

Diagnostic Value of Cerebrospinal Fluid Neurofilament Light Protein in Neurology

A Systematic Review and Meta-analysis

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 Supplemental content

IMPORTANCE Neurofilament light protein (NfL) is elevated in cerebrospinal fluid (CSF) of a number of neurological conditions compared with healthy controls (HC) and is a candidate biomarker for neuroaxonal damage. The influence of age and sex is largely unknown, and levels across neurological disorders have not been compared systematically to date.

OBJECTIVES To assess the associations of age, sex, and diagnosis with NfL in CSF (cNfL) and to evaluate its potential in discriminating clinically similar conditions.

DATA SOURCES PubMed was searched for studies published between January 1, 2006, and January 1, 2016, reporting cNfL levels (using the search terms *neurofilament light* and *cerebrospinal fluid*) in neurological or psychiatric conditions and/or in HC.

STUDY SELECTION Studies reporting NfL levels measured in lumbar CSF using a commercially available immunoassay, as well as age and sex.

DATA EXTRACTION AND SYNTHESIS Individual-level data were requested from study authors. Generalized linear mixed-effects models were used to estimate the fixed effects of age, sex, and diagnosis on log-transformed NfL levels, with cohort of origin modeled as a random intercept.

MAIN OUTCOME AND MEASURE The cNfL levels adjusted for age and sex across diagnoses.

RESULTS Data were collected for 10 059 individuals (mean [SD] age, 59.7 [18.8] years; 54.1% female). Thirty-five diagnoses were identified, including inflammatory diseases of the central nervous system (n = 2795), dementias and predementia stages (n = 4284), parkinsonian disorders (n = 984), and HC (n = 1332). The cNfL was elevated compared with HC in a majority of neurological conditions studied. Highest levels were observed in cognitively impaired HIV-positive individuals (iHIV), amyotrophic lateral sclerosis, frontotemporal dementia (FTD), and Huntington disease. In 33.3% of diagnoses, including HC, multiple sclerosis, Alzheimer disease (AD), and Parkinson disease (PD), cNfL was higher in men than women. The cNfL increased with age in HC and a majority of neurological conditions, although the association was strongest in HC. The cNfL overlapped in most clinically similar diagnoses except for FTD and iHIV, which segregated from other dementias, and PD, which segregated from atypical parkinsonian syndromes.

CONCLUSIONS AND RELEVANCE These data support the use of cNfL as a biomarker of neuroaxonal damage and indicate that age-specific and sex-specific (and in some cases disease-specific) reference values may be needed. The cNfL has potential to assist the differentiation of FTD from AD and PD from atypical parkinsonian syndromes.

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Identifying neuroaxonal damage and quantifying the intensity of this process is a critical step in patient care because it may support diagnosis and help estimate the prognosis of neurological conditions. In addition, it is essential for the evaluation of drug candidates with disease-modifying potential. Neurofilament light protein (NfL) is an abundant cytoskeletal protein exclusively expressed by central and peripheral neurons. Elevated levels of NfL in cerebrospinal fluid (CSF) were first reported in neurodegenerative conditions more than 20 years ago,¹ sparking interest in the potential of this neuron-specific protein as a biomarker. Since then, elevated levels of NfL in CSF (cNfL) have been described in a number of neurological and psychiatric conditions. The magnitude of the increase in inflammatory, degenerative, infectious, ischemic, and traumatic neurological conditions, as well as in psychiatric disorders, varies between conditions and studies. To date, cNfL levels have not been compared systematically between neurological disorders, and patient numbers in individual studies are often low. A positive association between cNfL and age has been reported in healthy controls (HC)² but was not systematically investigated in neurological conditions and may alter the performance of this biomarker across age categories. Together, these questions limit clinical implementation of cNfL. To compare cNfL levels between diagnoses, assess the association of age and sex with these variables, and evaluate the potential of cNfL level as a diagnostic biomarker, we performed a systematic review and meta-analysis on individual data collected from studies reporting cNfL levels in diseases and controls.

Methods

Search Strategy

This systematic review and meta-analysis followed Preferred Reporting Items for Systematic Reviews and Meta-analyses (PRISMA) reporting guidelines.³ We searched PubMed for articles published in English between January 1, 2006, and January 1, 2016, reporting cNfL levels (using the search terms *neurofilament light* and *cerebrospinal fluid*) in neurological or psychiatric conditions and/or in HC. Titles and abstracts were reviewed, and relevant studies were selected. The quality of primary articles was assessed using relevant criteria from the Strengthening of Reporting of Observational Studies in Epidemiology (STROBE) guidelines⁴ and the QUADAS-2 guidelines.⁵ All studies were approved by local ethics committees.

Inclusion Criteria

Studies were included if lumbar cNfL was reported for neurological patients and/or HC and/or individuals with subjective neurological or cognitive complaints and/or a psychiatric condition and/or a systemic disease that may affect the central nervous system (CNS). A reference method for the measurement of cNfL is lacking to date. To limit between-cohort heterogeneity due to the measurement tool, we included only those studies that used the same commercially available immunoassay (NF-light ELISA [enzyme-linked immunosorbent assay]; UmanDiagnostics) on the market since 2006. This assay

Key Points

Question How do levels of neurofilament light in cerebrospinal fluid (cNfL) compare between neurological conditions and with healthy controls?

Findings Among 10 059 individuals in this systematic review and meta-analysis, cNfL was elevated in most neurological conditions compared with healthy controls, and the magnitude of the increase varies extensively. Although cNfL overlaps between most clinically similar conditions, its distribution did not overlap in frontotemporal dementia and other dementias or in Parkinson disease and atypical parkinsonian syndromes.

Meaning The cNfL is a marker of neuronal damage and may be useful to differentiate some clinically similar conditions, such as frontotemporal dementia from Alzheimer disease and Parkinson disease from atypical parkinsonian syndromes.

was selected because it was used in a majority of publications (71 of 112) since 2006 and was reported to be sensitive and robust.⁶

Data Collection

We contacted the corresponding authors to request access to individual-level cNfL, age at CSF sampling, sex, and diagnosis. An individual's data were included only if all of those variables were available. For patients with multiple sclerosis (MS) and HIV-positive individuals, treatment status was also collected.^{7,8} Information on study procedures was extracted from the publication or requested from the corresponding author.

Diagnostic Categories

Diagnosis was established by the original study authors according to published criteria when applicable (Table 1). Information about the clinical subtype of neurodegenerative conditions was not retained, and all clinical subtypes of a condition were pooled in a single diagnostic group. Stroke, cardiac arrest, HIV infection, chronic inflammatory demyelinating polyradiculoneuropathy (CIDP), Guillain-Barré syndrome (GBS), Cushing disease in remission, and optic neuritis (ON) were diagnosed according to clinical guidelines. Presymptomatic genetic frontotemporal dementia (pgFTD), Huntington disease (HD), and premanifest HD (pHD) were diagnosed by genetic testing. The HIV-infected individuals with cognitive impairment (iHIV) included individuals with mild neurocognitive impairment and individuals with HIV-associated dementia.

Individuals with subjective neurological complaint (SNC) or subjective cognitive decline (SCD) had complaints but no objectifiable neurological condition after extensive workup. Inflammatory neurological diseases (IND) were inflammatory diseases of the CNS, excluding MS, clinically isolated syndrome (CIS), and ON. Noninflammatory neurological diseases (NID) were any CNS disease that was not of inflammatory nature. Mixed dementia (MD) was dementia of assumed mixed pathology, and dementia not specified (DNS) was dementia of uninvestigated origin. Healthy controls were individuals who did not have neurological complaints or signs of a neurological condition.

Diagnostic Groups

We clustered a subset of frequent neurological conditions into 3 groups of clinically similar disorders. These included the following: (1) untreated relapsing-remitting MS (uRRMS), individuals with relapsing-remitting MS treated with disease-modifying therapy (tRRMS), CIS, ON, primary progressive MS (PPMS), secondary progressive MS (SPMS), and IND; (2) Alzheimer disease (AD), FTD, combined FTD and amyotrophic lateral sclerosis (FTD/ALS), vascular dementia (VaD), dementia with Lewy bodies (DLB), idiopathic normal-pressure hydrocephalus (iNPH), mild cognitive impairment of suspected AD pathology (MCI), SCD, and iHIV; and (3) Parkinson disease (PD), PD dementia (PDD), DLB, multiple system atrophy (MSA), progressive supranuclear palsy (PSP), and corticobasal syndrome of suspected tau underlying pathology (CBS).

cNfL Measurement

The cNfL was measured at 17 different centers using the commercially available kit (NF-light ELISA assay). The cNfL values were reported in picograms per milliliter or nanograms per liter. A systematic error in the reported concentration of cNfL was identified at 8 centers due to a misinterpretation of the assay's protocol. The protocol indicated to perform a 1:1 dilution of CSF before performing the assay. However, because this dilution is included a priori in the value assignment of the standard curve, this initial dilution should not be corrected for at calculation of the concentration. Raw NfL values obtained from the 8 implicated centers were corrected for the systematic error (divided by 2).

