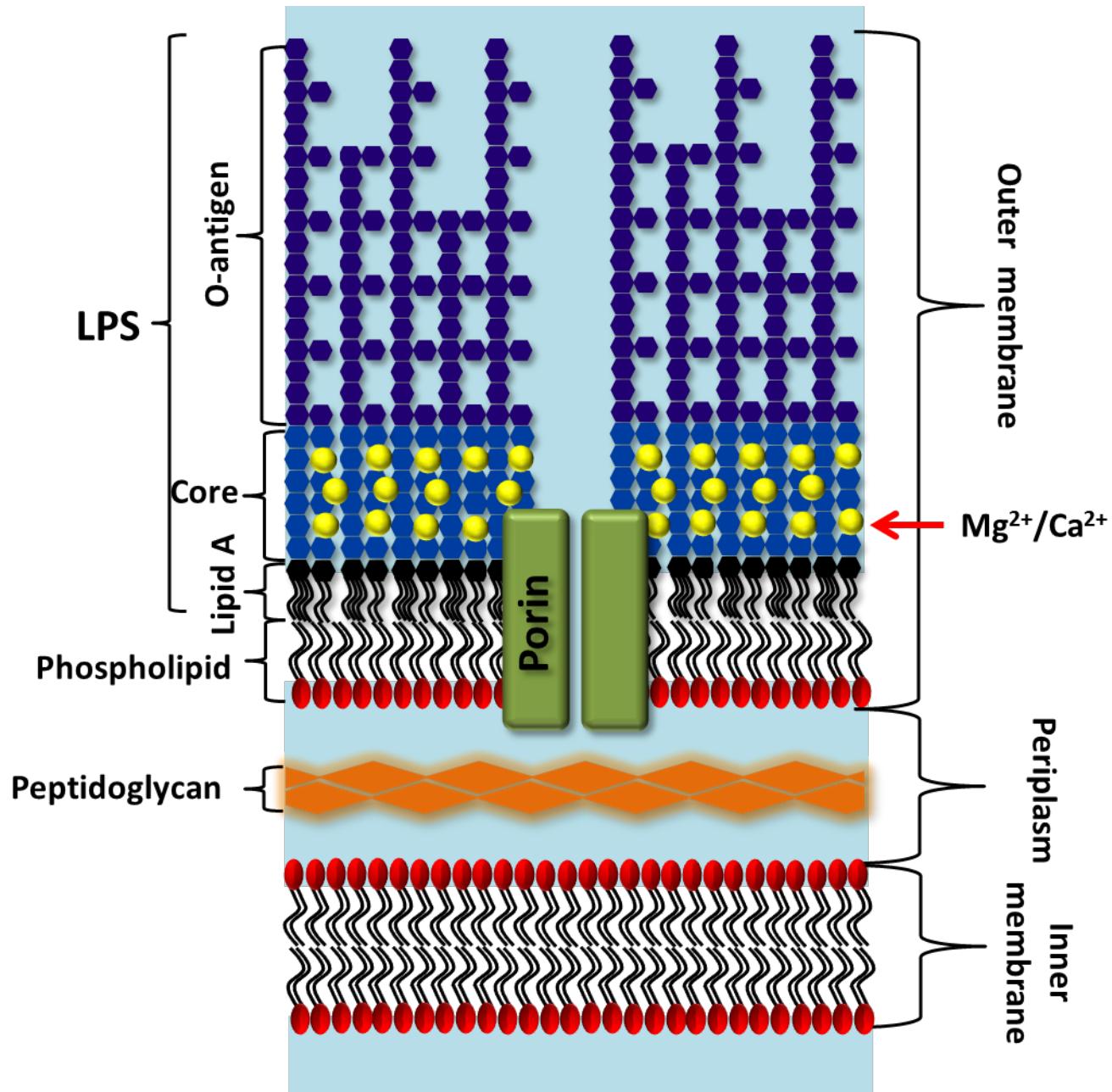
Sample deuterium labelling is often required : SANS



