

MS02star annotation, Human PD-1

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```
## read the original network
network <- read.csv("../Data/human.pin.csv", header=T, stringsAsFactors=F)
geneA <- network$geneA
geneB <- network$geneB

## the GO terms for biological processes (BP)
## In this slim, each GO terms was shared by at least 20 genes
GOcategory.file <- read.csv("../Data/human.bp.term.csv",header=TRUE, stringsAsFactors=F)
bp.go.cat <- GOcategory.file$GO.term
bp.dim <- length(bp.go.cat)

# Gene-GO term file
# Note in human PIN the standard gene names have been used
GOterm.file <- read.csv("../Data/human.bp.gene.term.csv", header=T, stringsAsFactors=F)
bp.GO.gene <- GOterm.file$gene #it should be changed to System for yeast pin
bp.GO.term <- GOterm.file$GO.term

#gene list, only a single gene is specified here for annotation
gene.list <- c("PDCD1")

vec <- numeric(length=bp.dim)

for (i in 1:length(gene.list)) {
  orf <- as.character(gene.list[i])
  intA <- geneB[which(geneA %in% orf)]
  for (j in 1:length(intA)) {
    bpA <- bp.GO.term[which(bp.GO.gene %in% intA[j])]
    for (k in 1:length(bpA)) {
      na <- which(bp.go.cat %in% bpA[k])
      vec[na] <- vec[na] + 1
    }
  }

  intB <- geneA[which(geneB %in% orf)]
  for (s in 1:length(intB)) {
    bpB <- bp.GO.term[which(bp.GO.gene %in% intB[s])]
    for (t in 1:length(bpB)) {
      nb <- which(bp.go.cat %in% bpB[t])
      vec[nb] <- vec[nb] + 1
    }
  }
}

write.table(vec, file="human.pdc1.bp.txt", col.names=F, row.names=F, quote=F)

# Now the ms02star permutations
for (p in 1:100) {
```

```

permutation.file <- paste("../ms02star/human/", "ms02.", p, ".csv", sep="")
permutation <- read.csv(permutation.file, header=T, stringsAsFactors = F)
geneA <- permutation$id1
geneB <- permutation$id2

vecp <- numeric(length = bp.dim)
for (i in 1:length(gene.list)) {
  orf <- as.character(gene.list[i])
  intA <- geneB[which(geneA %in% orf)]
  for (j in 1:length(intA)) {
    bpA <- bp.GO.term[which(bp.GO.gene %in% intA[j])]
    for (k in 1:length(bpA)) {
      na <- which(bp.go.cat %in% bpA[k])
      vecp[na] <- vecp[na] + 1
    }
  }

  intB <- geneA[which(geneB %in% orf)]
  for (s in 1:length(intB)) {
    bpB <- bp.GO.term[which(bp.GO.gene %in% intB[s])]
    for (t in 1:length(bpB)) {
      nb <- which(bp.go.cat %in% bpB[t])
      vecp[nb] <- vecp[nb] + 1
    }
  }
}

output <- paste("ms02.human", "/", "pdcd1", "/", "ms02.", p, ".bp.matrix.csv", sep="")

write.table(vecp, file = output, col.names=F, row.names=F, quote=F)
}

```

```

library("microbenchmark")
library("matrixStats")

conn.dim <- 1

hspin <- matrix(as.numeric(unlist(read.table("human.pdcd1.bp.txt", header=F, sep=","))), nrow=bp.dim, ncol=conn.dim)
obs <- c(hspin)

perm <- c()
for (i in 1:100) {
  name <- paste("ms02.human", "/", "pdcd1", "/", "ms02.", i, ".bp.matrix.csv", sep="")
  mat <- matrix(as.numeric(unlist(read.table(name, header=F, sep=","))), nrow=bp.dim, ncol=conn.dim)
  perm <- rbind(perm, c(mat))
}

mean <- colMeans(perm)
std <- colSds(perm)

zscore <- round((obs - mean)/std, 3)

z <- matrix(zscore, nrow=bp.dim, ncol=conn.dim)

```

```
write.table(z, file="human.pdcd1,bp.z.csv", sep=",", row.names=F, col.names=F, quote=F)
```

```
library('gplots')
```

```
##
## Attaching package: 'gplots'
## The following object is masked from 'package:stats':
##
##      lowess
```

