



# **ACEMFS FUT Minna**

# **Bioinformatics Workshop**

## **Introduction to Bioinformatics Training**

**Itunuoluwa Isewon PhD**  
Covenant University



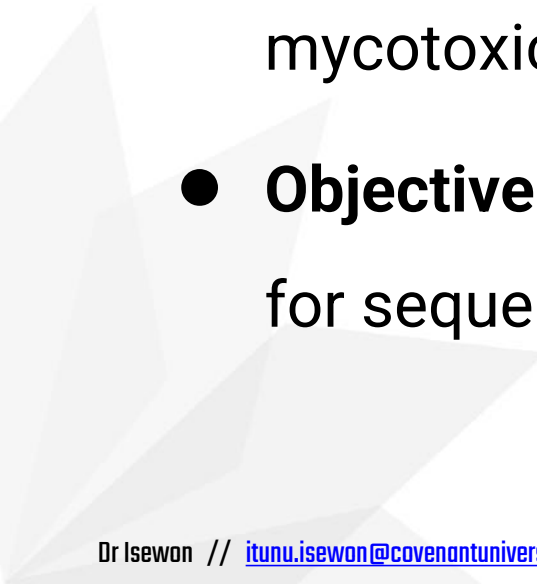
# CAPIC-ACE

- Covenant Applied Informatics and Communication Africa Centre of Excellence (CAPIC-ACE) domiciled at Covenant University
- Mission: dedicated to building indigenous capacity through training in Computer Science, Bioinformatics, Chemistry, Biology and ICT to reduce the disease burden in Africa.
- Role in this workshop: Providing support, guidance, and networking for participants.



# Workshop Overview



- 3-day hands-on training
  - **Focus:** Bioinformatics for mycology and mycotoxicology research
  - **Objective:** Equip participants with foundational skills for sequence analysis and structural bioinformatics.
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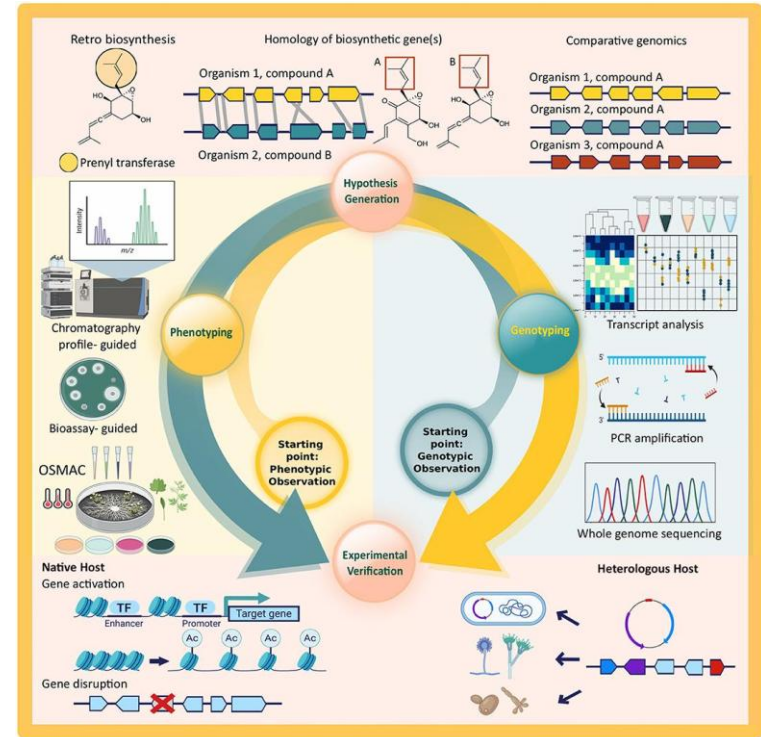


[https://bit.ly/ACE\\_IBT25\\_PreCourse](https://bit.ly/ACE_IBT25_PreCourse)

## **Pre Workshop Survey**

# What is Bioinformatics?

- **Definition:** Application of computational and statistical techniques to understand biological data.
- **Importance:** Handles large-scale genomic and molecular data.
- **Example:** Analyzing fungal genomes for mycotoxin biosynthesis genes.