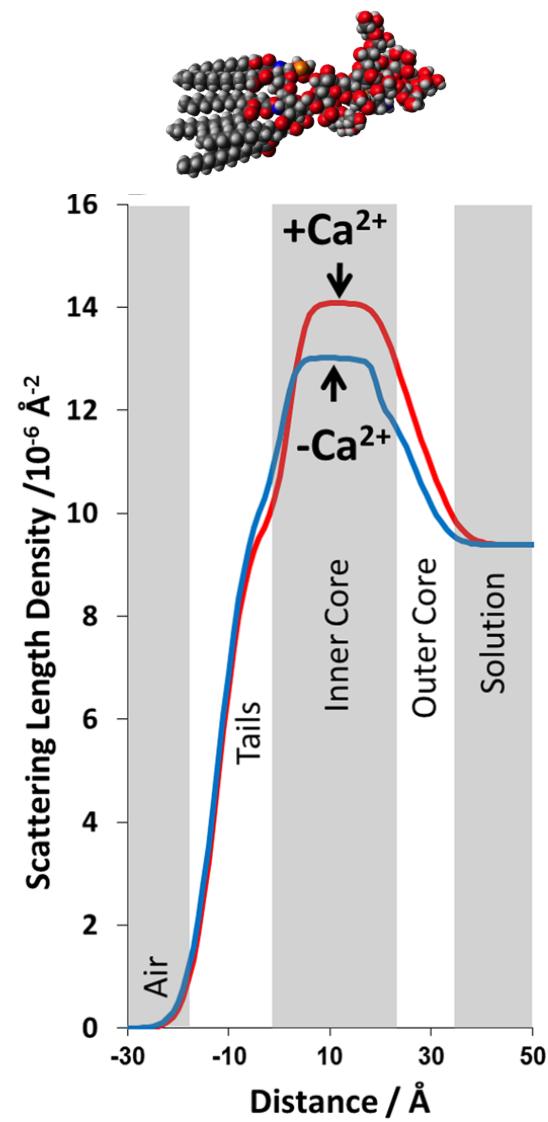
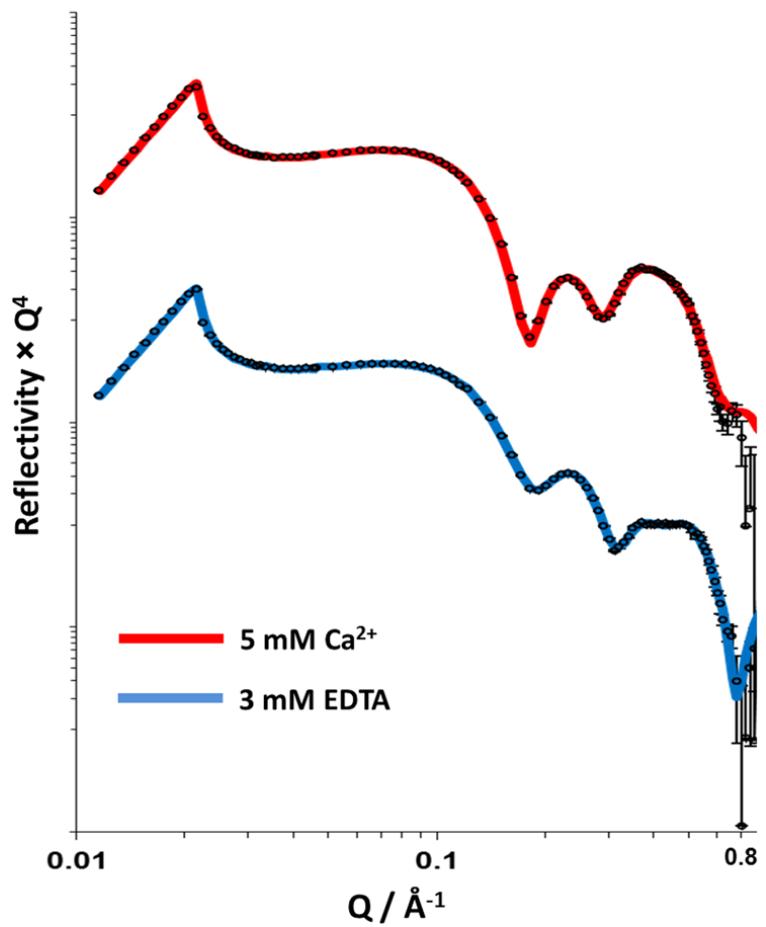
Sample deuterium labelling is often required : SANS



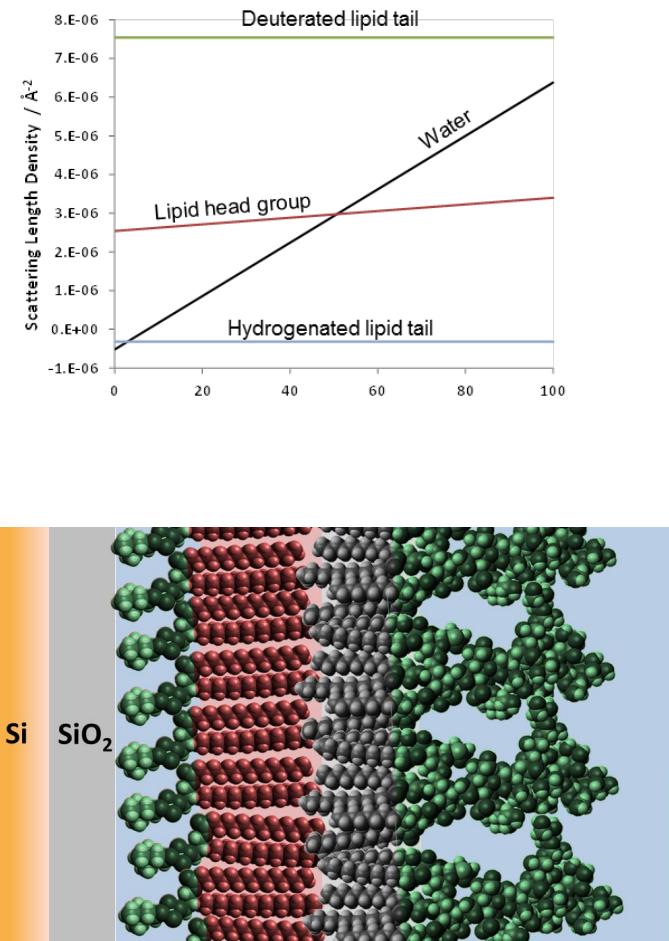
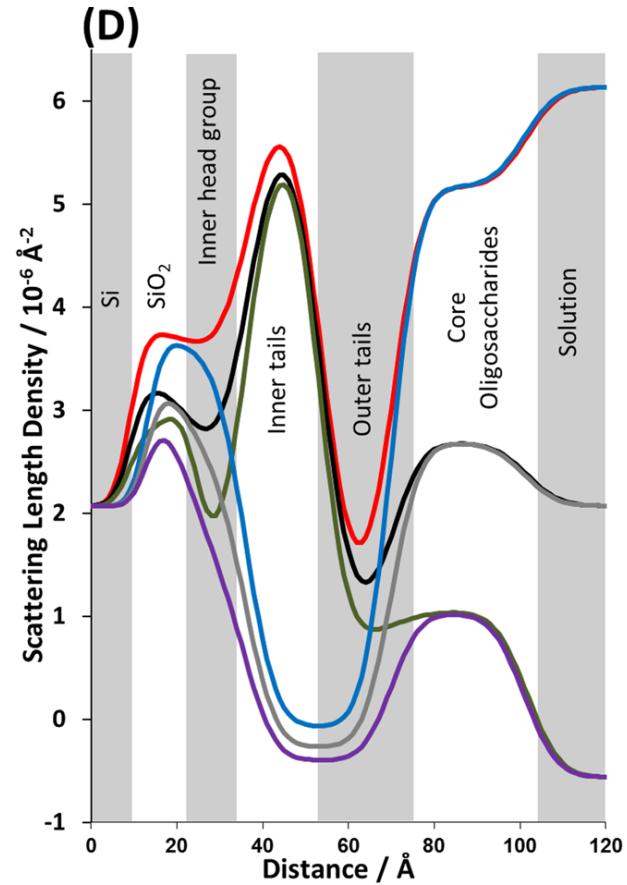
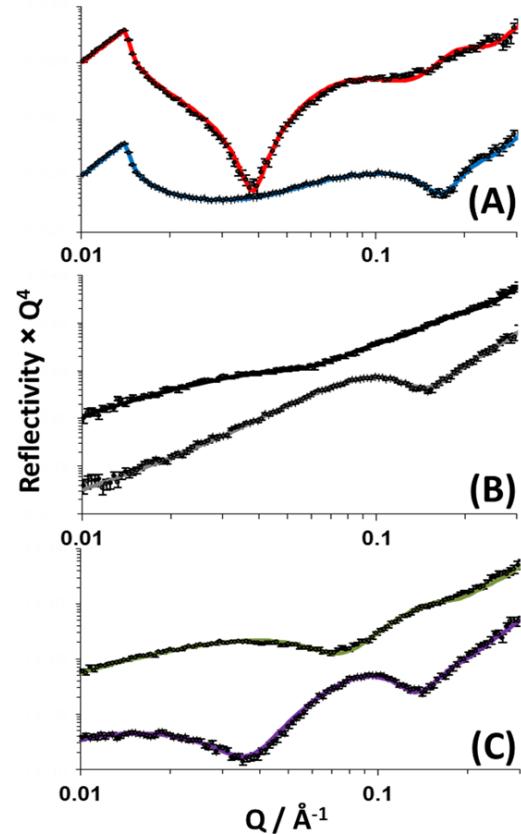
Sample labelling is often required : NR