Statistical Analysis

We performed an individual-level meta-analysis based on cNfL measurements provided by the corresponding authors. Linear mixed-effects models were used to estimate the fixed effects of age, sex, and diagnosis on log-transformed NfL levels, with cohort of origin modeled as a random intercept, using the R packages “lme4” and “lmerTest” (R Project for Statistical Computing). Age was centered according to the mean. First, we tested all 2-way and 3-way interaction terms between all fixed effects, which were retained in the model when statistically significant. No 2-way interaction of age and sex or 3-way interaction of age, sex, and diagnosis on cNfL was observed, and the best-fitting model included all fixed effects and interaction terms for diagnosis by age and diagnosis by sex. Next, we used the R package “emmeans” to obtain marginalized change folds and 95% CI cNfL and cNfL-age slope estimates for all diagnoses and to perform post hoc pairwise comparisons between diagnoses in the mean cNfL levels and in the strength of the associations between cNfL age, adjusting *P* values for multiple testing with the Tukey procedure. Finally, we calculated point estimates of fold-change increases for each diagnostic group compared with controls for specific ages. The consequences of study variability on the results was assessed using the intraclass correlation coefficient, which reflects the proportion of variance that can be attributed to between-study variation, for the total sample and per diagnostic group (analyses for the latter were performed on models the included the fixed effects of age and sex). Values higher than 0.60

Table 1. Diagnostic Criteria Used by the Original Study Authors

Diagnosis	Abbreviation	Diagnostic Criteria
Multiple sclerosis and clinically isolated syndrome	MS and CIS	McDonald criteria, ⁹ 2005 revisions, ¹⁰ and 2010 revisions ¹¹
Alzheimer disease and mild cognitive impairment	AD and MCI	Criteria by McKhann et al ¹² and IWG-2 criteria ¹³
Parkinson disease	PD	United Kingdom Parkinson Disease Society Brain Bank criteria ¹⁴ and National Institute of Neurological Disorders and Stroke criteria ¹⁵
Parkinson disease dementia	PDD	Movement Disorder Task Force ¹⁶
Progressive supranuclear palsy	PSP	Criteria by Litvan et al ¹⁷
Multiple system atrophy	MSA	Criteria by Gilman et al ¹⁸
Corticobasal syndrome	CBS	Criteria by Lee et al, ¹⁹ criteria by Litvan et al, ¹⁷ and criteria by Mathew et al ²⁰
Dementia with Lewy bodies	DLB	Criteria by McKeith et al ²¹
Frontotemporal dementia (including all clinical subtypes)	FTD	Criteria by Neary et al ²² and The Lund and Manchester Groups ²³
Amyotrophic lateral sclerosis	ALS	Revised El Escorial criteria ²⁴
Combined frontotemporal dementia and amyotrophic lateral sclerosis	FTD/ALS	
Vascular dementia	VaD	Criteria by Erkinjuntti et al ²⁵ and National Institute of Neurological Disorders and Stroke
Idiopathic normal-pressure hydrocephalus	iNPH	Criteria by Relkin et al ²⁶
Bipolar disorder	BD	<i>Diagnostic and Statistical Manual of Mental Disorders</i> (Fourth Edition)
HIV positive with cognitive impairment (including entire spectrum of cognitive impairment)	iHIV	Global Deficit Score ²⁷

Abbreviation: IWG-2, International Working Group 2.

were considered to be indicative of substantial heterogeneity. The results were considered statistically significant when they had an adjusted 2-sided *P* value below .05. All analyses were performed in R version 3.4.2.

Results

Data Set Characteristics, Population, and Demographics

The literature search resulted in 153 records. On the basis of title and abstract, 112 publications were selected for full-text review, and 44 data sets met our selection criteria and were included in the meta-analysis. In addition, 3 data sets unpublished at the time of data collection were provided by study authors, resulting in a total of 47 data sets (Table 2 and eFigure 1 in the Supplement). Data were obtained for 10 059 individuals (mean [SD] age, 59.7 [18.8] years; 54.1% female), and 35 diagnoses were identified, including control groups (HC [n = 1332], SNC [n = 45], and SCD [n = 24] [eTable 1 in the Supplement]), inflammatory diseases of the CNS (CIS, ON, RRMS, SPMS, PPMS, and IND [n = 2795]) (eTable 1 in the

Table 2. Data Sets Included in the Meta-analysis

Source	Contributed Diagnostic Categories (No. of Individuals)	Diagnostic Criteria	Healthy Controls Contributed, No.
Anckarsäter et al, ²⁸ 2014	None	NA	34
Axelsson et al, ²⁹ 2014	SPMS (n = 30), PPMS (n = 5)	McDonald criteria 2010 revisions ¹¹	14
Bäckström et al, ³⁰ 2015	PD (n = 99), MSA (n = 11), PSP (n = 12)	PD: United Kingdom Parkinson Disease Society Brain Bank criteria ¹⁴ MSA: Criteria by Gilman et al ¹⁸ PSP: Criteria by Litvan et al ¹⁷	30
Bjerke et al, ³¹ 2011	AD (n = 30), VaD (n = 26)	AD: Criteria by McKhann et al ¹² VaD: Criteria by Erkinjuntti et al ²⁵	30
Bjerke et al, ³² 2014 and Jonsson et al, ³³ 2012	MCI (n = 31)	Criteria by McKhann et al ¹²	15
Bruno et al, ³⁴ 2012	None	NA	19
Burman et al, ³⁵ 2014	RRMS (n = 43), SPMS (n = 20), National Institute of Neurological Disorders and Stroke (n = 7), SNC (n = 6)	McDonald criteria 2010 revisions ¹¹	2
Fialová et al, ³⁶ 2013	CIS (n = 32), RRMS (n = 18)	McDonald criteria 2005 revisions ¹⁰	24
Fialová et al, ³⁷ 2017	AD (n = 25), DNS (n = 13), IND (n = 17)	AD: Criteria by McKhann et al ¹²	25
Gunnarsson et al, ³⁸ 2011	RRMS (n = 92)	McDonald criteria 2010 revisions ¹¹	0
Hall et al, ³⁹ 2012 and Hall et al, ⁴⁰ 2015	AD (n = 48), PD (n = 196), PDD (n = 56), PSP (n = 53), MSA (n = 67), CBS (n = 15), DLB (n = 69)	AD: Criteria by McKhann et al ¹² PD: National Institute of Neurological Disorders and Stroke criteria ¹⁵ PDD: Movement Disorder Task Force ¹⁶ MSA: Criteria by Gilman et al ¹⁸ PSP and CBS: Criteria by Litvan et al ¹⁷ DLB: Criteria by McKeith et al ²¹ CBS: Criteria by Mathew et al ²⁰	150
Herbert et al, ⁴¹ 2015	PD (n = 64), MSA (n = 50)	PD: United Kingdom Parkinson Disease Society Brain Bank criteria ¹⁴ MSA: Criteria by Gilman et al ¹⁸	70
Hjalmarsson et al, ⁴² 2014	Stroke (n = 20)	Clinical	20
Jakobsson et al, ⁴³ 2014 and Rolstad et al, ⁴⁴ 2015	BD (n = 133)	<i>Diagnostic and Statistical Manual of Mental Disorders</i> (Fourth Edition)	38
Jeppsson et al, ⁴⁵ 2013	iNPH (n = 27)	Criteria by Relkin et al ²⁶	20
Jessen Krut et al, ⁴⁶ 2014	iHIV (n = 13)	Global Deficit Score ²⁷	152
Khademi et al, ² 2013 Aeinehband et al, ⁴⁷ 2015, and unpublished data	CIS (n = 203), RRMS (n = 682), IND (n = 387), National Institute of Neurological Disorders and Stroke (n = 370)	McDonald criteria ⁹	30
Khalil et al, ⁴⁸ 2013	CIS (n = 47), NID (n = 15)	McDonald criteria 2010 revisions ¹¹	0
Kuhle et al, ⁴⁹ 2013	CIS (n = 62), RRMS (n = 38), SPMS (n = 25), PPMS (n = 23)	McDonald criteria 2005 revisions ¹⁰	72
Kuhle et al, ⁵⁰ 2013	RRMS (n = 30)	McDonald criteria 2005 revisions ¹⁰	0
Kuhle et al, ⁵¹ 2015	RRMS (n = 36)	McDonald criteria 2005 revisions ¹⁰	0
Magdalinou et al, ⁵² 2015 and unpublished data	AD (n = 26), CBS (n = 16), FTD (n = 16), MSA (n = 30), PD (n = 10), PSP (n = 29)	AD: Criteria by McKhann et al ¹² CBS: Criteria by Mathew et al ²⁰ FTD: The Lund and Manchester Groups ²³ MSA: Criteria by Gilman et al ¹⁸ PD: United Kingdom Parkinson Disease Society Brain Bank criteria ¹⁴ PSP: Criteria by Litvan et al ¹⁷	28
Martínez et al, ⁵³ 2015 and unpublished data	PPMS (n = 17), SPMS (n = 6), RRMS (n = 192), CIS (n = 109)	McDonald criteria ⁹	0
Martínez et al unpublished data	CIS (n = 51), RRMS (n = 46)	McDonald criteria ⁹	0
Martínez et al unpublished data	NID (n = 6), IND (n = 2), stroke (n = 4), GBS (n = 1), ON (n = 1)	Clinical	0
Meeter et al, ⁵⁴ 2016	pgFTD (n = 42), FTD (n = 90)	Not specified	49

