Reliability and limits of transport-ventilators to safely ventilate severe patients in special surge situations.

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Abstract 299 words

Background: Several Intensive Care Units (ICU) have been overwhelmed by the surge of COVID-19

patients thus necessitating to extend ventilation capacity outside the ICU where air and oxygen

pressure are not always available. Transport ventilators requiring only O₂ source may be used to deliver

volume-controlled ventilation.

Objective: To evaluate the performances of four transport ventilators compared to an ICU ventilator

simulating severe respiratory conditions.

Materials and methods: Two pneumatic transport ventilators, (Oxylog 3000, Draeger; Osiris 3, Air

Liquide Medical Systems) and two turbine transport ventilators (Elisee 350, ResMed; Monnal T60, Air

Liquide Medical Systems) were compared to an ICU ventilator (Engström Carestation – GE Healthcare)

using a Michigan training test lung. We tested each ventilator with different set volumes Vt_{set} (350,

450, 550 ml) and different compliances (20 or 50 ml/cmH₂O) and a resistance of 15 cmH₂ O/L/sec based

on values recently described in COVID-19 Acute Respiratory Distress Syndrome. Volume error was

measured, as well as the trigger time delay during assist-control ventilation simulating spontaneous

breathing activity with a $P_{0.1}$ of 4 cm H_2O .

Results: Grouping all conditions, the volume error was 2.9 ± 2.2 % for Engström Carestation; 3.6 ± 3.9

% for Osiris 3; 2.5 ± 2.1 % for Oxylog 3000; 5.4 ± 2.7 % for Monnal T60 and 8.8 ± 4.8 % for Elisee 350.

Grouping all conditions, trigger delay was 42 ± 4 ms, 65 ± 5 ms, 151 ± 14 ms, 51 ± 6 and 64 ± 5 ms for

Engström Carestation, Osiris 3, Oxylog 3000, Monnal T60 and Elisee 350, respectively.

Conclusions: In special surge situations such as COVID-19 pandemic, most transport ventilators may

be used to safely deliver volume-controlled ventilation in locations where only oxygen pressure supply

is available with acceptable volume accuracy. Performances regarding triggering function are generally

acceptable but vary across ventilators.

Introduction

During the COVID 19 pandemic, some hospitals experienced the greatest shortage of ventilators ever seen since the heroic times of the polio epidemic in the 1950s. In this context, different alternative solutions including ventilator sharing and use of anesthesia ventilators have been considered to manage intubated patients with severe lung failure outside the walls of the ICU [1,2]. For this purpose, ventilators must be relatively easy for the users, able to deliver reliable volume-controlled ventilation in difficult mechanical conditions adapted to the principles of protective lung ventilation. Importantly, they must allow to vary FiO₂ without requiring two pressurized sources of gas, (i.e. wall air and oxygen at 50 psi). Several transport ventilators are based on pneumatic systems and Venturi systems for gas mixing; others use an internal turbine for pressurization; in this case, a low pressure source of oxygen is needed for increasing oxygen concentration, but not as a pressure source. They all represent interesting solutions in this context and could fulfill the requirements mentioned. Pneumatic transport ventilators have been used for decades both for in- and out-of-hospital transport. Their robustness and their relative technological simplicity could potentially facilitate massive industrial production. The general view on these ventilators is, however, that their limitations make them acceptable for a short period like transport but make them incompatible with the safe delivery of difficult ventilation for very sick patients over prolonged periods. Undoubtedly, they have limited capacities regarding ventilation modes and monitoring, but we questioned whether their performance for delivering lung protective ventilation in patients with ARDS merited to be tested again with these objectives in mind. Indeed, discarding their use in a context of surge could limit the extension of offering ICU beds possibilities for mechanically ventilated patients. The performance and limits of these ventilators have not been specifically tested with the appropriate settings and realistic conditions simulating the respiratory mechanics of patients with COVID-19 induced ARDS [3-6].

The aim of the present study was to evaluate the safety, reliability and limitations of ventilation provided by these different technologies in simulated bench conditions of severe respiratory mechanics mimicking patients with COVID-19 induced ARDS.

Materials and Methods

1. Ventilators

Brands

Four portable ventilators necessitating only one O₂ pressurized gas source were included in the study.

Two pneumatic transport ventilators using Venturi systems to mix air to oxygen were tested: the

Oxylog 3000 (Draeger, Lubeck, Germany) and the Osiris 3 (Air Liquide Medical Systems, Antony,

France). Two turbine transport ventilators necessitating additional oxygen only to increase FiO₂ were

also tested: the Elisee 350 (ResMed, San-Diego, USA) and the Monnal T60 (Air Liquide Medical Systems,

Antony France). The objective was to assess their capability to deliver acceptable ventilation by

comparing their performances to a standard ICU ventilator: Engström Carestation (GE healthcare,

Madison, USA). The characteristics of the five ventilators are given in Table 1.

Working principle and settings

In the two pneumatic transport ventilators tested (Oxylog 3000 and Osiris 3), the working pressure

that generates ventilation comes from the high-pressure oxygen supply. These ventilators based on

"Venturi-distributor" technology work as flow generator.