<https://doi.org/10.3389/fbioe.2021.649906>

# Bioinformatics in Mycology



- Bioinformatics as a tool to study fungi
- Applications in fungal identification and genome analysis
- Detecting pathogenic and industrially important fungi



Article | Published: 24 November 2021

# ***Aspergillus fumigatus* pan-genome analysis identifies genetic variants associated with human infection**

[Amelia E. Barber](#), [Tongta Sae-Ong](#), [Kang Kang](#), [Bastian Seelbinder](#), [Jun Li](#), [Grit Walther](#), [Gianni Panagiotou](#)

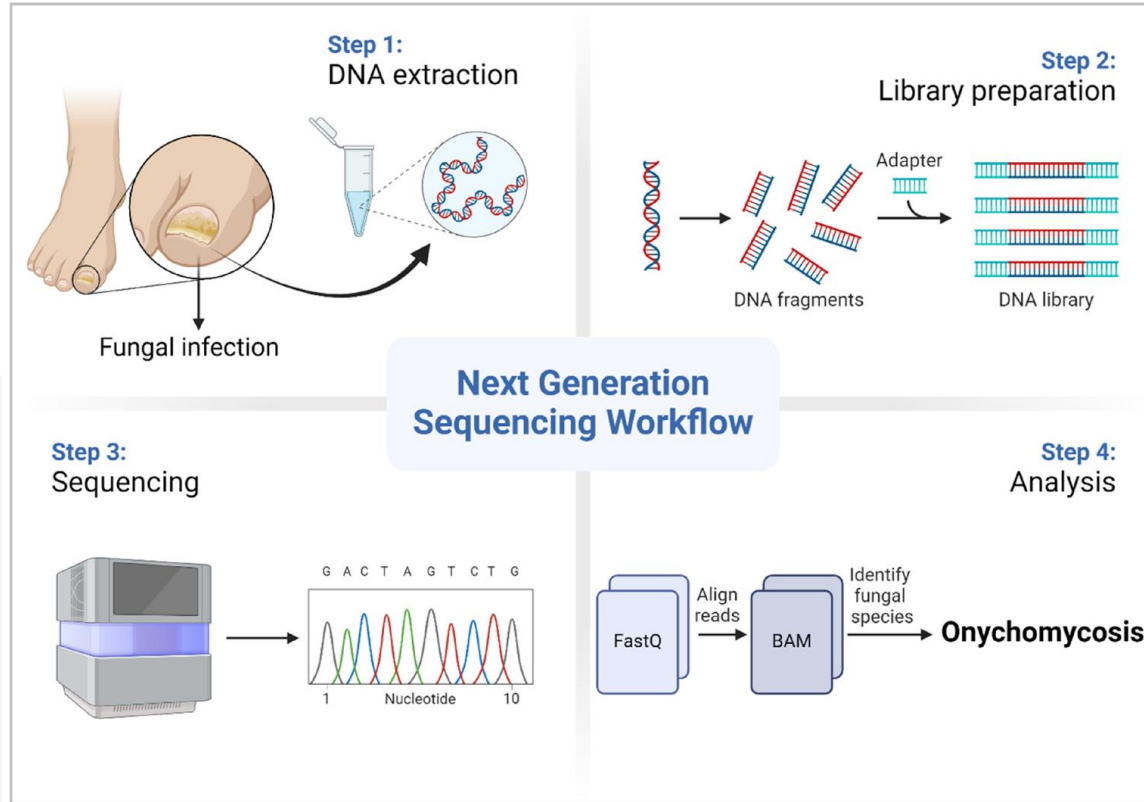
✉ & [Oliver Kurzai](#) ✉

[Nature Microbiology](#) **6**, 1526–1536 (2021) | [Cite this article](#)