(continued)

Table 2. Data Sets Included in the Meta-analysis (continued)

Source	Contributed Diagnostic Categories (No. of Individuals)	Diagnostic Criteria	Healthy Controls Contributed, No.
Menke et al, ⁵⁵ 2015 and Lu et al, ⁵⁶ 2015	ALS (n = 38)	Revised El Escorial criteria ²⁴	20
Modvig et al, ⁵⁷ 2013 and Modvig et al, ⁵⁸ 2016	ON (n = 56)	Clinical	27
Modvig et al, ⁵⁹ 2015	ON (n = 85)	Clinical	0
Paterson et al, ⁶⁰ 2015	AD (n = 94)	IWG-2 criteria ¹³	30
Pérez-Santiago et al, ⁶¹ 2016	iHIV (n = 14), HIV (n = 14)	Global Deficit Score ²⁷	0
Pijnenburg et al, ⁶² 2015	FTD/ALS (n = 26), FTD (n = 4), AD (n = 25), SCD (n = 24)	ALS: Revised El Escorial criteria ²⁴ AD: Criteria by McKhann et al ¹² Neuropathological confirmation (11 of 25 for AD, 15 of 23 for FTD) Genetic confirmation (12 of 23 for FTD)	0
Pyykkö et al, ⁶³ 2014	iNPH (n = 29), MD (n = 3), AD (n = 8)	AD: Criteria by McKhann et al ¹² iNPH: Clinical	0
Ragnarsson et al, ⁶⁴ 2013	Cushing disease (n = 12)	Clinical	6
Romme Christensen et al, ⁶⁵ 2014	PPMS (n = 12), SPMS (n = 12)	McDonald criteria 2005 revisions ¹⁰	0
Rosén et al, ⁶⁶ 2014	Cardiac arrest (n = 21)	Clinical	20
Sandberg et al, ⁶⁷ 2016	RRMS (n = 97), SPMS (n = 44), PPMS (n = 12)	McDonald criteria 2010 revisions ¹¹	0
Scherling et al, ⁶⁸ 2014	FTD (n = 83), PSP (n = 23), CBS (n = 16), PD (n = 6), AD (n = 45)	FTD: Criteria by Neary et al ²² PSP: Criteria by Litvan et al ¹⁷ AD: Criteria by McKhann et al ¹² CBS: Criteria by Lee et al ¹⁹	54
Skillbäck et al, ⁶⁹ 2014	AD (n = 1417), PDD (n = 45), FTD (n = 146), LBD (n = 114), MD (n = 517), VaD (n = 465), DNS (n = 545)	AD: IWG-2 criteria ¹³ DNS: <i>International Statistical Classification of Diseases, 10th Revision</i> PDD: Movement Disorder Task Force ¹⁶ FTD: The Lund and Manchester Groups ²³ DLB: Criteria by McKeith et al ²¹ VaD: National Institute of Neurological Disorders and Stroke	107
Stilund et al, ⁷⁰ 2015	RRMS (n = 44), PPMS (n = 15), CIS (n = 27), SNC (n = 39)	McDonald criteria 2010 revisions ¹¹	0
Tortelli et al, ⁷¹ 2015 and Tortelli et al, ⁷² 2012	CIDP (n = 25), ALS (n = 37), MCI (n = 3), AD (n = 15), MSA (n = 1), CBS (n = 2), NID (n = 5)	ALS: Revised El Escorial criteria ²⁴ CIDP: Clinical AD and MCI: Criteria by McKhann et al ¹² CBS: Criteria by Lee et al ¹⁹ MSA: Criteria by Gilman et al ¹⁸	0
Tortorella et al, ⁷³ 2015	CIS (n = 21)	McDonald criteria 2005 revisions ¹⁰	0
Trentini et al, ⁷⁴ 2014	PPMS (n = 21), SPMS (n = 10), National Institute of Neurological Disorders and Stroke (n = 15)	McDonald criteria ⁹	0
Vågberg et al, ⁷⁵ 2015	None	NA	53
Villar et al, ⁷⁶ 2015	RRMS (n = 98), CIS (n = 29)	McDonald criteria 2010 revisions ¹¹	37
Wild et al, ⁷⁷ 2015	HD (n = 30), pHD (n = 13)	Genetic testing	14
Zetterberg et al, ⁷⁸ 2016	MCI (n = 193), AD (n = 95)	Criteria by McKhann et al ¹²	111

Abbreviations: AD, Alzheimer disease; ALS, amyotrophic lateral sclerosis; BD, bipolar disorder; CBS, corticobasal syndrome; CIDP, chronic inflammatory demyelinating polyradiculopathy; CIS, clinically isolated syndrome; DLB, dementia with Lewy bodies; DNS, dementia not specified; FTD, frontotemporal dementia; FTD/ALS, combined frontotemporal dementia and amyotrophic lateral sclerosis; GBS, Guillain-Barré syndrome; HD, Huntington disease; iHIV, HIV positive with cognitive impairment; IND, inflammatory neurological disorders other than multiple sclerosis; iNPH, idiopathic normal-pressure hydrocephalus; IWG-2, International Working

Group 2; MCI, mild cognitive impairment; MD, mixed dementia; MSA, multiple system atrophy; NA, not applicable; NID, noninflammatory neurological disorders; ON, optic neuritis; PD, Parkinson disease; PDD, Parkinson disease dementia; pgFTD, presymptomatic genetic frontotemporal dementia; pHD, premanifest Huntington disease; PPMS, primary progressive multiple sclerosis; PSP, progressive supranuclear palsy; SCD, subjective cognitive decline; SNC, subjective neurological complaint; SPMS, secondary progressive multiple sclerosis; RRMS, relapsing-remitting multiple sclerosis; VaD, vascular dementia.

Supplement), dementias and predementia stages (MCI, AD, pgFTD, FTD, VaD, DLB, iNPH, DNS, MD, pHD, HD, iHIV, and FTD/ALS [n = 4339]) (eTable 1 in the Supplement), and parkinsonian syndromes (PD, PDD, MSA, PSP, CBS, and DLB [n = 984]) (eTable 1 in the Supplement). Three diagnostic categories were excluded from the statistical models because they had fewer than 5 observations per sex (Cushing disease, cardiac arrest, and HIV), resulting in 32 diagnostic categories and 10 012 individuals included in the analysis.

cNfL Distribution Across Diagnoses

We first examined the distribution of cNfL across diagnostic categories (Figure 1). The cNfL was increased compared with HC in most neurological conditions (Figure 1A). The fold changes compared with HC varied extensively between individual conditions, with the largest effect sizes observed in iHIV (21.36; 95% CI, 9.86-46.30), FTD/ALS (10.48; 95% CI, 4.85-22.67), ALS (7.58; 95% CI, 4.49-12.81), and HD (5.88; 95% CI, 2.43-14.27) (Figure 1B; eTable 2 in the Supplement).

Association of cNfL With Age and Sex

In HC, we observed a yearly increase of 3.30% (95% CI, 2.98%-3.62%) in cNfL levels (eTable 2 in the Supplement). A positive association between cNfL and age was also observed in individuals with subjective complaints, BD, and in most neurodegenerative conditions (eTable 2 in the Supplement). In MS, iHIV, and rapidly progressive neurodegenerative conditions (FTD, ALS, FTD/ALS, MSA, PSP, CBS, and HD), no such association was observed (eTable 2 in the Supplement). In HC, cNfL was higher in men (26.0%, 95% CI, 16.0%-37.0%) (eTable 3 in the Supplement). This was also the case in a minority of neurological conditions, including MS, AD, VaD, and PD (eTable 3 in the Supplement).

cNfL Levels Within 3 Groups of Clinically Similar Disorders

We next compared cNfL between neurological conditions within 3 groups of clinically similar disorders. In inflammatory conditions of the CNS, the mean cNfL levels were similar in ON, CIS, and MS subtypes (eTable 4A in the Supplement). The association between cNfL and age was positive in ON, CIS, and IND but was negative in uRRMS (Figure 2A; eFigure 2 and eTable 2 in the Supplement). The ratio of cNfL between ON and CIS, ON and IND, and CIS and IND remained stable across the age range of the study, while the ratio between uRRMS and CIS decreased with increasing age (eTable 5A in the Supplement). No association between cNfL and age was observed in tRRMS and PPMS (Figure 2A and eTable 4 in the Supplement). The ratio of cNfL between uRRMS and tRRMS and between uRRMS and PPMS remained stable across the age range of the study (eTable 5B in the Supplement). No association between cNfL and age was observed in SPMS (Figure 2A; and eTable 2 in the Supplement). Although cNfL levels tended to be higher in young uRRMS compared with age-corresponding SPMS, this did not reach statistical significance (eTable 5C in the Supplement). In dementias and related disorders, the mean cNfL levels were statistically significantly higher in FTD compared with other causes of dementia, such as AD (2.08; 95% CI, 1.72-2.56 [eTable 4B in the Supplement]), VaD (1.56;

95% CI, 1.25-1.96 [eTable 4B in the Supplement]), and DLB (2.50; 95% CI, 1.89-3.33 [eTable 4B in the Supplement]). An association of cNfL with age was positive in AD, VaD, and DLB but was absent in FTD (Figure 2B; eFigure 2B and eTable 4B in the Supplement). The ratio of cNfL between AD and FTD increased with age; in individuals 90 years and older, the distribution of cNfL in both conditions overlapped (eTable 5D in the Supplement). An association between cNfL and age was absent in FTD and FTD/ALS, while it was present in pgFTD (eFigure 2 and eTable 2 in the Supplement). A positive association with age was observed in AD, MCI, and SCD (Figure 2B; eTable 2 in the Supplement), and the ratio of cNfL between AD and MCI remained stable across the age range (eTable 5E in the Supplement). In parkinsonian syndromes, the mean cNfL levels did not differ between PD and PDD and between PDD and DLB, while they were higher in MSA, PSP, and CBS compared with PD (eTable 4C in the Supplement). In MSA, PSP, and CBS, no association with age was observed, while a positive association was found in PD, PDD, and DLB (Figure 2C and eTable 2 in the Supplement). The ratio of cNfL between MSA and PD, PSP and PD, and CBS and PD decreased with age but remained high across the age range of the study (Figure 2C and eTable 5G in the Supplement).