Sample labelling : X-rays find electron rich elements!



Sample deuterium labelling is often required : NR



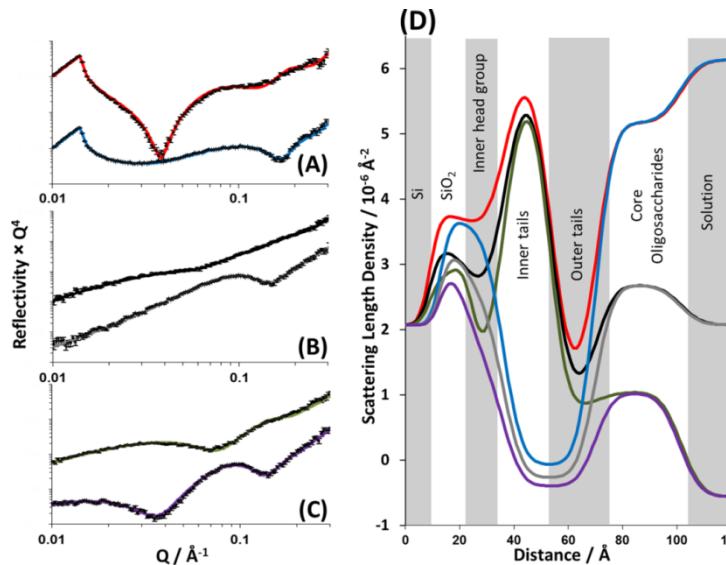
Determining Coverage and Asymmetry

$$\rho_{fitted\ D2O} = (\varphi_{h-lipid} \cdot \rho_{h-lipid}) + (\varphi_{d-lipid} \cdot \rho_{d-lipid}) + (\varphi_{D2O} \cdot \rho_{D2O})$$

