# Initial Analysis

## Boyd Tarlinton

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```
library(phyloseq)
library(qiime2R)
library(tidyverse)
## -- Attaching packages
## v ggplot2 3.3.2
                       v purrr
                                  0.3.4
## v tibble 3.0.1
                       v dplyr
                                  1.0.0
## v tidyr
            1.1.0
                       v stringr 1.4.0
                       v forcats 0.5.0
## v readr
             1.3.1
## -- Conflicts -----
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()
                     masks stats::lag()
library(magrittr)
##
## Attaching package: 'magrittr'
## The following object is masked from 'package:purrr':
##
##
       set_names
## The following object is masked from 'package:tidyr':
##
##
       extract
library(microbiome)
##
## microbiome R package (microbiome.github.com)
##
##
##
##
    Copyright (C) 2011-2020 Leo Lahti,
##
       Sudarshan Shetty et al. <microbiome.github.io>
##
## Attaching package: 'microbiome'
  The following object is masked from 'package:ggplot2':
##
##
       alpha
## The following object is masked from 'package:base':
```

##

```
## transform
library(vegan)

## Loading required package: permute
## Loading required package: lattice
## This is vegan 2.5-6

##
## Attaching package: 'vegan'

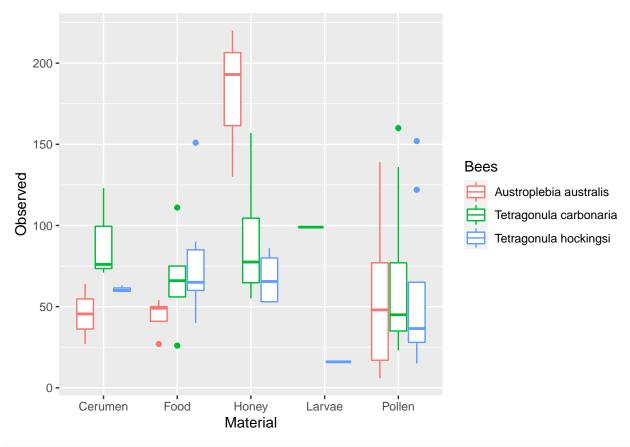
## The following object is masked from 'package:microbiome':
##
## diversity
library(symbioteR)
```

```
hiveMB_1 <- qza_to_phyloseq(features = "QIIME_Processing_1/feat_tab.qza",
   taxonomy = "QIIME_Processing_1/taxonomy.qza")
tax_1 <- read_qza("QIIME_Processing_1/taxonomy.qza")</pre>
tax_table(hiveMB_1) <- as.character(tax_1$data$Taxon) %>%
  strsplit(";") %>%
  do.call(what = qpcR:::rbind.na) %>%
  set_colnames(c("Kingdom", "Phylum", "Class", "Order", "Family",
                             "Genus", "Species")) %>%
  data.frame() %>%
  lapply(str_replace_all, "D_.__", "") %>%
  as.data.frame() %>%
  set_rownames(tax_1$data$Feature.ID) %>%
  as.matrix()
hiveMB_2 <- qza_to_phyloseq(features = "QIIME_Processing_2/feat_tab.qza",
   taxonomy = "QIIME_Processing_2/taxonomy.qza")
tax_2 <- read_qza("QIIME_Processing_2/taxonomy.qza")</pre>
tax_table(hiveMB_2) <- as.character(tax_2$data$Taxon) %>%
  strsplit(";") %>%
  do.call(what = qpcR:::rbind.na) %>%
  set_colnames(c("Kingdom", "Phylum", "Class", "Order", "Family",
                             "Genus", "Species")) %>%
  data.frame() %>%
  lapply(str_replace_all, "D_.__", "") %>%
  as.data.frame() %>%
  set_rownames(tax_2$data$Feature.ID) %>%
  as.matrix()
hiveMB <- merge_phyloseq(hiveMB_1, hiveMB_2)</pre>
sample_data(hiveMB) <- read.csv("Metadata_03082020.csv") %>%
 column to rownames("ID")
```

### hiveMB <- removeNonbacterial(hiveMB)</pre> makeObservationsRelative(hiveMB) %>% prevalenceFilter(threshold = 0.05) %>% plot\_bar(fill = "Class") + facet\_wrap(~ Bees + Material, scales = "free\_x", nrow = 1) ebia ebia ebia ebia ila c ila l ila l ila l ila l ila l Food Ione 'olle rum Food lone arva folle rum Food lone arva folle rum 1.00 -Class Actinobacteria 0.75 Alphaproteobacteria Abundance Bacilli 0.50 Bacteroidia Clostridia Erysipelotrichia 0.25 Gammaproteobacteria Mollicutes 0.00 2000 2000 11111 0000 0000 0000 11111 1111 ,,,,,,,,,, ..... FlaviaM-16S-69 FlaviaM-16S-70 000000 000000 111111 (00000) מששט מששט הוחוד

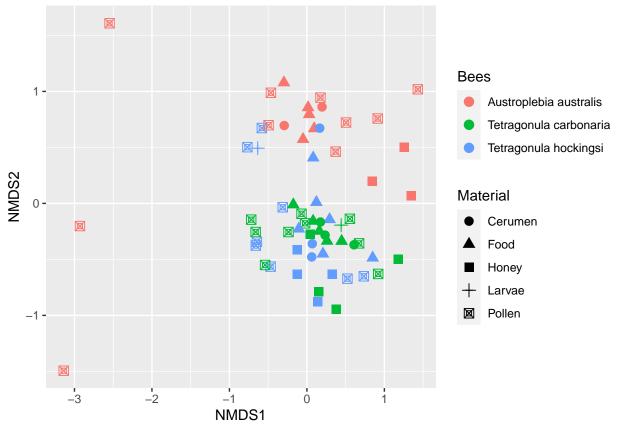
```
alphaDiv <- estimate_richness(hiveMB, measures = c("Observed")) %>%
  cbind(data.frame(sample_data(hiveMB)))
ggplot(alphaDiv, aes(x = Material, y = Observed, color = Bees)) + geom_boxplot()
```

Sample



