Integrative Network Analysis using R and RCytoscape

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10th May 2017 Sydney Users of R (SURF) Meetup SMSA Townhall

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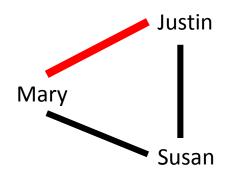


Outline

- Why is network being used?
- R and RCytoscape
- What are negative genetic interactions?
- Applications of negative genetic interactions
- Investigating the network basis of genetic interactions
- Results & Code Examples
- Summary

Where is network being used?

Friends recommendation on facebook

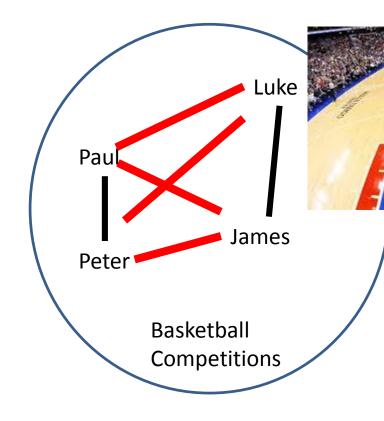


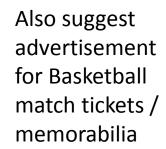
Friends of Friends

Red = relationships predicted/suggested by Facebook



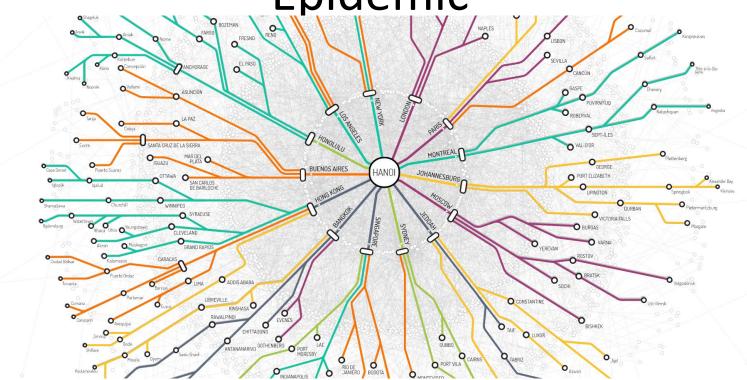






Same Social Group

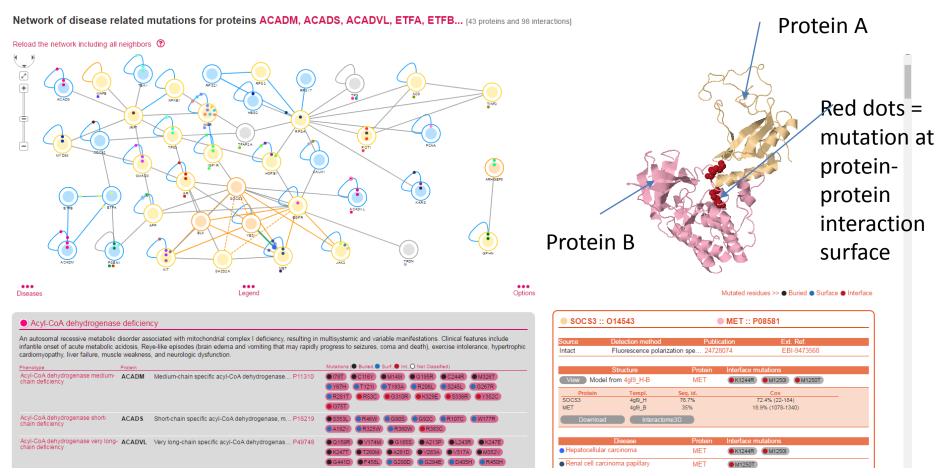
Modelling the Spread of Viruses in an Epidemic



EPIDEMIC RAPID TRANSIT MAP

http://www.mobs-lab.org/epi-rail.html





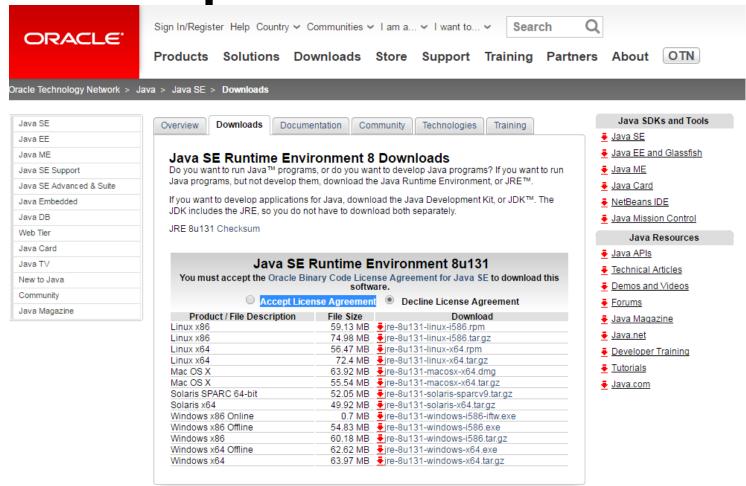
http://dsysmap.irbbarcelona.org/

Why would you want to drive Cytoscape using R scripts?