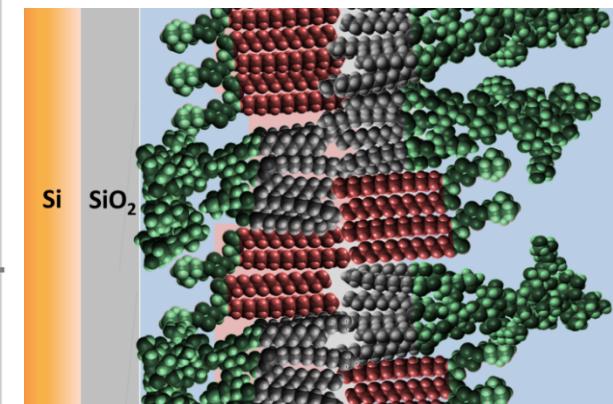
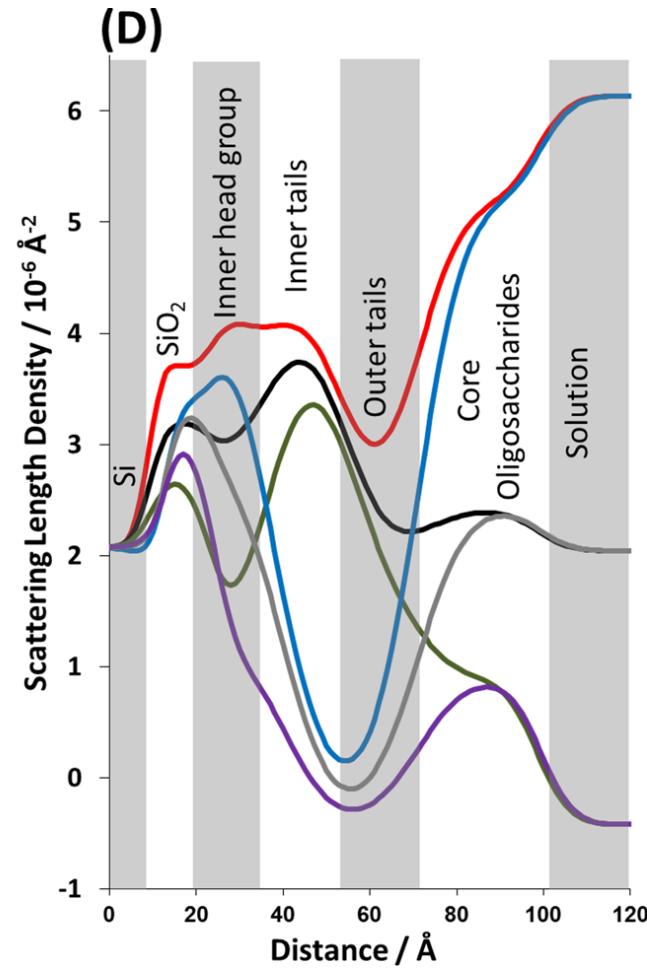
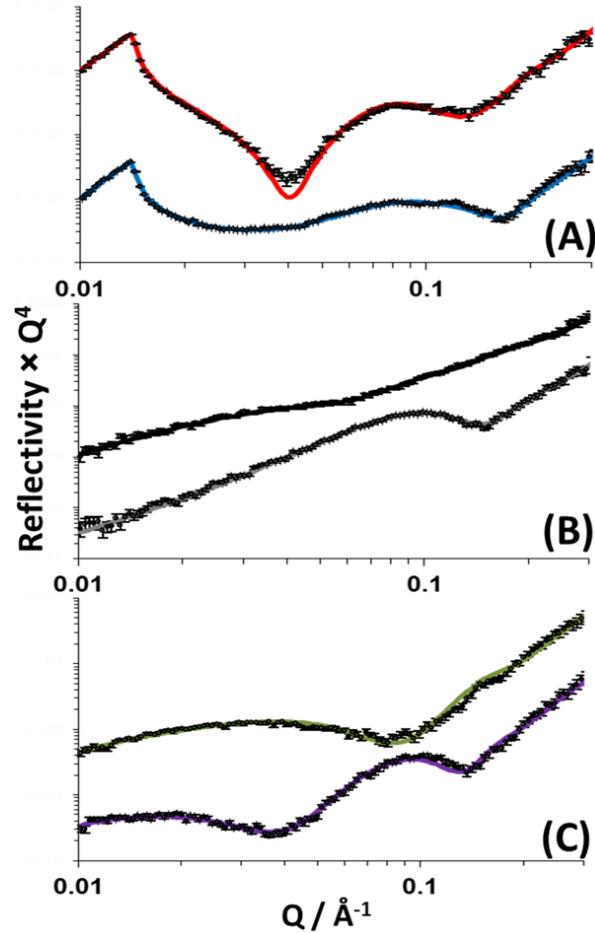
$$\rho_{fitted\ H2O} = (\varphi_{h-lipid} \cdot \rho_{h-lipid}) + (\varphi_{d-lipid} \cdot \rho_{d-lipid}) + (\varphi_{H2O} \cdot \rho_{H2O})$$

