# 1 Basic linux commands

Here will be briefly explained some basic linux commands. For information about more commands, feel free to consult with the internet.

## 1.1 Getting around

Command	Explanation
ls	list contents of current directory
$ls\ dirname$	list contents of directory named dirname
$\mathtt{cd}\ dirname$	change current directory to dirname
cd ~	go to home directory (default directory)
cd	go one directory up
pwd	print name of current directory
${\tt mkdir} \ dirname$	make a new directory named $dirname$
${\tt man}\ command$	show manual for <i>command</i>

# 1.2 File manipulation

Command	Explanation
cp filename1 filename2	copy file
cp -r $dirname1$ $dirname2$	copy directory
mv name1 name2	move file or directory (can be used for renaming)
rm filename	remove file (you won't be able to recover removed file)
rm -r $dirname$	remove directory and its contents
${ t more} \ filename$	command for paging through text file one screenful at a time (q t
${\tt less}~filename$	similar as more. Better for viewing large text files (q to quit)
${\tt cat}\ filename1\ filename2\ filenameN$	concatenate text files in print output to terminal window
$\mathtt{head}\;filename$	print first 10 lines of a text file to terminal window
head -n 20 $filename$	print first 20 lines of a text file to terminal window
tail	opposite of head
wc filename	show the number of lines, words and characters in a text file
<pre>cut -f 2 tabDelimitedFilename</pre>	extract 2nd column from a tab delimited file
${ t cut}$ -f 3 -d , $comaSeperatedFilename$	extract 3rd column from a coma seperated file
nano $filename$	open filename in text editor nano

# 1.3 Archiving and unarchiving of files

Note that for the tar utility, option c stands for compress, x - for uncompress or extract, z - for dealing with tar.gz, and j - for dealing with tar.bz2

## 1.3.1 Compressing

Command	Explanation
tar -cvf filename.tar filename	compress file to .tar format
tar -zcvf filename.tar.gz filename	compress file to .tar.gz format 11
tar -jcvf filename.tar.bz2 filename	compress file to .tar.bz2 format
$ exttt{zip} \ filename.zip \ filename$	$\mathbf{compress}$ file to $\mathbf{.zip}$ format
$ exttt{gzip} \ filename$	$\mathbf{compress}$ file to $\mathbf{.gz}$ format

### 1.3.2 Uncompressing

Command	Explanation
tar -xvf filename.tar	uncompress from .tar format
$ exttt{tar}$ -zxvf $filename.tar.gz$	$\mathbf{uncompress}$ from $\mathbf{.tar.gz}$ format
ar -jxvf $filename.tar.bz2$	uncompress from .tar.bz2 format
${ t unzip} \ filename.zip$	uncompress from .zip format
$ exttt{gzip}$ -d $file.gz$	$\mathbf{uncompress}$ from $\mathbf{.gz}$ format

# 1.4 Input/Output redirection

Command	Explanation
command > filename	Output of <i>command</i> is saved to <i>filename</i> , overwriting it
command >> filename	Output of command is appended at the end of filename
command < filename	command reads input from filename
$command1 \mid command2$	command2 takes the output of command1 and produces result

## 1.5 Filters

Command	Explanation
grep text filename	Prints every line in <i>filename</i> containing text
<pre>sed 's/red/green/' filename</pre>	Prints every line in filename substituting word red with word green

# 1.6 Pattern matching

Pattern	Explanation
*	matches zero or more characters
?	matches one character
~	refers to user's default (home) directory

# 1.7 Miscellaneous

Command	Explanation
echo text	display a line of text
history	view your command line history
${\tt wget}\ some WebAddress$	download contents of $some WebAddress$ to current directory
make	tool that is used to compile source code creating executables
export $name = value$	sets value to name. Type echo \$name to view value

# 2 Setup of the working environment

Since we do not have the administrator's rights on this server, we can't install software on the system. However, we can still install software locally in our home directories. We will create a special directory where all our executables will be stored. Let's create this directory and name it binaries. In linux terminal type:

#### mkdir binaries

Let's make this directory special - every executable file we put there, we will be able to easily execute, just by typing the executable's file name from anywhere in the system. To achieve this, we will be adding ~/binaries folder to the \$PATH system's environment variable by editing a text file named .bashrc using text editor nano. In linux terminal type:

nano ~/.bashrc

Text editor will open the file and you can edit it. Navigate to the bottom of the text file using arrows and type in following text:

export PATH=~/binaries:\$PATH

Hit Ctrl o and Enter to save file and Ctrl x to exit. Reload .bashrc:

source .bashrc

\$PATH variable contains a list of directories that the system will look in, when we are entering a command. To view contents of \$PATH, type in terminal:

echo \$PATH

To install software we will simply have to copy executable to our special directory binaries.