**9711** Accesses | **100** Citations | **240** Altmetric | [Metrics](#)

<https://doi.org/10.1038/s41564-021-00993-x>

# Fungal Genome Sequencing

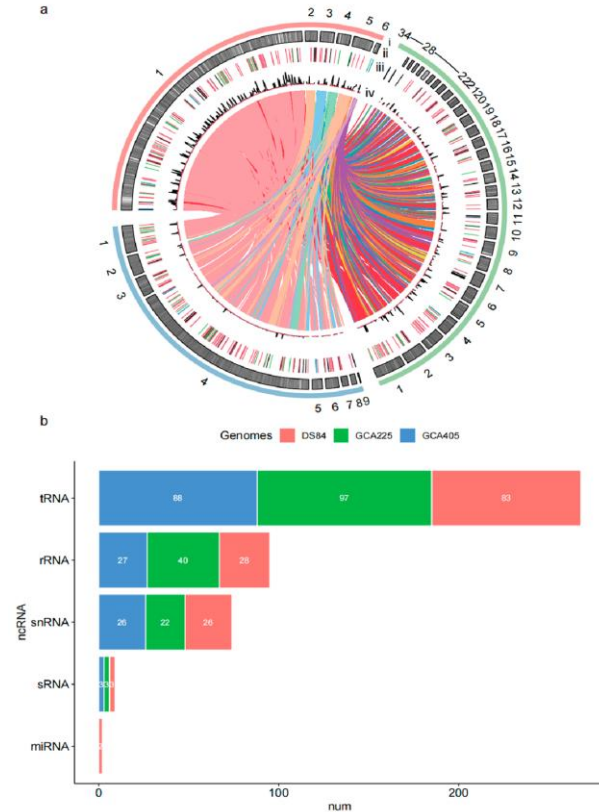


<https://doi.org/10.3390/diagnostics14151664>



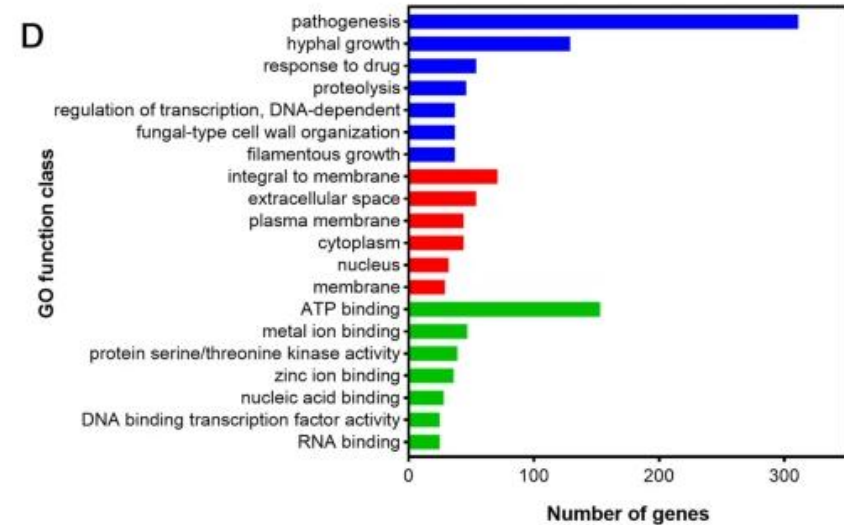
# Comparative Genomics in Fungi

- Comparing genomes across strains/species
- Identification of conserved and variable regions
- Detecting genes involved in toxin biosynthesis
- Case study: *Aspergillus flavus* vs. *Aspergillus parasiticus*