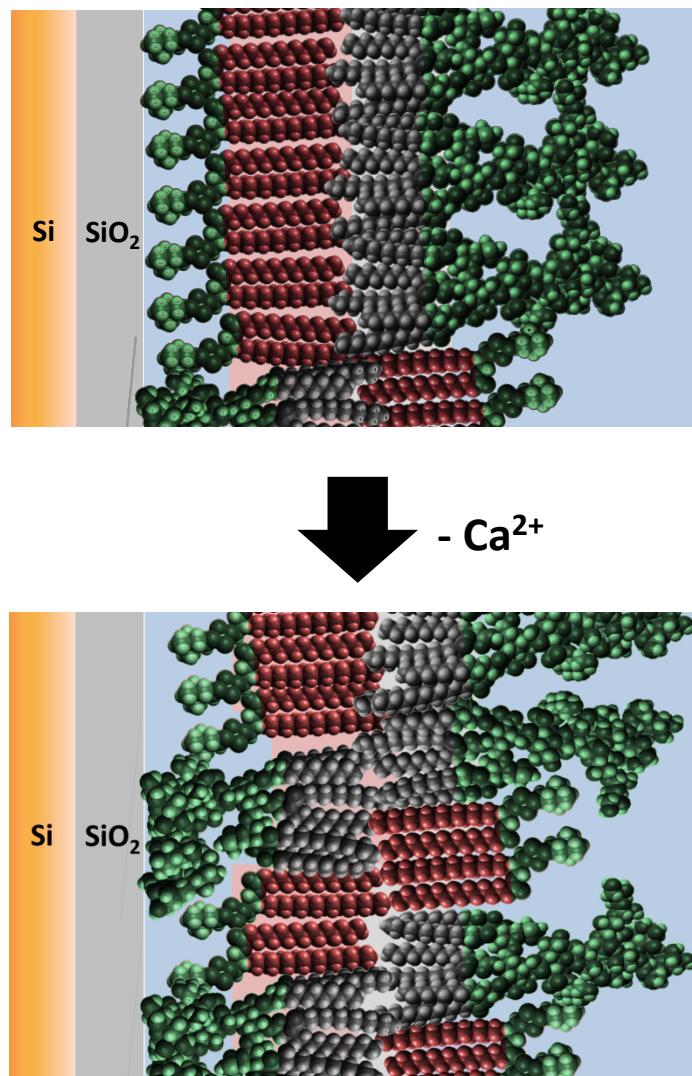
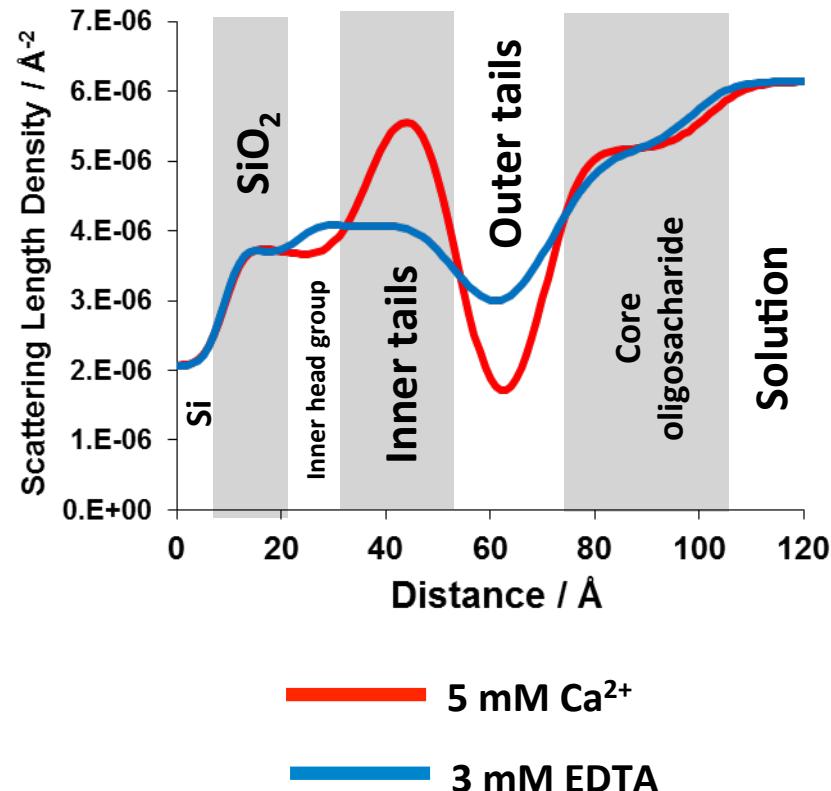
$$\varphi_{water} = \frac{(\rho_{fitted-D2O} - \rho_{fitted-H2O})}{(\rho_{D2O} - \rho_{H2O})}$$

$$\varphi_{d-lipid} = \varphi_{lipid} \times \left(\frac{((\rho_{fitted} - (\rho_{D2O} \varphi_{D2O}) / \varphi_{lipid}) - \rho_{h-lipid\ tails})}{(\rho_{d-lipid\ tails} - \rho_{h-lipid\ tails})} \right)$$