- Animate your network, make the network change dynamically, visualize time series data
- Support reproducibility, sharing of methods via code
- Another way of developing applications, without the difficulty of developing Cytoscape apps in Java
- No need to click buttons

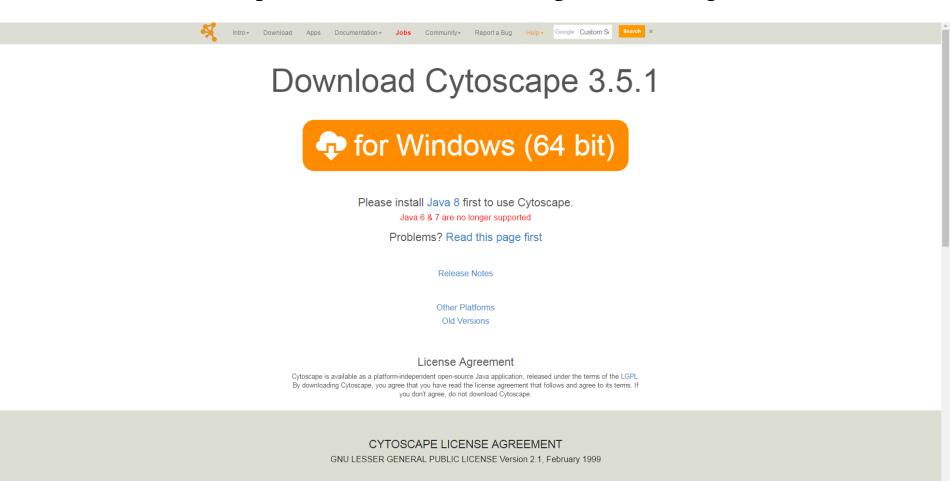
Installation Guide – How to get R and Cytoscape to talk to each other?

Step 1: Install Java 8



 http://www.oracle.com/technetwork/java/javase/downloads/jre8downloads-2133155.html

Step 2: Install Cytoscape



http://www.cytoscape.org/download.php

Step 3: Understand what the Cytoscape app CyREST is for?

 A Cytoscape Core app for driving Cytoscape from R/Python/Julia/Node.js/etc.

 IMPORTANT: Now this app is a part of the Cytoscape Core!

Ono, Keiichiro, et al. "<u>CyREST: Turbocharging</u>
 <u>Cytoscape Access for External Tools via a</u>
 <u>RESTful API</u>." F1000Research 4 (2015).

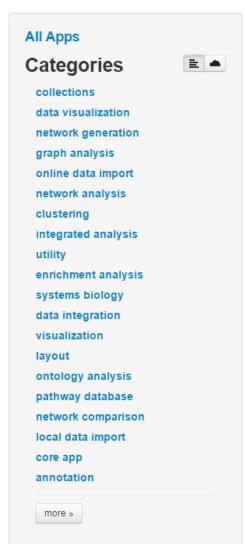
Step 4: Rstudio and libraries

- Install R and/or Rstudio
- Install igraph
- install.packages("igraph")

- Install Bioconductor download manager (lite version)
- source("https://bioconductor.org/biocLite.R")
- Install RCy3 in R (which connects R to Cytoscape, R side application).
- biocLite("RCy3") # previously called RCytoscape

Step 5: Running R and Cytoscape together

- In the same desktop environment
- Open Rstudio and load the following packages:
- library (igraph)
- library(RCy3)
- Open Cytoscape and have it side by side. CyREST is automatically loaded once you start Cytoscape. Now you're ready to draw some networks!



http://apps.cytoscape.org/

Newest Releases

Get Started with the App Store »



PSFC

\$1.015 tor:

Pathway Signal Flow Calculator: Calculates pathway activity based



CYREST



Core App: Language-agnostic RESTful API



Rene



Regulatory Network Enhancing Plugin - ReNE allows users to



MetScape



Maps human, mouse and rat metabolomics and gene



Omics Analysis Collection 3.0

network analysis of omic data. By

Collection of apps to facilitate



Functional Enrichment Collection



Collection of apps to facilitate functional enrichment analysis. By

more newest releases »

