

⚠ This page is being updated, so some sections will be incomplete. Please feel free to use the instructions that are present, and we will update the page shortly. The prior version of this page is provided here: [tSNE protocol 2017-04-04.pdf](#).

Introduction

This protocol describes how to perform Spectre's 'discovery workflow' using FlowJo – including data preparation, clustering with FlowSOM, downsampling, dimensionality reduction with UMAP, creating plots, annotating clusters, and performing quantitative and statistical analysis. For more information on this process, please see the main '[discovery workflow](#)' page.



Citation

If you use Spectre, or Spectre-themed FlowJo workflows in your research, please consider citing [Ashhurst TM, Marsh-Wakefield F, Putri GH et al. \(2020\). bioRxiv. 2020.10.22.349563](#). To continue providing open-source tools such as Spectre, it helps us if we can demonstrate that our efforts are contributing to analysis efforts in the community. Please also consider citing the authors of the individual packages or tools (e.g. [CytoNorm](#), [FlowSOM](#), [tSNE](#), [UMAP](#), etc) that are critical elements of your analysis work. We have provided some generic text that you can use for your methods section with each protocol and on the '[about](#)' page.

ⓘ Sample methods blurb

Here is a sample methods blurb for this workflow. You may need to adapt this text to reflect any changes made in your analysis.

Computational analysis of data was performed using the FlowJo using the analysis workflows from the Spectre R package (Ashhurst et al., 2020). The FlowSOM algorithm (Van Gassen et al., 2015) was then run on a merged dataset to cluster the data, where every cell is assigned to a specific cluster and metacluster. Subsequently, the data was downsampled and analysed by the dimensionality reduction algorithm Uniform Manifold Approximation and Projection (UMAP) (McInnes, Healy, Melville, 2018) for cellular visualisation.

Setup FlowJo and plugins

For this workflow, you will need to install [FlowJo](#) and have access to a [licence or dongle](#).

You will also need to ensure you have the following plugins installed:

- Downsample
- FlowSOM
- UMAP (or tSNE, Flt-SNE, etc)

To check your packages, go to the 'Applications' folder (on Mac), and find the folder called 'plugins'.

FlowJo discovery	plugins	
Favourites		
OneDrive - The...	▶ Adobe Acrobat DC	▶ ClassyDL_v1.5
Dropbox (Sydne...)	▶ Adobe Creative Cloud	▶ CytoNorm_v0.6
Dropbox (Perso...)	▶ Anaconda-Navigator.app	▶ EmbedSOM_v0.2
Google Drive	▶ Cisco	▶ FlowSOM_v2.5
AirDrop	▶ EN EndNote X9	▶ Monocle_v4.1.1
Recent	▶ FileMaker Pro 18 Advanced	▶ UMAP_v2.2
Desktop	▶ histoCAT	▶ X-Shift_v1.3
Applications	▶ MATLAB	▶ ClassyDL_v1.5.jar
Documents	▶ plugins	▶ CytoNorm_v0.6.jar
Downloads	▶ Utilities	▶ EmbedSOM_v0.2.jar
Movies	▶ 1Password 7.app	▶ FlowSOM_v2.5.jar
Music	▶ Anaconda-N...gator.app alias	▶ Monocle_v4.1.jar
	▶ App Store.app	▶ UMAP_v2.2.jar
	▶ Atom.app	▶ XShift_v1.3.jar
	▶ Automator.app	
	▶ Backup and s...Google.app	
	▶ Books.app	
	▶ Calculator.app	
	▶ Mac SSD > Applications > plugins	

For each plugin you should see a '.jar' file, and possibly a folder of the same name.

▶ ClassyDL_v1.5
▶ CytoNorm_v0.6
▶ DownSample_v3.3
▶ EmbedSOM_v0.2
▶ FlowSOM_v2.5
▶ Monocle_v4.1.1
▶ UMAP_v2.2
▶ X-Shift_v1.3
▶ ClassyDL_v1.5.jar
▶ CytoNorm_v0.6.jar
▶ DownSample_v3.3.jar
▶ EmbedSOM_v0.2.jar
▶ FlowSOM_v2.5.jar
▶ Monocle_v4.1.jar
▶ UMAP_v2.2.jar
▶ XShift_v1.3.jar

Within the each folder will be documentation related to that plugin.

ClassyDL_v1.5	►	MD License.md
CytoNorm_v0.6	►	MD README.md
DownSample_v3.3	►	
EmbedSOM_v0.2	►	
FlowSOM_v2.5	►	
Monocle_v4.1.1	►	
UMAP_v2.2	►	
X-Shift_v1.3	►	
ClassyDL_v1.5.jar		
CytoNorm_v0.6.jar		
DownSample_v3.3.jar		
EmbedSOM_v0.2.jar		
FlowSOM_v2.5.jar		
Monocle_v4.1.jar		
UMAP_v2.2.jar		
XShift_v1.3.jar		

To download new plugins, go to the [FlowJo exchange](#) site. Download the following (if you don't already have them), unzip them, and move the .jar files (and optionally the associated folder) into the 'plugins' folder as above.

DownSample

v3.3 published March 2nd, 2020
Subset your sample in a specified event count

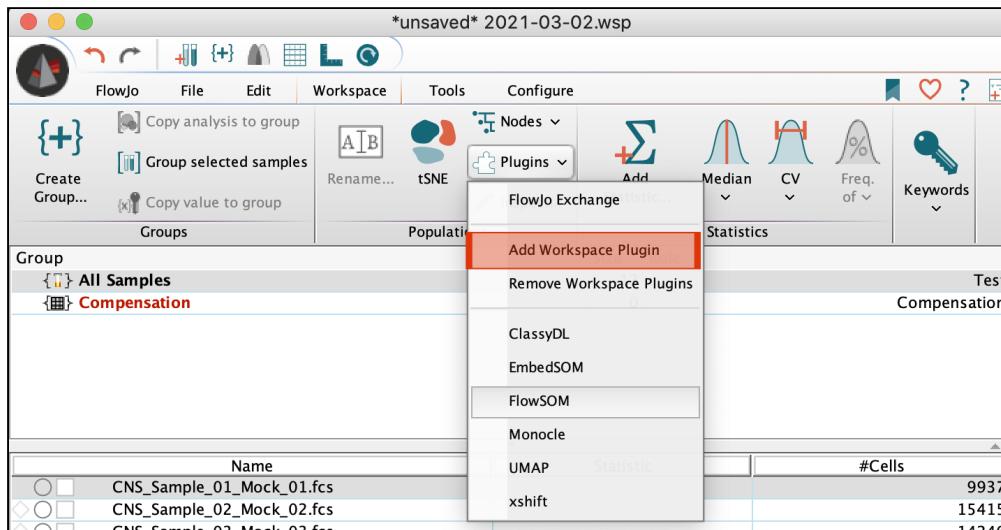
FlowSOM

v2.9 published November 20th, 2020
Cluster using Self-Organizing Maps

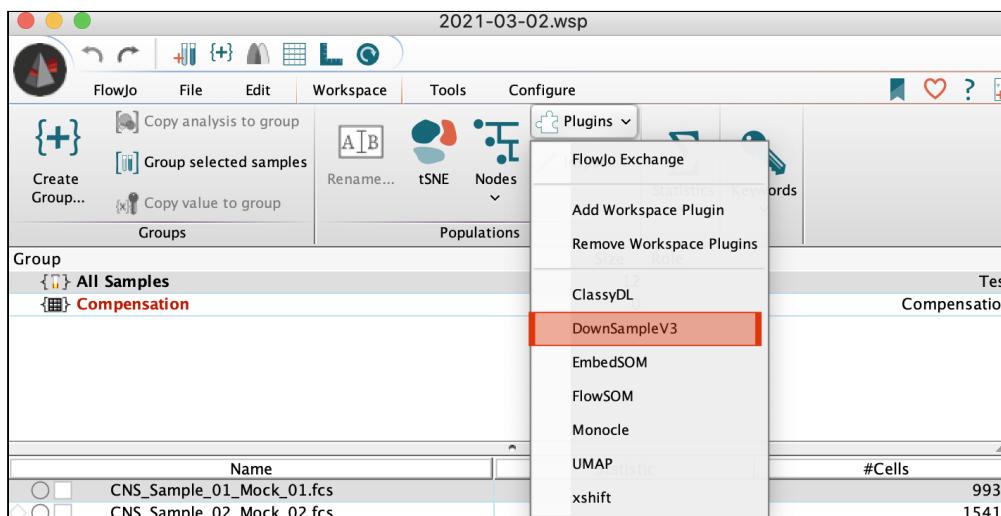
UMAP

v3.1 published March 24th, 2020
A dimensionality reduction technique similar to t-SNE

To activate them in FlowJo, go to Workspace/Plugins and select 'Add Workspace plugin'. You can select the plugins from your 'plugins' folder.