Many of the open source tools are deposited in the https://github.com repository. To download software from https://github.com easily, we will use a tool called git. git is already preinstalled on our servers, however, on your own Ubuntu servers you can install it by typing:

sudo apt-get install git

# 3 De-novo assembly of sequenced reads

### 3.1 Installation of *de-novo* assembler

For de-novo assembly we will use mira assembler. You can download it from http://sourceforge.net/projects/mira-assembler/. Click on Files  $\rightarrow$  MIRA  $\rightarrow$  stable. Rightclick on mira\_4.0.2\_linux-gnu\_x86\_64\_static.tar.bz2 and Copy link address. To download it on our linux server, we will be using command wget. In linux terminal type:

```
cd ~/programs
wget -0 mira.tar.bz2
```

paste the copied location and hit Enter. The program will be downloaded in our ~/programs directory. The downloaded software is archived in .tar.bz2 format, therefore we need to extract it from archive. To extract it from archive, type in terminal:

```
tar -jxvf mira.tar.bz2
```

A new folder named mira\_4.0.2\_linux-gnu\_x86\_64\_static will appear. This is our extracted software. MIRA is already precompiled for us, so we just need to find the compiled executable files in the folder and install them locally (or plainly speaking - copy them to our binaries):

```
cd mira_4.0.2_linux-gnu_x86_64_static
cd bin
cp mira ~/binaries
cd ~
```

Check if MIRA was installed successfully:

mira

If you see

The program 'mira' is currently not installed.

then reload .bashrc again:

source .bashrc

and check the installation again.

Let's create a seperate directory for our *de-novo* assembly project and copy the reads in it:

```
cd ngs_work
mkdir denovo
cd denovo
cp ~/data/day1/readsForDenovo.fastq .
```

MIRA needs a manifest file for performing *de-novo* assembly. We will create a basic manifest file. Our manifest file will consist of 5 entries. From MIRA's manual, these entries are:

- **project** name of our assemblies project. The project name will be used by MIRA in project's directory naming
- $\bullet$  **job** tells the assembler whether
  - 1. we want to perform de-novo assembly or map reads against reference genome
  - 2. genomic DNA or transcripts were sequenced
  - 3. we want accurate (slow) or draft (fast) assembly
- readgroup tells assembler which reads can be pooled together when assembling reads from multiple sequencing technologies
- data tells the assembler where are our reads
- technology tells the assembler what sequencing technology was used for generating reads

To create manifest file, we will use text editor nano. To start the text editor, type

#### nano

and the editor will open. Now, to create the manifest file, in the text editor type:

```
project=readsForDenovo_Assembly
job=denovo,genome,draft
readgroup
data=readsForDenovo.fastq
technology=iontor
```

To save the text file hit Ctrl o, enter the name of the file (e.g. readsForDenovo.mnfst), hit Enter to save and Ctrl x to quit nano. To launch MIRA, type:

#### mira readsForDenovo.mnfst

If you wish to gain finer control of some aspects of the assembling process, then, please, do refer to the MIRA's manual.

# 4 Building variant calling pipeline

We have *IonTorrent* targeted resequencing data from human chromosomes 13., 17. and 20. Our task is to find all nonsynonymous and stop mutations present in the data and to automatize this process by building data analysis pipeline. To accomplish this task we can divide our work in following subtasks:

- Installation of relevant tools
- Obtaining of reference sequences
- Read mapping against reference sequence
- SNP calling
- SNP annotation
- Filtering of nonsynonymous and stop mutation variants
- Pipeline building

### 4.1 Tool installation

### 4.1.1 Installation of *IonTorrent* mapping software tmap

To detect variants present in the data we need to map sequencing reads against reference sequence. For reads generated with *IonTorrent* sequencing platform we will use program tmap.

tmap and its installation instructions can be found at https://github.com/iontorrent/TS/tree/master/Analysis/TMAP. Since we do not have the administrator's rights on this server, we can't install software on the server. However, we can still use it locally. Compilation instructions:

```
cd ~/programs
git clone git://github.com/iontorrent/TMAP.git
cd TMAP
git submodule init
git submodule update
sh autogen.sh
./configure
make
```

Lets test the program to confirm that it was compiled successfully:

```
./tmap
```

If you see the programs interface, the program was compiled successfully. Move the compiled binary file to our binaries folder:

#### 4.1.2 Installation of samtools and bcftools

We will also need two tools named samtools and bcftools which are used for manipulation of mapped reads and variation calling. You can obtain these tools from http://sourceforge.net/projects/samtools/. Click on Files  $\rightarrow$  samtools  $\rightarrow$  1.2 Rightclick on samtools-1.2.tar.bz2 and choose Copy link address. We will download these tools in directory programs:

```
cd ~/programs
wget -0 samtools.tar.bz2
```

paste the copied location and hit Enter. Repeat this process for bcftools:

```
wget -0 bcftools.tar.bz2
paste the copied location and hit Enter.
```

The tools are compressed in .tar.bz2 format, so we need to extract them:

```
tar -jxvf samtools.tar.bz2
tar -jxvf bcftools.tar.bz2
```

Note that if we had a lot more files to extract and it would be too time consuming to manually extract them, we could use a for loop and pattern matching to extract archives automatically:

```
for archive in *.tar.bz2; do
   tar -jxvf $archive
done
```