With the Oxylog 3000, the air-O₂ mixing is regulated from 40% to 100% via a Venturi system coupled

with a proportional inspiratory valve that also permits to directly set the volume (Vt_{set}). The inspiratory

flow depends on both the respiratory rate (RR) and the Inspiratory: Expiratory (I:E) ratio. In other

words, for a given set volume, changing RR and/or I:E ratio keep the set Vt but affects inspiratory flow.

The monitoring of the expired Vt is available via a specific flow sensor inserted between the

endotracheal tube and the patient circuit.

With the Osiris 3, the pneumatic system is based on the Venturi effect to entrain ambient air into the

ventilator for FiO₂ management coupled with a distributor for inspiratory flow setting. Only two

positions are available for FiO₂: 100% or an FiO₂ around 70%. For a given combination of I:E ratio and

respiratory rate, the Vt is set by directly adjusting a Vt knob that regulates the inspiratory flow. The

monitoring of the expired Vt is available via a specific flow sensor inserted between the endotracheal

tube and the patient circuit.

The Elisee 350 and T60 are two turbine-based ventilators which need oxygen only to adjust FiO₂. On

those ventilators, the Vt and the inspiratory flow are directly set. Changing the respiratory rate does

not affect neither Vt nor inspiratory flow.

The Engström Carestation is a classical high-quality ICU ventilator requiring two sources of pressurized

gas for oxygen and air (usually wall pressure at 50-55 psi).

Performances during volume control (VC) ventilation and assist volume control ventilation (ACV) were

evaluated with different conditions of simulated respiratory mechanics reproducing patients with

COVID-19 induced ARDS. All experiments have been performed in the Ventilatory Laboratory (Vent-

Lab) of the Angers Hospital, medical ICU.

2. Volume delivered and PEEP with different respiratory mechanics

We assessed the volume effectively delivered (Vtemeasured) by the ventilators in different conditions of

respiratory mechanics simulated on a Michigan test lung (Michigan Instruments, Kentwood, MI, USA).

A linear pneumotachograph (PNT 3700 series, Shawnee, USA) and a pressure transducer (SD160 series:

Biopac systems, Goleta, CA, USA) were used to measure flow and airway pressure. Signals were

converted with an analog digital converter (MP150; Biopac systems, Goleta, CA, USA) at a sample rate

of 200Hz, and stored in a laptop using a dedicated software (Acknowledge, Biopac Systems). Volume

was obtained from numerical integration of the flow signal.

Three set volumes (Vt_{set}) were tested: 350 mL, 450 mL and 550 mL, which approximately cover 6ml/kg

of Predicted Body Weight (PBW) for most adult patients. The different respiratory mechanics

conditions tested were: compliance of 50 ml/cmH₂O and 20 ml/cmH₂O, both combined with a resistance of 15 cmH₂O/L/s. The combinations of compliance and resistance tested were based on recently described COVID-19 respiratory mechanics [3-6].

Assist Control Ventilation (ACV) mode was selected and similar ventilator settings were applied for each ventilator (respiratory rate 30 cycles/min).

The pneumatic transport ventilators were set with an Inspiratory: Expiratory ratio of 1:3 (I:E) whereas a flow of 60L/min was adjusted on the Engström Carestation, Elisee 350 and Monnal T60.

The three set volumes were tested with FiO₂ 100% and 70% (air-O₂ mix for Osiris 3) as follows: FiO₂ was selected, Vt_{set} was adjusted on the ventilator and Vte_{measured} was recorded and averaged over 5 cycles after stabilization. FiO₂ was measured on Osiris 3 when air-O₂ mix was selected with a PF300 gas analyzer (IMT Medical, Buchs, Switzerland) in different conditions (Vt = 350 - 450 - 550 mL and Compliance = $20 - 50 \text{ mL/cmH}_2\text{O}$).

The performances of Venturi-based ventilation in terms of volume delivery could be altered by set inspiratory flow values [7]. To assess the impact of the inspiratory flow, Vte_{error} was calculated on the Osiris 3 ventilator. We set a volume of 450 ml using a wide range of inspiratory flows achieved by changing respiratory rate (RR). ACV mode with air-O₂ mix was selected, a resistance of 15 cmH₂O/L/s and a compliance of 20ml/cmH2O were applied and we set a I:E ratio of 1:3. The lowest RR (6 cycles/min) was chosen and was progressively increased by 4 cycles/min until reaching the maximum RR of 40 cycles/min. Vte_{set} had to be adjusted in consequence at each RR increment to keep its value at 450 ml. Vte_{error} was estimated at each step.

Two levels of PEEP were applied (10 cmH₂O and 15 cmH₂O) and the accuracy of the effective PEEP $(PEEP_{measured})$ was assessed.

Volume error and PEEP end-points:

The relative volume error (Vte_{error}), which is the difference between the effective expired volume (Vte_{measured}) and the set volume (Vte_{set}) was calculated and averaged as previously described over the four different conditions [8,9]:

Resistance = 15 cmH₂O/L/s, Compliance = 20 or 50 ml/cmH₂O, PEEP = 10 or 15 cmH₂O

The relative volume error was expressed in percentage and defined as follows:

$$Vte_{error} = \frac{|Vte_{measured} - Vte_{set}|}{Vte_{set}} \times 100$$

End-point for VT_{error}: The three tidal volumes tested were chosen to cover theoretical "6 ml/kg PBW" in adult male or female patients (350, 450 and 550 ml correspond to 6ml/kg PBW for respectively 58, 75 and 92 kg PBW).