Top Downloaded Apps



ClueGO

3.0

Creates and visualizes a functionally grouped network of



BiNGO



Calculates overrepresented GO terms in the network and display



GeneMANIA



Imports interaction networks from public databases from a list of



CluePedia



CluePedia: A ClueGO plugin for pathway insights using integrated



Code example 1 – Drawing a triplet in Cytoscape

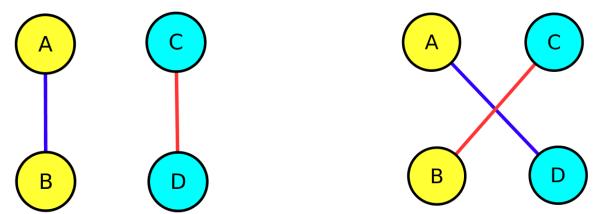
Notes:

- Please refer to the script 'draw_a_single_triplet.Rmd' in the Github page.
- I've used the Bioconductor RCy3 vignette extensively for this example. Here is the reference: https://bioconductor.org/packages/release/bioc/vignette s/RCy3/inst/doc/RCy3.R

Generating Randomized Networks but Keeping the Degree Distribution

Before Edge Swap

After Edge Swap



- The node 'degree' is the number of interaction partners each node has
- Establishing the background level of triplets
- The number of times the edges were swapped was equal to the size of the network.
- Each type of biological network (e.g. protein-protein interaction network)
 was randomized independently of other types of networks.
- These were combined to form the randomized integrated network.

Code example 2 – Performing Network Randomization

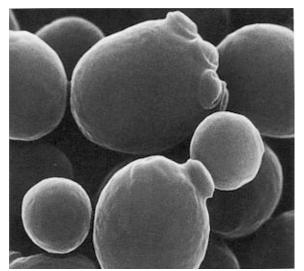
Notes:

- Please refer to the script 'rewire_network.Rmd' in the Github page.
- This script show you how to perform network randomization, by edge swapping, while keeping the degree distribution the same. The number of interaction partners for every node should remain the same after this type of network randomization. The script also show you what happens if we randomize the network without conserving the degree distribution.

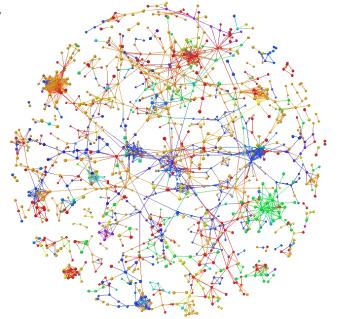
Saccharomyces cerevisiae

The world's most characterized eukaryote

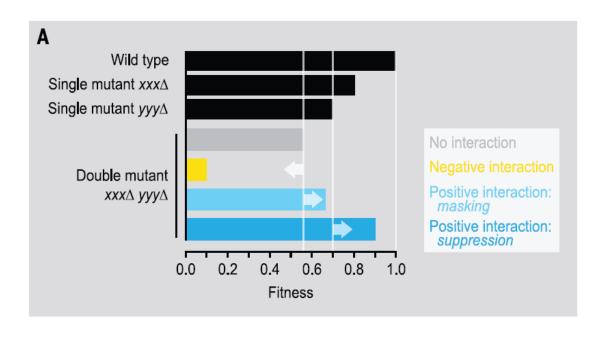
- Vast genome, transcriptome and proteome resources
- Knock out and over-expression mutants available for many genes
- Databases for protein-protein, proteir
 DNA, protein-RNA interactions
- → Start with *Saccharomyces*
- → Testing grounds for algorithms
- → Map results to genes that are evolutionarily conserved in humans (e.g. essential genes, drug targets)



http://tinyurl.com/q36n6dh



Genetic Interactions



Expected Score If there is No Interaction = $F(wild type) - F(xxx\Delta) * F(yyy\Delta)$

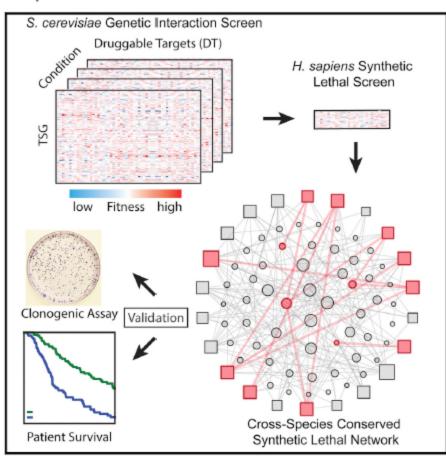
van Leeuwen et al. (2016) Science 354(6312):aag0839

