# Exploration of landscapes of phylogenetic trees

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# treescape: exploration of landscapes of phylogenetic trees

treescape implements new methods for the exploration and analysis of distributions of phylogenetic trees for a given set of taxa.

#### Installing treescape

To install the development version from github:

```
library(devtools)
install_github("thibautjombart/treescape")
```

The stable version can be installed from CRAN using:

```
install.packages("treescape")
```

Then, to load the package, use:

```
library("treescape")
```

#### Content overview

The main functions implemented in *treescape* are:

treescape: explore landscapes of phylogenetic trees

treescapeServer: open up an application in a web browser for an interactive exploration of the diversity in a set of trees

findGroves: identify clusters of similar trees

plotGroves: scatterplot of groups of trees

medTree: find geometric median tree(s) to summarise a group of trees

Other functions are central to the computations of distances between trees: treeVec: characterise a tree by a vector

treeDist: find the distance between two tree vectors

multiDist: find the pairwise distances of a list of trees

Distributed datasets include:

woodmiceTrees: illustrative set of 201 trees built using the neighbour-joining and bootstrapping example from the woodmice dataset in the ape documentation.

## Exploring trees with treescape

## class: pco dudi

We first load *treescape*, and packages required for graphics:

```
library("treescape")
library("ade4")
library("adegenet")
library("adegraphics")
library("ggplot2")
```

The function treescape defines typologies of phylogenetic trees using a two-step approach:

- 1. perform pairwise comparisons of trees using various (Euclidean) metrics; by default, the comparison uses the Kendall and Colijn metric (Kendall & Colijn, 2015) which is described in more detail below; other metrics rely on tips distances implemented in *adephylo* (Jombart *et al.* 2010).
- 2. use Metric Multidimensional Scaling (MDS, aka Principal Coordinates Analysis, PCoA) to summarise pairwise distances between the trees as well as possible into a few dimensions; the output of the MDS is typically visualised using scatterplots of the first few Principal Components (PCs); this step relies on the PCoA implemented in ade4 (Dray & Dufour 2007).

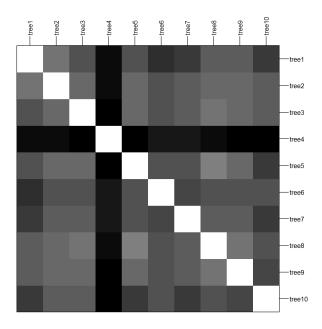
The function treescape performs both tasks, returning both the matrix of pairwise tree comparisons (\$D), and the PCoA (\$pco). This can be illustrated using randomly generated trees:

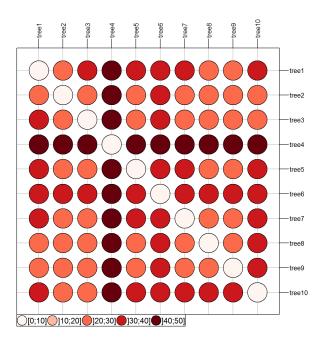
```
## generate list of trees
set.seed(1)
x \leftarrow rmtree(10, 20)
names(x) <- paste("tree", 1:10, sep = "")</pre>
## use treescape
res <- treescape(x, nf=3)
names(res)
## [1] "D"
             "pco"
res
## $D
##
          tree1 tree2 tree3 tree4 tree5 tree6 tree7 tree8 tree9
## tree2 26.00
## tree3 31.06 26.74
## tree4 42.85 42.12 44.44
## tree5 30.66 27.71 27.37 44.79
## tree6 36.50 31.18 30.18 41.81 31.59
## tree7 34.64 28.71 29.48 40.35 31.11 32.37
## tree8 28.97 26.29 24.45 43.74 23.47 30.41 29.00
## tree9 29.63 27.42 27.48 45.61 26.31 30.89 29.77 24.60
## tree10 34.87 30.00 29.44 44.97 34.06 31.05 34.41 31.54 32.59
##
## $pco
## Duality diagramm
```

```
## $call: dudi.pco(d = D, scannf = is.null(nf), nf = nf)
##
## $nf: 3 axis-components saved
## $rank: 9
## eigen values: 142.1 76.52 62.69 49.88 41.07 ...
   vector length mode
                          content
## 1 $cw
                  numeric column weights
## 2 $1w
                  numeric row weights
            10
## 3 $eig
            9
                  numeric eigen values
##
##
     data.frame nrow ncol content
## 1 $tab
                         modified array
               10
                     9
## 2 $li
                10
                        row coordinates
                     3
## 3 $11
               10
                   3
                       row normed scores
## 4 $co
               9
                     3
                         column coordinates
## 5 $c1
                9
                     3
                          column normed scores
## other elements: NULL
```