Ventilation was considered safe and acceptable when $Vte_{measured}$ was within \pm 0.5 ml/kg PBW, which covers a volume between 5.5 and 6.5 ml/kg PBW. This corresponds to an 8% difference between set and measured Vt.

End-point for PEEP: a difference between measured PEEP and set PEEP was acceptable when less than 2 cmH₂O.

3. Synchronization and trigger performances

Assist control ventilation (ACV) with the inspiratory trigger function "on" was tested by connecting ventilators to a double chamber Michigan test lung (Michigan Instruments, Kentwood, MI, USA) to simulate spontaneous breathing. One chamber of the test lung was defined as the driving lung while the other chamber was connected to the ventilator being tested. A lung-coupling clip allowed a

connection between the two chambers, so that a positive pressure created in the driving lung induced

a negative pressure in the experimental lung, leading to trigger the ventilator tested.

The driving lung was connected to an Evita XL ventilator (Draeger, Lubeck, Germany) and set in volume-

controlled mode with constant flow. The respiratory rate was set at 25 breaths/ min. The ventilatory

settings were chosen to achieve a decrease in airway pressure 100 ms after occlusion (P_{0:1}) of 4 cmH₂0

(consistent with P_{0:1} values recently described in COVID patients [10]), measured at the airway opening

of the lung model [11]. A level of PEEP was applied to the driving lung to obtain a perfect contact of

the lung-coupling clip between the two chambers at the end of expiration.

For each ventilator tested, volume assist-control ventilation (ACV) mode was selected, with a tidal

volume of 450 mL, a respiratory rate of 20 cycles/min and a PEEP of 10 cmH₂O. I:E ratio was set at 1:3

for Osiris 3 and Oxylog 3000, while a flow of 60L/min was set on Elisee 350, Monnal T60 and the

Engström Carestation ICU ventilator. We selected air O_2 -mix for Osiris 3, and Fi O_2 of 70% on the other

ventilators. Inspiratory triggers were set at their most responsive position while avoiding auto-

triggering. The trigger of the Osiris 3 was set at -0.5 cmH₂O. Flow-triggered ventilators were set at 1

L/min for Engström Carestation and Oxylog 3000 and 2 L/min for Monnal T60 and Elisee 350.

For each configuration, trigger performance was assessed by measuring airway pressure changes using

the flow trace to determine the start of inspiration [9,12]. Negative pressure drop (ΔP , cmH₂0),

Triggering Delay (TD, ms) and Pressurization Delay (PD, ms) as defined on figure 1 were computed. The

overall Inspiratory Delay (ID) corresponds to the addition of TD and PD.

End-point: triggering function was considered as "safe and acceptable" when TD was less than 100 ms.

4. Statistical analysis

Continuous variables were expressed as mean ± SD values averaged from 5 consecutive breaths. These

variables were compared using an ANOVA test. The type I significance level was set at 0.05. When the

global F was significant, post hoc tests were computed using a student t-test with Bonferroni correction, which sets the level of significance for pairwise differences between the five ventilators at 0.005.

Results

1. Volume delivered and PEEP measured with different respiratory mechanics

Results are displayed in figure 2 and mean volume errors (Vte_{error}) for each ventilator are shown in

Table 2. When all conditions and set volumes were included, the Engström Carestation was the most

precise ventilator, and the Oxylog 3000 was comparable. Each ventilator performance was considered

as acceptable (delta Vt ± 0.5ml/kg PBW) except for one turbine ventilator (Elisee 350). The impact of

 FiO_2 selection (FiO2 100% or 70%) on volume error was significant considering all ventilators (p < 0.05),

reaching a maximum of 11.9 % for Elisee 350 with FiO₂ 100% (changes for each ventilator are available

on table 2). The impact of compliance on volume error was not significant considering all ventilators

(p > 0.05) and changes for each ventilator are available on table 2. FiO₂ measured on Osiris 3 reached

 $72.3 \pm 1.7\%$ regarding all the conditions tested.

When all conditions were put together, differences between measured PEEP and set PEEP were less

than 2 cmH₂O as shown on table 2.

2. Impact of inspiratory flow on pneumatic ventilators

The effect of inspiratory flow rates on Vte_{error} for Osiris 3 is shown on figure 3. The lowest values of

inspiratory flow are associated with a Vte_{error} above 8% (delta Vt ± 0.5ml/kg PBW). Performances are

acceptable when inspiratory flow (resulting from the combination of Vt, I:E ratio and respiratory rate)

was strictly above 30 L/min, which corresponds to a respiratory rate higher than 20 cycles/min.

3. Synchronization and trigger performances during ACV

Inspiratory trigger was evaluated for each ventilator and results are displayed on figure 4. All simulated

efforts triggered a ventilatory cycle. The Triggering Delay was 42 ± 4 ms, 65 ± 5 ms, 151 ± 14 ms, 51 ± 14

6 ms and 64 ± 5 ms for Engström Carestation, Osiris 3, Oxylog 3000, Monnal T60 and Elisee 350,

respectively (all conditions grouped). Each ventilator performance was considered as acceptable (TD < 100ms) except for one pneumatic ventilator (Oxylog 3000).