```
hiveMB <- transform(hiveMB, "clr")</pre>
otu_table(hiveMB)[otu_table(hiveMB) <= 0] <- 0</pre>
ord <- ordinate(hiveMB, distance = "bray", method = "NMDS")</pre>
## Wisconsin double standardization
## Run 0 stress 0.1975073
## Run 1 stress 0.1792445
## ... New best solution
## ... Procrustes: rmse 0.08878056 max resid 0.3129829
## Run 2 stress 0.1984643
## Run 3 stress 0.1767967
## ... New best solution
## ... Procrustes: rmse 0.02579431 max resid 0.08901046
## Run 4 stress 0.1768018
## ... Procrustes: rmse 0.000583197 max resid 0.002737982
## ... Similar to previous best
## Run 5 stress 0.1824837
## Run 6 stress 0.1763814
## ... New best solution
## ... Procrustes: rmse 0.01552371 max resid 0.05670576
## Run 7 stress 0.1988439
## Run 8 stress 0.1763114
## ... New best solution
## ... Procrustes: rmse 0.008162403 max resid 0.03649033
## Run 9 stress 0.1965415
## Run 10 stress 0.176388
```

```
## ... Procrustes: rmse 0.005753602 max resid 0.03233328
## Run 11 stress 0.1798948
## Run 12 stress 0.179392
## Run 13 stress 0.1791805
## Run 14 stress 0.1768018
## ... Procrustes: rmse 0.01053242 max resid 0.05014851
## Run 15 stress 0.1988445
## Run 16 stress 0.1794633
## Run 17 stress 0.1976074
## Run 18 stress 0.1767494
## ... Procrustes: rmse 0.007332822 max resid 0.04213818
## Run 19 stress 0.2040346
## Run 20 stress 0.1763949
## ... Procrustes: rmse 0.00599557 max resid 0.03361826
## *** No convergence -- monoMDS stopping criteria:
##
        1: no. of iterations >= maxit
       19: stress ratio > sratmax
plot_ordination(hiveMB, ord, color = "Bees", shape = "Material") +
  geom_point(size = 3)
```



```
adonis(dist ~ Bees + Material + Hive_ID, as(sample_data(hiveMB), "data.frame"))

##
## Call:
## adonis(formula = dist ~ Bees + Material + Hive_ID, data = as(sample_data(hiveMB), "data.frame")
```

dist <- phyloseq::distance(hiveMB, method = "bray")</pre>

```
##
## Permutation: free
## Number of permutations: 999
##
## Terms added sequentially (first to last)
##
             Df SumsOfSqs MeanSqs F.Model
##
                                               R2 Pr(>F)
                   3.4246 1.71230 6.7357 0.15940 0.001 ***
## Bees
                   2.7422 0.68555 2.6967 0.12763
## Material
                                                   0.001 ***
                   3.3700 0.33700 1.3257 0.15686 0.002 **
## Hive_ID
             10
## Residuals 47
                  11.9480 0.25421
                                          0.55611
## Total
             63
                  21.4848
                                          1.00000
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