Pairwise distances can be visualised using adegraphics:

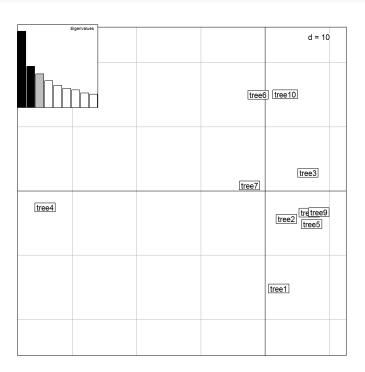
```
## table.image
table.image(res$D, nclass=30)
```





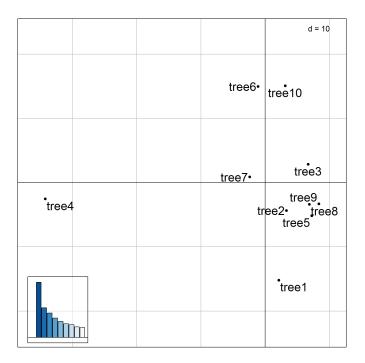
The best representation of these distances in a 2-dimensional space is given by the first 2 PCs of the MDS. These can be visualised using *adegraphics*'s function scatter:

## scatter(res\$pco)



Alternatively, the function plotGroves can be used:

#### plotGroves(res\$pco, lab.show=TRUE, lab.cex=1.5)



The functionality of treecsape can be further illustrated using *ape*'s dataset *woodmouse*, from which we built the 201 trees supplied in woodmiceTrees using the neighbour-joining and bootstrapping example from the *ape* documentation.

```
data(woodmiceTrees)
wm.res <- treescape(woodmiceTrees,nf=3)</pre>
## this is the PCoA / MDS:
wm.res$pco
## Duality diagramm
## class: pco dudi
## $call: dudi.pco(d = D, scannf = is.null(nf), nf = nf)
##
## $nf: 3 axis-components saved
## $rank: 54
## eigen values: 32.69 24.41 6.952 6.348 4.363 ...
    vector length mode
##
                          content
## 1 $cw
            54
                  numeric column weights
## 2 $1w
            201
                  numeric row weights
## 3 $eig
           54
                  numeric eigen values
##
##
    data.frame nrow ncol content
## 1 $tab
               201 54 modified array
## 2 $li
               201 3
                         row coordinates
## 3 $11
               201 3
                         row normed scores
## 4 $co
               54 3
                         column coordinates
## 5 $c1
                         column normed scores
## other elements: NULL
```

```
## PCs are stored in:
head(wm.res$pco$li)
```

```
## A1 A2 A3

## 1 -0.9949 -1.363 -0.7918

## 2 -0.6137 -1.014 -0.6798

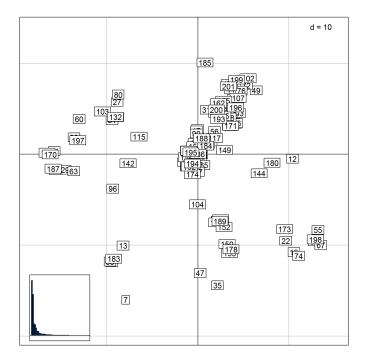
## 3 2.6667 4.219 -2.9293

## 4 -13.6081 1.854 1.0947

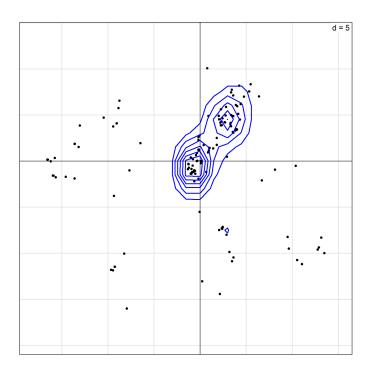
## 5 2.1980 4.176 -3.1960

## 6 3.6013 4.865 2.9853
```