We have downloaded and extracted source code of the tools, but to make these tools usable, we need to compile the source code. Source code compiling is performed with command make:

```
cd samtools-1.2 make
```

If (hopefully) no errors were encountered, then samtools was compiled correctly. type:

```
./samtools
```

to test the tool. If you see the program's interface, then the program was compiled successfully. Move compiled binary file to a directory where we are storing our compiled software:

```
cp samtools ~/binaries
```

Repeat the same process for bcftools (go to bcftools source code directory, compile it and copy resulting binary file to ~/binaries)

### 4.2 Obtaining reference sequences

We will download reference sequences in a seperate directory to avoid file cluttering. Let's make a new directory in our ngs\_work directory named reseq, and there we will create a seperate folder ref for our reference sequences:

```
cd ~
cd ngs_work
mkdir reseq
cd reseq
mkdir ref
cd ref
```

Our reference sequences can be accessed from a database made by University of California, Santa Cruz. The web address of the database is http://genome.ucsc.edu/. To find the necessary references sequences for chromosomes 13., 17. and 20. click on Downloads  $\rightarrow$  human  $\rightarrow$  Data set by chromosome. Right click on chr13.fa.gz and choose Copy link location. In terminal type

#### wget

paste the copied location and hit Enter. The reference sequence is compressed in .gz format, so we need to extract it:

```
gzip -d chr13.fa.gz
```

File chr13.fa will appear in our folder.

Repeat the process for chromosomes 17. and 20.

After we have obtained and extracted our reference sequences, we need to concatenate them. To accomplish this task we will use command cat:

```
cat chr13.fa chr17.fa chr20.fa > chr_merged.fa
```

And we have the needed reference for further data processing steps.

## 4.3 Read mapping against reference sequence

The sequenced reads we obtain from sequencing platform usually are in FASTQ format. Each record in FASTQ file consists of four entries:

- 1. read ID, beginning with symbol @
- 2. DNA sequence of read
- 3. symbol +
- 4. ASCII encoded quality of each nucleotide in read

To enable read mapper to efficiently read and process reference sequence, we need to index reference sequence using mapping software's provided indexing function. Often generated indexes are incompatible between different read mappers. As a consequence, almost every read mapping software has their own indexing algorithms.

To perform reference sequence indexing with tmap software, type in linux terminal:

```
tmap index -f ref/chr_merged.fa
```

To perform the actual read mapping against our reference, type:

```
tmap map3 -f ref/chr_merged.fa \
   -r reseq_reads.fastq \
   -i fastq \
   -o 2 \
   -s reseq_reads_mapped.bam
```

We have obtained BAM file. BAM stands for Binary SAM file. SAM, in turn, stands for Sequence Alignment/Map format.

Let's find out what is in the BAM file using samtools:

```
samtools view reseq_reads_mapped.bam | head -n 20
```

Detailed information about SAM/BAM file format, see http://samtools.github.io/hts-specs/SAMv1.pdf.

Alternatively, we could have defined the sample name during mapping process, using option -R with the complete tab delimited RG line of bam header:

```
tmap map3 -f ref/chr_merged.fa \
  -r reseq_reads.fastq \
  -R "@RG\tID:SomeID\tSM:Sample1" \
  -i fastq \
  -o 2 \
  -s reseq_reads_mapped.reheaded.bam
```

Note that the tab is denoted as \t in our header line.

samtools sort reseq\_reads\_mapped.reheaded.bam reseq\_reads\_mapped.reheaded.sorted.bam

### 4.4 SNP calling

```
samtools mpileup -uf ref/chr_merged.fa \
   reseq_reads_mapped.reheaded.sorted.bam \
   | bcftools call -cv \
   | vcfutils.pl varFilter > reseq_reads.variants.samtools.vcf
```

### 4.5 SNP annotation with snpEff

After generating a list of SNPs in *VCF* format, we will use a tool snpEff to predict whether they are causing amino acid change.

### 4.5.1 Setting up and configuring snpEff

snpEff is available at http://snpeff.sourceforge.net. To download and install it, type in linux terminal:

```
cd ~/programs
wget http://sourceforge.net/projects/snpeff/files/snpEff_latest_core.zip
   paste the copied location and hit Enter. To unzip snpEff type:
unzip snpEff_latest_core.zip
```

Unfortunately we wil not be able to launch snpEff from binaries folder and we can leave the software in programs directory.

snpEff requires databases to predict effects of SNPs. The needed database is already downloaded for us and resides in ~/data/day1/snpEffData. Our task is to configure snpEff to tell it where this database can be found.

After downloading and unzipping of snpEff we will need to edit snpEff's configuration file named snpEff.config and change entry data.dir = ./data/to data.dir = ~/data/day1/snpEffData/

### Annotate called variants:

```
java -jar ~/programs/snpEff/snpEff.jar \
   ann \
   -c ~/programs/snpEff/snpEff.config \
   GRCh38.76 \
   reseq_reads.variants.samtools.vcf \
   > reseq_reads.variants.samtools.annotated.vcf
```