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Molecular Cell

A Network of Conserved Synthetic Lethal Interactions for Exploration of Precision Cancer Therapy

Graphical Abstract



Authors

Rohith Srivas, John Paul Shen, Chih Cheng Yang, ..., Pedro Aza-Blanc, Robert W. Sobol, Trey Ideker

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In Brief

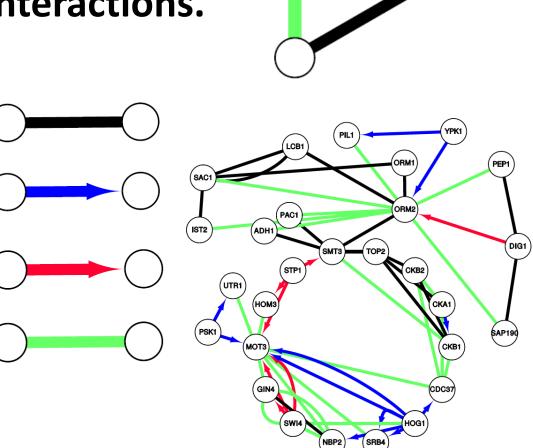
An emerging strategy for precision cancer therapy is to induce synthetic lethality in a tumor based on its pattern of genetic mutations. Here we map a large network of conserved synthetic lethal interactions between selective drugs and tumor suppressor genes somatically mutated in human cancer. All of these interactions are mirrored in budding yeast, an accessible model for researching drug mode of action.

Applications

- Searching for Synergistic drug targets
- One drug target Protein A
- Another drug target Protein B
- Their combined effect is greater than expected (i.e. kills cancer cells or microbes quicker)
- Limit the chance of drug resistance developing
- New drugs are hard to develop, use pairs of existing drugs

A triplet consists of one negative genetic interaction and two other interactions.

- Protein-protein interactions
- Kinase-substrate interactions
- Transcription factor-target gene interactions
- Negative genetic interactions



Identification of Triplet Motifs

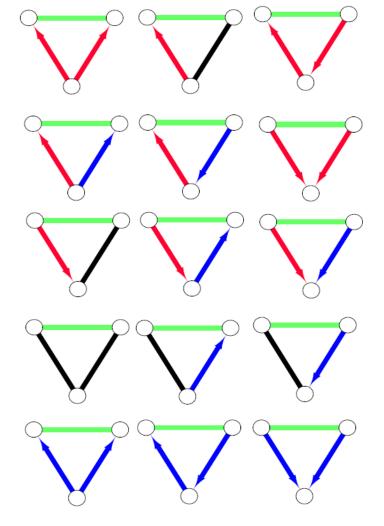
- There are 15 possible types of motifs that could occur, from the different combinations of the edges.
- We have identified all 15
 possible types of motifs, with
 30,850 triplets, 3,293 proteins
 and 23,225 interactions in total.
 - Protein-protein interactions
 - Kinase-substrate interactions
 - Transcription factortarget gene interactions
 - Negative genetic interactions











Edge list table - caveat

Node A	Node B
Adam	Xavier
Xavier	Adam

^{*} The edges can be entered into the edge list table in two different directions. Sometimes you need to filter / clean the table so that you only have one of these entry (e.g. Node A string > Node B string, "Adam" is alphabetically before "Xavier" etc...). Other times you need to keep both entries. I've done these filtering several times in the script 'form_triplet_motifs.Rmd'.

Code example 3 – Setting random number seeds and distributing tasks to multiple cores

Notes:

- Please refer to the script 'random_number_generator.Rmd' in the Github page.
- This script show you how to set the seed so that the results are replicable each time you run it. It also show you how to do this in a setting where you send the execution to multicores (in a Linux / Unix environment).
- I did not show this example in the actual talk, mainly due to time constraint, but it is included here as a useful resource.