```
library('GO.db')
```

```
## Loading required package: AnnotationDbi
## Loading required package: stats4
## Loading required package: BiocGenerics
## Loading required package: parallel
##
## Attaching package: 'BiocGenerics'
## The following objects are masked from 'package:parallel':
##
##      clusterApply, clusterApplyLB, clusterCall, clusterEvalQ,
##      clusterExport, clusterMap, parApply, parCapply, parLapply,
##      parLapplyLB, parRapply, parSapply, parSapplyLB
## The following objects are masked from 'package:stats':
##
##      IQR, mad, sd, var, xtabs
## The following objects are masked from 'package:base':
##
##      anyDuplicated, append, as.data.frame, basename, cbind,
##      colMeans, colnames, colSums, dirname, do.call, duplicated,
##      eval, evalq, Filter, Find, get, grep, grepl, intersect,
##      is.unsorted, lapply, lengths, Map, mapply, match, mget, order,
##      paste, pmax, pmax.int, pmin, pmin.int, Position, rank, rbind,
##      Reduce, rowMeans, rownames, rowSums, sapply, setdiff, sort,
##      table, tapply, union, unique, unsplit, which, which.max,
##      which.min
## Loading required package: Biobase
## Welcome to Bioconductor
##
##      Vignettes contain introductory material; view with
##      'browseVignettes()'. To cite Bioconductor, see
##      'citation("Biobase")', and for packages 'citation("pkgname")'.
##
## Attaching package: 'Biobase'
## The following objects are masked from 'package:matrixStats':
##
##      anyMissing, rowMedians
## Loading required package: IRanges
```

```

## Warning: package 'IRanges' was built under R version 3.5.1
## Loading required package: S4Vectors
##
## Attaching package: 'S4Vectors'
## The following object is masked from 'package:gplots':
##
##     space
## The following object is masked from 'package:base':
##
##     expand.grid
##
order <- order(z)
bp.go.cat <- bp.go.cat[order]
z <- z[order]

z <- t(z)
colnames(z) <- bp.go.cat

enriched.list <- bp.go.cat[which(z >= 3)]
enriched <- c("GO.ID", "GO.Term", "Z-score")
for (i in 1:length(enriched.list)) {
  term <- Term(GOID(as.character(enriched.list[i])))
  enriched <- rbind(enriched, c(as.character(enriched.list[i]),term,
                                z[which(bp.go.cat %in% enriched.list[i])]))
}

print(enriched)

##
## enriched "GO.ID"
##          "GO:0001932"
##          "GO:0043407"
##          "GO:0048008"
##          "GO:0006958"
##          "GO:0002244"
##          "GO:0006911"
##          "GO:0009755"
##          "GO:0042267"
##          "GO:0050829"
##          "GO:0070372"
##          "GO:0006470"
##          "GO:0021549"
##          "GO:0007163"
##          "GO:0034332"
##          "GO:0046627"
##          "GO:0071222"
##          "GO:0006910"
##          "GO:0048709"
##          "GO:0048873"
##          "GO:0035335"
##          "GO:0050731"

```

```

##      "GO:0051260"
##      "GO:0006641"
##      "GO:0032753"
##      "GO:0045879"
##      "GO:0050871"
##      "GO:0046854"
##      "GO:0001816"
##      "GO:0046676"
##      "GO:0048839"
##      "GO:0051209"
##      "GO:0051924"
##      "GO:0060325"
##      "GO:0050900"
##      "GO:0030168"
##      "GO:0071345"
##      "GO:0006882"
##      "GO:0042110"
##      "GO:0050890"
##      "GO:0007169"
##      "GO:0050853"
##      "GO:0007166"
##      "GO:0032729"
##      "GO:0032689"
##      "GO:0050690"
##      "GO:0042130"
##      "GO:0050776"
##      "GO:0031295"
##      "GO:0006955"
##      "GO:0042102"
##      "GO:0030217"
##      "GO:0002250"
##      "GO:0019886"
##      "GO:0060333"
##      "GO:0050852"
##      "GO:0070207"
##      GO:0001932
## enriched "GO.Term"
##      "regulation of protein phosphorylation"
##      "negative regulation of MAP kinase activity"
##      "platelet-derived growth factor receptor signaling pathway"
##      "complement activation, classical pathway"
##      "hematopoietic progenitor cell differentiation"
##      "phagocytosis, engulfment"
##      "hormone-mediated signaling pathway"
##      "natural killer cell mediated cytotoxicity"
##      "defense response to Gram-negative bacterium"
##      "regulation of ERK1 and ERK2 cascade"
##      "protein dephosphorylation"
##      "cerebellum development"
##      "establishment or maintenance of cell polarity"
##      "adherens junction organization"
##      "negative regulation of insulin receptor signaling pathway"
##      "cellular response to lipopolysaccharide"
##      "phagocytosis, recognition"

```