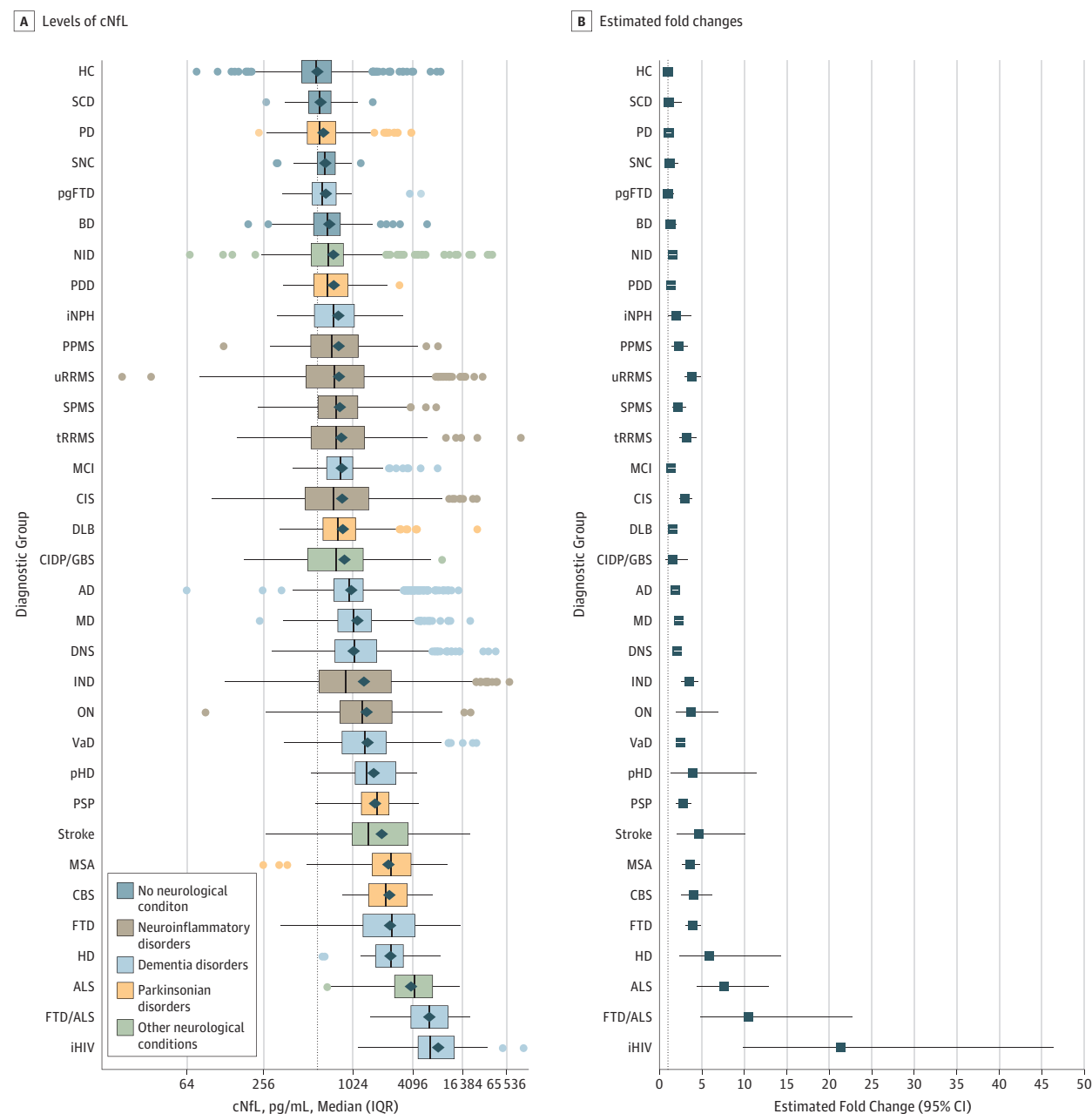
Assessment of Cohort Heterogeneity

In this meta-analysis, we pooled individual patient data originating from 42 different data sets. To estimate the proportion of the total variance of cNfL accounted for by the data set (cohort) of origin, we calculated the intraclass coefficient for cohort-related random intercepts. Across the total sample (n = 10 012), the intraclass coefficient was low at 0.15. Likewise, in a majority of diagnostic categories, the intraclass coefficient was low to moderate (<0.60). However, in 7 of the 32 diagnostic categories (MD, DNS, PDD, DLB, NID, iHIV, and stroke), the intraclass coefficients were high (>0.60), indicating that a large proportion of the variance in cNfL was due to the data set of origin (eTable 6 in the Supplement).

Discussion

In this meta-analysis that included 10 012 individuals, we found that cNfL was increased compared with HC in most neurological conditions studied. The largest effect sizes were observed in iHIV, FTD/ALS, ALS, and HD, while the effect sizes in inflammatory conditions of the CNS were low. Other neurological disorders showed much subtler increases that failed to reach statistical significance (PD and CIDP/GBS). However, the effect sizes in these conditions were positive, and larger sample sizes may allow for more robust estimates. In HC, we observed a positive association between cNfL and age. A positive association, albeit weaker, was also present in a majority of neurological conditions. An association with sex was absent in most diagnostic categories except for HC, PPMS, AD, VaD, and PD, where levels were higher in men. In clinically similar disorders, the distribution of cNfL relative to age mostly overlapped, suggesting limited use for differential diagnosis. Exceptions were FTD, which segregated from other common

Figure 1. Neurofilament Light in Cerebrospinal Fluid (cNfL) Levels Across Diagnostic Categories



A, Levels of cNfL are shown corrected for age and sex. B, Estimated fold changes are compared with healthy controls (HC). AD indicates Alzheimer disease; ALS, amyotrophic lateral sclerosis; BD, bipolar disorder; CBS, corticobasal syndrome; CIDP/GBS, chronic inflammatory demyelinating polyradiculopathy and Guillain-Barré syndrome; CIS, clinically isolated syndrome; DLB, dementia with Lewy bodies; DNS, dementia not specified; FTD, frontotemporal dementia; FTD/ALS, combined frontotemporal dementia and amyotrophic lateral sclerosis; HD, Huntington disease; iHIV, HIV positive with cognitive impairment; IND, inflammatory neurological disorders other than multiple sclerosis; iNPH, idiopathic normal-pressure hydrocephalus; MCI, mild