The Inspiratory Delay (ID) was measured at 54 ± 5 ms for Engström Carestation, 95 ± 5 ms for Osiris 3, 217 \pm 21 ms for Oxylog 3000, 72 \pm 6 ms for Monnal T60 and 85 \pm 7 ms for Elisee 350 and (p < 0.05; pairwise differences between ventilators were all significant with a p-value < 0.005). A similar trend was observed for triggering delay.

The airway pressure drop was much larger for Oxylog 3000 (- 4.1 ± 0.3 cmH₂O), than for the others:-0.9 ± 0.3 cmH₂O for Engström Carestation, -1.9 ± 0.1 cmH₂O for Osiris 3, , -0.6 ± 0.1 cmH₂O for Monnal T60 and -0.9 ± 0.1 for Elisee 350 (p < 0.05; pairwise differences between ventilators were all significant with a p-value < 0.005, except between Osiris 3 and Elisee 350).

Discussion

The results of the present bench test study comparing turbine and pneumatic transport ventilators to an ICU ventilator, can be summarized as follows: 1. Turbine ventilators show performance in VC and ACV very close to the ICU ventilator tested for most of the settings. 2. In severe respiratory mechanics conditions, the volume error exceeds 0.5 ml/kg PBW only for one turbine ventilator. 3. Inspiratory trigger reactivity is less than 100 ms except for one pneumatic transport ventilator. 4. Volume error delivered by the simplest pneumatic ventilator significantly increases when inspiratory flow is less than 30 L/min indicating a technological limit of the Venturi system.

The increasing number of patients requiring mechanical ventilation in the context of the COVID-19 worldwide crisis, and the ventilators shortage reported in some severely affected countries, has led to discuss the possibilities to manage intubated patients outside the walls of the ICU [2]. According to this dire scenario, simple and easy to set ventilators that only require one oxygen pressure source to function and able to deliver lung protective ventilation could be considered. In addition, an assisted volume mode that ensures the set Vt with PEEP up to 15 cmH₂O and FiO₂ up to 100% is required to manage high elastic load and severe shunt that characterize potentially severe COVID-19 ARDS [1,2]. Recent turbine transport and emergency ventilators have performances which are very close to conventional ICU ventilators [13,14]. In the context of "mass casualty", as experienced with the COVID-19 crisis, pneumatic transport ventilators could be used to extend the possibility to manage intubated patients in case of ICU beds shortage. The working principle of these pneumatic ventilators is based on a "Venturi system" which is a simple technological solution that permits to manage ventilation generated by the oxygen pressurized source when a position called air-O₂ mix is selected. Interestingly, the simplicity of such pneumatic systems permits to consider massive industrialization faster and at a lower cost. On the opposite, the Venturi system explains the limits observed with low inspiratory flow previously described with this technology [7].

Performances during controlled ventilation:

In case of high impedance, a low inspiratory flow may increase significantly volume error when the O₂

air mix position is selected "on". In turn, manipulating I:E ratio, respiratory rate and increasing

inspiratory flow above 30 L/min permits to reverse the Vt error that is directly explained by the working

principle of this ventilator (see figure 3). The technological adaptations available on Oxylog 3000

(Venturi coupled with proportional inspiratory valve) solve this problem while expired Vt monitoring

available on Osiris 3 simplifies settings adaptation if required. Previous bench test studies have

reported a Vt error with pneumatic basic transport ventilators that reached 20% of set Vt with resistive

load [7,15]. These experiments were performed with very low set inspiratory flow thus explaining the

Vt reduction observed. For this purpose, an essential recommendation is to set the I:E ratio at 1:3

(minimal available value), and set the RR above 20 /min before adjusting the knob (that controls the

inspiratory flow) to reach the set Vt as indicated on the ventilator. With these recommendations,

volume error measured on pneumatic transport ventilators at low compliance are closed from turbine

performances and acceptable since it did not exceed 0.5 ml/kg PBW.

Of note, only the ICU and turbine ventilators tested compensate for the loss in Vt due to the

compression of gas inside the circuit. Nevertheless, this effect previously quantified in ICU ventilators

with inspiratory-expiratory circuits is significantly less in basic transport ventilators since they are

equipped with a single limb circuit [8]. Of note, an HEPA filter can be easily adjusted on the expiratory

limb and they should not expose to higher risks of viral contamination.

Performances during assisted ventilation:

Recent experience with COVID-19 induced ARDS reports that these patients often exhibit high

respiratory drive and asynchrony that may require deep sedation and sometimes paralysis [4]. We

therefore evaluated the behavior of these two pneumatic transport ventilators during triggered

breaths since performances of their trigger have been questioned [14,15]. The synchronization was

overall respected except for the Oxylog 3000 exhibiting the poorest triggering performances. The

triggering time delay was consistently longer in pneumatic ventilators but acceptable except on the

Oxylog 3000, compared to the ICU ventilator [14].