# Functional Annotation of Fungal Genes

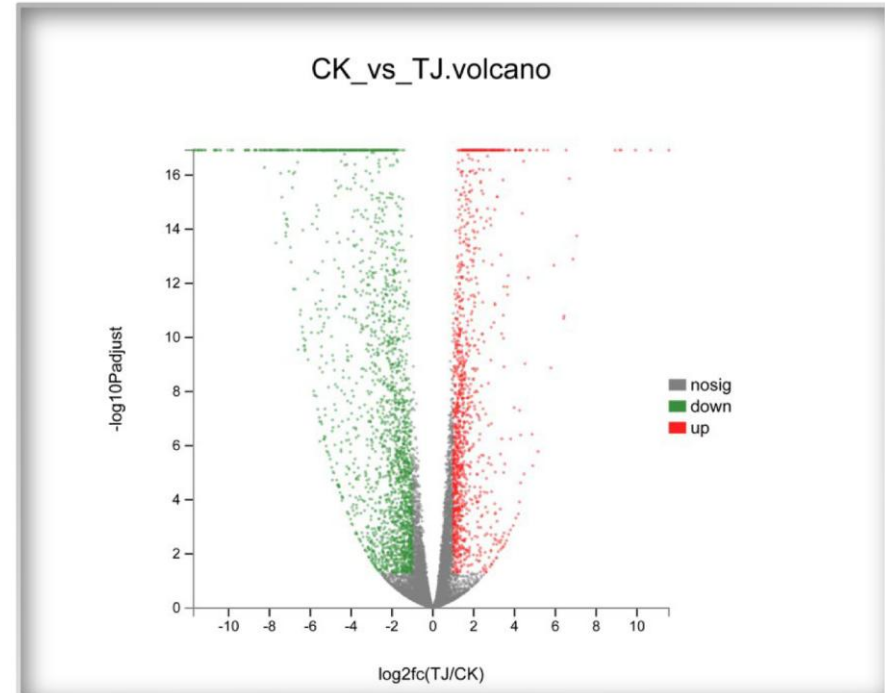
- Predicting gene function using bioinformatics
- Gene ontology and pathway mapping
- Tools: InterProScan, BLAST, KEGG
- Relevance to mycotoxin pathway discovery



<https://doi.org/10.1186/s40246-023-00512-5>

# Transcriptomics in Mycology

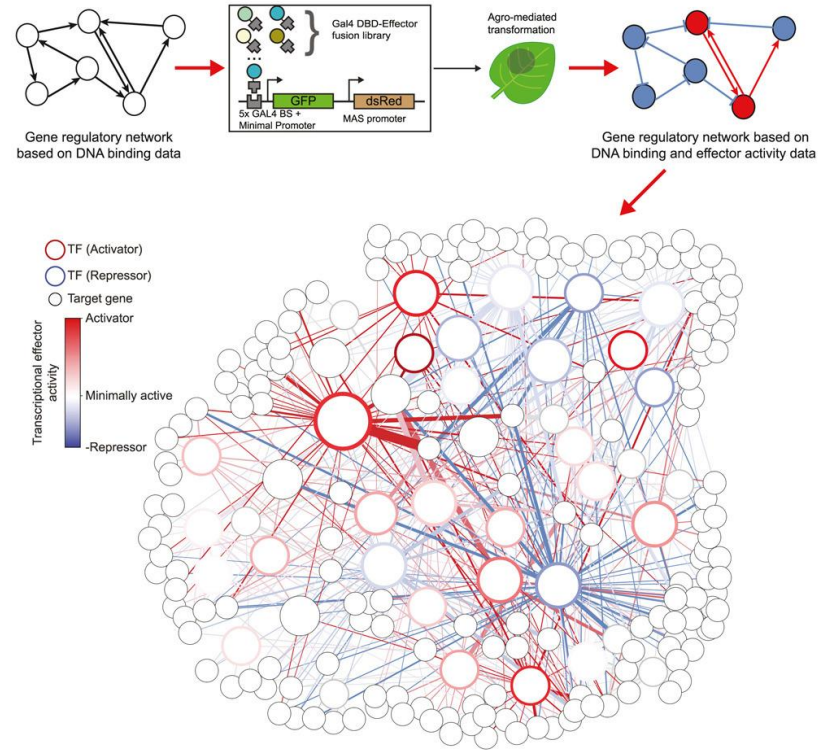
- RNA-seq for studying gene expression
- Detecting genes activated under stress or toxin-producing conditions
- Example: Upregulation of aflatoxin biosynthesis genes
- Tools: HISAT2, featureCounts, DESeq2