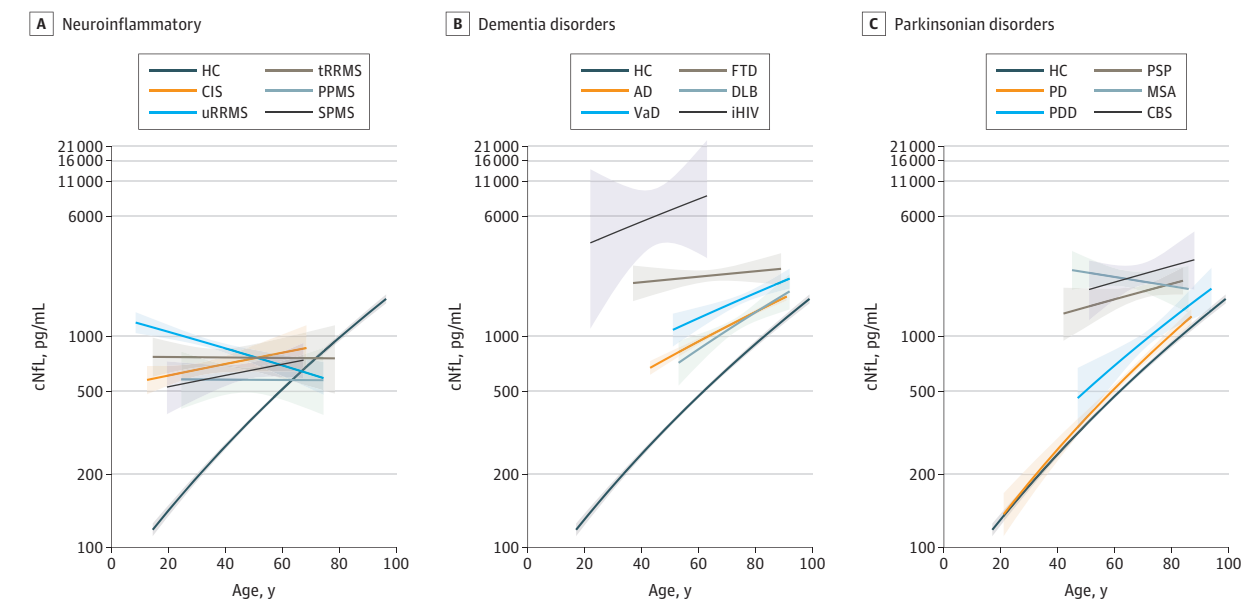
cognitive impairment; MD, mixed dementia; MSA, multiple system atrophy; NID, noninflammatory neurological disorders; ON, optic neuritis; PD, Parkinson disease; PDD, Parkinson disease dementia; pgFTD, presymptomatic genetic frontotemporal dementia; pHd, premanifest Huntington disease; PPMS, primary progressive multiple sclerosis; PSP, progressive supranuclear palsy; SCD, subjective cognitive decline; SNC, subjective neurological complaint; SPMS, secondary progressive multiple sclerosis; tRRMS, treated relapsing-remitting multiple sclerosis; uRRMS, untreated relapsing-remitting multiple sclerosis; and VaD, vascular dementia.

causes of dementia (including AD and VaD), and PD, which segregated from atypical parkinsonian syndromes. These data indicate that cNfL may contribute to the differentiation of these conditions, particularly in younger individuals.

cNfL and Age

In about two-thirds of the diagnoses, including HC, we observed a positive association between cNfL and age. In the control groups (HC, SNC, and SCD), as well as in pgFTD and BD,

Figure 2. Neurofilament Light in Cerebrospinal Fluid (cNfL) in Neurological Conditions According to Age



A-C, Log cNfL values are shown according to age across diagnoses. Shading around regression lines represents standard errors. AD indicates Alzheimer disease; CBS, corticobasal syndrome; CIS, clinically isolated syndrome; DLB, dementia with Lewy bodies; FTD, frontotemporal dementia; HC, healthy controls; iHIV, HIV positive with cognitive impairment; MSA, multiple system

atrophy; PD, Parkinson disease; PDD, Parkinson disease dementia; PPMS, primary progressive multiple sclerosis; PSP, progressive supranuclear palsy; SPMS, secondary progressive multiple sclerosis; tRRMS, treated relapsing-remitting multiple sclerosis; uRRMS, untreated relapsing-remitting multiple sclerosis; and VaD, vascular dementia.

the association of cNfL with age was strongest. This positive association in diagnostic categories without an overt neurological condition may reflect a decrease in CSF clearance with age, the presence of a preclinical age-related neurological condition, or age-related neuronal loss.⁷⁹ The association of cNfL with age in HC implies that age-specific reference values may be needed and that the diagnostic potential of cNfL may decrease with age. In neurological conditions with substantially elevated levels of cNfL, such as FTD, ALS, FTD/ALS, HD, and iHIV, as well as in atypical parkinsonian syndromes, no association with age was observed, suggesting that neuropathological processes may cause plateau levels or mask age associations. In MS, an association with age was absent or negative, which may reflect the observation that younger patients with MS have more active diseases.²

cNfL and Sex

In a minority of diagnoses, including HC, cNfL was higher in men than women. The clinical relevance of these findings is uncertain, but the results suggest that sex-specific reference values may be needed.

Other Determinants of cNfL Levels

Age, sex, and the random (cohort) association explained 46% of the variance of cNfL in the best-fitting model, indicating that many determinants of cNfL remain to be identified. Disease duration and severity could influence cNfL levels. However, these data were not available in the data sets that were included in this meta-analysis, and studies designed specifi-

cally to evaluate the association of these variables and others (eg, smoking, physical activity, and body size) are ongoing.

cNfL in Inflammatory Conditions of the CNS, Including MS

The cNfL was increased in all inflammatory conditions of the CNS examined in this meta-analysis, but the effect sizes were small. The distribution of cNfL in CIS, ON, and RRMS overlapped, which may be expected because CIS and a proportion of ON are initial manifestations of RRMS. Neurodegeneration has a central role in MS, contributing to disease progression and long-term disability.⁸⁰ Poor understanding of the processes driving neurodegeneration, together with the lack of biomarkers allowing dynamic measurement of its rate, hampers the development of specific treatments.⁸¹ The cNfL has been reported to correlate with brain atrophy,^{50,82} which is considered a marker of neurodegeneration.^{83,84} We found that levels of cNfL did not differ statistically significantly between RRMS, PPMS, and SPMS, indicating that on a population level cNfL may not differentiate acute inflammation-induced neuronal damage in the context of relapses from progressive neurodegeneration if the consequences of recent relapses or novel lesion formation are not considered. In individual patients, cNfL has been reported to reflect acute neuronal and axonal damage in MS, with levels transiently increasing during relapse.⁸⁵⁻⁸⁷ We found that cNfL levels in uRRMS and tRRMS did not differ statistically significantly. However, patients with the most active RRMS with potentially highest cNfL levels are also those who are most likely to be treated, and cNfL has been reported to decrease after treatment initiation in individual patients.^{38,50,51}

cNfL in Dementia and ALS

The higher levels of cNfL observed in FTD compared with other frequent causes of dementias, including AD, VaD and DLB, may be related to the anatomical location of neurodegeneration or the rate of neuronal death. This finding suggests that cNfL may support the differentiation of FTD from other dementias, in line with a recent study⁸⁸ not included in this meta-analysis, which reported that in combination with YKL40 and A β 42 cNfL assists in the differentiation between FTD and AD with high accuracy. In iHIV, which included both mild cognitive impairment due to HIV and HIV-associated dementia, we observed highest levels of cNfL, setting it apart from neurodegenerative and vascular causes of dementia. This may reflect a high rate of neuroaxonal damage due to the presence of HIV and the inflammatory response to it in the CNS, or it may indicate additional peripheral nervous system damage contributing to the elevation of cNfL. In prodementia stages, such as MCI and pgFTD, cNfL values were similar to levels in HC, suggesting that CNS damage must reach a certain extent before it is reflected by increased cNfL. However, the pgFTD cohort was small ($n = 42$); therefore, a small effect size could have been missed. The cNfL levels were highly elevated in ALS and FTD/ALS compared with HC. These results are in line with single-center studies not included in this meta-analysis that used different assays to measure NfL in CSF.⁸⁹ Together with the high levels of cNfL observed in stroke, these findings indicate that the rate of neuroaxonal damage may be an important determinant of the magnitude of NfL increase in CSF, possibly by overriding CSF clearance mechanisms.

cNfL in Degenerative Parkinsonian Syndromes

In degenerative parkinsonian syndromes, cNfL clustered into 2 groups. The first group consisted of PD, PDD, and DLB, in which cNfL levels were similar to those in HC, and the second group consisted of atypical parkinsonian syndromes MSA, PSP, and CBS, with elevated levels of cNfL compared with HC and the absence of association with age. This finding is in line with the results of another meta-analysis⁹⁰ that focused on parkinsonian disorders, examining data sets not included in the present meta-analysis, further underscoring the robustness of our findings. These data have important clinical implications because they suggest a potential for cNfL in supporting the differentiation of PD from atypical parkinsonian syndromes. Accurate and early differential diagnosis of these conditions is crucial because their prognosis and management differ substantially.

Serum NfL

A few years ago, an ultrasensitive assay was developed that allows measurement of NfL in serum (sNfL). This assay uses the same antibody pair as the immunoassay used in the studies included in this meta-analysis, and studies^{91,92} have reported high correlations between serum and CSF levels. These findings indicate that sNfL may replace cNfL. In addition, it may

likely be that the findings of the present meta-analysis, which collected data over 10 years, can be readily translated to sNfL.

Limitations of the Study

Our systematic review and meta-analysis has some limitations. In all studies included in the meta-analysis except one,⁹³ diagnosis was based on clinical criteria. This limitation is mostly a concern for dementias and parkinsonian syndromes, for which definitive diagnosis requires postmortem examination. However, the agreement between clinical and pathological diagnoses was reported to be high when diagnoses were established in specialized centers using consensus criteria.^{94,95} For AD and MCI, 2 consensus criteria were applied (criteria by McKhann et al¹² and the International Working Group 2 [IWG-2] criteria¹³), for which a high concordance rate was reported.⁹⁶ For VaD, the 2 consensus diagnostic criteria used (criteria by Erkinjuntti et al²⁵ and the National Institute of Neurological Disorders and Stroke criteria) were also reported to have a high agreement.⁹⁷ For PD, 2 consensus criteria were applied, for which concordance evaluation is not available. For ALS, FTD, PSP, MSA, PDD, DLB, and iHIV, the same consensus criteria were applied in all studies. In MS, the McDonald criteria were revised over time, and this may have influenced classification of RRMS and CIS. A further limitation is the inability to capture dementia of multifactorial origin, which may have increased heterogeneity in the dementia diagnostic categories and blurred the difference in cNfL distributions between dementia subtypes. Further classification of neurodegenerative conditions into clinical phenotypes could not be performed because this information was absent in a majority of studies. Therefore, the specific value of cNfL in subphenotypes could have been missed in this meta-analysis. In addition, for some conditions, data and age ranges were limited, resulting in large standard errors and low statistical power, and conclusions for these conditions should be interpreted with caution. Finally, we included only those studies that used a specific immunoassay for cNfL in an attempt to reduce heterogeneity due to the analytical procedure. However, the range of conditions that were explored in the studies not included in the meta-analysis for the same reason did not differ from those included.