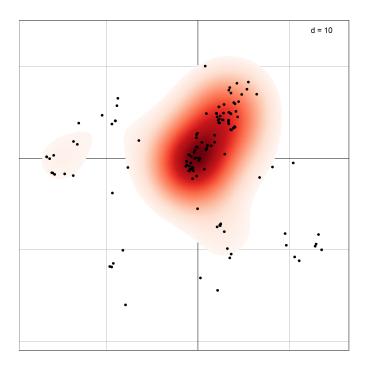
```
## plot results
plotGroves(wm.res$pco, lab.show=TRUE, lab.optim=FALSE)
```



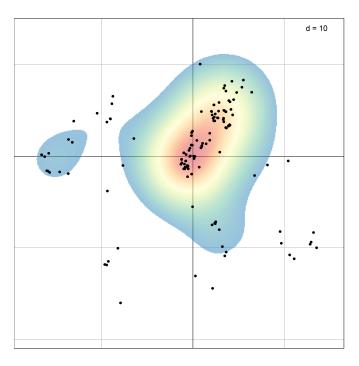
```
## visualising density of points
s.kde2d(wm.res$pco$li)
```



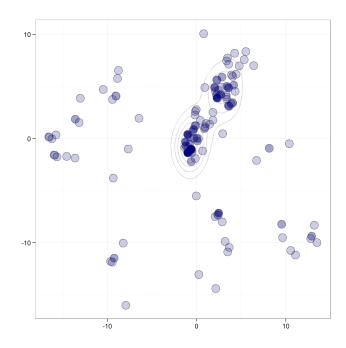
```
## alternative visualisation
s.density(wm.res$pco$li, col=redpal(100), bandwidth=3)
```



```
## same, other palette
s.density(wm.res$pco$li, col=rev(transp(spectral(100),.5)), bandwidth=3)
```



```
## alternative using ggplot2
woodmiceplot <- ggplot(wm.res$pco$li, aes(x=A1, y=A2)) # create plot
woodmiceplot + geom_density2d(colour="gray80") + # contour lines
geom_point(size=6, shape=1, colour="gray50") + # grey edges
geom_point(size=6, alpha=0.2, colour="navy") + # transparent blue points
xlab("") + ylab("") + theme_bw(base_family="") # remove axis labels and grey background</pre>
```



Note that alternatively, the function multiDist simply performs the pairwise comparison of trees and outputs a distance matrix. This function may be preferable for large datasets, and when principal co-ordinate analysis is not required. It includes an option to save memory at the expense of computation time.

# Identifying clusters of trees

Once a typology of trees has been derived using the approach described above, one may want to formally identify clusters of similar trees. One simple approach is:

- 1. select a few first PCs of the MDS (retaining signal but getting rid of random noise)
- 2. derive pairwise Euclidean distances between trees based on these PCs
- 3. use hierarchical clustering to obtain a dendrogram of these trees
- 4. cut the dendrogram to obtain clusters

In treescape, the function findGroves implements this approach, offering various clustering options (see ?findGroves). Here we supply the function with our treescape output wm.res since we have already calculated it, but it is also possible to skip the steps above and directly supply findGroves with a multiPhylo list of trees.

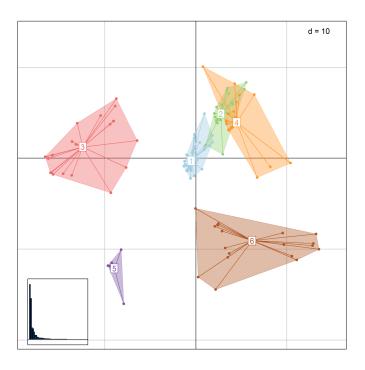
```
wm.groves <- findGroves(wm.res, nclust=6)
names(wm.groves)</pre>
```

#### ## [1] "groups" "treescape"

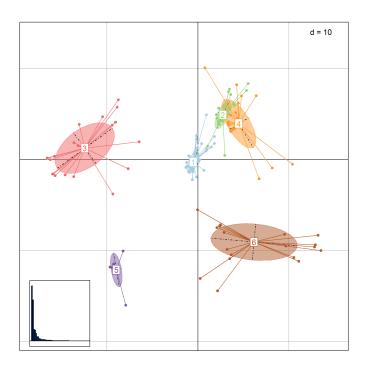
Note that when the number of clusters (nclust) is not provided, the function will display a dendrogram and ask for a cut-off height.