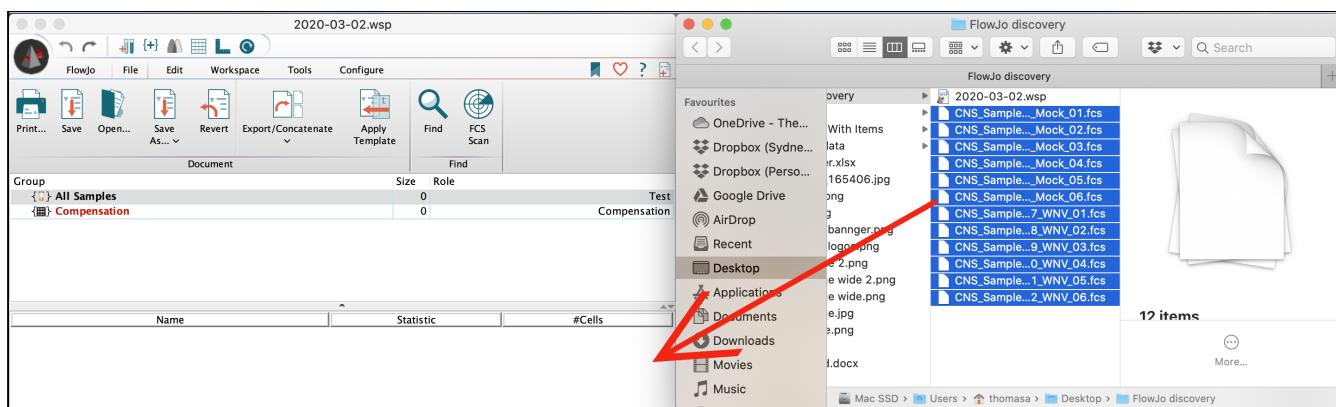


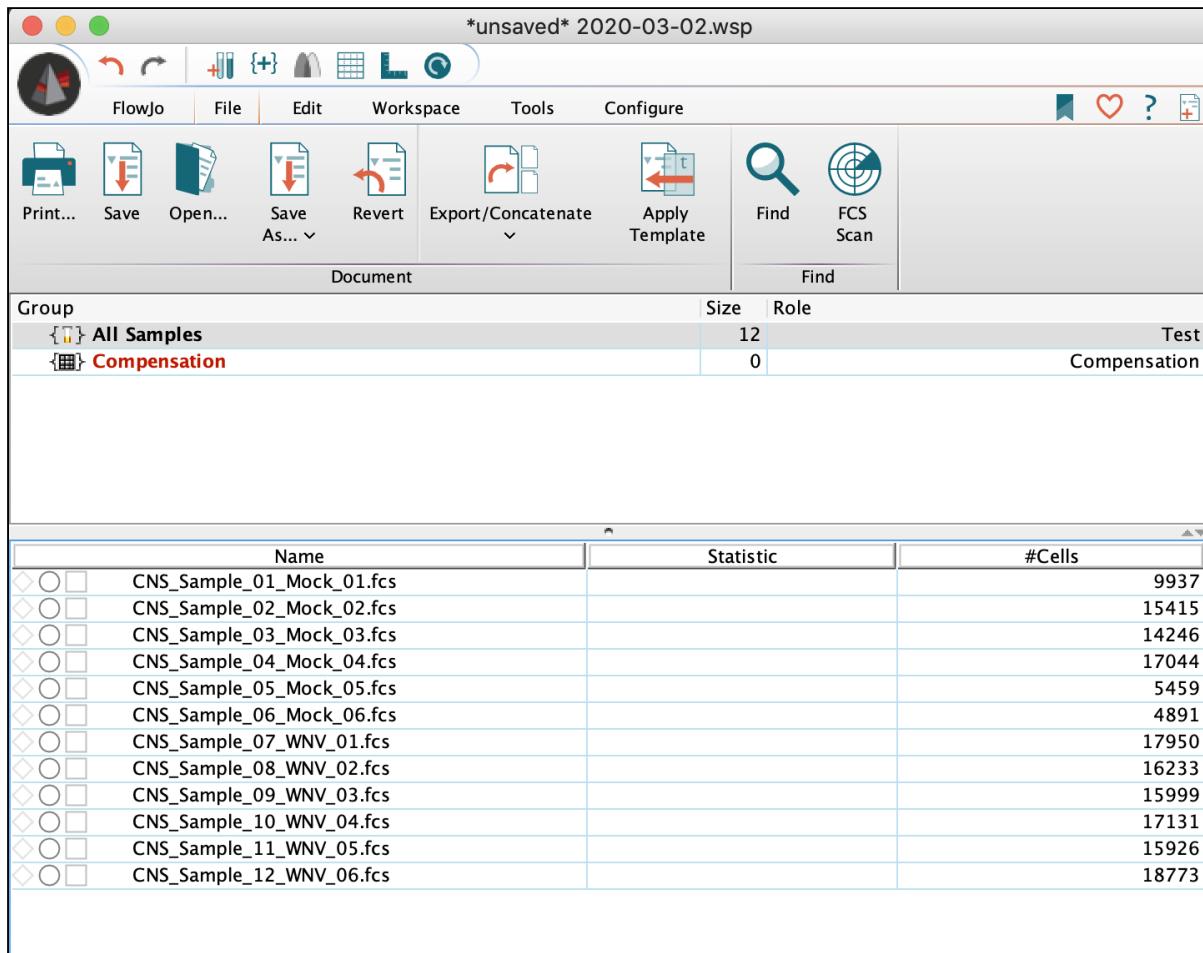
Once they have been 'activated' with FlowJo, they should show up in this list.



1. Data preparation and organisation

To start with, we want to pull our FCS files into FlowJo.





Gate to your population of interest (POI) in the first sample.

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Add the gates to the group, and adjust for all samples, as per a normal analysis in FlowJo.

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Select your POI and use the 'select equivalent nodes' tool under the 'edit' menu.

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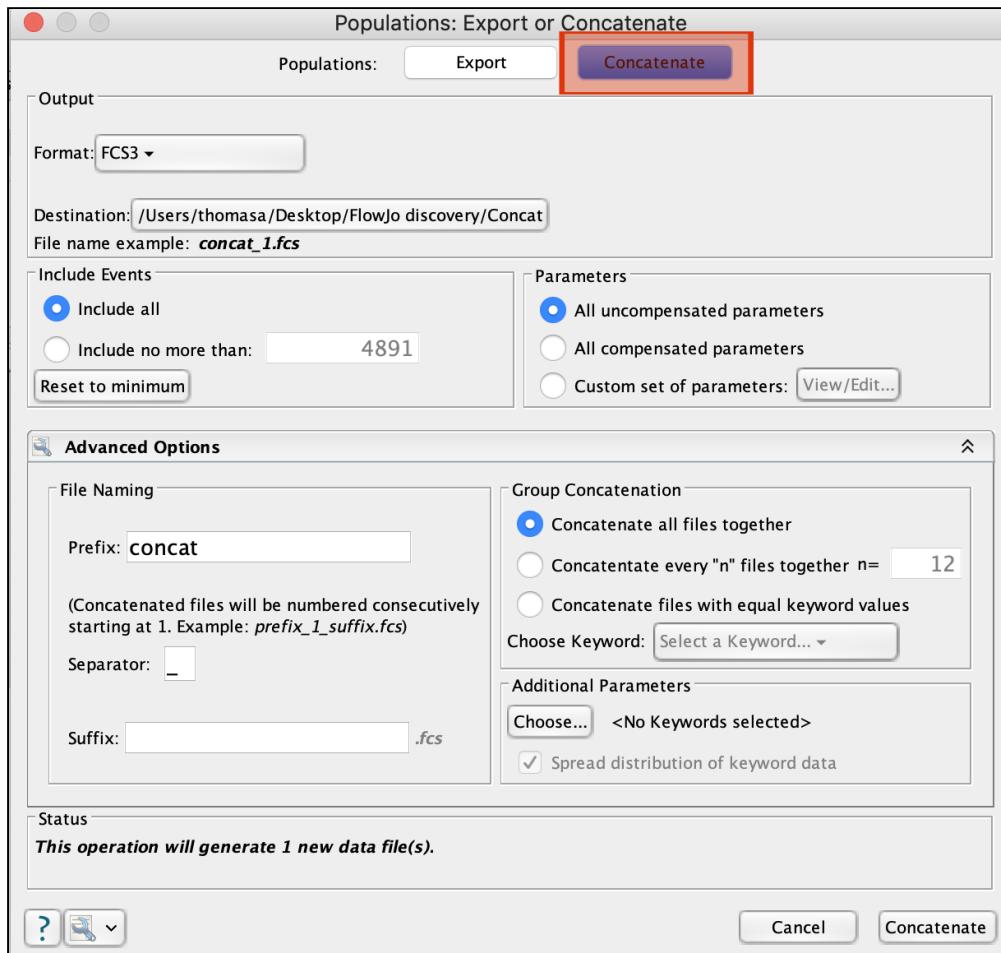
Right click on one of the nodes, and select 'Export / Concatenate Populations'.

Name	Statistic	#Cells
CNS_Sample_01_Mock_01.fcs	⌘C	9937
CNS_Sample_02_Mock_02.fcs	⌘V	15415
CNS_Sample_03_Mock_03.fcs	⌘X	14246
CNS_Sample_04_Mock_04.fcs	⌘I	17044
CNS_Sample_05_Mock_05.fcs	⌘B	5459
CNS_Sample_06_Mock_06.fcs		4891
CNS_Sample_07_WNV_01.fcs		17950
CNS_Sample_08_WNV_02.fcs		16233
CNS_Sample_09_WNV_03.fcs		15999
CNS_Sample_10_WNV_04.fcs		17131
CNS_Sample_11_WNV_05.fcs		15926
CNS_Sample_12_WNV_06.fcs		18773

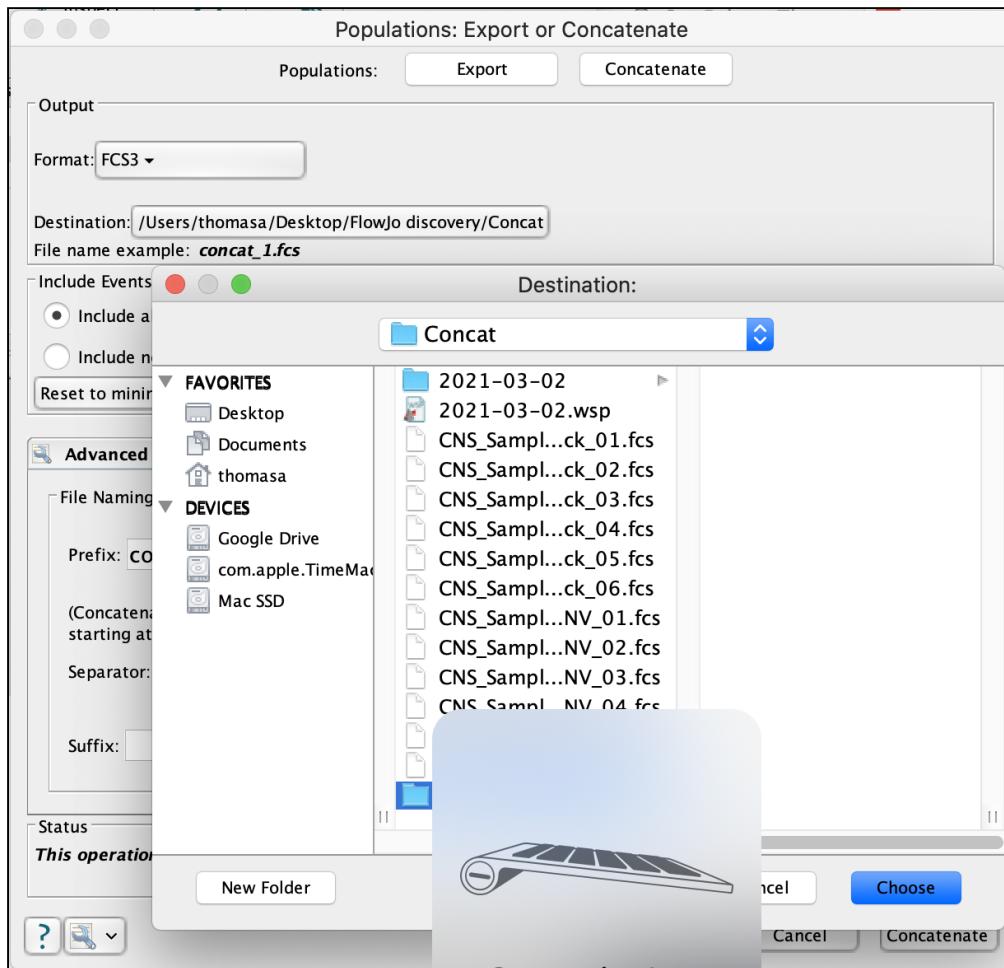
Right-click context menu for CNS_Sample_12_WNV_06.fcs:

- Copy ⌘C
- Paste ⌘V
- Clear ⌘X
- Add Keyword ⌘I
- Add Statistic... ⌘B
- Inspect... ⌘I
- Copy value to group
- Copy analysis to group ⌘G
- Select Equivalent Nodes ⌘E
- Reset Column Widths
- Search for FCS files...
- Export to FACSDiva
- Import from FACSDiva
- Export / Concatenate Populations ⌘E** (highlighted)
- BifurGate
- Derive Parameters... ⌘D
- Consensus Gate
- Cluster Frequency
- Open Parent Folder(s)...