Conclusions

Our study was designed to compare cNfL levels across neurological conditions and controls, assess the association of age and sex with these variables, and evaluate the potential of cNfL to differentiate clinically similar conditions. Our meta-analysis found that cNfL was elevated in a majority of the neurological conditions included in this study. Although cNfL overlapped between most clinically similar conditions, its distribution did not overlap in FTD compared with other dementia subtypes or in PD compared with atypical parkinsonian syndromes, indicating clinical potential in differentiating these conditions.

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REFERENCES

- Rosengren LE, Karlsson JE, Karlsson JO, Persson LI, Wikkelso C. Patients with amyotrophic lateral sclerosis and other neurodegenerative diseases have increased levels of neurofilament protein in CSF. *J Neurochem*. 1996;67(5):2013-2018. doi:10.1046/j.1471-4159.1996.67052013.x
- Khademi M, Dring AM, Gilthorpe JD, et al. Intense inflammation and nerve damage in early multiple sclerosis subsides at older age: a reflection by cerebrospinal fluid biomarkers. *PLoS One*. 2013;8(5):e63172. doi:10.1371/journal.pone.0063172
- Moher D, Liberati A, Tetzlaff J, Altman DG; PRISMA Group. Preferred Reporting Items for Systematic Reviews and Meta-analyses: the PRISMA statement. *Ann Intern Med*. 2009;151(4):264-269. W64. doi:10.7326/0003-4819-151-4-200908180-00135
- Vandenbroucke JP, von Elm E, Altman DG, et al; STROBE Initiative. Strengthening the Reporting of Observational Studies in Epidemiology (STROBE): explanation and elaboration. *Epidemiology*. 2007;18(6):805-835. Medline:18049195 doi:10.1097/EDE.0b013e3181577511
- Whiting PF, Rutjes AW, Westwood ME, et al; QUADAS-2 Group. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med*. 2011;155(8):529-536. doi:10.7326/0003-4819-155-8-20110180-00009
- Petzold A, Altintas A, Andreoni L, et al. Neurofilament ELISA validation. *J Immunol Methods*. 2010;352(1-2):23-31. doi:10.1016/j.jim.2009.09.014
- Teunissen CE, Khalil M. Neurofilaments as biomarkers in multiple sclerosis. *Mult Scler*. 2012;18(5):552-556. doi:10.1177/1352458512443092
- Yilmaz A, Blennow K, Hagberg L, et al. Neurofilament light chain protein as a marker of neuronal injury: review of its use in HIV-1 infection and reference values for HIV-negative controls. *Expert Rev Mol Diagn*. 2017;17(8):761-770. doi:10.1080/14737159.2017.1341313
- McDonald WI, Compston A, Edan G, et al. Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the diagnosis of multiple sclerosis. *Ann Neurol*. 2001;50(1):121-127. doi:10.1002/ana.1032
- Polman CH, Reingold SC, Edan G, et al. Diagnostic criteria for multiple sclerosis: 2005 revisions to the "McDonald Criteria". *Ann Neurol*. 2005;58(6):840-846. doi:10.1002/ana.20703
- Polman CH, Reingold SC, Banwell B, et al. Diagnostic criteria for multiple sclerosis: 2010 revisions to the McDonald Criteria. *Ann Neurol*. 2011;69(2):292-302. doi:10.1002/ana.22366
- McKhann GM, Knopman DS, Chertkow H, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement*. 2011;7(3):263-269. doi:10.1016/j.jalz.2011.03.005
- Dubois B, Feldman HH, Jacova C, et al. Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria [published correction appears in *Lancet Neurol*. 2014;13(8):757]. *Lancet Neurol*. 2014;13(6):614-629. doi:10.1016/S1474-4422(14)70090-0
- Hughes AJ, Daniel SE, Kilford L, Lees AJ. Accuracy of clinical diagnosis of idiopathic Parkinson's disease: a clinico-pathological study of 100 cases. *J Neurol Neurosurg Psychiatry*. 1992;55(3):181-184. doi:10.1136/jnnp.55.3.181
- Gelb DJ, Oliver E, Gilman S. Diagnostic criteria for Parkinson disease. *Arch Neurol*. 1999;56(1):33-39. doi:10.1001/archneur.56.1.33
- Emre M, Aarsland D, Brown R, et al. Clinical diagnostic criteria for dementia associated with Parkinson's disease. *Mov Disord*. 2007;22(12):1689-1707. doi:10.1002/mds.21507
- Litvan I, Agid Y, Jankovic J, et al. Accuracy of clinical criteria for the diagnosis of progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome). *Neurology*. 1996;46(4):922-930. doi:10.1212/WNL.46.4.922
- Gilman S, Low PA, Quinn N, et al. Consensus statement on the diagnosis of multiple system atrophy. *J Neurol Sci*. 1999;163(1):94-98. doi:10.1016/S0022-510X(98)00304-9
- Lee SE, Rabinovici GD, Mayo MC, et al. Clinicopathological correlations in corticobasal degeneration. *Ann Neurol*. 2011;70(2):327-340. doi:10.1002/ana.22424
- Mathew R, Bak TH, Hodges JR. Diagnostic criteria for corticobasal syndrome: a comparative study. *J Neurol Neurosurg Psychiatry*. 2012;83(4):405-410. doi:10.1136/jnnp-2011-300875
- McKeith IG, Boeve BF, Dickson DW, et al. Diagnosis and management of dementia with Lewy bodies: Fourth consensus report of the DLB Consortium. *Neurology*. 2017;89(1):88-100. doi:10.1212/WNL.0000000000004058