<https://doi.org/10.3390/jof9121193>

# Regulatory Networks in Fungi

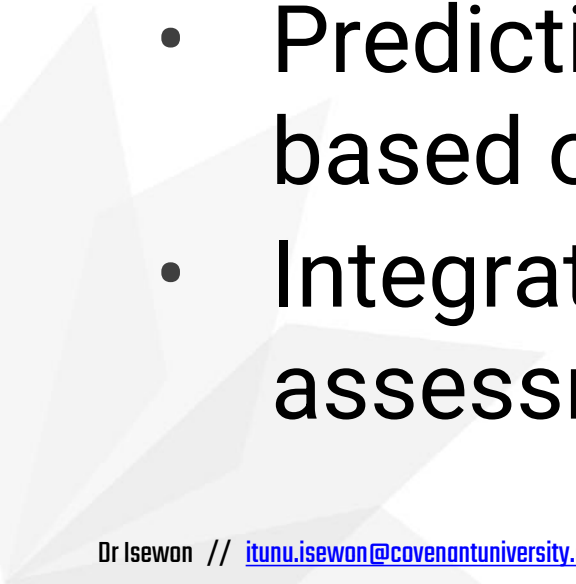
- Gene regulatory networks controlling toxin production
- Transcription factors and secondary metabolite clusters
- Visualizing networks: Cytoscape and co-expression analysis



<https://doi.org/10.1016/j.cels.2023.05.002>

# Bioinformatics in Mycotoxicology: Overview



- Linking fungal genomics to toxin production
  - Predicting mycotoxin potential based on gene content
  - Integrating omics data for risk assessment
- 

# Detection of Mycotoxin-Producing Fungi

- Using genome and transcriptome data to identify toxin producers
- Marker genes for aflatoxins, fumonisins, ochratoxins
- Case study: *Aspergillus flavus* detection in stored grains

## Machine Learning for Predicting Mycotoxin Occurrence in Maize



Marco Camardo Leggieri



Marco Mazzoni



Paola Battilani\*

Department of Sustainable Crop Production, Università Cattolica del Sacro Cuore, Piacenza, Italy


<https://doi.org/10.3389/fmicb.2021.661132>

# Predicting Toxin Production

- Correlating gene presence with observed toxin levels
- Using bioinformatics pipelines for secondary metabolite prediction
- Tools: antiSMASH, SMURF, PRISM

Use of predictive modelling as tool for prevention of fungal spoilage at different points of the food chain

[sonia Marín<sup>1</sup>](#) , [Luísa Freire<sup>2</sup>](#), [Antoni Femenias<sup>1</sup>](#), [Anderson S Sant'Ana<sup>2</sup>](#)

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<https://doi.org/10.1016/j.cofs.2021.02.006>

## Bioinformatic Analysis of Secondary Metabolite Biosynthetic Potential in Pathogenic *Fusarium*

by Chao Lin<sup>1</sup>, Xi-long Feng<sup>1</sup> , Yu Liu<sup>1</sup>, Zhao-chen Li<sup>1</sup>, Xiu-Zhang Li<sup>2</sup> and Jianzhao Qi<sup>1,\*</sup>  

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\* Author to whom correspondence should be addressed.

<https://doi.org/10.3390/jof9080850>

# Transcriptomic Analysis for Toxin Regulation

- RNA-seq to monitor expression of toxin biosynthesis genes
- Differential expression under different environmental conditions
- Visualizing results: heatmaps, volcano plots, PCA

Transcriptome analysis reveals the mechanism of tolerance to copper toxicity in the white rot fungus *Trametes hirsuta* AH28-2

Chenkai Wang<sup>a b c</sup>, Kun Wu<sup>a b c</sup>, Na Pang<sup>a b c</sup>, Huifang Zhao<sup>a b c</sup>, Shenglong Liu<sup>a b c</sup>, Xinlei Zhang<sup>a b c</sup>, Yazhong Xiao<sup>a b c</sup>  , Zemin Fang<sup>a b c</sup>  , Juanjuan Liu<sup>a b c</sup>  