#### Session Info

#### sessionInfo()

```
## R version 4.0.2 (2020-06-22)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 18363)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=English_Australia.1252 LC_CTYPE=English_Australia.1252
## [3] LC_MONETARY=English_Australia.1252 LC_NUMERIC=C
## [5] LC TIME=English Australia.1252
##
## attached base packages:
## [1] stats
                 graphics grDevices utils
                                               datasets methods
                                                                    base
##
## other attached packages:
## [1] symbioteR_0.0.0.9000 vegan_2.5-6
                                                  lattice 0.20-41
## [4] permute_0.9-5
                             microbiome_1.11.0
                                                  magrittr_1.5
## [7] forcats_0.5.0
                             stringr_1.4.0
                                                  dplyr_1.0.0
## [10] purrr_0.3.4
                                                  tidyr_1.1.0
                             readr_1.3.1
## [13] tibble_3.0.1
                             ggplot2_3.3.2
                                                  tidyverse_1.3.0
## [16] qiime2R_0.99.13
                             phyloseq_1.33.0
## loaded via a namespace (and not attached):
##
     [1] Rtsne 0.15
                                 colorspace_1.4-1
                                                         ellipsis_0.3.1
                                 XVector_0.29.3
##
     [4] htmlTable_2.0.0
                                                         base64enc_0.1-3
                                 rstudioapi_0.11
##
     [7] fs_1.4.1
                                                         farver_2.0.3
##
    [10] fansi 0.4.1
                                 lubridate 1.7.9
                                                         xml2 1.3.2
## [13] codetools 0.2-16
                                 splines_4.0.2
                                                         robustbase_0.93-6
## [16] knitr 1.29
                                 ade4 1.7-15
                                                         Formula 1.2-3
                                                         cluster_2.1.0
## [19] jsonlite_1.6.1
                                 broom_0.5.6
                                 png_0.1-7
## [22] dbplyr_1.4.4
                                                         shiny_1.5.0
## [25] compiler_4.0.2
                                 httr_1.4.1
                                                         backports_1.1.6
## [28] fastmap_1.0.1
                                 assertthat_0.2.1
                                                         Matrix_1.2-18
## [31] cli_2.0.2
                                 later_1.1.0.1
                                                         acepack_1.4.1
## [34] htmltools_0.5.0
                                 tools_4.0.2
                                                         igraph_1.2.5
```

##	[37]	gtable_0.3.0	glue_1.4.1	reshape2_1.4.4
##	[40]	Rcpp_1.0.4.6	Biobase_2.49.0	cellranger_1.1.0
##	[43]	vctrs_0.3.1	Biostrings_2.57.2	rhdf5filters_1.1.0
##	[46]	multtest_2.45.0	ape_5.4	nlme_3.1-148
##	[49]	iterators_1.0.12	crosstalk_1.1.0.1	xfun_0.15
##	[52]	rvest_0.3.5	miniUI_0.1.1.1	mime_0.9
##	[55]	lifecycle_0.2.0	DEoptimR_1.0-8	zlibbioc_1.35.0
##	[58]	MASS_7.3-51.6	scales_1.1.1	hms_0.5.3
##	[61]	promises_1.1.1	parallel_4.0.2	biomformat_1.17.0
##	[64]	rhdf5_2.33.3	RColorBrewer_1.1-2	yaml_2.2.1
##	[67]	gridExtra_2.3	qpcR_1.4-1	rpart_4.1-15
##	[70]	latticeExtra_0.6-29	stringi_1.4.6	S4Vectors_0.27.12
##	[73]	foreach_1.5.0	checkmate_2.0.0	BiocGenerics_0.35.4
##	[76]	<pre>manipulateWidget_0.10.1</pre>	rlang_0.4.6	pkgconfig_2.0.3
##	[79]	rgl_0.100.54	evaluate_0.14	Rhdf5lib_1.11.2
##	[82]	labeling_0.3	htmlwidgets_1.5.1	tidyselect_1.1.0
##	[85]	plyr_1.8.6	R6_2.4.1	IRanges_2.23.10
##	[88]	generics_0.0.2	$Hmisc_4.4-0$	DBI_1.1.0
##	[91]	pillar_1.4.4	haven_2.3.1	foreign_0.8-80
##	[94]	withr_2.2.0	mgcv_1.8-31	survival_3.1-12
##	[97]	nnet_7.3-14	modelr_0.1.8	crayon_1.3.4
##	[100]	rmarkdown_2.3	jpeg_0.1-8.1	grid_4.0.2
##	[103]	readxl_1.3.1	minpack.lm_1.2-1	data.table_1.12.8
##	[106]	blob_1.2.1	webshot_0.5.2	reprex_0.3.0
##	[109]	digest_0.6.25	xtable_1.8-4	httpuv_1.5.4
##	[112]	stats4_4.0.2	munsell_0.5.0	