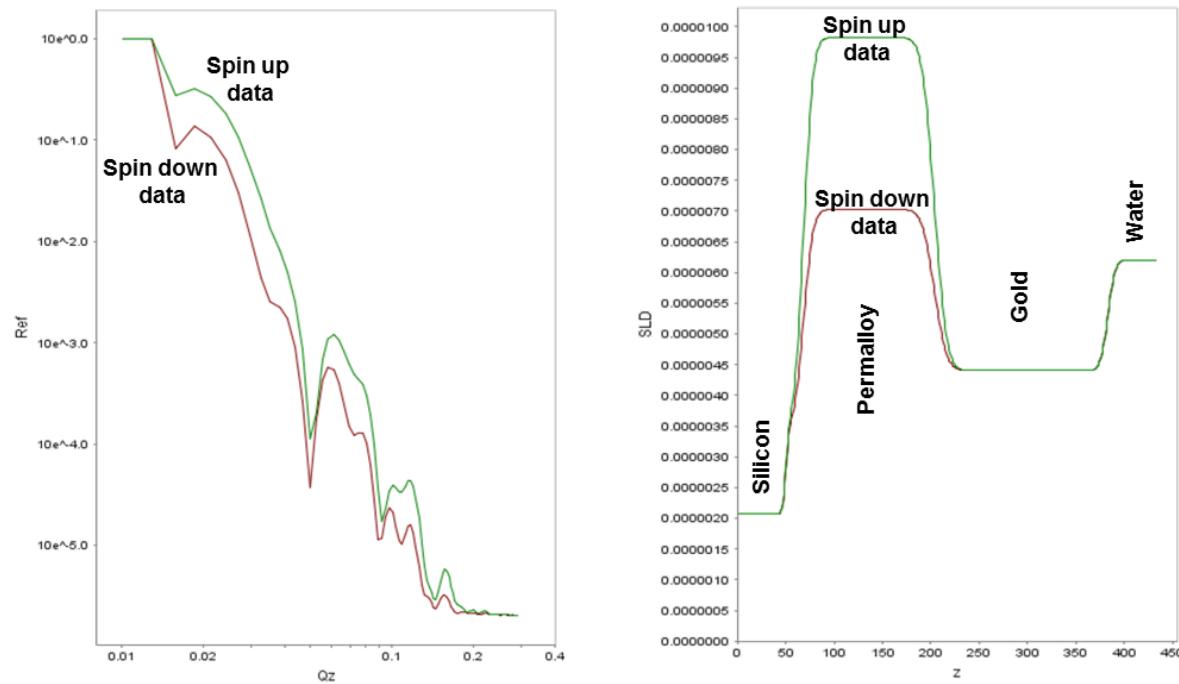


**Asymmetric DPPC (inner leaflet) : Ra-LPS (outer leaflet) bilayer deposited
on Silicon in EDTA containing buffer**

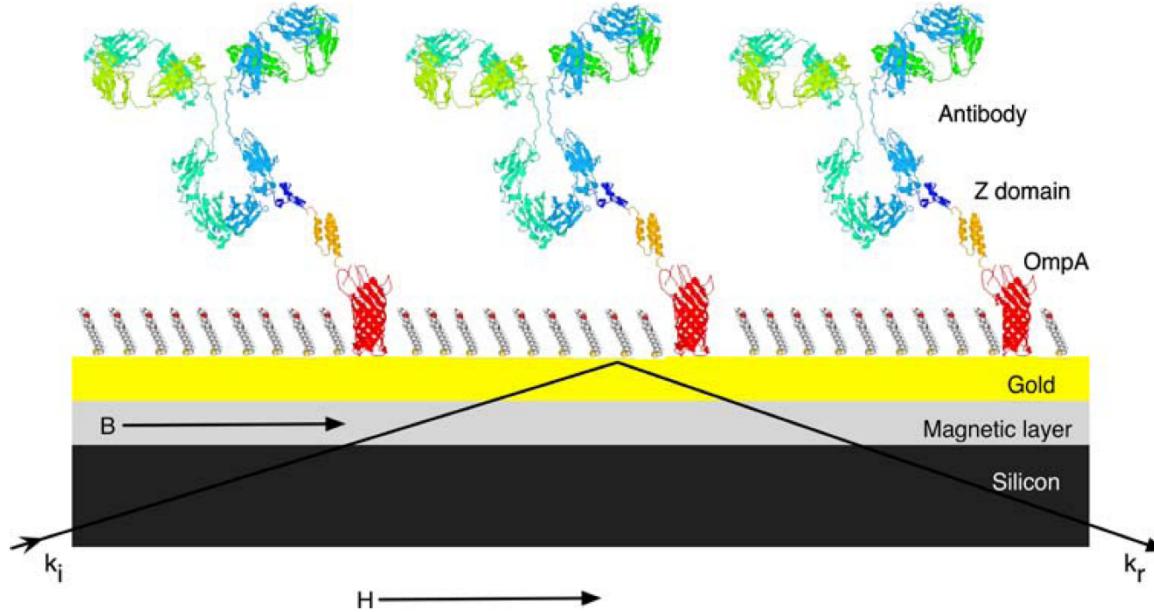




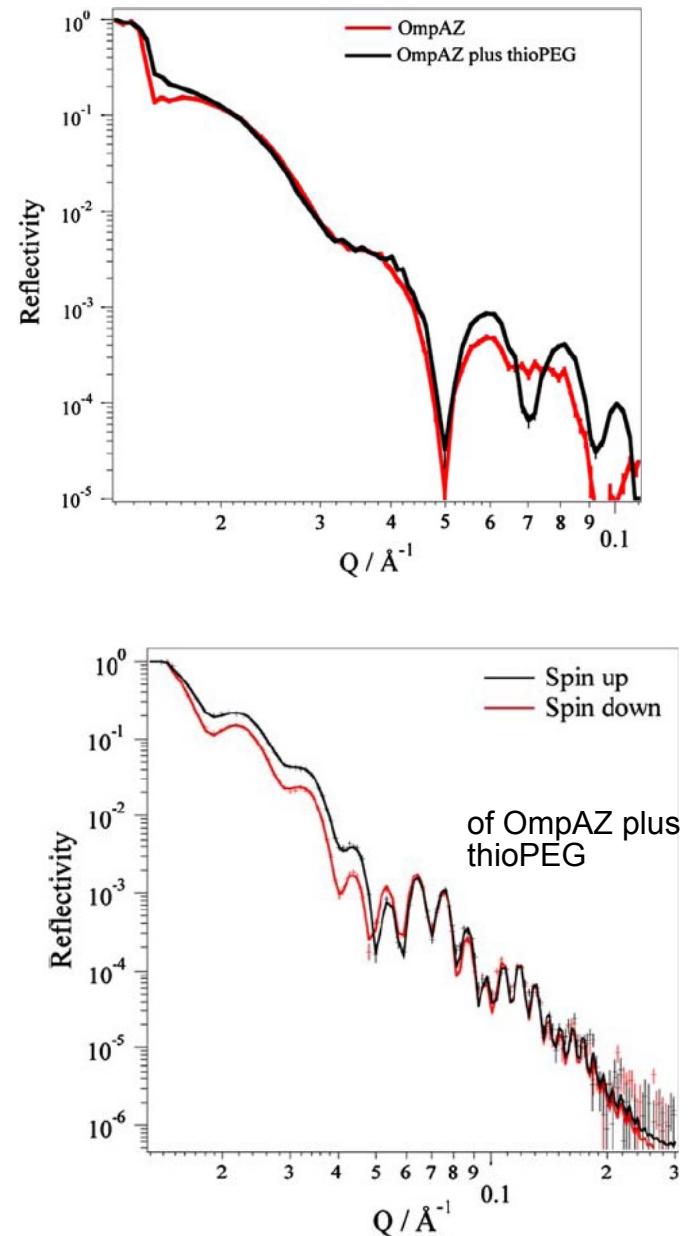
Magnetic Contrast

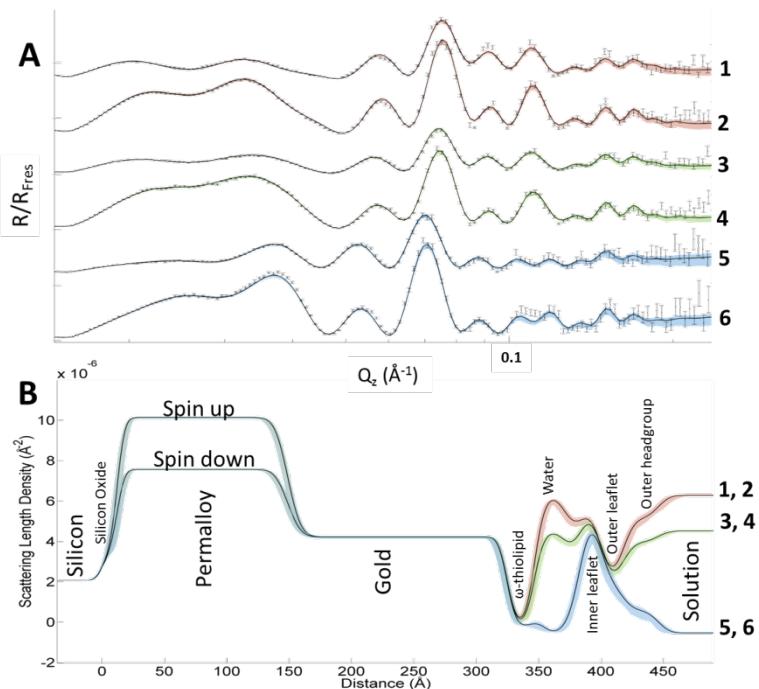


Magnetic Contrast

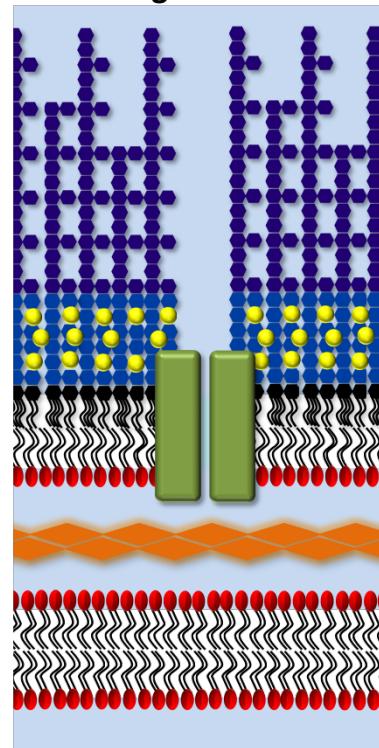