In the new window, select 'Concatenate' at the top. We will be concatenating FCS files, so we can just use 'all uncompensated parameters'. By default, each sample will be separated in a new parameter called 'SampleID', where samples are organised alphabetically (**I think it's alphabetical**). Alternatively, you can add additional keywords, and add them as additional parameters to the concatenated file.



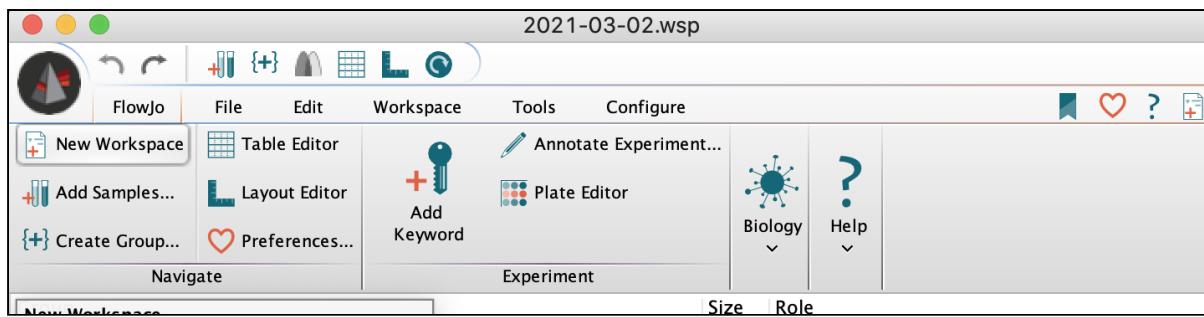
Choose an output location – best to create a folder within your existing experiment folder.



Click 'Concatenate' on the bottom right, and wait for the new FCS file to be created.

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Open a NEW FlowJo workspace.



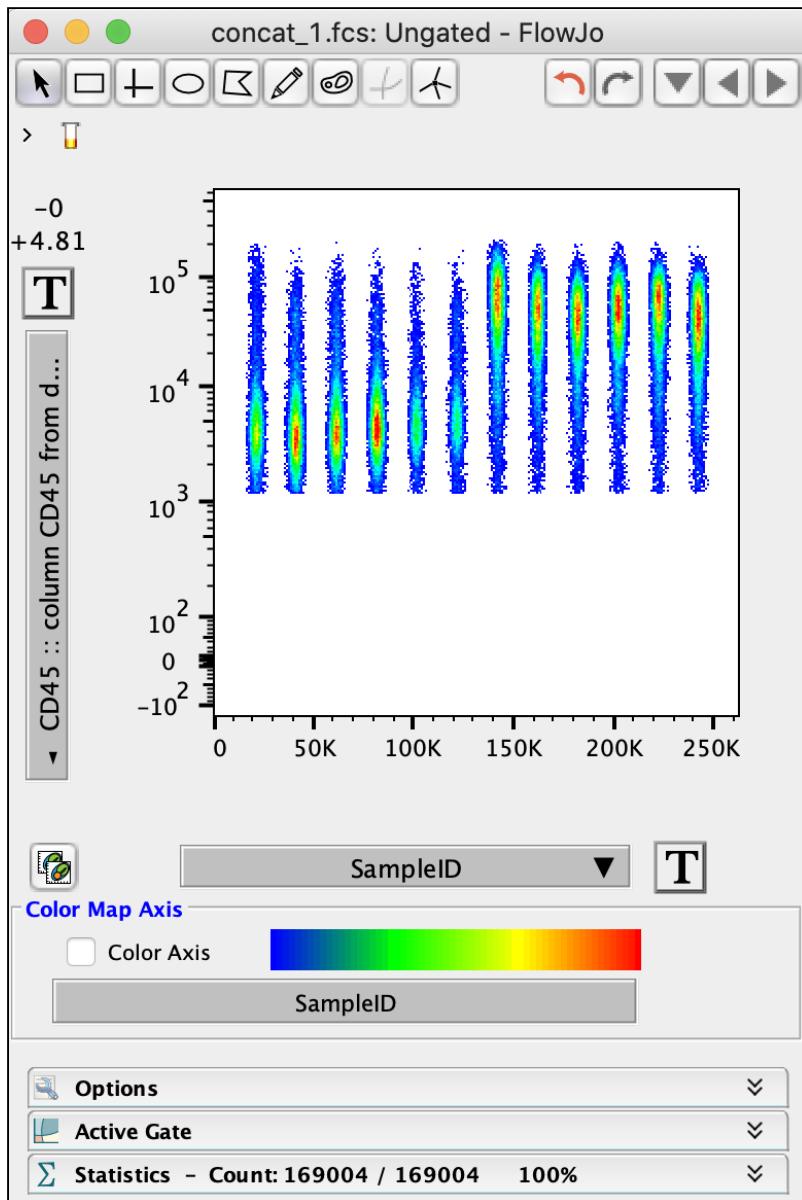
Drag the new FCS file into that workspace, and save the workspace in that folder.

The screenshot shows the FlowJo software interface. The title bar reads "*unsaved* 02-Mar-2021". The menu bar includes File, Edit, Workspace, Tools, Configure, and a set of icons for Annotate Experiment..., Plate Editor, Biology, and Help. The Workspace menu is open, displaying options: New Workspace, Table Editor, Add Samples..., Layout Editor, Create Group..., Preferences..., Annotate Experiment..., Plate Editor, Biology, and Help. Below the menu is a table titled "Experiment" with columns Group, Size, and Role. It contains two rows: "All Samples" (Size 1, Role Test) and "Compensation" (Size 0, Role Compensation). At the bottom, a table lists a single file: concat_1.fcs, with columns Name, Statistic, and #Cells, showing values Name: concat_1.fcs, Statistic: , and #Cells: 169004.

Group	Size	Role
All Samples	1	Test
Compensation	0	Compensation

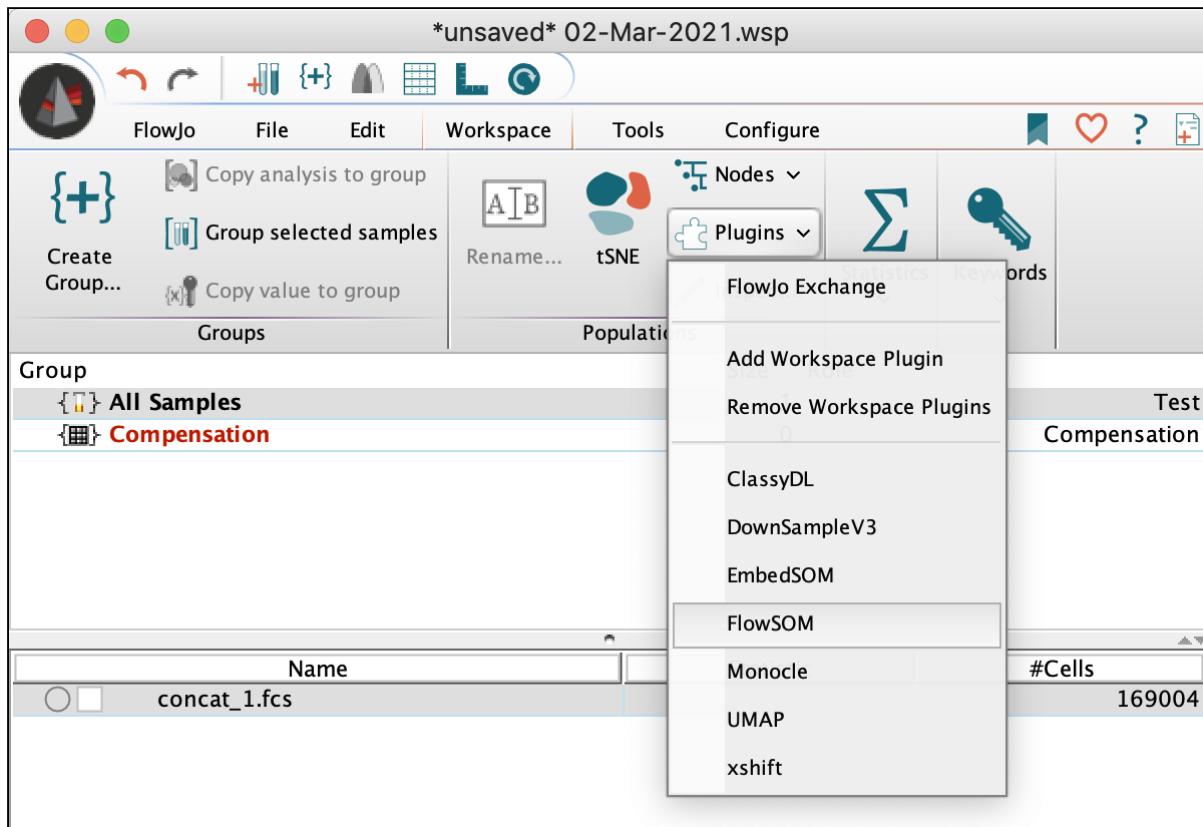
Name	Statistic	#Cells
concat_1.fcs		169004

You can check to see that all your samples have been included by opening the file, and plotting some parameter against 'SampleID' (or your custom parameter).