Limitations

The results obtained in vitro necessitate some caution to be translated to the clinical practice, but

previous studies showed that this type of simulation predicts the results observed in clinical situations

with a high fidelity [8,16,17]. The lung model gives the unique opportunity to compare ventilator

performances according to several simulated but standardized clinical conditions. Bench experiment

also permits to accurately depict and understand advantages and limits of the different ventilator's

technologies as previously done [7]. Our experiment reported performances of only two pneumatic

and two turbine ventilators while several other ventilators with similar technology are available

worldwide. We did not evaluate pressure support ventilation while this approach can be useful to

manage weaning of COVID-19 patients. Previous studies already showed that turbine-based ventilator

significantly outperform pneumatic transport ventilators during pressure mode ventilation [13,14].

Conclusion

The present bench study suggests that turbine technologies may replace ICU ventilators to extend ICU

beds where only oxygen pressure supply is available, in special surge situations such as COVID-19 crisis.

Pneumatic transport ventilators provide acceptable volume accuracy even in severe simulated

conditions. For this purpose, attention is required to maintain sufficient inspiratory flow in the Osiris 3

(or any other pneumatic ventilator using a venturi system). A monitoring of expired Vt available on the

two pneumatic transport ventilators tested greatly facilitates settings. Performances regarding

triggering function are non-acceptable in one of the pneumatic transport ventilator thus rendering

hazardous its use in assist control ventilation.

References:

- [1] Beitler JR, Mittel AM, Kallet R, Kacmarek R, Hess D, Branson R, et al. Ventilator Sharing during an Acute Shortage Caused by the COVID-19 Pandemic. Am J Respir Crit Care Med. 15 août 2020;202(4):600-4.
- [2] Emanuel EJ, Persad G, Upshur R, Thome B, Parker M, Glickman A, et al. Fair Allocation of Scarce Medical Resources in the Time of Covid-19. N Engl J Med. 21 mai 2020;382(21):2049-55.
- [3] Pan C, Chen L, Lu C, Zhang W, Xia J-A, Sklar MC, et al. Lung Recruitability in COVID-19—associated Acute Respiratory Distress Syndrome: A Single-Center Observational Study. Am J Respir Crit Care Med. 15 mai 2020;201(10):1294-7.
- [4] Gattinoni L, Coppola S, Cressoni M, Busana M, Rossi S, Chiumello D. Covid-19 Does Not Lead to a "Typical" Acute Respiratory Distress Syndrome. Am J Respir Crit Care Med. 30 mars 2020;rccm.202003-0817LE.
- [5] Gattinoni L, Chiumello D, Caironi P, Busana M, Romitti F, Brazzi L, et al. COVID-19 pneumonia: different respiratory treatments for different phenotypes? Intensive Care Med. 14 avr 2020;s00134-020-06033-2.
- [6] Grasselli G, Zangrillo A, Zanella A, Antonelli M, Cabrini L, Castelli A, et al. Baseline Characteristics and Outcomes of 1591 Patients Infected With SARS-CoV-2 Admitted to ICUs of the Lombardy Region, Italy.

 JAMA. 28 avr 2020;323(16):1574.
- [7] Breton L, Minaret G, Aboab J, Richard J-C. Fractional inspired oxygen on transport ventilators: an important determinant of volume delivery during assist control ventilation with high resistive load. Intensive Care Med. août 2002;28(8):1181-1181.
- [8] Lyazidi A, Thille AW, Carteaux G, Galia F, Brochard L, Richard J-CM. Bench test evaluation of volume delivered by modern ICU ventilators during volume-controlled ventilation. Intensive Care Med. déc 2010;36(12):2074-80.

- [9] Boussen S, Gainnier M, Michelet P. Evaluation of Ventilators Used During Transport of Critically III Patients: A Bench Study. Respiratory Care. 1 nov 2013;58(11):1911-22.
- [10] Gattinoni L, Marini JJ, Camporota L. The Respiratory Drive: An Overlooked Tile of COVID-19 Pathophysiology. Am J Respir Crit Care Med. 3 sept 2020;rccm.202008-3142ED.
- [11] Telias I, Junhasavasdikul D, Rittayamai N, Piquilloud L, Chen L, Ferguson ND, et al. Airway Occlusion Pressure As an Estimate of Respiratory Drive and Inspiratory Effort during Assisted Ventilation. Am J Respir Crit Care Med. 1 mai 2020;201(9):1086-98.
- [12] Richard J-C, Carlucci A, Breton L, Langlais N, Jaber S, Maggiore S, et al. Bench testing of pressure support ventilation with three different generations of ventilators. Intensive Care Med. août 2002;28(8):1049-57.
- [13] Thille AW, Lyazidi A, Richard J-CM, Galia F, Brochard L. A bench study of intensive-care-unit ventilators: new versus old and turbine-based versus compressed gas-based ventilators. Intensive Care Med. août 2009;35(8):1368-76.
- [14] L'Her E, Roy A, Marjanovic N. Bench-test comparison of 26 emergency and transport ventilators. Crit Care. oct 2014;18(5):506.
- [15] Zanetta G, Robert D, Guérin C. Evaluation of ventilators used during transport of ICU patients a bench study. Intensive Care Med. avr 2002;28(4):443-51.
- [16] Carteaux G, Lyazidi A, Cordoba-Izquierdo A, Vignaux L, Jolliet P, Thille AW, et al. Patient-Ventilator Asynchrony During Noninvasive Ventilation. Chest. août 2012;142(2):367-76.
- [17] Vignaux L, Tassaux D, Jolliet P. Performance of noninvasive ventilation modes on ICU ventilators during pressure support: a bench model study. Intensive Care Medicine. 23 juill 2007;33(8):1444-51.