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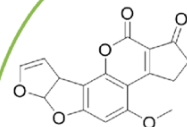
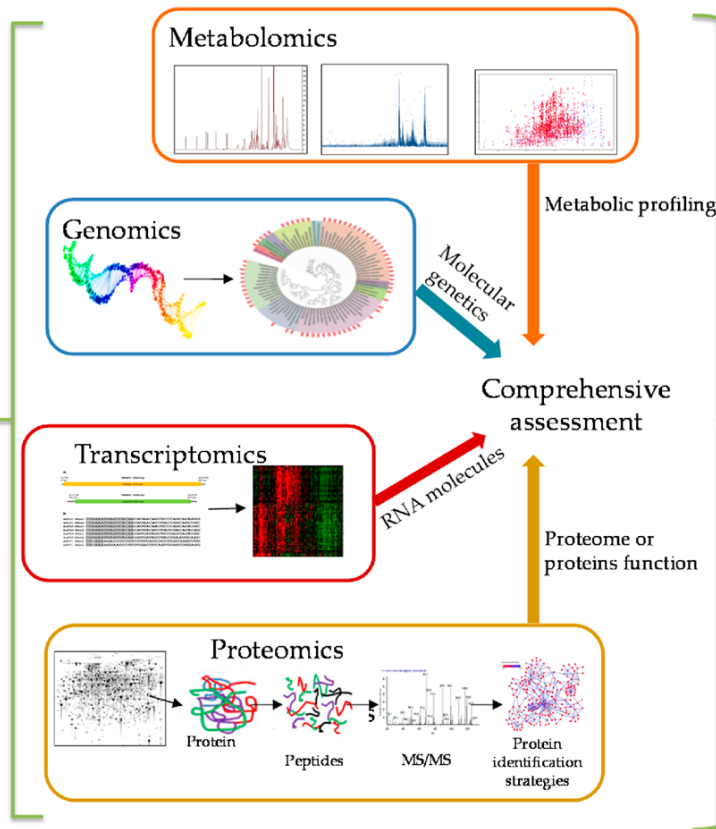
<https://doi.org/10.1016/j.ecoenv.2025.118194>



# Integrating Multi-Omics Data



Mycotoxin contamination



Develop an affordable real time solution to mitigate mycotoxin issue pre and post harvesting.

<https://doi.org/10.3390/toxins10110433>

# Key Benefits of Bioinformatics in Mycotoxin Research



- Rapid identification of fungal strains and toxin potential
- Predicting secondary metabolite production pathways
- Understanding regulatory networks for toxin biosynthesis
- Designing targeted interventions or detection assays

# Training Objectives



Understand  
bioinformatics concepts  
and databases



Perform sequence  
quality control and  
alignment



Conduct differential  
gene expression analysis



Visualize genomic and  
structural data



Apply computational  
tools to mycotoxin  
research

# Facilitators



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Emmanuella  
Matumamboh  
**Covenant University**



Dorcas Omonigbehin  
**Covenant University**



Temitayo Ogundimu  
**Covenant University**

# Day 1 Plan: Genomics & Sequence Analysis

Time	Session	Details
9:00 – 9:30	<b>Welcome &amp; Overview</b>	Workshop objectives, introduction to bioinformatics, relevance to mycotoxin research
9:30 – 10:30	<b>Introduction to Biological Databases</b>	NCBI, ENA, UniProt, PDB, Ensembl, KEGG – how to search, download, and interpret
10:30 – 11:00	Break	
11:00 – 12:30	<b>Sequence Retrieval &amp; Quality Control (Galaxy)</b>	Tool: FastQC; Dataset: Mycotoxin-related fungal genome from ENA
12:30 – 1:30	Lunch	
1:30 – 3:00	<b>Primer Design for Gene Targeting</b>	Tool: Primer3 (via Galaxy or Primer-BLAST); Task: Design primers for a conserved gene
3:00 – 4:30	<b>Variant Calling &amp; Functional Annotation</b>	Tool: Galaxy, EnsemblFungi

# Day 2 Plan: Phylogenetics, Biostatistics & Transcriptomics



Time	Session	Details
9:00 – 09:30	Recap of Day 1 Activities	
9:30 – 10:30	<b>Phylogenetic Tree Construction</b>	Tool: Clustal Omega , MEGA
10:30 – 11:00	Break	
11:00 – 12:30	<b>Biostatistics in R for Omics Data</b>	Tool: RStudio ; Task: Statistical tests and Visualizations
12:30 – 1:30	Lunch	
1:30 – 3:00	<b>Transcriptomic Data Analysis: RNA-Seq</b>	Tools: Galaxy RNA-seq workflow – HISAT2, featureCounts
3:00 – 4:30	<b>Differential Expression Analysis in Galaxy</b>	Tools: DESeq2; Task: Fold change and volcano plots using example dataset