The results can be plotted directly using plotGroves (see ?plotGroves for options):

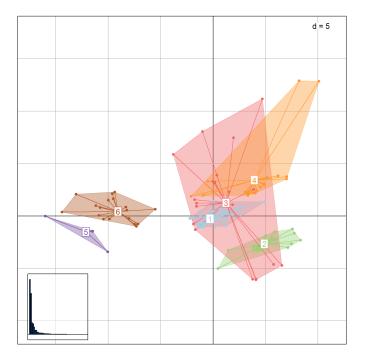
```
## basic plot
plotGroves(wm.groves)
```



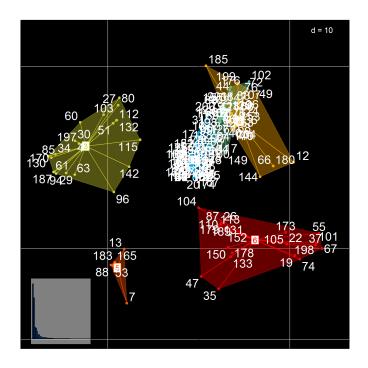
## alternative with inertia ellipses
plotGroves(wm.groves, type="ellipse")



## plot axes 2-3
plotGroves(wm.groves, xax=2, yax=3)



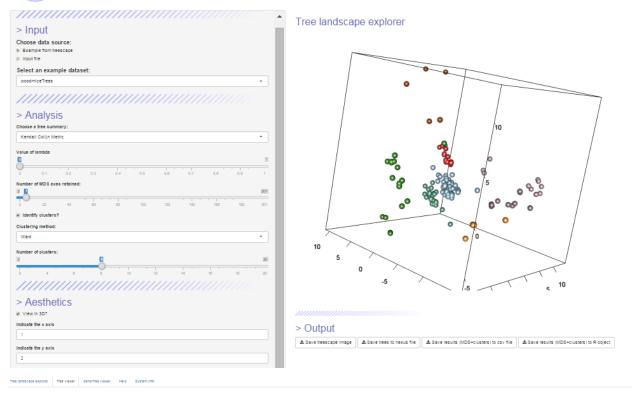
## customize graphics
plotGroves(wm.groves, bg="black", col.pal=lightseasun, lab.show=TRUE, lab.col="white", lab.cex=1.5)



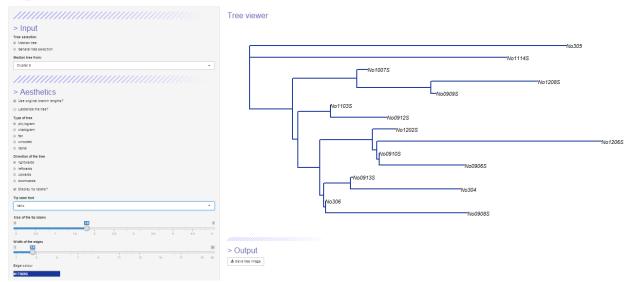
# treescapeServer: a web application for treescape

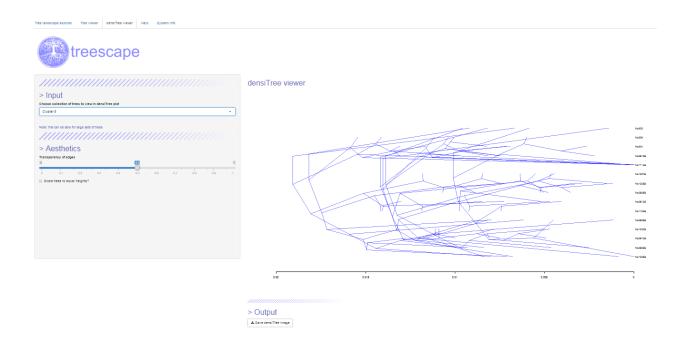
The functionalities of treescape are also available via a user-friendly web interface, running locally on the default web browser. It can be started by simply typing treescapeServer(). The interface allows you to import trees and run treescape to view and explore the tree space in 2 or 3 dimensions. It is then straightforward to analyse the tree space by varying lambda, looking for clusters using findGroves and saving results in various formats. Individual trees can be easily viewed including median trees per cluster, and collections of trees can be seen together using densiTree from the package phangorn. It is fully documented in the *help* tab.