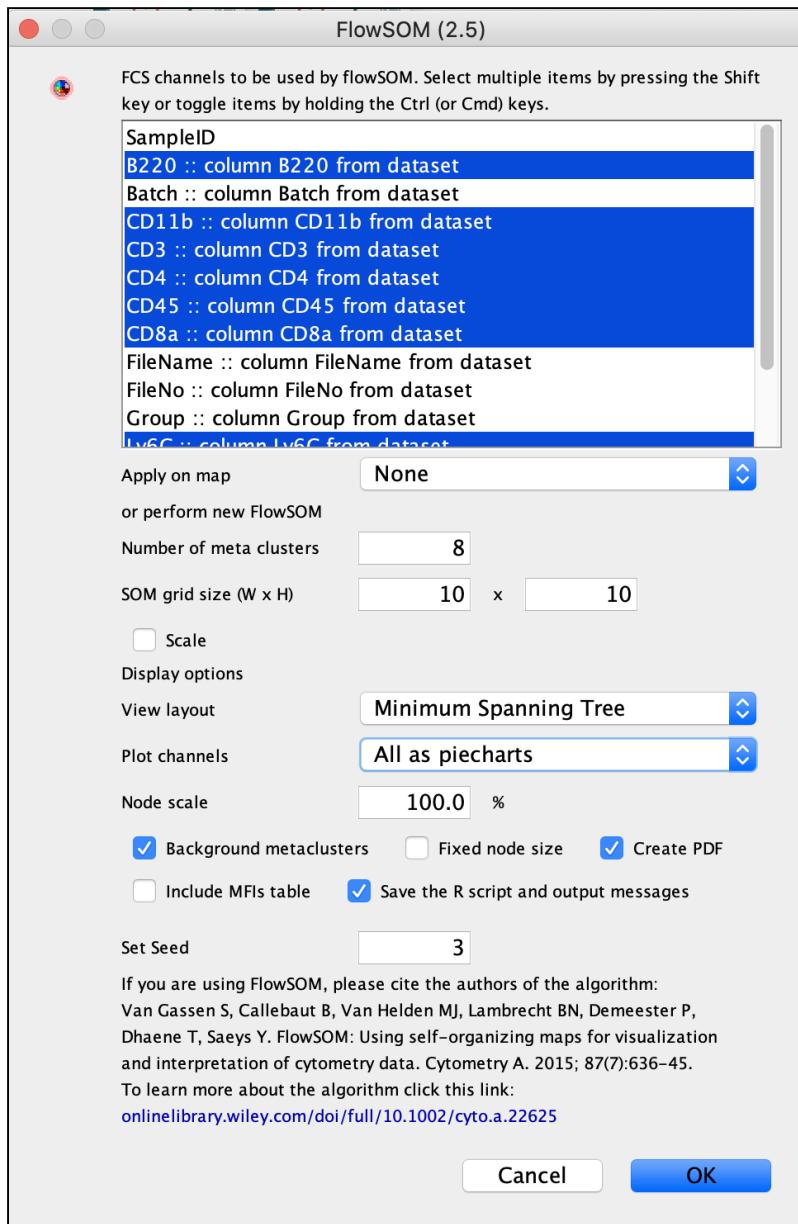


2. Clustering and dimensionality reduction

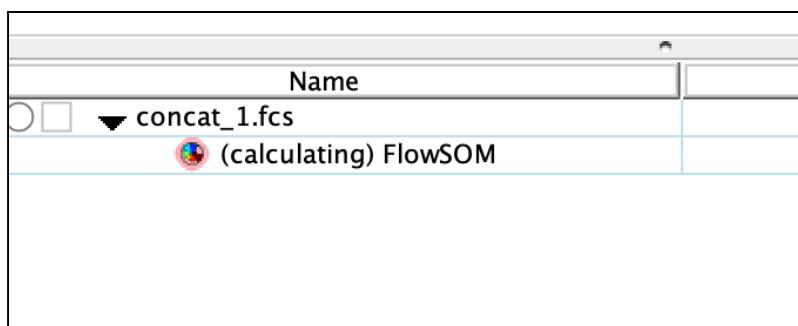
Select the file, and go to Workspace / Plugins / FlowSOM.

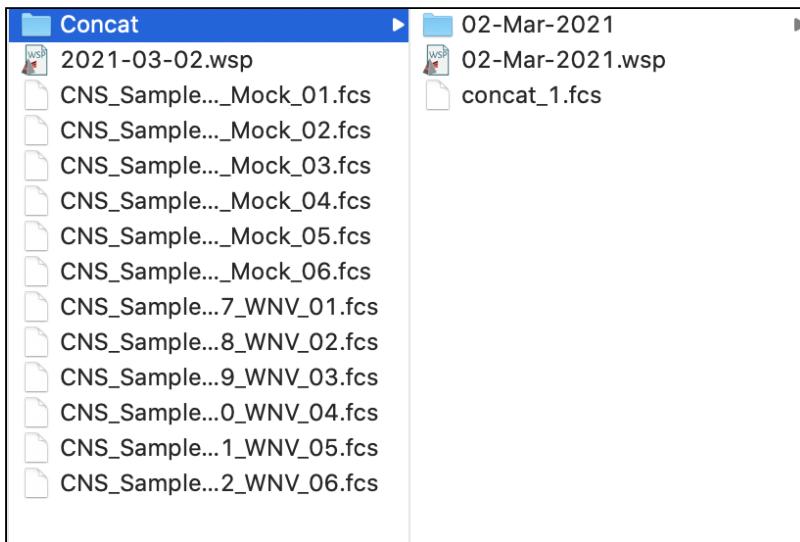


Choose the parameters you wish to use for FlowSOM clustering. You should also choose a target number of metaclusters. Click 'OK' when you are done.



You can see that FlowSOM is running by the appearance of this node below the file in FlowJo. You can also see some folders generated within your experiment folder.

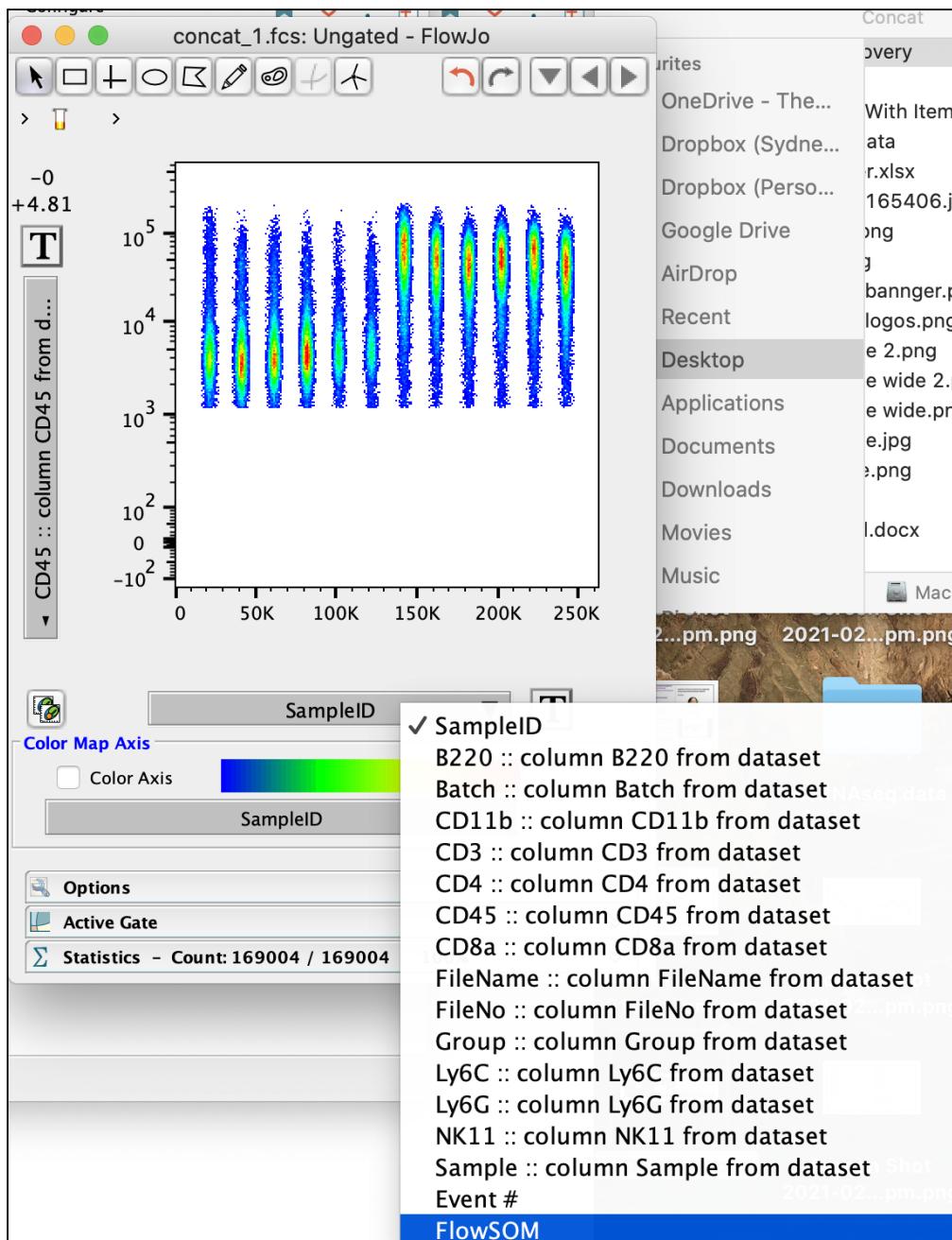


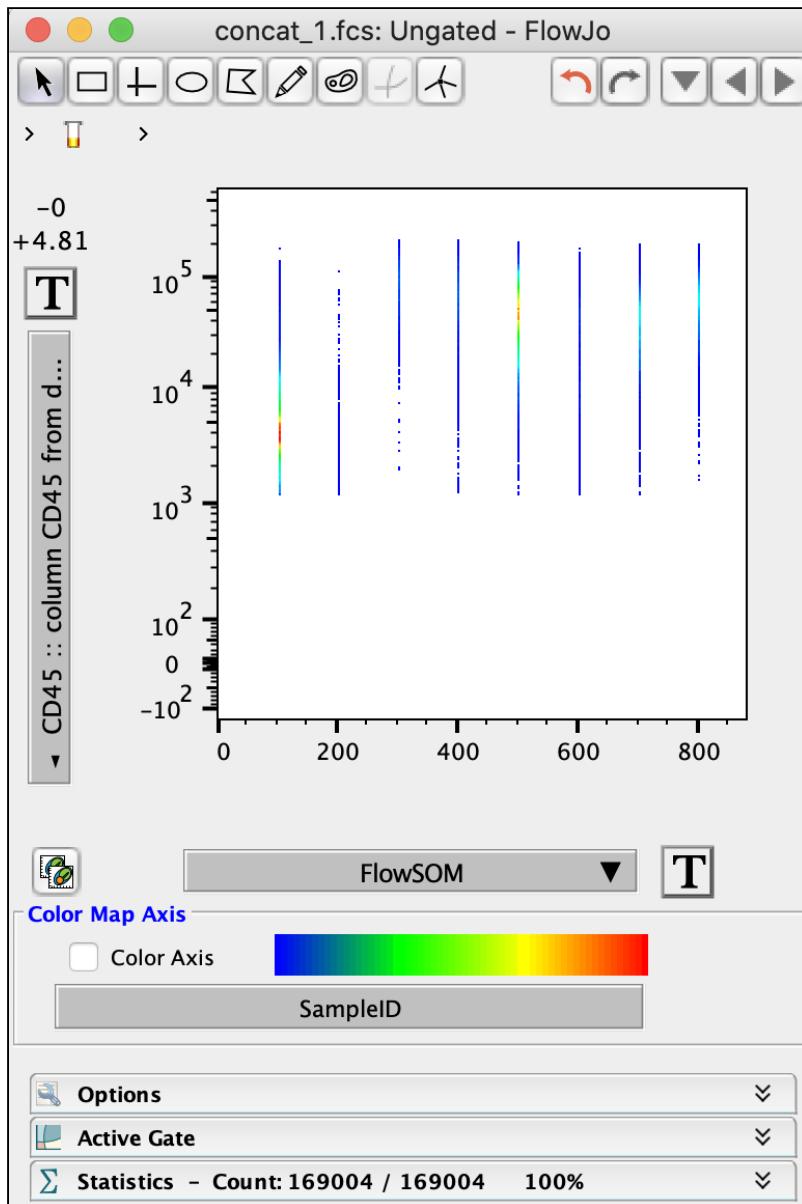


When FlowSOM has finished running, you will see the a/b node (containing the FlowSOM settings), and a virtual gate for each FlowSOM metaclusters.