# Day 3 Plan: Structural Bioinformatics



Time	Session	Details
9:00 – 10:30	Recap of Day 1 and Day 2 Activities	
9:30 – 10:30	<b>Introduction to Protein Structure &amp; Databases</b>	Tools: RCSB PDB, UniProt, AlphaFold; Visualizers: PyMOL or ChimeraX
10:30 – 12:00	<b>Ligand Preparation and Protein-Ligand Binding</b>	Tools: AutoDock Vina, Chimera; Dataset: Mycotoxin + fungal enzyme PDB structure
12:00 – 1:00	Lunch	
1:00 – 2:50	<b>Mini - Project Implementation</b>	
2:50 – 3:30	<b>Presentation of Mini-Projects</b>	
3:30 – 4:00	<b>Wrap-up: Tools Integration &amp; Project Ideas</b>	Group discussion: Apply tools to participants' own datasets or design mini-projects
4:00 – 4:30	<b>Certificates &amp; Feedback</b>	Evaluation, group photo



# Group Mini Projects



- Genomics
- Phylogenetics
- Transcriptomics





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**Course Page**

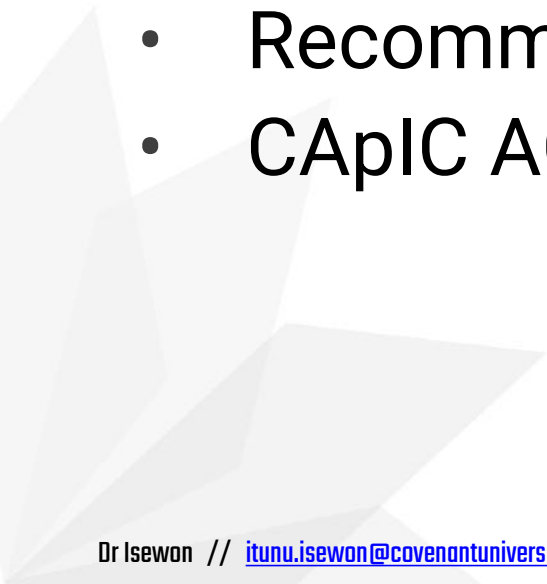
# Expected Outcomes for Participants



- Practical skills in sequence and transcriptomic analysis
- Ability to use Galaxy , MEGA and R for mycotoxin research
- Understanding structural bioinformatics basics
- Confidence to start independent projects

# Post-Training Resources



- Access to Galaxy workflows and R scripts
  - Recommended readings and databases
  - CApIC ACE support and networking
- 

# Selected References

- Bertani, F. R., Businaro, L., Gambacorta, L., Mencattin, A., Brenda, D., Di Giuseppe, D., De Ninno, A., Solfrizzo, M., Martinelli, E., & Gerardino, A. (2020). Optical detection of Aflatoxins B in grained almonds using fluorescence spectroscopy and machine learning algorithms. *arXiv*. <https://doi.org/10.48550/arXiv.2003.04096>
- Bhatnagar, D., Rajasekaran, K., Gilbert, M., & Payne, G. (2008). The “omics” tools: Genomics, proteomics, metabolomics and their potential for solving the aflatoxin contamination problem. *World Mycotoxin Journal*, 1(1), 3–12. <https://doi.org/10.3920/wmj2008.x001>
- Bhatnagar, D., Rajasekaran, K., Gilbert, M., et al. (2018). Advances in molecular and genomic research to safeguard food and feed supply from aflatoxin contamination. *World Mycotoxin Journal*, 11(1), 47–72. <https://doi.org/10.3920/wmj2017.2283>
- Chen, Z. Y., Rajasekaran, K., Brown, R. L., et al. (2015). Discovery and confirmation of genes/proteins associated with maize aflatoxin resistance. *World Mycotoxin Journal*, 8(2), 211–224. <https://doi.org/10.3920/wmj2014.1732>
- Dhakal, R., Chai, C., Karan, R., Windham, G. L., Williams, W. P., & Subudhi, P. K. (2017). Expression profiling coupled with in-silico mapping identifies candidate genes for reducing aflatoxin accumulation in maize. *Frontiers in Plant Science*, 8, Article 503. <https://doi.org/10.3389/fpls.2017.00503>
- Foster, Z. S., Sharpton, T. J., & Grunwald, N. J. (2017). Metacoder: An R package for visualization and manipulation of community taxonomic diversity data. *PLoS Computational Biology*, 13(2), e1005404. <https://doi.org/10.1371/journal.pcbi.1005404>
- Frontiers in Microbiology. (2020). Whole-genome sequencing and bioinformatics analysis of *Aspergillus mycotoxinivorans*: Predicting putative Zearalenone-degradation enzymes. *Frontiers in Microbiology*. <https://doi.org/10.3389/fmicb.2020.01866>
- Kõljalg, U., et al. (2013). Towards a unified paradigm for sequence-based identification of fungi. *Molecular Ecology*, 22, 5271–5277. <https://doi.org/10.1111/mec.12481>
- Kõljalg, U., et al. (2021). The airborne mycobiome and associations with mycotoxins and inflammatory markers in the Norwegian grain industry. *Scientific Reports*. <https://doi.org/10.1038/s41598-021-88252-1>
- Liew, W.-P. P., & Sabran, M.-R. (2022). Recent advances in immunoassay-based mycotoxin analysis and toxicogenomic technologies. *Journal of Food and Drug Analysis*, 30(4), 549–561. <https://doi.org/10.38212/2224-6614.3430>