- Polarised neutron reflection used to probe the structure of an antibody on gold (separated by a thioPEG monolayer).
- Polarised neutrons are used as this provides a means of achieving extra contrast in samples having a magnetic metal layer (Fe or Ni) under the gold surface.
- This contrast is attained without resorting to hydrogen/deuterium exchange in the biological layer.

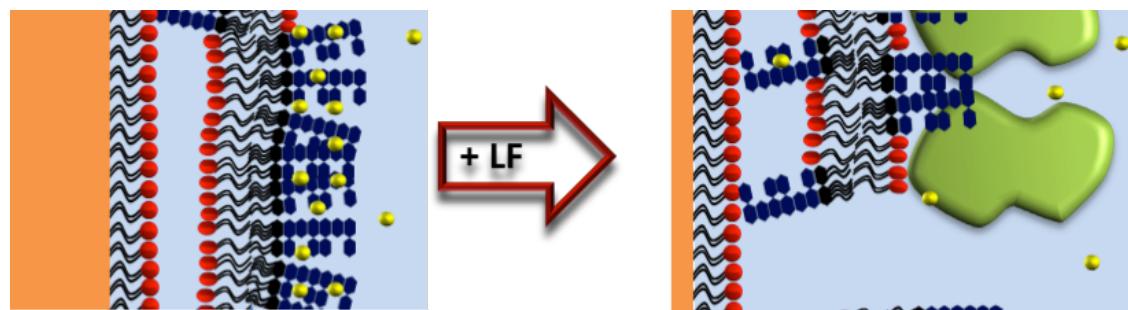
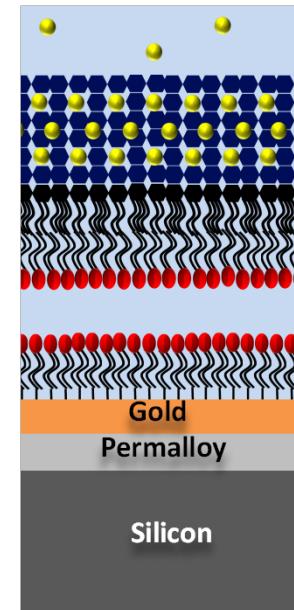