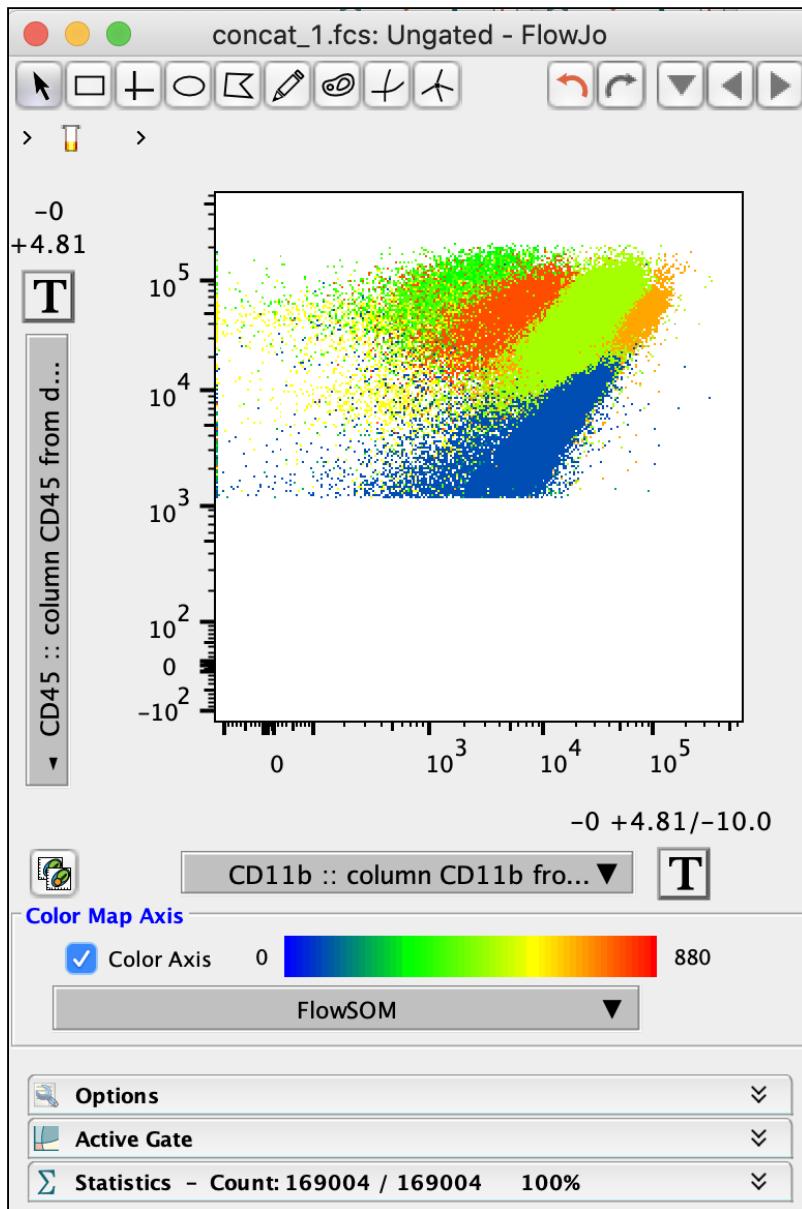
Name	Statistic	#Cells
concat_1.fcs		169004
FlowSOM		
FlowSOM		
FlowSOM.Pop0	40.9	69185
FlowSOM.Pop1	0.31	522
FlowSOM.Pop2	2.13	3594
FlowSOM.Pop3	2.32	3919
FlowSOM.Pop4	38.7	65439
FlowSOM.Pop5	1.50	2530
FlowSOM.Pop6	7.67	12955
FlowSOM.Pop7	6.43	10860

You can also see the metaclusters as a new 'parameter' – these will be distributed on a numerical scale between 0 and 1024.





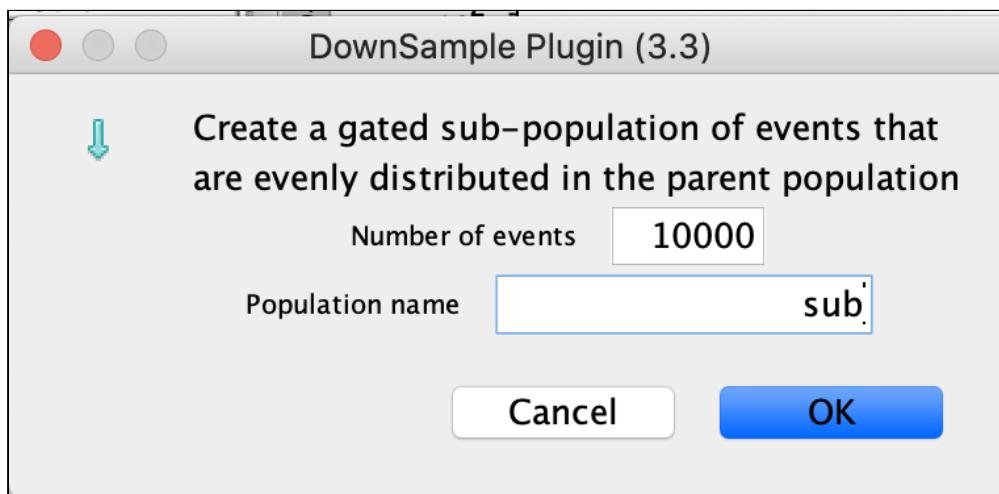
You can also use this as a 'colour axis' parameter when plotting two cellular parameters against each other.



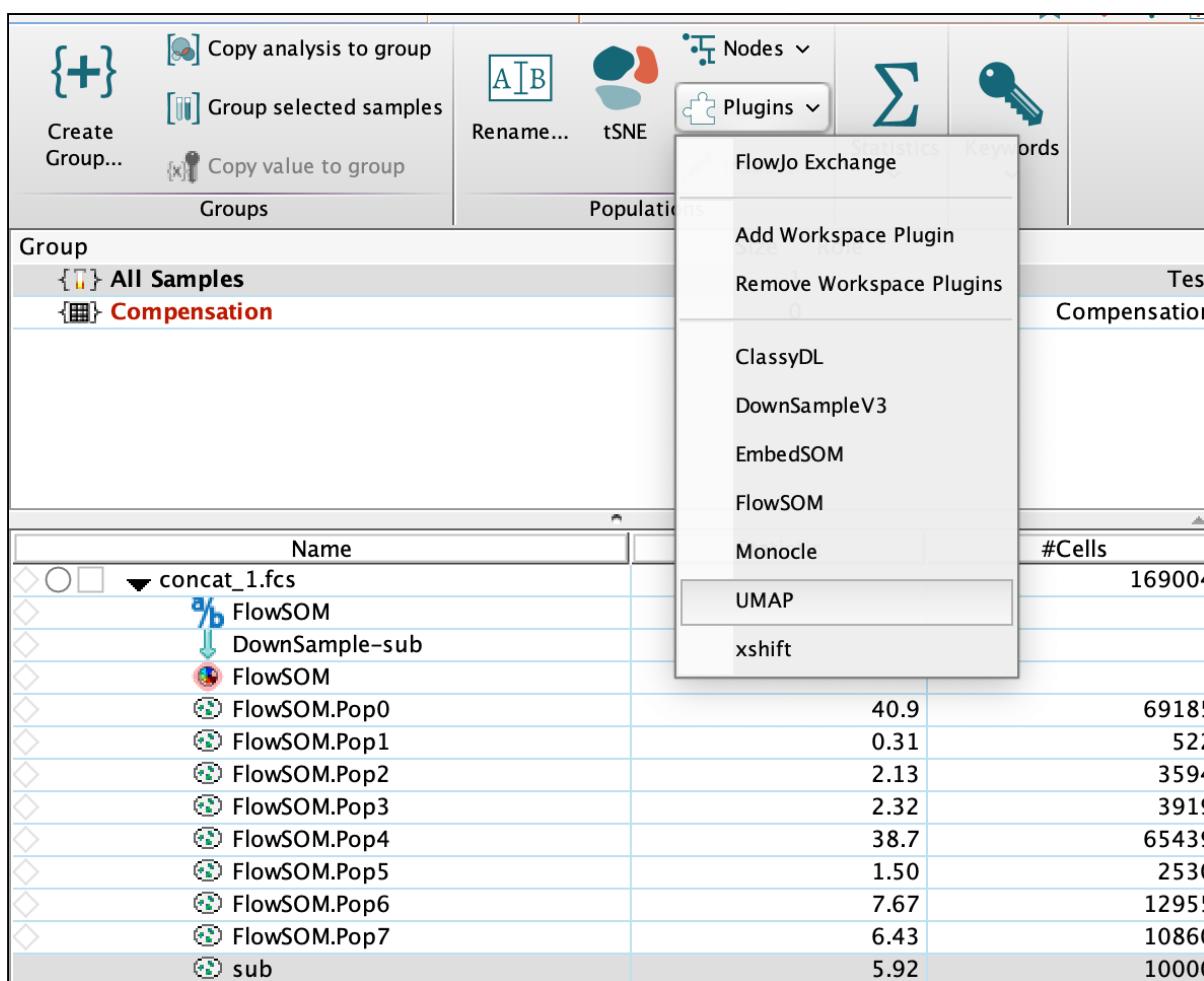
3. Dimensionality reduction and plotting