# treescape



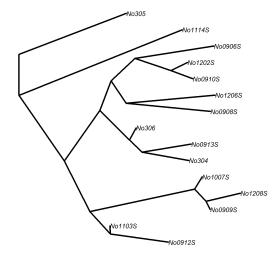


# Finding median trees

When a set of trees have very similar structures, it makes sense to summarize them into a single 'consensus' tree. In treescape, this is achieved by finding the *median tree* for a set of trees according to the Kendall and Colijn metric. That is, we find the tree which is closest to the centre of the set of trees in the tree landscape defined in treescape. This procedure is implemented by the function medTree:

```
## get first median tree
tre <- medTree(woodmiceTrees)$trees[[1]]

## plot tree
plot(tre,type="cladogram",edge.width=3, cex=0.8)</pre>
```



However, a more complete and accurate summary of the data can be given by finding a summary tree from each cluster. This is achieved using the <code>groups</code> argument of <code>medTree</code>:

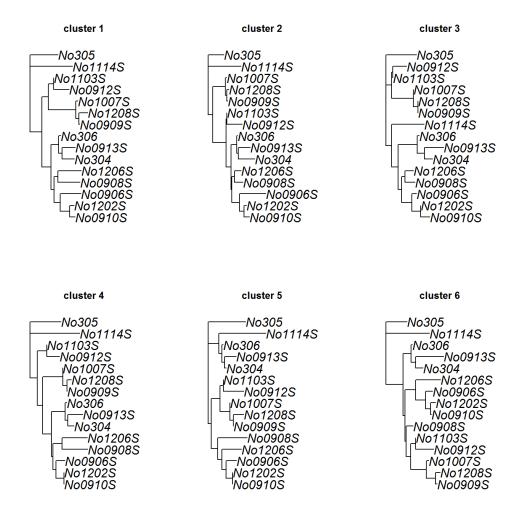
```
## find median trees for the 6 clusters identified earlier:
res <- medTree(woodmiceTrees, wm.groves$groups)

## there is one output per cluster
names(res)

## [1] "1" "2" "3" "4" "5" "6"

## get the first median of each
med.trees <- lapply(res, function(e) ladderize(e$trees[[1]]))

## plot trees
par(mfrow=c(2,3))
for(i in 1:length(med.trees)) plot(med.trees[[i]], main=paste("cluster",i),cex=1.5)</pre>
```



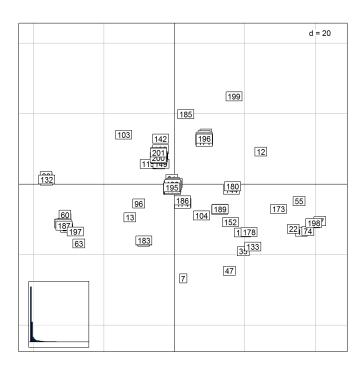
These trees exhibit a number of topological differences, e.g. in the placement of the (1007S,1208S,0909S) clade. Performing this analysis enables the detection of distinct representative trees supported by data.

Note that we supplied the function medTree with the multiPhylo list of trees. A more computationally efficient process (at the expense of using more memory) is to use the option return.tree.vectors in the initial treescape call, and then supply these vectors directly to medTree. In this case, the tree indices are returned by medTree but the trees are not (since they were not supplied).

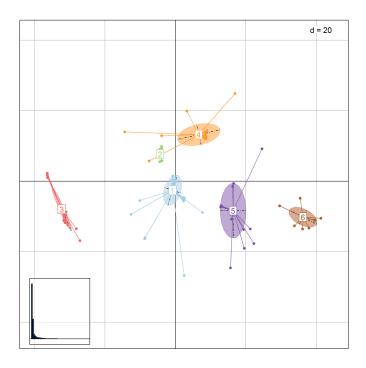
## Emphasising the placement of certain tips or clades

In some analyses it may be informative to emphasise the placement of particular tips or clades within a set of trees. This can be particularly useful in large trees where the study is focused on a smaller clade. Priority can be given to a list of tips using the argument emphasise.tips, whose corresponding values in the vector comparison will be given a weight of emphasise.weight times the others (the default is 2, i.e. twice the weight).