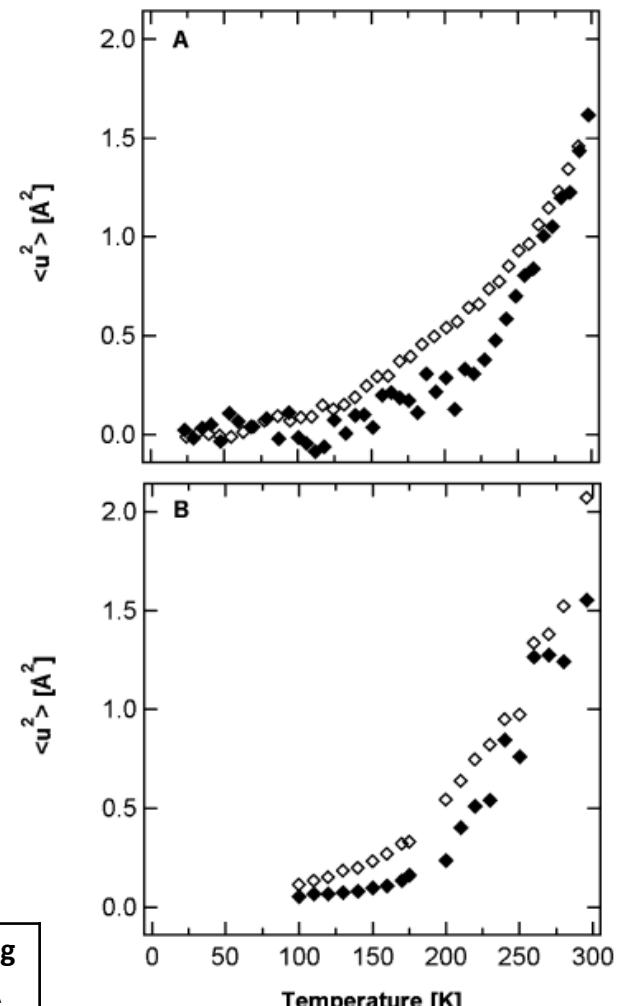
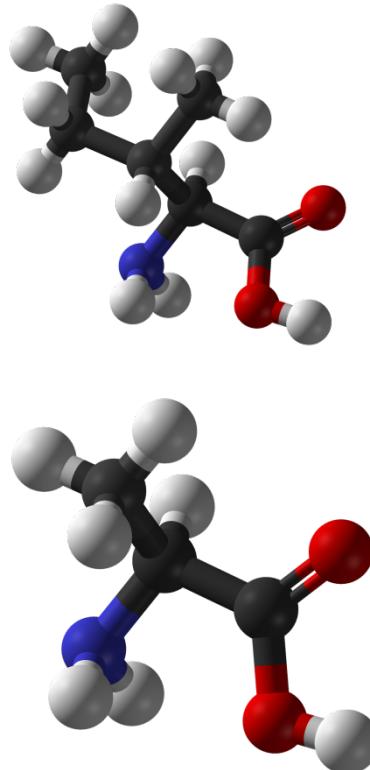
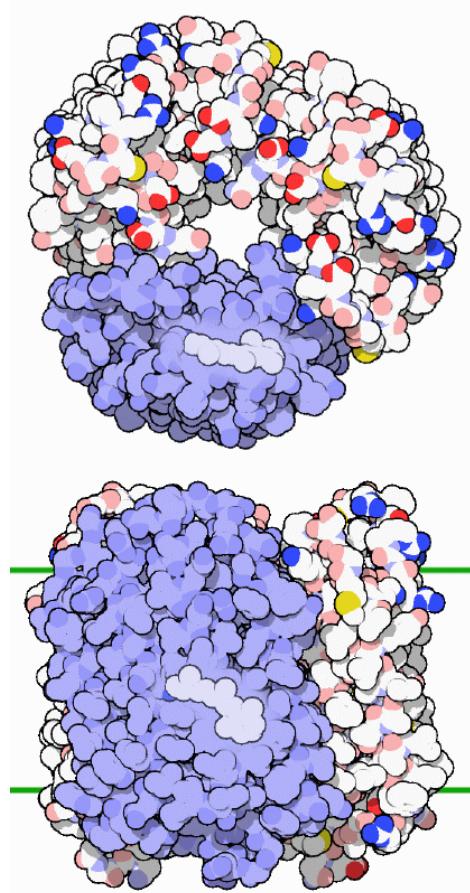
Gram Negative bacteria



Model



Sample deuterium labelling is often required : QENS



| Element | Coherent Scattering Length (b_{coh})/ 10^{-5} \AA | Incoherent Scattering Length (b_{inc})/ 10^{-5} \AA |
|-----------|--|--|
| Hydrogen | -3.74 | 25.274 |
| Deuterium | 6.671 | 4.04 |

Conclusions

- Isotopic Labelling is a powerful tool to examine complex biological structures with NS
- Solution labelling is by far the easiest way to match out components of a complex.
- Deuterium labelling of the samples is often required – if so label the cheapest part!
- Checks should be made to ensure labelling does not change the physiochemical properties of the samples.

Further Reading

- Small angle neutron and X-ray scattering in structural biology, recent examples from the literature, 2008, Cameron Neylon, Eur Biophys J, DOI 10.1007/s00249-008-0259-2.
- Neutrons for biologists: a beginner's guide, or why you should consider using neutrons., 2009, J. Lakey, J. R. Soc. Interface, 6, Supp 5, S567-73.
- Examining protein-lipid complexes using neutron scattering. L Clifton, C. Neylon and J. H. Lakey, Lipid-Protein Interactions, Methods in Molecular Biology, 2013.
- Small Angle X-ray and Neutron Scattering from Solution of Biological Macromolecules, D. I. Svergun, M. H. Koch, P. A. Timmins, R. P. May.
- Small-angle scattering for structural biology-Expanding the frontier while avoiding the pitfalls. David Jacques and Jill Trewella. Protein Science, 19, 642-657