# Selected References

- Nekrasov, N., Jaric, S., Kireev, D., Emelianov, A. V., Orlov, A. V., Gadjanski, I., Nikitin, P. I., Akinwande, D., & Bobrinetskiy, I. (2021). Real-time detection of Ochratoxin A in wine through insight of aptamer conformation in conjunction with graphene field-effect transistor. arXiv. <https://doi.org/10.48550/arXiv.2104.10551>
- Soni, P., Gangurde, S. S., Ortega-Beltran, A., et al. (2020). Functional biology and molecular mechanisms of host-pathogen interactions for aflatoxin contamination in groundnut (*Arachis hypogaea* L.) and maize (*Zea mays* L.). *Frontiers in Microbiology*, 11. <https://doi.org/10.3389/fmicb.2020.00227>
- Wikipedia. (2025). Toxicogenomics. In Wikipedia. Retrieved August 24, 2025, from <https://en.wikipedia.org/wiki/Toxicogenomics>
- Wu, B., Hussain, M., Zhang, W., Stadler, M., Liu, X., & Xiang, M. (2019). Current insights into fungal species diversity and perspective on naming the environmental DNA sequences of fungi. *Mycology*, 10, 127–140. <https://doi.org/10.1080/21501203.2019.1574393>
- Xie, M., Wu, J., An, F., Yue, X., Tao, D., Wu, R., & Lee, Y. (2019). An integrated metagenomic/metaproteomic investigation of microbiota in dajiang-meju, a traditional fermented soybean product in Northeast China. *Food Research International*, 115, 414–424. <https://doi.org/10.1016/j.foodres.2018.10.003>
- Xu, X., Jiang, Y., Ma, L., Ma, X., Liu, Y., Shan, J., Ma, K., & Xing, F. (2020). Comprehensive transcriptome and proteome analyses reveal the modulation of aflatoxin production by *Aspergillus flavus* on different crop substrates. *Frontiers in Microbiology*, 11. <https://doi.org/10.3389/fmicb.2020.01497>
- Zhang, F., Guo, Z., Zhong, H., Wang, S., Yang, W., Liu, Y., & Wang, S. (2014). RNA-Seq-based transcriptome analysis of aflatoxigenic *Aspergillus flavus* in response to water activity. *Toxins*, 6, 3187–3208. <https://doi.org/10.3390/toxins6113187>
- Zielezinski, A., Girgis, H. Z., Bernard, G., Leimeister, C.-A., Tang, K., Dencker, T., Lau, A. K., Röhling, S., Choi, J. J., & Waterman, M. S. (2019). Benchmarking of alignment-free sequence comparison methods. *Genome Biology*, 20, Article 144. <https://doi.org/10.1186/s13059-019-1773-7>
- Zieliński, B., Sroka-Oleksiak, A., Rymarczyk, D., Piekarczyk, A., & Brzychczy-Włoch, M. (2020). Deep learning approach to describe and classify fungi microscopic images. arXiv. <https://doi.org/10.48550/arXiv.2005.11772>

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# **Closing & Questions**

**Recap of Objectives  
and Day Plans**

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