Code example 4 – counting the number of each type of triplet motif

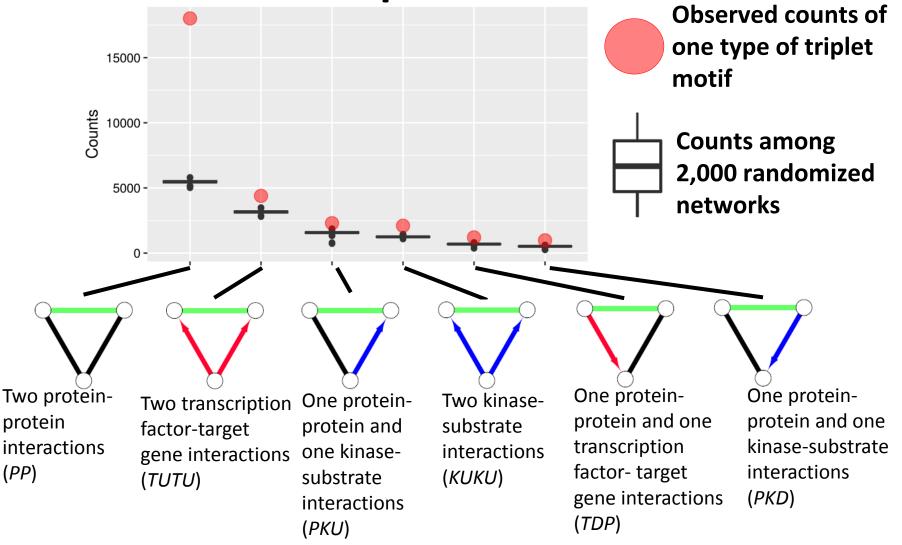
Notes:

- Please refer to the script 'form triplet motifs.Rmd' in the Github page.
- It counts the number of times each type of triplet motif has been identified in the network.

Calculating the Significance of Triplet Motifs

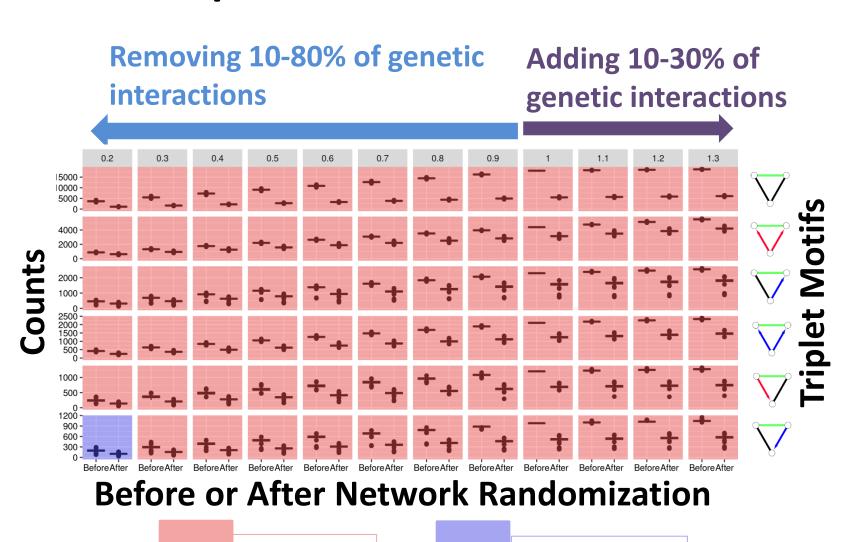
- p-value calculated by the number of times 2000 randomized networks has triplet motif counts greater than counts of the same motif in the actual network
- The Bonferroni method was used for multiple testing correction.
- A triplet motif is significant if:
 - The adjusted p-value is < 0.05, and
 - Triplet motifs with observed counts less 2% of the total number of triplets is deemed un-reliable under a 2% false discovery rate.

Overrepresentation of 6 out of 15 Types of Triplet Motifs



P= protein-protein, T= transcription factor-target gene, K=kinase-substrate, U=up, D=down

Triplet Motifs are Robust



Non-significant

Significant

Known Limitations

- Network is only a reference map, static not dynamic
- Lacking information on whether interaction is present or absent in a specific condition
- Condition-specific interactions need to be tested for under various stresses, drug treatments, genetic backgrounds etc...
- False positive or false negative interactions

Summary

- Cytoscape can be used to visualize networks
- RCytoscape can be used to drive network analyses from R, providing a programmable workflow
- Network randomization can be used to generate many randomized networks. For these background set, we can calculate the 'null' or expected distribution. We can then compared our observed data with the expected distribution and infer the significance of the differences.

Resources and Databases

- Github page for today's talk
 - https://github.com/IgnatiusPang/Network_SURF
- Biological Networks Databases
 - https://thebiogrid.org/
 - http://string-db.org/
 - http://www.ebi.ac.uk/intact/
- Cytoscape, Rcytoscape, igraph
 - http://www.cytoscape.org
 - https://bioconductor.org/packages/release/bioc/html/RCy3.html
 - http://igraph.org/r/
 - Igraph tutorial: http://kateto.net/networks-r-igraph

5/11/2017

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5/11/2017

Acknowledgements

- We would like to acknowledge funding from the Bioplatforms Australia and the Australian Research Council (ARC).
- We would also like to thank people from the Wilkins lab and Edwards lab for feedback and suggestions.