22. Neary D, Snowden JS, Gustafson L, et al. Frontotemporal lobar degeneration: a consensus on clinical diagnostic criteria. *Neurology*. 1998;51(6):1546-1554. doi:10.1212/WNL.51.6.1546
23. The Lund and Manchester Groups. Clinical and neuropathological criteria for frontotemporal dementia. *J Neurol Neurosurg Psychiatry*. 1994;57(4):416-418. doi:10.1136/jnnp.57.4.416
24. Ludolph A, Drory V, Hardiman O, et al; WFN Research Group on ALS/MND. A revision of the El Escorial criteria: 2015. *Amyotroph Lateral Scler Frontotemporal Degener*. 2015;16(5-6):291-292. doi:10.3109/21678421.2015.1049183
25. Erkinjuntti T, Haltia M, Palo J, Sulkava R, Paetau A. Accuracy of the clinical diagnosis of vascular dementia: a prospective clinical and post-mortem neuropathological study. *J Neurol Neurosurg Psychiatry*. 1988;51(8):1037-1044. doi:10.1136/jnnp.51.8.1037
26. Relkin N, Marmarou A, Klinge P, Bergsneider M, Black PM. Diagnosing idiopathic normal-pressure hydrocephalus. *Neurosurgery*. 2005;57(3)(suppl):S4-S16.
27. Gonzalez R, Heaton RK, Moore DJ, et al; HIV Neurobehavioral Research Center Group. Computerized reaction time battery versus a traditional neuropsychological battery: detecting HIV-related impairments. *J Int Neuropsychol Soc*. 2003;9(1):64-71. doi:10.1017/S155617703910071
28. Anckarsäter R, Anckarsäter H, Bromander S, Blennow K, Wass C, Zetterberg H. Non-neurological surgery and cerebrospinal fluid biomarkers for neuronal and astroglial integrity. *J Neural Transm (Vienna)*. 2014;121(6):649-653. doi:10.1007/s00702-013-1156-0
29. Axelsson M, Malmeström C, Gunnarsson M, et al. Immunosuppressive therapy reduces axonal damage in progressive multiple sclerosis. *Mult Scler*. 2014;20(1):43-50. doi:10.1177/1352458513490544
30. Bäckström DC, Eriksson Domellöf M, Linder J, et al. Cerebrospinal fluid patterns and the risk of future dementia in early, incident Parkinson disease. *JAMA Neurol*. 2015;72(10):1175-1182. doi:10.1001/jamaneurol.2015.1449
31. Bjerke M, Zetterberg H, Edman Å, Blennow K, Wallin A, Andreasson U. Cerebrospinal fluid matrix metalloproteinases and tissue inhibitor of metalloproteinases in combination with subcortical and cortical biomarkers in vascular dementia and Alzheimer's disease. *J Alzheimers Dis*. 2011;27(3):665-676. doi:10.3233/JAD-2011-110566
32. Bjerke M, Jonsson M, Nordlund A, et al. Cerebrovascular biomarker profile is related to white matter disease and ventricular dilation in a LADIS substudy. *Dement Geriatr Cogn Dis Extra*. 2014;4(3):385-394. doi:10.1159/000366119
33. Jonsson M, Zetterberg H, Rolstad S, et al. Low cerebrospinal fluid sulfatide predicts progression of white matter lesions: the LADIS study. *Dement Geriatr Cogn Disord*. 2012;34(1):61-67. doi:10.1159/000341576
34. Bruno D, Pomara N, Nierenberg J, et al. Levels of cerebrospinal fluid neurofilament light protein in healthy elderly vary as a function of TOMM40 variants. *Exp Gerontol*. 2012;47(5):347-352. doi:10.1016/j.exger.2011.09.008
35. Burman J, Zetterberg H, Fransson M, Loskog AS, Raininko R, Fagius J. Assessing tissue damage in multiple sclerosis: a biomarker approach. *Acta Neurol Scand*. 2014;130(2):81-89. doi:10.1111/ane.12239
36. Fialová L, Bartos A, Švarcová J, Zimová D, Kotoučová J. Serum and cerebrospinal fluid heavy neurofilaments and antibodies against them in early multiple sclerosis. *J Neuroimmunol*. 2013;259(1-2):81-87. doi:10.1016/j.jneuroim.2013.03.009
37. Fialová L, Bartos A, Švarcová J. Neurofilaments and tau proteins in cerebrospinal fluid and serum in dementias and neuroinflammation. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub*. 2017;161(3):286-295. doi:10.5507/bp.2017.038
38. Gunnarsson M, Malmeström C, Axelsson M, et al. Axonal damage in relapsing multiple sclerosis is markedly reduced by natalizumab. *Ann Neurol*. 2011;69(1):83-89. doi:10.1002/ana.22247
39. Hall S, Öhrfelt A, Constantinescu R, et al. Accuracy of a panel of 5 cerebrospinal fluid biomarkers in the differential diagnosis of patients with dementia and/or parkinsonian disorders. *Arch Neurol*. 2012;69(11):1445-1452. doi:10.1001/archneurol.2012.1654
40. Hall S, Surova Y, Öhrfelt A, Zetterberg H, Lindqvist D, Hansson O. CSF biomarkers and clinical progression of Parkinson disease. *Neurology*. 2015;84(1):57-63. doi:10.1212/WNL.0000000000001098
41. Herbert MK, Aerts MB, Beenes M, et al. CSF neurofilament light chain but not FLT3 ligand discriminates parkinsonian disorders. *Front Neurol*. 2015;6(May):91. doi:10.3389/fneur.2015.00091
42. Hjalmarsson C, Bjerke M, Andersson B, et al. Neuronal and glia-related biomarkers in cerebrospinal fluid of patients with acute ischemic stroke. *J Cent Nerv Syst Dis*. 2014;6:51-58. doi:10.4137/JCNSD.S13821
43. Jakobsson J, Bjerke M, Ekman CJ, et al. Elevated concentrations of neurofilament light chain in the cerebrospinal fluid of bipolar disorder patients. *Neuropsychopharmacology*. 2014;39(10):2349-2356. doi:10.1038/npp.2014.81
44. Rolstad S, Jakobsson J, Sellgren C, et al. Cognitive performance and cerebrospinal fluid biomarkers of neurodegeneration: a study of patients with bipolar disorder and healthy controls. *PLoS One*. 2015;10(5):e0127100. doi:10.1371/journal.pone.0127100
45. Jeppsson A, Zetterberg H, Blennow K, Wikkelso C. Idiopathic normal-pressure hydrocephalus: pathophysiology and diagnosis by CSF biomarkers. *Neurology*. 2013;80(15):1385-1392. doi:10.1212/WNL.0b013e31828c2fda
46. Jessen Krut J, Mellberg T, Price RW, et al. Biomarker evidence of axonal injury in neuroasymptomatic HIV-1 patients. *PLoS One*. 2014;9(2):e88591. doi:10.1371/journal.pone.0088591
47. Aeinehband S, Lindblom RPF, Al Nimer F, et al. Complement component C3 and butyrylcholinesterase activity are associated with neurodegeneration and clinical disability in multiple sclerosis. *PLoS One*. 2015;10(4):e0122048. doi:10.1371/journal.pone.0122048
48. Khalil M, Enzinger C, Langkammer C, et al. CSF neurofilament and N-acetylaspartate related brain changes in clinically isolated syndrome. *Mult Scler*. 2013;19(4):436-442. doi:10.1177/1352458512458010
49. Kuhle J, Plattner K, Bestwick JP, et al. A comparative study of CSF neurofilament light and heavy chain protein in MS. *Mult Scler*. 2013;19(12):1597-1603. doi:10.1177/1352458513482374
50. Kuhle J, Malmeström C, Axelsson M, et al. Neurofilament light and heavy subunits compared as therapeutic biomarkers in multiple sclerosis. *Acta Neurol Scand*. 2013;128(6):e33-e36. doi:10.1111/ane.12151
51. Kuhle J, Disanto G, Lorscheider J, et al. Fingolimod and CSF neurofilament light chain levels in relapsing-remitting multiple sclerosis. *Neurology*. 2015;84(16):1639-1643. doi:10.1212/WNL.0000000000001491
52. Magdalinou NK, Paterson RW, Schott JM, et al. A panel of nine cerebrospinal fluid biomarkers may identify patients with atypical parkinsonian syndromes. *J Neurol Neurosurg Psychiatry*. 2015;86(11):1240-1247. doi:10.1136/jnnp-2014-309562
53. Martínez MAM, Olsson B, Bau L, et al. Glial and neuronal markers in cerebrospinal fluid predict progression in multiple sclerosis. *Mult Scler*. 2015;21(5):550-561. doi:10.1177/1352458514549397
54. Meeter LH, Doppler EG, Jiskoot LC, et al. Neurofilament light chain: a biomarker for genetic frontotemporal dementia. *Ann Clin Transl Neurol*. 2016;3(8):623-636. doi:10.1002/acn3.325
55. Menke RA, Gray E, Lu CH, et al. CSF neurofilament light chain reflects corticospinal tract degeneration in ALS. *Ann Clin Transl Neurol*. 2015;2(7):748-755. doi:10.1002/acn3.212
56. Lu CH, Macdonald-Wallis C, Gray E, et al. Neurofilament light chain: a prognostic biomarker in amyotrophic lateral sclerosis [published correction appears in *Neurology*. 2015;85(10):921]. *Neurology*. 2015;84(22):2247-2257. doi:10.1212/WNL.0000000000001642
57. Modvig S, Degen M, Horwitz H, et al. Relationship between cerebrospinal fluid biomarkers for inflammation, demyelination and neurodegeneration in acute optic neuritis. *PLoS One*. 2013;8(10):e77163. doi:10.1371/journal.pone.0077163
58. Modvig S, Degen M, Sander B, et al. Cerebrospinal fluid neurofilament light chain levels predict visual outcome after optic neuritis. *Mult Scler*. 2016;22(5):590-598. doi:10.1177/1352458515599074
59. Modvig S, Degen M, Roed H, et al. Cerebrospinal fluid levels of chitinase 3-like 1 and neurofilament light chain predict multiple sclerosis development and disability after optic neuritis. *Mult Scler*. 2015;21(14):1761-1770. doi:10.1177/1352458515574448
60. Paterson RW, Toombs J, Slattery CF, et al. Dissecting IWG-2 typical and atypical Alzheimer's disease: insights from cerebrospinal fluid analysis. *J Neurol*. 2015;262(12):2722-2730. doi:10.1007/s00415-015-7904-3
61. Pérez-Santiago J, Schrier RD, de Oliveira MF, et al. Cell-free mitochondrial DNA in CSF is associated with early viral rebound, inflammation, and severity of neurocognitive deficits in HIV infection. *J Neurovirol*. 2016;22(2):191-200. doi:10.1007/s13365-015-0384-5
62. Pijnenburg YA, Verwey NA, van der Flier WM, Scheltens P, Teunissen CE. Discriminative and prognostic potential of cerebrospinal fluid phosphoTau/tau ratio and neurofilaments for frontotemporal dementia subtypes. *Alzheimers Dement (Amst)*. 2015;1(4):505-512. doi:10.1016/j.dadm.2015.11.001
63. Pykkö OT, Lumela M, Rummukainen J, et al. Cerebrospinal fluid biomarker and brain biopsy findings in idiopathic normal pressure hydrocephalus. *PLoS One*. 2014;9(3):e91974. doi:10.1371/journal.pone.0091974