For example, if we wanted to emphasise where the woodmice trees agree and disagree on the placement of the (1007S,1208S,0909S) clade, we can simply emphasise that clade as follows:



It can be seen from the scale of the plot and the density of clustering that the trees are now separated into more distinct clusters.



Conversely, where the structure of a particular clade is not of interest (for example, lineages within an outgroup which was only included for rooting purposes), those tips can be given a weight less than 1 so as to give them less emphasis in the comparison. We note that although it is possible to give tips a weighting of 0, we advise caution with this as the underlying function will no longer be guaranteed to be a metric. That is, a distance of 0 between two trees will no longer necessarily imply that the trees are identical. In most cases it would be wiser to assign a very small weighting to tips which are not of interest.

#### Method: characterising a tree by a vector

Kendall and Colijn proposed a metric for comparing rooted phylogenetic trees. Each tree is characterised by a vector which notes the placement of the most recent common ancestor (MRCA) of each pair of tips. Specifically, it records the distance between the MRCA of a pair of tips (i, j) and the root in two ways: the number of edges  $m_{i,j}$ , and the path length  $M_{i,j}$ . It also records the length  $p_i$  of each 'pendant' edge between a tip i and its immediate ancestor. This procedure results in two vectors for a tree T:

$$m(T) = (m_{1,2}, m_{1,3}, \dots, m_{k-1,k}, \underbrace{1, \dots, 1}_{k \text{ times}})$$

and

$$M(T) = (M_{1,2}, M_{1,3}, \dots, M_{k-1,k}, p_1, \dots, p_k).$$

In m(T) we record the pendant lengths as 1, as each tip is 1 step from its immediate ancestor. We combine m and M with a parameter lambda between zero and one to weight the contribution of branch lengths, characterising each tree with a vector

$$v_{\lambda}(T) = (1 - \lambda)m(T) + \lambda M(T).$$

This is implemented as the function treeVec. For example,

```
## generate a random tree:
tree <- rtree(6)</pre>
## topological vector of mrca distances from root:
treeVec(tree)
    [1] 1 2 1 0 1 1 3 0 2 1 0 1 0 2 0 1 1 1 1 1 1
## vector of mrca distances from root when lambda=0.5:
treeVec(tree, 0.5)
## [1] 0.7196 1.3042 0.7196 0.0000 0.7196 0.7196 2.4325 0.0000 1.6221 0.7196
## [11] 0.0000 0.7196 0.0000 1.6221 0.0000 0.8311 0.9432 0.8928 0.6913 0.7264
## [21] 0.7287
## vector of mrca distances as a function of lambda:
vecAsFunction <- treeVec(tree,return.lambda.function=TRUE)</pre>
## evaluate the vector at lambda=0.5:
vecAsFunction(0.5)
## [1] 0.7196 1.3042 0.7196 0.0000 0.7196 0.7196 2.4325 0.0000 1.6221 0.7196
## [11] 0.0000 0.7196 0.0000 1.6221 0.0000 0.8311 0.9432 0.8928 0.6913 0.7264
## [21] 0.7287
```

The metric – the distance between two trees – is the Euclidean distance between these vectors:

$$d_{\lambda}(T_a, T_b) = ||v_{\lambda}(T_a) - v_{\lambda}(T_b)||.$$

This can be found using treeDist:

```
## generate random trees
tree_a <- rtree(6)
tree_b <- rtree(6)

## topological (lambda=0) distance:
treeDist(tree_a, tree_b)</pre>
```

```
## [1] 6.083
```

```
## branch-length focused (lambda=1) distance:
treeDist(tree_a,tree_b,1)
```

## [1] 3.186

#### References

- Dray S & Dufour AB (2007): The ade4 package: implementing the duality diagram for ecologists. Journal of Statistical Software 22(4): 1-20.
- Jombart R, Balloux F & Dray S (2010) adephylo: new tools for investigating the phylogenetic signal in biological traits. Bioinformatics 26: 1907-1909. Doi: 10.1093/bioinformatics/btq292
- Kendall M & Colijn C (Preprint 2015) A tree metric using structure and length to capture distinct phylogenetic signals. arXiv 1507.05211

# Authors / Contributors

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