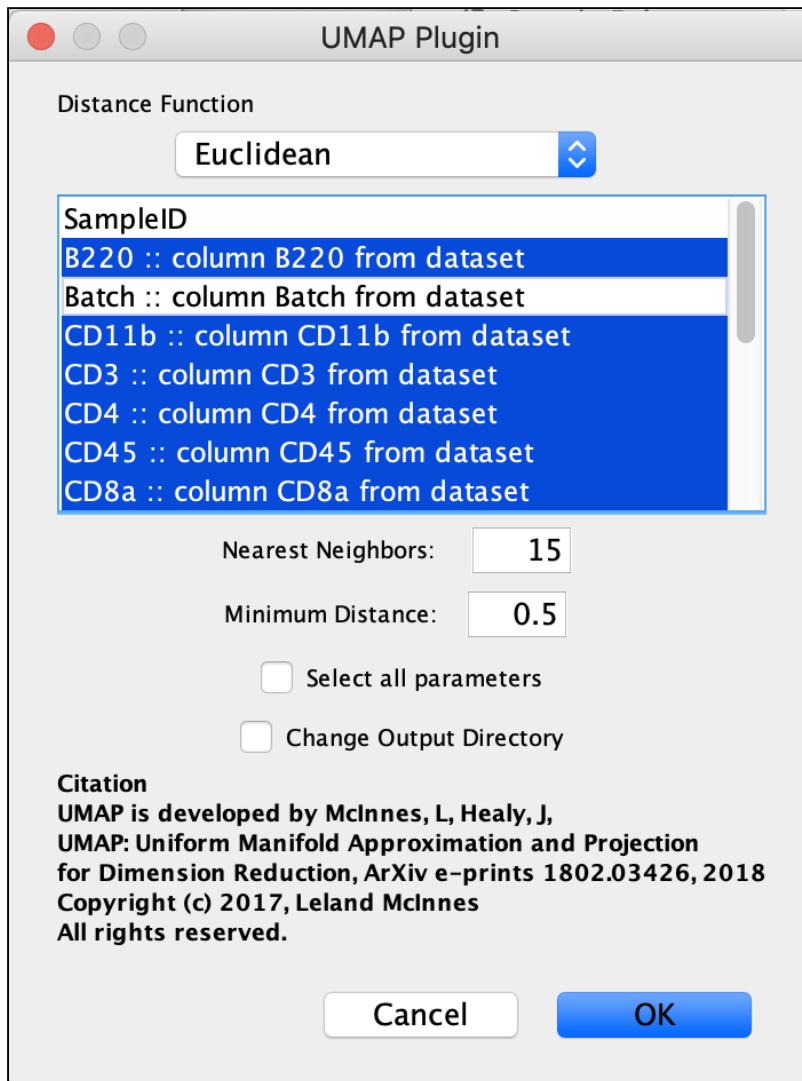
02-Mar-2021.wsp

Name	#Cells
FlowSOM	169004
FlowSOM	69185
FlowSOM.Pop0	0.31
FlowSOM.Pop1	522
FlowSOM.Pop2	2.13
FlowSOM.Pop3	3594
FlowSOM.Pop4	2.32
FlowSOM.Pop5	3919
FlowSOM.Pop6	38.7
FlowSOM.Pop7	65439
FlowSOM.Pop8	1.50
FlowSOM.Pop9	2530
FlowSOM.Pop10	7.67
FlowSOM.Pop11	12955
FlowSOM.Pop12	6.43
FlowSOM.Pop13	10860

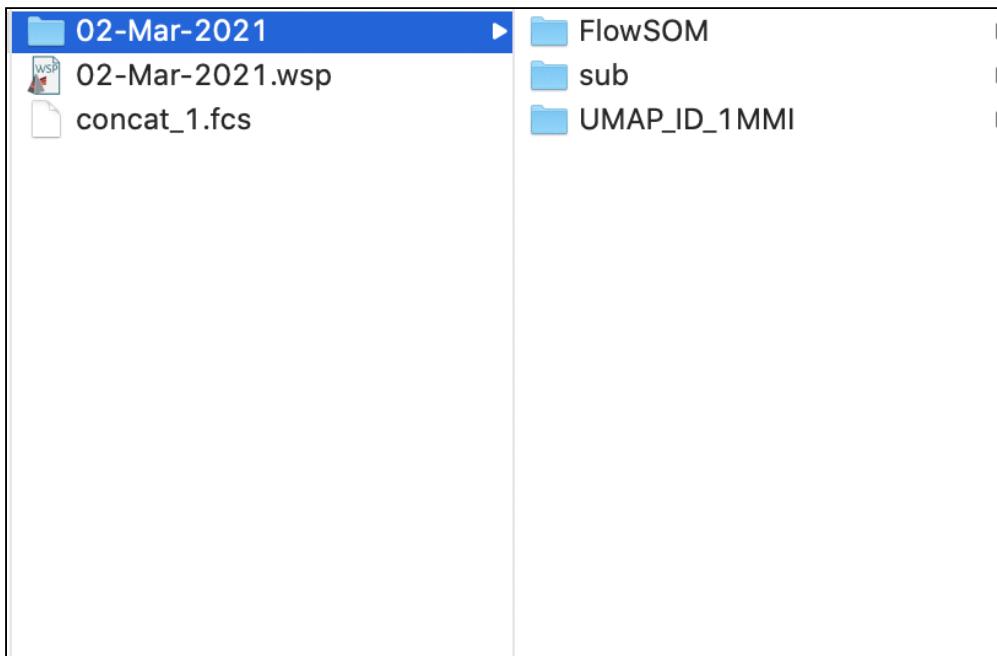


Name	Statistic	#Cells
concat_1.fcs		169004
FlowSOM		
DownSample-sub		
FlowSOM		
FlowSOM.Pop0	40.9	69185
FlowSOM.Pop1	0.31	522
FlowSOM.Pop2	2.13	3594
FlowSOM.Pop3	2.32	3919
FlowSOM.Pop4	38.7	65439
FlowSOM.Pop5	1.50	2530
FlowSOM.Pop6	7.67	12955
FlowSOM.Pop7	6.43	10860
sub	5.92	10000