64. Ragnarsson O, Berglund P, Eder DN, et al. Neurodegenerative and inflammatory biomarkers in cerebrospinal fluid in patients with Cushing's syndrome in remission. *Eur J Endocrinol*. 2013;169(2):211-215. doi:10.1530/EJE-13-0205
65. Romme Christensen J, Ratzer R, Börnsen L, et al. Natalizumab in progressive MS: results of an open-label, phase 2A, proof-of-concept trial. *Neurology*. 2014;82(17):1499-1507. doi:10.1212/WNL.0000000000000361
66. Rosén C, Rosén H, Andreasson U, et al. Cerebrospinal fluid biomarkers in cardiac arrest survivors. *Resuscitation*. 2014;85(2):227-232. doi:10.1016/j.resuscitation.2013.10.032
67. Sandberg L, Biström M, Salzer J, Vågberg M, Svenningsson A, Sundström P. Vitamin D and axonal injury in multiple sclerosis. *Mult Scler*. 2016;22(8):1027-1031. doi:10.1177/1352458515606986
68. Scherling CS, Hall T, Berisha F, et al. Cerebrospinal fluid neurofilament concentration reflects disease severity in frontotemporal degeneration. *Ann Neurol*. 2014;75(1):116-126. doi:10.1002/ana.24052
69. Skillbäck T, Farahmand B, Bartlett JW, et al. CSF neurofilament light differs in neurodegenerative diseases and predicts severity and survival. *Neurology*. 2014;83(21):1945-1953. doi:10.1212/WNL.0000000000001015
70. Stilund M, Gjelstrup MC, Petersen T, Møller HJ, Rasmussen PV, Christensen T. Biomarkers of inflammation and axonal degeneration/damage in patients with newly diagnosed multiple sclerosis: contributions of the soluble CD163 CSF/serum ratio to a biomarker panel. *PLoS One*. 2015;10(4):e0119681. doi:10.1371/journal.pone.0119681
71. Tortelli R, Copetti M, Ruggieri M, et al. Cerebrospinal fluid neurofilament light chain levels: marker of progression to generalized amyotrophic lateral sclerosis. *Eur J Neurol*. 2015;22(1):215-218. doi:10.1111/ene.12421
72. Tortelli R, Ruggieri M, Cortese R, et al. Elevated cerebrospinal fluid neurofilament light levels in patients with amyotrophic lateral sclerosis: a possible marker of disease severity and progression. *Eur J Neurol*. 2012;19(12):1561-1567. doi:10.1111/j.1468-1331.2012.03777.x
73. Tortorella C, Drenzo V, Taurisano P, et al. Cerebrospinal fluid neurofilament tracks fMRI correlates of attention at the first attack of multiple sclerosis. *Mult Scler*. 2015;21(4):396-401. doi:10.1177/1352458514546789
74. Trentini A, Comabella M, Tintoré M, et al. N-acetylaspartate and neurofilaments as biomarkers of axonal damage in patients with progressive forms of multiple sclerosis. *J Neurol*. 2014;261(12):2338-2343. doi:10.1007/s00415-014-7507-4
75. Vågberg M, Norgren N, Dring A, et al. Levels and age dependency of neurofilament light and glial fibrillary acidic protein in healthy individuals and their relation to the brain parenchymal fraction. *PLoS One*. 2015;10(8):e0135886. doi:10.1371/journal.pone.0135886
76. Villar LM, Picón C, Costa-Frossard L, et al. Cerebrospinal fluid immunological biomarkers associated with axonal damage in multiple sclerosis. *Eur J Neurol*. 2015;22(8):1169-1175. doi:10.1111/ene.12579
77. Wild EJ, Boggio R, Langbehn D, et al. Quantification of mutant huntingtin protein in cerebrospinal fluid from Huntington's disease patients. *J Clin Invest*. 2015;125(5):1979-1986. doi:10.1172/JCI80743
78. Zetterberg H, Skillbäck T, Mattsson N, et al; Alzheimer's Disease Neuroimaging Initiative. Association of cerebrospinal fluid neurofilament light concentration with Alzheimer disease progression. *JAMA Neurol*. 2016;73(1):60-67. doi:10.1001/jamaneurol.2015.3037
79. Pakkenberg B, Gundersen HJ. Neocortical neuron number in humans: effect of sex and age. *J Comp Neurol*. 1997;384(2):312-320. doi:10.1002/(SICI)1096-9861(19970728)384:2<312::AID-CNE10>3.0.CO;2-K
80. Cohen JA, Reingold SC, Polman CH, Wolinsky JS; International Advisory Committee on Clinical Trials in Multiple Sclerosis. Disability outcome measures in multiple sclerosis clinical trials: current status and future prospects. *Lancet Neurol*. 2012;11(5):467-476. doi:10.1016/S1474-4422(12)70059-5
81. Stadelmann C, Wegner C, Brück W. Inflammation, demyelination, and degeneration - recent insights from MS pathology. *Biochim Biophys Acta*. 2011;1812(2):275-282. doi:10.1016/j.bbdis.2010.07.007
82. Gaiottino J, Norgren N, Dobson R, et al. Increased neurofilament light chain blood levels in neurodegenerative neurological diseases. *PLoS One*. 2013;8(9):e75091. doi:10.1371/journal.pone.0075091
83. Tortorella C, Drenzo V, Ruggieri M, et al. Cerebrospinal fluid neurofilament light levels mark grey matter volume in clinically isolated syndrome suggestive of multiple sclerosis [published correction appears in *Mult Scler*. 2019;25(2):302]. *Mult Scler J*. 2018;24(8):1039-1045.
84. De Stefano N, Matthews PM, Filippi M, et al. Evidence of early cortical atrophy in MS: relevance to white matter changes and disability. *Neurology*. 2003;60(7):1157-1162. doi:10.1212/01.WNL.0000055926.69643.03
85. Malmström C, Haghighi S, Rosengren L, Andersen O, Lycke J. Neurofilament light protein and glial fibrillary acidic protein as biological markers in MS. *Neurology*. 2003;61(12):1720-1725. doi:10.1212/01.WNL.0000098880.19793.B6
86. Norgren N, Sundström P, Svenningsson A, Rosengren L, Stigbrand T, Gunnarsson M. Neurofilament and glial fibrillary acidic protein in multiple sclerosis. *Neurology*. 2004;63(9):1586-1590. doi:10.1212/01.WNL.0000142988.49341.D1
87. Giovannoni G, Nath A. After the storm: neurofilament levels as a surrogate endpoint for neuroaxonal damage. *Neurology*. 2011;76(14):1200-1201. doi:10.1212/WNL.Ob013e3182143345
88. Alcolea D, Vilaplana E, Suárez-Calvet M, et al. CSF sAPPβ, YKL-40, and neurofilament light in frontotemporal lobar degeneration. *Neurology*. 2017;89(2):178-188. doi:10.1212/WNL.00000000000004088
89. Reijn TS, Abdo WF, Schelhaas HJ, Verbeek MM. CSF neurofilament protein analysis in the differential diagnosis of ALS. *J Neurol*. 2009;256(4):615-619. doi:10.1007/s00415-009-0131-z
90. Hu X, Yang Y, Gong D. Cerebrospinal fluid levels of neurofilament light chain in multiple system atrophy relative to Parkinson's disease: a meta-analysis. *Neurol Sci*. 2017;38(3):407-414. doi:10.1007/s10072-016-2783-7
91. Soylu-Kucharz R, Sandelius Å, Sjögren M, et al. Neurofilament light protein in CSF and blood is associated with neurodegeneration and disease severity in Huntington's disease R6/2 mice. *Sci Rep*. 2017;7(1):14114. doi:10.1038/s41598-017-14179-1
92. Bergman J, Dring A, Zetterberg H, et al. Neurofilament light in CSF and serum is a sensitive marker for axonal white matter injury in MS. *Neurol Neuroimmunol Neuroinflamm*. 2016;3(5):e271. doi:10.1212/NXI.0000000000000271
93. Teunissen CE, Elias N, Koel-Simmelink MJ, et al. Novel diagnostic cerebrospinal fluid biomarkers for pathologic subtypes of frontotemporal dementia identified by proteomics. *Alzheimers Dement (Amst)*. 2016;2:86-94. doi:10.1016/j.dadm.2015.12.004
94. Hughes AJ, Daniel SE, Ben-Shlomo Y, Lees AJ. The accuracy of diagnosis of parkinsonian syndromes in a specialist movement disorder service. *Brain*. 2002;125(pt 4):861-870. doi:10.1093/brain/awf080
95. Plassman BL, Khachaturian AS, Townsend JJ, et al. Comparison of clinical and neuropathologic diagnoses of Alzheimer's disease in 3 epidemiologic samples. *Alzheimers Dement*. 2006;2(1):2-11. doi:10.1016/j.jalz.2005.11.001
96. Visser PJ, Vos S, van Rossum I, Scheltens P. Comparison of International Working Group criteria and National Institute on Aging-Alzheimer's Association criteria for Alzheimer's disease. *Alzheimers Dement*. 2012;8(6):560-563. doi:10.1016/j.jalz.2011.10.008
97. Pohjasvaara T, Mäntylä R, Ylikoski R, Kaste M, Erkinjuntti T; National Institute of Neurological Disorders and Stroke-Association Internationale pour la Recherche et l'Enseignement en Neurosciences. Comparison of different clinical criteria (DSM-III, ADDTC, ICD-10, NINDS-AIREN, DSM-IV) for the diagnosis of vascular dementia. *Stroke*. 2000;31(12):2952-2957. doi:10.1161/01.STR.31.12.2952