```

## "oligodendrocyte differentiation"
## "homeostasis of number of cells within a tissue"
## "peptidyl-tyrosine dephosphorylation"
## "positive regulation of peptidyl-tyrosine phosphorylation"
## "protein homooligomerization"
## "triglyceride metabolic process"
## "positive regulation of interleukin-4 production"
## "negative regulation of smoothened signaling pathway"
## "positive regulation of B cell activation"
## "phosphatidylinositol phosphorylation"
## "cytokine production"
## "negative regulation of insulin secretion"
## "inner ear development"
## "release of sequestered calcium ion into cytosol"
## "regulation of calcium ion transport"
## "face morphogenesis"
## "leukocyte migration"
## "platelet activation"
## "cellular response to cytokine stimulus"
## "cellular zinc ion homeostasis"
## "T cell activation"
## "cognition"
## "transmembrane receptor protein tyrosine kinase signaling pathway"
## "B cell receptor signaling pathway"
## "cell surface receptor signaling pathway"
## "positive regulation of interferon-gamma production"
## "negative regulation of interferon-gamma production"
## "regulation of defense response to virus by virus"
## "negative regulation of T cell proliferation"
## "regulation of immune response"
## "T cell costimulation"
## "immune response"
## "positive regulation of T cell proliferation"
## "T cell differentiation"
## "adaptive immune response"
## "antigen processing and presentation of exogenous peptide antigen via MHC class II"
## "interferon-gamma-mediated signaling pathway"
## "T cell receptor signaling pathway"
## "protein homotrimerization"
##
## enriched "Z-score"
## "3.164"
## "3.164"
## "3.164"
## "3.172"
## "3.374"
## "3.374"
## "3.374"
## "3.374"
## "3.374"
## "3.374"
## "3.374"
## "3.384"
## "3.627"
## "3.938"

```

```
##      "3.938"
##      "3.938"
##      "3.967"
##      "4.337"
##      "4.337"
##      "4.337"
##      "4.357"
##      "4.477"
##      "4.607"
##      "4.874"
##      "4.874"
##      "4.874"
##      "4.874"
##      "5.229"
##      "5.658"
##      "5.658"
##      "5.658"
##      "5.658"
##      "5.658"
##      "5.658"
##      "5.823"
##      "6.438"
##      "6.641"
##      "6.965"
##      "9.496"
##      "9.9"
##      "10.117"
##      "10.709"
##      "11.368"
##      "11.426"
##      "11.49"
##      "12.187"
##      "13.468"
##      "17.663"
##      "18.216"
##      "18.712"
##      "18.73"
##      "20.697"
##      "20.74"
##      "24.028"
##      "25.793"
##      "40.757"
##      "Inf"
```

```
write.table(enriched, file="human.pdcd1.bp.enriched.csv", row.names=F, col.names=F, quote=F, sep="\t")
```

```
###No suppressed terms have been found
#sup.list <- bp.go.cat[which(z <= -3)]
#sup <- c("GO.ID", "GO.Term", "Z-score")
#for (i in 1:length(sup.list)) {
#  term <- Term(GOID(as.character(sup.list[i])))
#  sup <- rbind(sup, c(as.character(sup.list[i]),term,
#                      z[which(bp.go.cat %in% sup.list[i])]))
#}
```

```

#print(sup)

#write.table(sup, file="human.pdcd1.bp.suppressed.csv", row.names=F, col.names=F, quote=F, sep="\t")
###

#Note that there may be "inf" Z-scores owing to lack of sampling (i.e., zero in standard deviations)
#We can also extract the GO-terms for the gene for comparison

terms <- bp.GO.term[which(bp.GO.gene %in% orf)]
extract <- c("GO.ID", "GO.Term")
for (i in 1:length(terms)) {
  go.id <- as.character(terms[i])
  go.con <- Term(GOID(go.id))
  extract <- rbind(extract, c(go.id, go.con))
}

print(extract)

##                                GO:0006915
## extract "GO.ID"                "GO.Term"
##          "GO:0006915" "apoptotic process"
##          "GO:0006959" "humoral immune response"
##          "GO:0007275" "multicellular organism development"
##          "GO:0031295" "T cell costimulation"
##          "GO:0043066" "negative regulation of apoptotic process"
write.table(extract, file="human.pdcd1.bp.original.csv", row.names=F, col.names=F, quote=F, sep="\t")

```