Table 1. General characteristics of the ventilators

	Engström Carestation	Osiris 3	Oxylog 3000	Monnal T60	Elisee 350
Manufacturer	GE Healthcare	Air Liquide Medical Systems	Draeger	Air Liquide Medical Systems	Resmed
Weight [kg]	31.0	5.0	5.4	3.7	4.0
Working pressure	Pressurized oxygen and air	Pressurized oxygen	Pressurized oxygen	Pressurized oxygen	Pressurized oxygen
Expired volume monitoring	Yes	Yes	Yes	Yes	Yes
Tidal volume (Vt) [mL]	20 - 2000	100 - 2000	50 - 2000	20 - 2000	50 - 2500
PEEP [cmH20]	1 - 50	0 - 15	0 - 20	0 - 20	0 - 25
Peak inspiratory pressure [cmH20]	7 - 100	10 - 80	PEEP + 3 - PEEP + 55	0 - 80	0 - 100
FiO ₂ [%]	21 - 100	70 or 100	40 - 100	21 - 100	21 - 100
Battery duration [h]	0.5 - 2	6 - 14	4	2.5 - 5	3 - 6

Table 2. Mean volume errors and Positive End Expiratory Pressure (PEEP) measured for each ventilator

	Engström Carestation	Osiris 3	Oxylog 3000	Monnal T60	Elisee 350
Vte _{error} global [%]	2.9 ± 2.2	3.6 ± 3.9	2.5 ± 2.1	5.4 ± 2.7	8.8 ± 4.8
Vte _{error} 100% FiO ₂ [%]	1.0 ± 0.7	3.7 ± 3.7	2.0 ± 1.2	3.3 ± 1.4	11.9 ± 4.1
Vte _{error} 70% FiO ₂ [%]	4.9 ± 1.3	3.5 ± 4.2	2.9 ± 2.7	7.5 ± 2.0	5.9 ± 3.5
Vte _{error} C50 [%]	3.3 ± 2.7	3.4 ± 4.0	1.6 ± 1.1	5.1 ± 3.1	10.4 ± 5.3
Vte _{error} C20 [%]	2.6 ± 1.6	3.8 ± 3.8	3.4 ± 2.6	5.7 ± 2.2	7.0 ± 3.7
mean PEEP 10 [cmH ₂ O]	9.9 ± 0.2	10.6 ± 0.6	11.5 ± 0.3	9.5 ± 0.5	10.3 ± 0.1
mean PEEP 15 [cmH ₂ O]	15.1 ± 0.2	14.9 ± 0.6	15.4 ± 2.2	14.7 ± 0.1	15.4 ± 0.2

Table 2. Abbreviations

- Vteerror global = mean volume error including all conditions of resistance (15 cm $H_2O/L/sec$), compliance (20 50 ml/cm H_2O) and PEEP (10 15 cm H_2O) for both 100% FiO2 and 70% FiO2.
- Vteerror 100% FiO2 = mean volume error including all conditions of resistance (15 cm $H_2O/L/sec$), compliance (20 50 ml/cm H_2O) and PEEP (10 15 cm H_2O) for 100% FiO2.
- Vteerror 70% FiO2 = mean volume error including all conditions of resistance (15 cm $H_2O/L/sec$), compliance (20 50 ml/cm H_2O) and PEEP (10 15 cm H_2O) for 70% FiO2.
- Vteerror C50 = mean volume error including all conditions of resistance (15 cm $H_2O/L/sec$), FiO2 (100 70 %) and PEEP (10 15 cm H_2O) for a compliance of 50 ml/cm H_2O .
- Vteerror C20 = mean volume error including all conditions of resistance (15 cm $H_2O/L/sec$), FiO2 (100 70%) and PEEP (10 15 cm H_2O) for a compliance of 20 ml/cm H_2O .
- Mean PEEP 10 = mean PEEP measured when PEEP was set at 10 cm H_2O including all conditions of resistance (15 cm $H_2O/L/sec$) and compliance (20 50 ml/cm H_2O) for both 100% FiO2 and 70% FiO2.
- Mean PEEP 15 = mean PEEP measured when PEEP was set at 15 cm H_2O including all conditions of resistance (15 cm $H_2O/L/sec$) and compliance (20 50 ml/cm H_2O) for both 100% FiO2 and 70% FiO2.