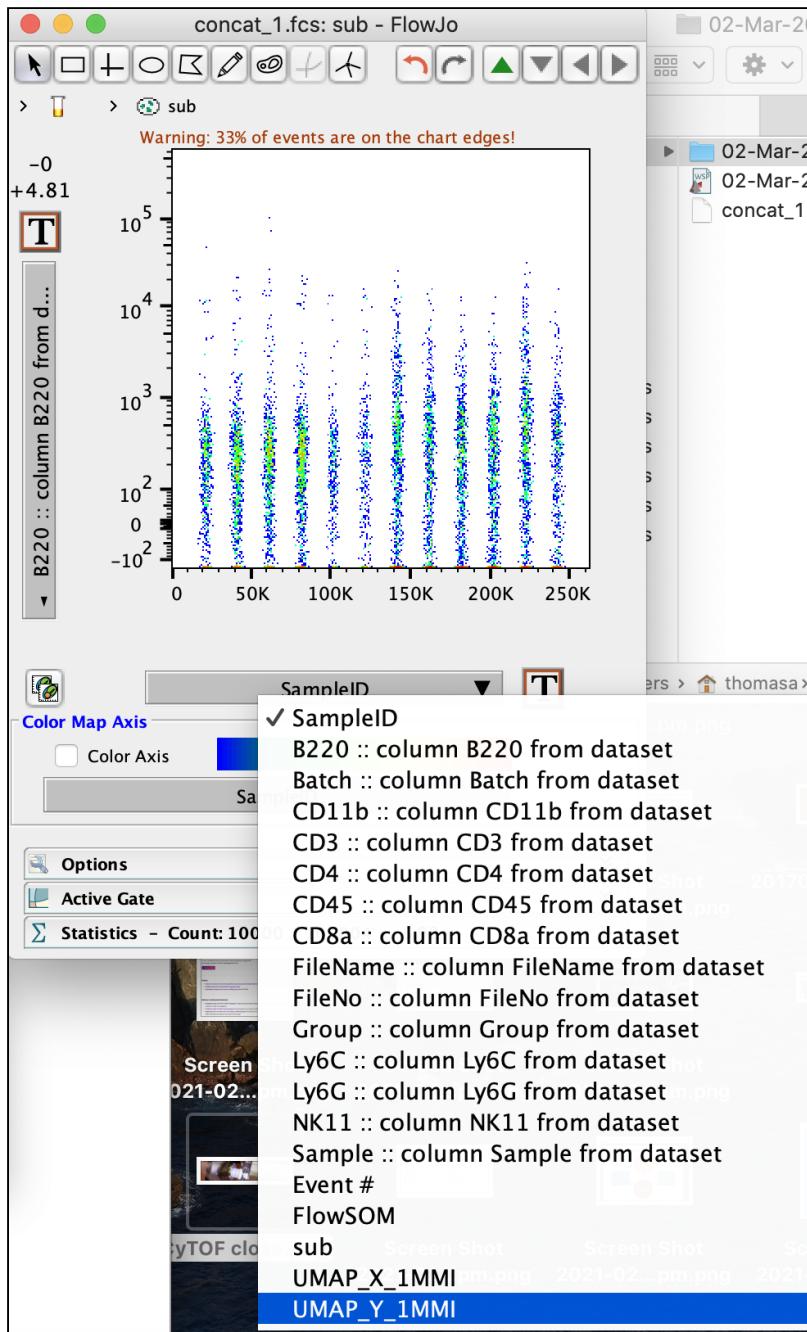


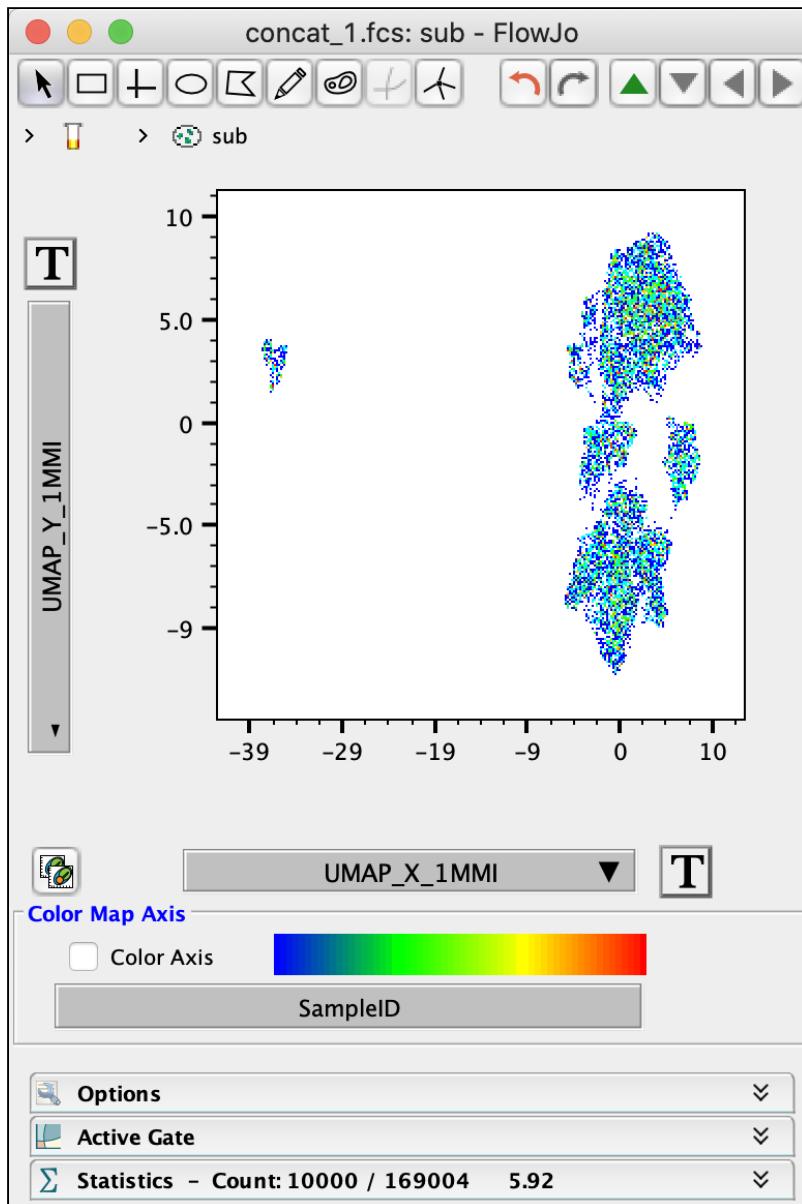


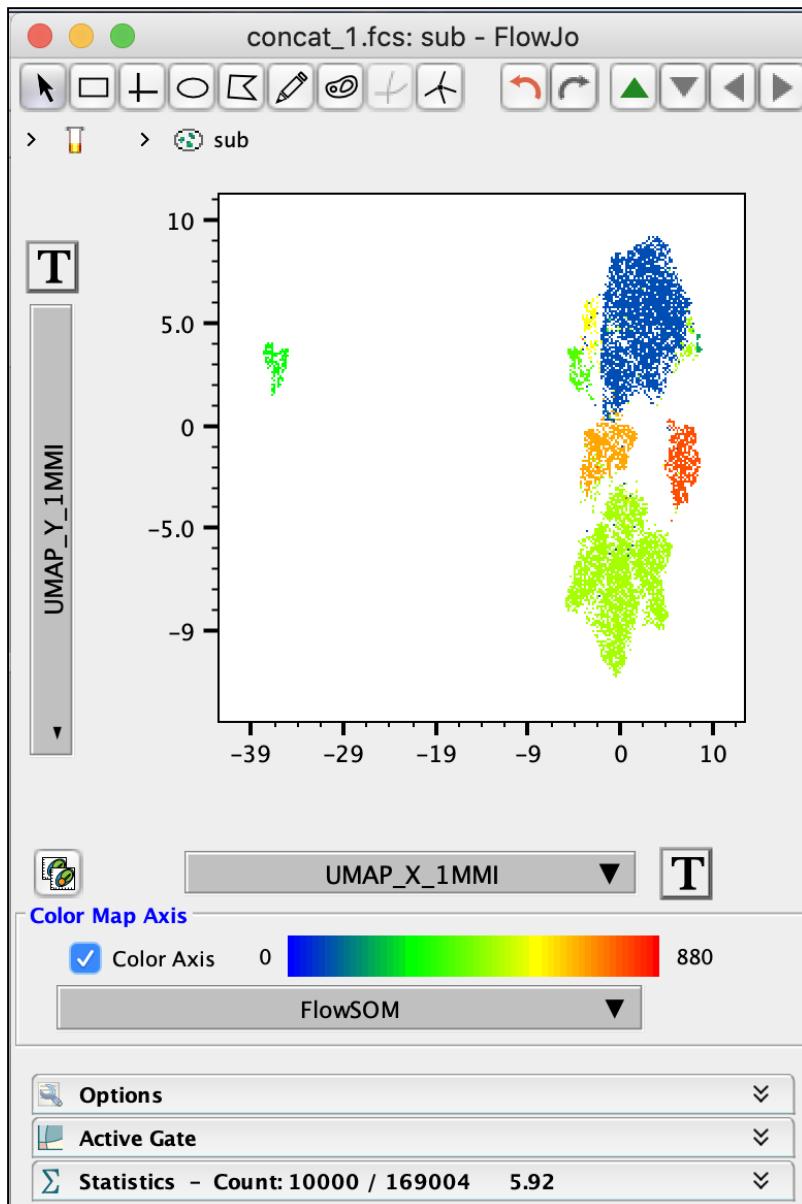
Name	Statistic	#Cells
concat_1.fcs		169004
FlowSOM		
DownSample-sub		
FlowSOM		
FlowSOM.Pop0	40.9	69185
FlowSOM.Pop1	0.31	522
FlowSOM.Pop2	2.13	3594
FlowSOM.Pop3	2.32	3919
FlowSOM.Pop4	38.7	65439
FlowSOM.Pop5	1.50	2530
FlowSOM.Pop6	7.67	12955
FlowSOM.Pop7	6.43	10860
sub	5.92	10000
(calculating) UMAP_ID_1MMI		

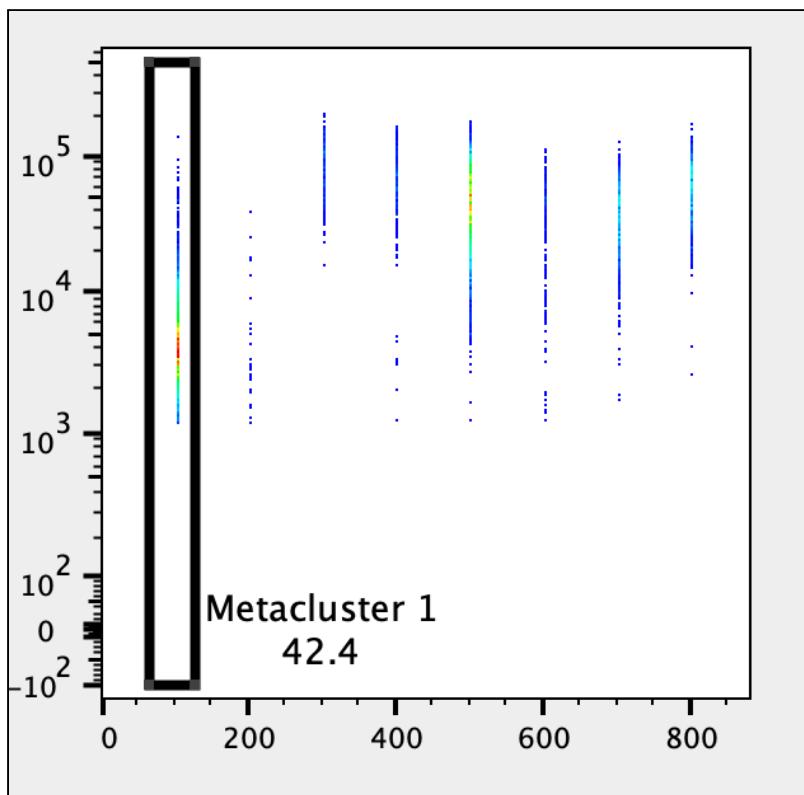


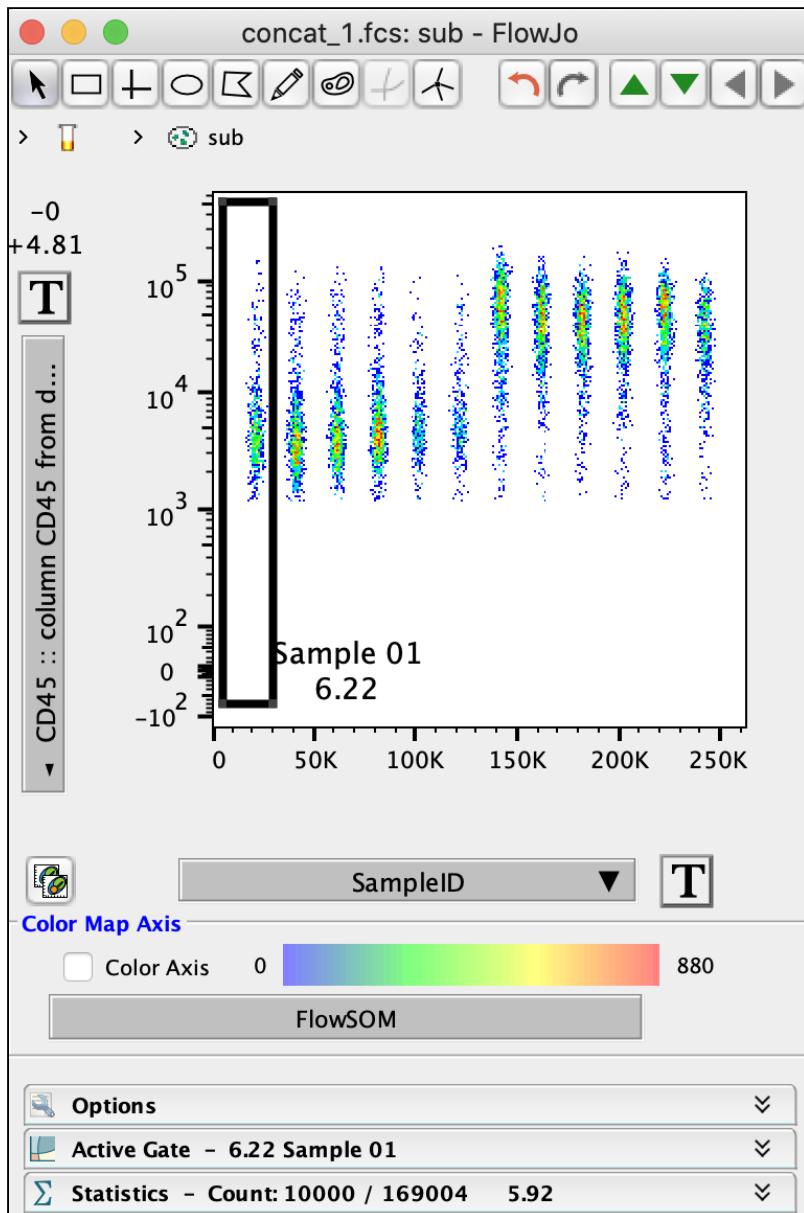
concat_1.fcs		
		169004
FlowSOM		
UMAP_X_1MMI		
UMAP_Y_1MMI		
DownSample-sub		
FlowSOM		
FlowSOM.Pop0	40.9	69185
FlowSOM.Pop1	0.31	522
FlowSOM.Pop2	2.13	3594
FlowSOM.Pop3	2.32	3919
FlowSOM.Pop4	38.7	65439
FlowSOM.Pop5	1.50	2530
FlowSOM.Pop6	7.67	12955
FlowSOM.Pop7	6.43	10860
sub	5.92	10000
UMAP_ID_1MMI of sub		UMAP completed

