

## Introduction

Spectre provides a number of options for performing quantitative, differential and statistical analysis of cytometry data after the initial analysis using clustering or similar methods. Here we provide a demonstration of a number these options.

For this tutorial, we will use one of the demo datasets included in Spectre: a dataset of cells isolated from murine brains, 7 days following mock infection, or infection with West Nile virus (WNV). The **demo.clustered** dataset has already been subject to arcsinh transformation, clustering, and population annotation..

```
library('Spectre')
package.load()

cell.dat <- Spectre::demo.clustered
cell.dat
```

	FileName	NK11	CD3	CD45	Ly6G	CD11b	B220	CD8a
Ly6C	CD4 NK11_asinh	CD3_asinh	CD45_asinh	Ly6G_asinh	...			
1:	CNS_Mock_01.csv	42.3719	40.098700	6885.08	-344.7830	14787.30	-40.2399	83.7175
958.7000	711.0720	0.04235923	0.040087962	2.627736	-0.33829345	...		
2:	CNS_Mock_01.csv	42.9586	119.014000	1780.29	-429.6650	5665.73	86.6673	34.7219
448.2590	307.2720	0.04294540	0.118734817	1.340828	-0.41743573	...		
3:	CNS_Mock_01.csv	59.2366	206.238000	10248.30	-1603.8400	19894.30	427.8310	285.8800
1008.8300	707.0940	0.05920201	0.204803270	3.022631	-1.25101677	...		
4:	CNS_Mock_01.csv	364.9480	-0.233878	3740.04	-815.9800	9509.43	182.4200	333.6050
440.0710	249.7840	0.35729716	-0.000233878	2.029655	-0.74509796	...		
5:	CNS_Mock_01.csv	440.2470	40.035200	9191.38	40.5055	5745.82	-211.6940	149.2200
87.4815	867.5700	0.42713953	0.040024513	2.914359	0.04049443	...		
---								
169000:	CNS_WNV_D7_06.csv	910.8890	72.856100	31466.20	-316.5570	28467.80	-7.7972	-271.8040
12023.7000	1103.0500	0.81693878	0.072791800	4.142314	-0.31149515	...		
169001:	CNS_WNV_D7_06.csv	-10.2642	64.188700	45188.00	-540.5140	22734.00	202.4110	-936.4920
4188.3300	315.9400	-0.01026402	0.064144703	4.504101	-0.51715205	...		
169002:	CNS_WNV_D7_06.csv	-184.2910	-9.445650	11842.60	-97.9383	17237.00	123.4760	-219.9320
8923.4000	-453.4640	-0.18326344	-0.009445510	3.166628	-0.09778240	...		
169003:	CNS_WNV_D7_06.csv	248.3860	229.986000	32288.20	-681.1630	19255.80	-656.0540	-201.5880
10365.7000	61.6765	0.24590035	0.228005328	4.168089	-0.63716643	...		
169004:	CNS_WNV_D7_06.csv	738.9810	95.470300	46185.10	-1004.6000	22957.80	-661.6280	72.3356
9704.4700	-31.8532	0.68430866	0.095325863	4.525922	-0.88462254	...		

We will also provide some 'cell count' data for each sample (i.e. number of total leukocytes in each sample). We expect

```
counts.dt <- data.frame('Sample' = unique(cell.dat[['Sample']]),
                        'Counts' = c(4.20E+05, 2.40E+05, 2.56E+05, 2.52E+05, 3.45E+05, 7.02E+05,
                                     5.07E+06, 2.94E+06, 2.12E+06, 4.32E+06, 4.08E+06, 1.83E+06)
                        )
counts.dt
```

	Sample	Counts
1	01_Mock_01	4.20E+05
2	02_Mock_02	2.40E+05
3	03_Mock_03	2.56E+05
4	04_Mock_04	2.52E+05
5	05_Mock_05	3.45E+05
6	06_Mock_06	7.02E+05
7	07_WNV_01	5.07E+06
8	08_WNV_02	2.94E+06

```

9    09_WNV_03  2.12E+06
10   10_WNV_04  4.32E+06
11   11_WNV_05  4.08E+06
12   12_WNV_06  1.83E+06

```

Key to the comparison of populations across samples, is the generation of 'summary' data. Where 'cellular' data consists of cells (rows) vs cell features (columns: e.g. CD4 expression, CD8 expression etc); 'summary' data consists of samples (rows) vs sample features (number of monocytes per sample, expression level of Ly6C on CD8 T cells, etc). This summary data can then be used to generate plots that compare these metrics between experimental groups.