Figure 1. Explicative figure of ventilator triggering assessment

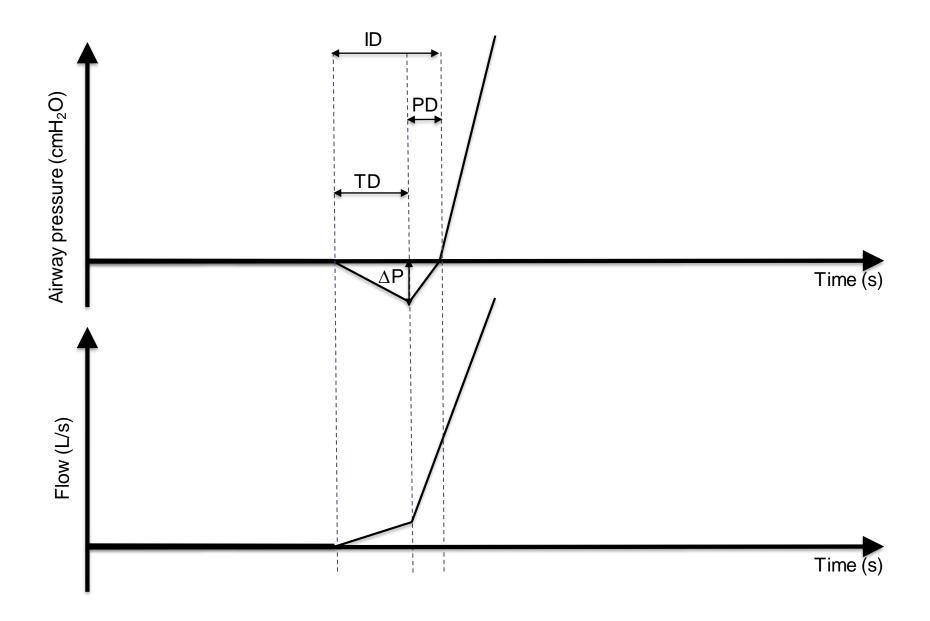
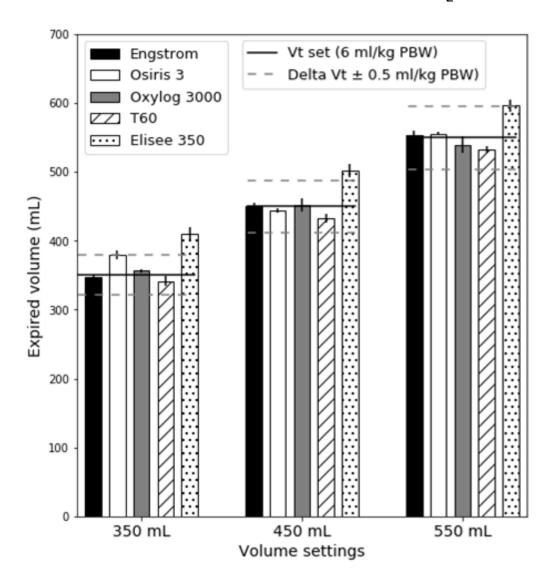


Figure 2. Tidal Volume delivery in volume control ventilation in static conditions

