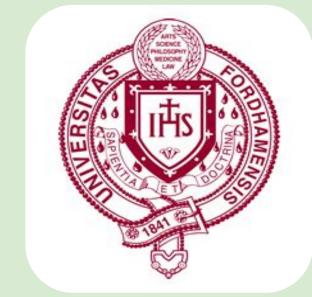
Developing a Streamlined Data Pipeline for Quantifying Ionic Liquid Effects on Locomotive Activity



Isaia Caluza, Jazmine Hudson, Miguel Pereira, Dr. Mark Botton, Dr. Alma Rodenas, Dr. Alma Rodenas-Ruano

Results

Lights OFF

emim-oAc: 20-1 (C) 25 versus 35 dpe

Lights ON

Background

This project aims to investigate how different types and concentrations of ionic liquids—emim-oAc, emim-dmp, and omim-Cl—affect the locomotion of horseshoe crab larvae across two developmental stages, 20-1 and 20-2. Animals were exposed to these chemicals for varying durations and tested using a behavioral assay with alternating **light and dark** visual stimuli to probe stimulus-responsive movement.

The goal of this project, from a computational neuroscience perspective, was to develop a **streamlined and efficient approach** to quantify and visualize the movement we consistently observe in video recordings and path traces. We sought to establish whether differences in locomotion across groups were **statistically meaningful**, and to build a reproducible framework that translates raw tracking data into interpretable movement metrics.

Methods

Behavioral Experiments (Video

and Traces of Horseshoe Crabs)

Euclidean

distance

 $(\mathbf{d}(\mathbf{p},\mathbf{q})^2 = (q_1 - p_1)^2 + (q_2 - p_2)^2$

Euclidean distance

Cumulative distance

over time

25dpe 0.4 25dpe 0.3 (SO.1 Lights ON Lights OFF --- 800 mg - Mean 35dpe 1600 mg - Mean 0.3 0.1 $\times 10^7$ Normalized Time (microsecond) emim-dmp: 20-1 (C) 25 versus 35 dpe Lights OFF Lights ON 25dpe 25dpe 800 mg - Mean 1600 mg - Mean Collecting raw 0.3 data (x, y coordinates) (S_{0.1} Lights OFF Lights ON 0 mg - Mean 35dpe 35dpe 800 mg - Mean 1600 mg - Mean 1600 mg - Mean 0.1 $\times 10^7$ $\times 10^7$ Normalized Time (microsecond) Cleaning data omim-Cl: 20-1 C vs. DC (26-35 dpe) (removing missing values) Lights OFF (Chorionated) Lights ON (Chorionated) 0 mg - Mean — 0 mg - Mean 400 mg - Mean 400 mg - Mean 0.4 800 mg - Mean 800 mg - Mean 0.3 (SO.: Velocity plot Lights OFF (Dechorionated) Lights ON (Dechorionated) - 0 mg - Mean 200 mg - Mean 200 mg - Mean 400 mg - Mean 400 mg - Mean OFF OFF

Normalized Time (microsecond)

Discussion

Mapping data automation

This project centered on developing a streamlined data pipeline to process behavioral tracking data from Zebralab. Due to inconsistent experimental setups—varying in concentration levels, ionic liquid types, and number of animals per dish—we implemented a flexible group-mapping system to automate assignment of animals to their respective conditions based on a custom file naming system.

Data validation via cross-reference approach

Raw x-y coordinates were preprocessed, and velocity was computed using Euclidean distance, followed by Gaussian smoothing and time normalization to align animals across light stimulus bins (**lights ON** vs. **lights OFF**). We validated movement traces by cross-referencing velocity graphs with complete path images and raw video footage, addressing false positives from the tracking system.

Statistical analysis

 $\times 10^7$

To support quantitative comparisons, we automated one-way ANOVA and post hoc Tukey HSD tests, applied at fixed time bins (e.g., 10 seconds). Significant comparisons to the control were visualized using color-coded asterisks directly on the velocity plots. This approach ensures rigorous, reproducible analysis while communicating findings effectively to non-technical audiences.

Acknowledgements

Mary G. Hamilton Natural Sciences Summer Research Award

Natural Science Department at FCLC

References

Flieger, J., & Flieger, M. (2020). Ionic Liquids Toxicity—Benefits and Threats. International Journal of Molecular Sciences, 21(17), Article17. https://doi.org/10.3390/ijms21176267

Botton, M. L., Tankersley, R. A., & Loveland, R. E. (2010). Developmental ecology of the American horseshoe crab Limulus polyphemus. Current Zoology, 56(5), 550–562. https://doi.org/10.1093/czoolo/56.5.550

Botton, M.L., Itow, T. (2009). The Effects of Water Quality on Horseshoe Crab Embryos and Larvae. In: Tanacredi, J., Botton, M., Smith, D. (eds) Biology and Conservation of Horseshoe Crabs. Springer, Boston, MA. https://doi.org/10.1007/978-0-387-89959-6_27