First, let's examine the columns in the cellular dataset.

```
as.matrix(names(cell.dat))
```

```

[,1]
[1,] "FileName"
[2,] "NK11"
[3,] "CD3"
[4,] "CD45"
[5,] "Ly6G"
[6,] "CD11b"
[7,] "B220"
[8,] "CD8a"
[9,] "Ly6C"
[10,] "CD4"
[11,] "NK11_asinh"
[12,] "CD3_asinh"
[13,] "CD45_asinh"
[14,] "Ly6G_asinh"
[15,] "CD11b_asinh"
[16,] "B220_asinh"
[17,] "CD8a_asinh"
[18,] "Ly6C_asinh"
[19,] "CD4_asinh"
[20,] "Sample"
[21,] "Group"
[22,] "Batch"
[23,] "FlowSOM_cluster"
[24,] "FlowSOM_metacluster"
[25,] "Population"
[26,] "UMAP_X"
[27,] "UMAP_Y"

```

We can choose any number of these to be measured as 'dynamic' columns (**dyn.cols**), where we will measure the median expression of these markers on each population in each sample. In this case we will choose CD11b (#15) and Ly6C (#18).

```

dyn.cols <- names(cell.dat)[c(15,18)]
dyn.cols

```

```
[1] "CD11b_asinh" "Ly6C_asinh"
```

To create the summary data, we can use the **create.sumtable** function.

```

sum.dat <- create.sumtable(dat = cell.dat, # The dataset to be summarised
                           sample.col = 'Sample', # The column that denotes the sample name/ID
                           pop.col = 'Population', # The column that denotes the population name/ID
                           use.cols = dyn.cols, # Columns (markers) whose expression we will measure on
                           each population in each sample

```

```

annot.cols = c('Group', 'Batch'), # Additional columns we would like to
include for annotation purposes (e.g. group names, batch names, etc)
counts = counts.dt) # A data.frame or data.table of the total cells per
sample, to generate counts of each cell type per sample.

```

Once the function is complete, we can review the data.

```
sum.dat
```

Each row represents a sample, and each column a feature of that sample (e.g. Percent of sample -- CD4 T cells, etc).

	Sample	Group	Batch	Percent of sample -- CD4 T cells	Percent of sample -- CD8 T cells	...
1:	01_Mock_01	Mock	A	0.7346282	2.6969910	...
2:	02_Mock_02	Mock	B	0.5060006	1.0379500	...
3:	03_Mock_03	Mock	B	0.5405026	1.0037905	...
4:	04_Mock_04	Mock	A	0.2522882	0.4048345	...
5:	05_Mock_05	Mock	A	0.2015021	0.5495512	...
6:	06_Mock_06	Mock	B	0.5315886	1.3085259	...
7:	07_WNV_01	WNV	A	5.4094708	5.0529248	...
8:	08_WNV_02	WNV	B	2.0636974	2.3901928	...
9:	09_WNV_03	WNV	A	2.6314145	3.4814676	...
10:	10_WNV_04	WNV	A	2.6910280	3.2397408	...
11:	11_WNV_05	WNV	B	3.1520784	3.6292854	...
12:	12_WNV_06	WNV	A	3.4251318	3.7500666	...

Review all of the sample 'features' that we have calculated.

```
as.matrix(names(sum.dat))
```

```

[,1]
[1,] "Sample"
[2,] "Group"
[3,] "Batch"
[4,] "Percent of sample -- CD4 T cells"
[5,] "Percent of sample -- CD8 T cells"
[6,] "Percent of sample -- Infil Macrophages"
[7,] "Percent of sample -- Microglia"
[8,] "Percent of sample -- Neutrophils"
[9,] "Percent of sample -- NK cells"
[10,] "Cells per sample -- CD4 T cells"
[11,] "Cells per sample -- CD8 T cells"
[12,] "Cells per sample -- Infil Macrophages"
[13,] "Cells per sample -- Microglia"
[14,] "Cells per sample -- Neutrophils"
[15,] "Cells per sample -- NK cells"
[16,] "MFI of CD11b_asinh -- CD4 T cells"
[17,] "MFI of CD11b_asinh -- CD8 T cells"
[18,] "MFI of CD11b_asinh -- Infil Macrophages"
[19,] "MFI of CD11b_asinh -- Microglia"
[20,] "MFI of CD11b_asinh -- Neutrophils"
[21,] "MFI of CD11b_asinh -- NK cells"
[22,] "MFI of Ly6C_asinh -- CD4 T cells"
[23,] "MFI of Ly6C_asinh -- CD8 T cells"
[24,] "MFI of Ly6C_asinh -- Infil Macrophages"
[25,] "MFI of Ly6C_asinh -- Microglia"
[26,] "MFI of Ly6C_asinh -- Neutrophils"
[27,] "MFI of Ly6C_asinh -- NK cells"

```