Panel A: Volume Control 100% FiO₂



Panel B: Volume Control 70% FiO₂

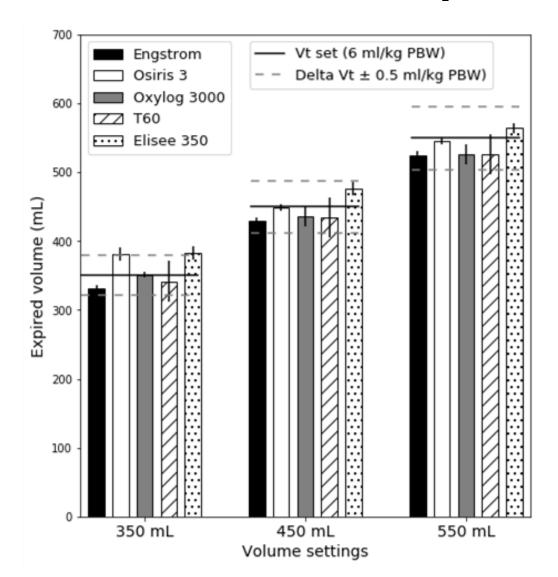


Figure 3. Impact of flow on effective volume with Osiris 3 ventilator

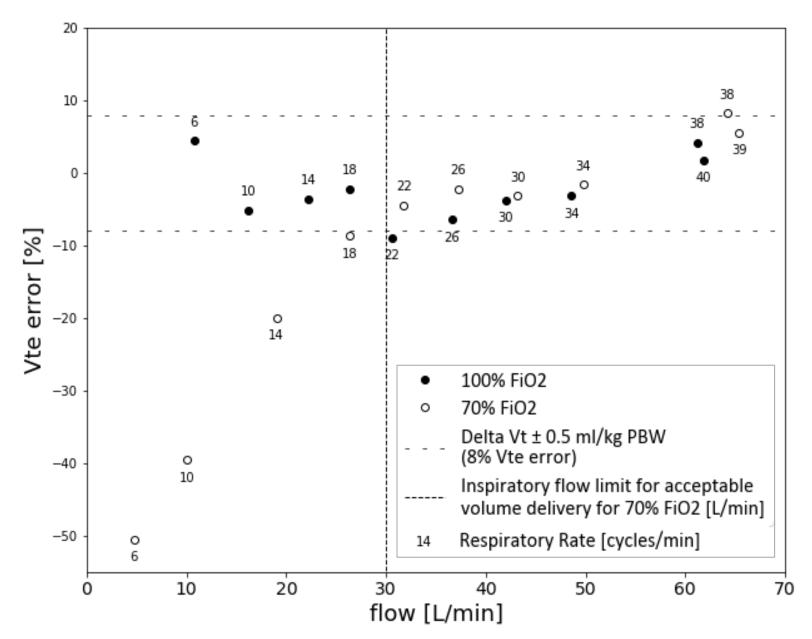


Figure 4. Triggering efficiency in volume assist-control ventilation

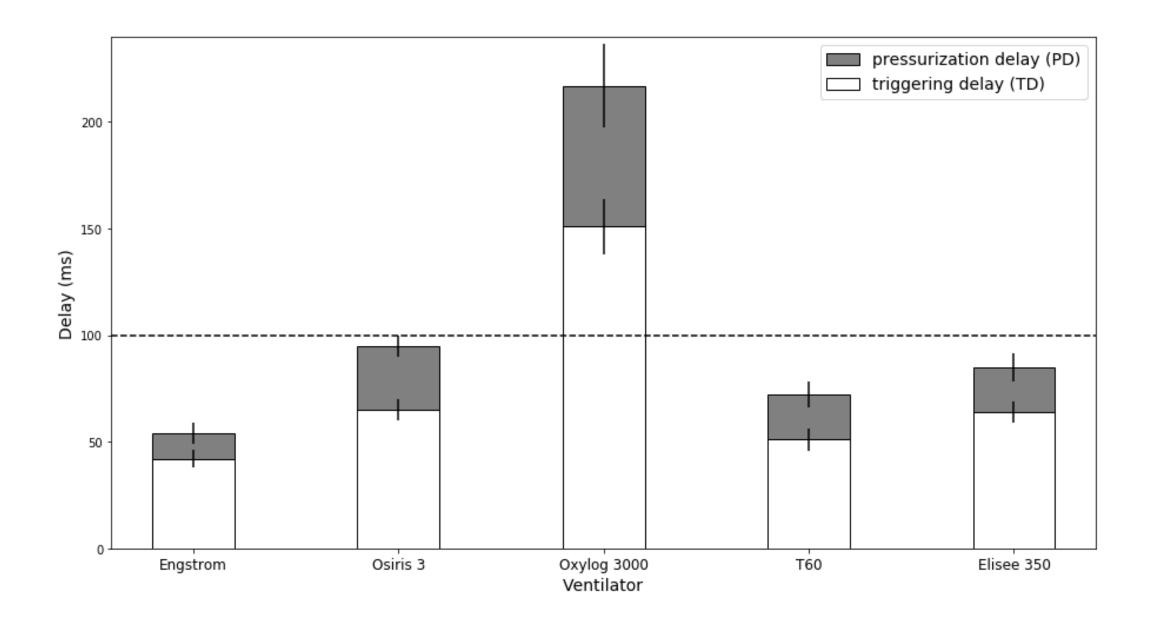


Figure 1.

The figure illustrates ventilator triggering assessment. Airway pressure (Paw) and flow are displayed. Triggering delay (TD) is the delay between the onset of airway pressure drop ("patient" effort) and flow delivery by the ventilator. Pressurization delay (PD) is defined by the time at which the airway pressure comes back to the level of PEEP. The addition of TD and PD gives the inspiratory delay (ID). The drop of airway pressure (ΔP) due to patient effort is also shown on the figure.

Figure 2.

Panel A. The histogram represents the mean expired volumes measured for each ventilator according to the three Vt set in 100% FiO₂. The average was computed over the four conditions of resistance (15 cmH₂O/L/sec), compliance (20 - 50 ml/cmH₂O) and PEEP (10 - 15 cmH₂O). The three tidal volumes tested were chosen to cover 6 ml/kg PBW, with 350, 450 and 550 ml corresponding to 6ml/kg PBW for respectively 58, 75 and 92 kg PBW. Limits of acceptable ventilation are displayed with dotted lines and defined as a volume change within \pm 0.5 ml/kg PBW, which corresponds to a Vt between 5.5 and 6.5 ml/kg PBW.

Panel B. The histogram represents the mean expired volumes measured for each ventilator according to the three Vt set in 70% FiO₂. The average was computed over the four conditions of resistance (15 cmH₂O/L/sec), compliance (20 - 50 ml/cmH₂O) and PEEP (10 - 15 cmH₂O).

Figure 3.

This figure shows the volume error of the Osiris 3 expressed in % of Vt set according to different inspiratory flows obtained at a constant 450 ml Vt set. Compliance, resistance and PEEP were set at 20 ml/cm H_2O , 15 cm H_2O /L/sec and 10 cm H_2O respectively. Black circles were obtained with 100% FiO₂ while the white circles were obtained with 70% FiO₂. Respiratory rate associated with each point is also displayed. This figure illustrates that for an inspiratory flow below 30 L/min, the Vt error is substantial with 70% FiO₂. The Vt error is within \pm 0.5 ml/kg PBW (which corresponds to an 8% difference between set and measured Vt) whatever the inspiratory flow when 100% FiO₂ is selected.

Figure 4.

The figure illustrates the triggering efficiency for each ventilator tested during assist-control ventilation using the Michigan test lung to simulate spontaneous breathing. A PEEP of $10 \text{ cmH}_2\text{O}$, a compliance of $20 \text{ and } 50 \text{ ml/cmH}_2\text{O}$ and a resistance of $15 \text{ cmH}_2\text{O/L/sec}$ were selected. Triggering Delay (TD, ms) and Pressurization Delay (PD, ms) were computed. A definition of TD and PD is available on figure 1. Triggering function was considered safe and acceptable when TD was less than 100 ms.