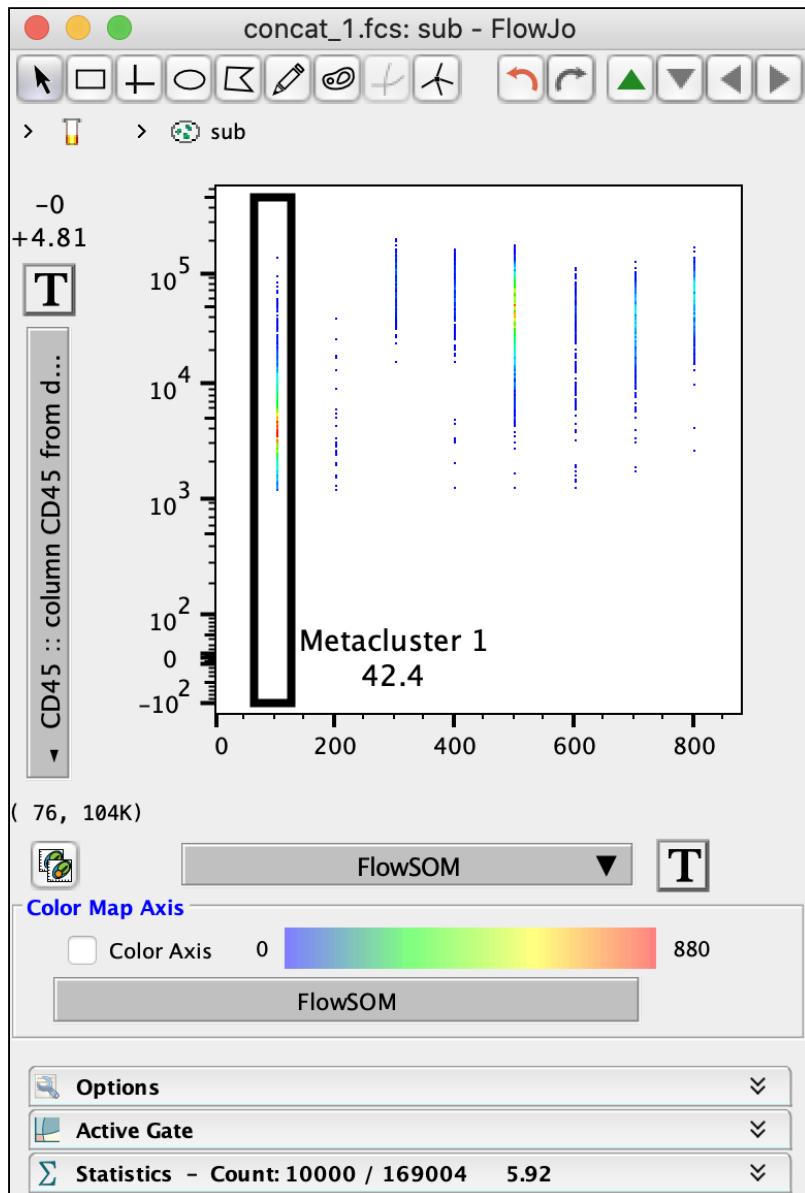


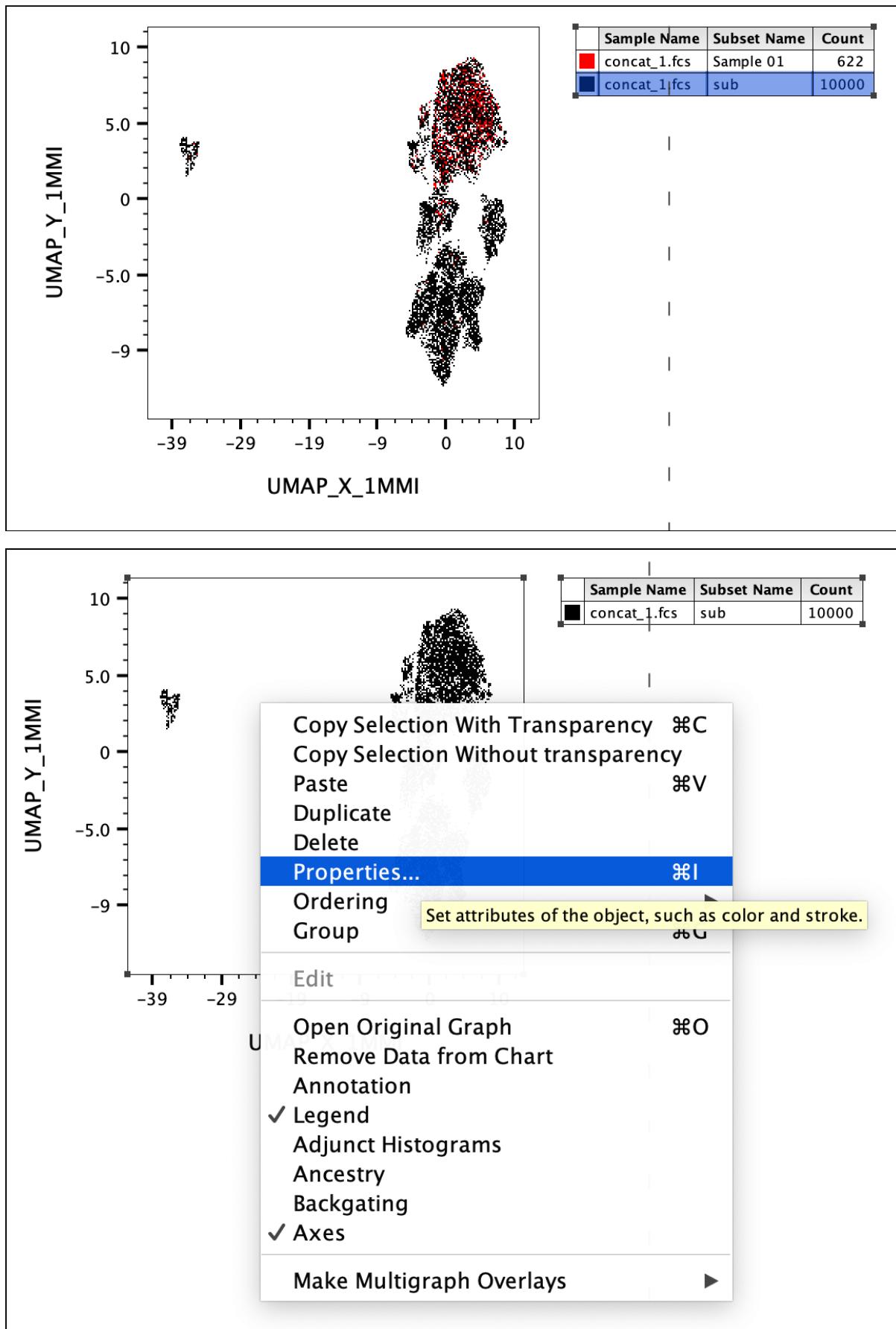


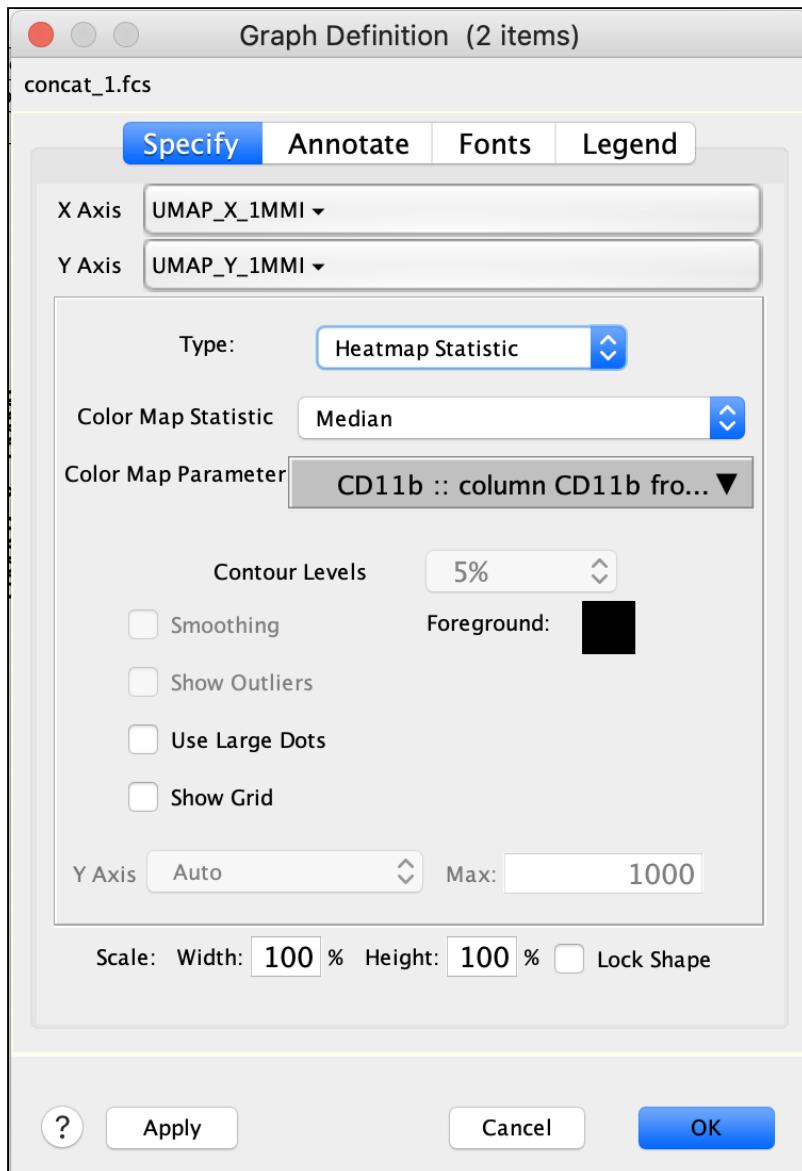


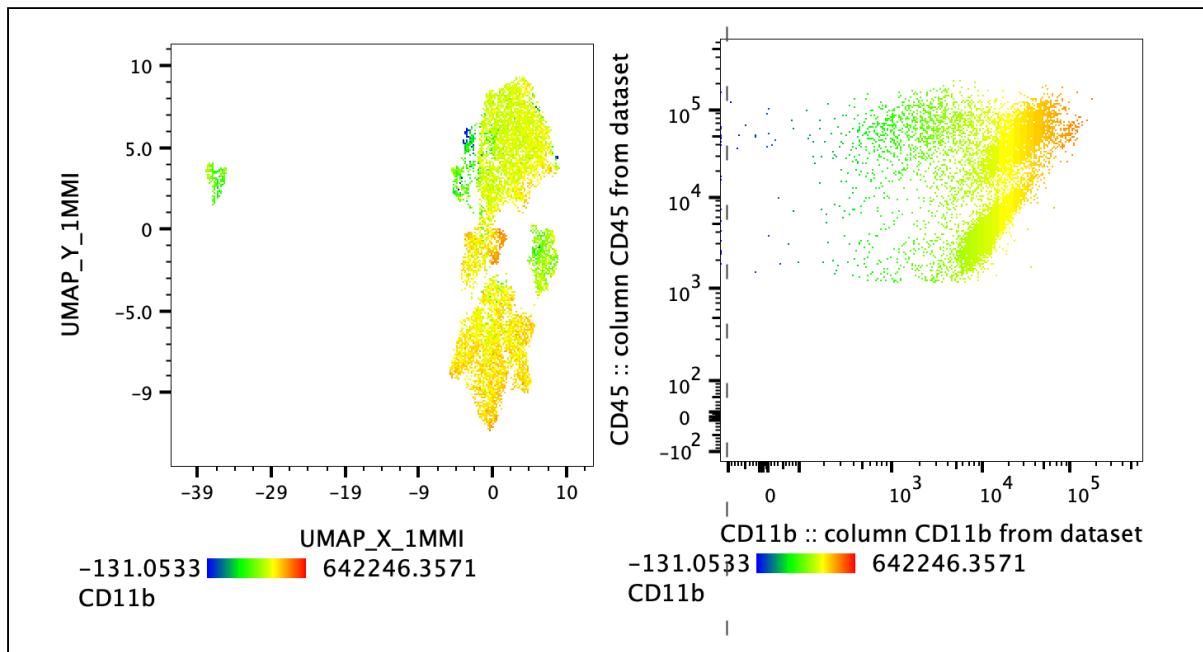












4. Quantitative and statistical analysis