R package rodeoFABM: Basic Use and Sample Applications

johannes.feldbauer@tu-dresden.de

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1 Main features of rodeoFABM

The R package rodeoFABM is a colletion of functions to help create water quality models that can be coupled to physical host models using the FABM interface (Bruggeman and Bolding (2014)). As the name suggests it is heaviely influenced by the R package rodeo (Kneis, Petzoldt, and Berendonk (2017)). The principle idea is to creat tools that:

- Decouple the "code writing" from the "model development" part of creating a model
- Make model adaptation, communikation, and maintenace easier

The main concept is writing the model equations in the standard Peterson matrix notation and storing them in text files or spread sheets. The package rodeoFABM then can be used to automatically generate FABM specific FORTRAN code from these files, create .yaml control files for the water quality model, automatically compile the 1D physical lake model GOTM (Burchard et al. (2006)) coupled with the model, and generate LATEX documentation of the model.

2 Installation and requirements

So far rodeoFABM was only tested with different Linux based operating systems. In order to use rodeoFABM and run the examples some software tools are neede:

- The GNU compilers ([gcc.gnu.org]{https://gcc.gnu.org})
- GNU Make ([gnu.org/software/make/]{https://gnu.org/software/make/})
- GNU CMake ([cmake.org]{https://cmake.org})
- R and devtools (R-project.org)
- git (git-scm.com)
- the netcdf libraries
- R packages: readODS, plot3D, ncdf4, reshape2, and RColorBrewer

The package rodeoFABM can be installed from github using:

```
library("devtools")
install_github("JFeldbauer/rodeoFABM")
```

3 Basic use

3.1 First example

To demonstrate the workflow we will create and compile a simple model. The files used in this example are contained in the package and can be copied to the current working directory using:

```
# copy example ods file
example_model <- system.file("extdata/simple_model.ods", package = "rodeoFABM")
file.copy(from = example_model, to = ".", recursive = TRUE)</pre>
```

This will copy the Libre Office spread sheet $simple_model.ods$ to your current working directory. Now we can read in the tables with the declarations of state variables, model parameters, used functions and external dependencies, process rate descriptions, and stoichiometry matrix.

```
library(readODS)

# read in example ods file
odf_file <- "simple_model.ods"
vars <- read_ods(odf_file, sheet = 1)
pars <- read_ods(odf_file, sheet = 2)
funs <- read_ods(odf_file, sheet = 3)
pros <- read_ods(odf_file, sheet = 4)
stoi <- read_ods(odf_file, sheet = 5)</pre>
```

We store the declarations in the five data frames vars, pars, funs, pros, and stoi. Using these we can now generate FORTRAN source files using the function gen_fabm_code()

```
library(rodeoFABM)

# generate fabm code
gen_fabm_code(vars,pars,funs,pros,stoi,"simple_model.f90",diags = TRUE)
```

This will create two new files: the FABM specific FORTRAN source code simple_model.f90 and the control file fabm.yaml which can be used to change model parameters and initial conditions. The function also checks if all parameter, functions, and state variables used are also decalred and issues a warning because the units decalred for the parameters are not in seconds, which is required by FABM.

Using the source code file *simple_model.f90* we can compile *GOTM-FABM*. Therfore we first need to clone the lake branche of *GOTM* from github and prepare the build process. This can automatically be done using

the function clone_GOTM():

```
# clone github repo
clone_GOTM(build_dir = "build", src_dir = "gotm_src")
```

This will take a moment and download the source code for GOTM and FABM as well as prepare the compilation using CMake. You can see that there are now two new folders in the working directory called $gotm_src$ and build. Now we can build GOTM-FABM with our simple model using the $simple_model.f90$ file and the build_gotm() function:

This will copy the *simple_model.f90* file we just created to the correct folder within the *gotm_src* folder and then compile *GOMT-FABM* using *make*. As a last step it will copy the created executable to the current working directory. You can see that there is now a *gotm* executable file in the working directory. In order to run our created model we will need a *gotm.yaml* file (the GOTM controll fille), an hypsograph file, and the meteorological forcing data. We will copy the example files provides in this package by using:

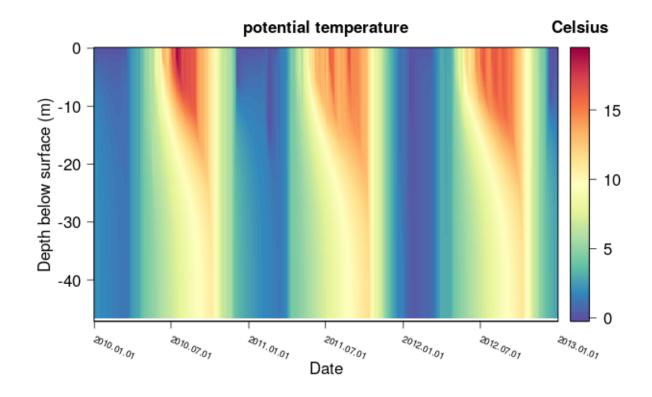
Now that we have the files *gotm.yaml*, *hypsograph.dat*, and *meteo_file.dat* in our working directory, we can run *GOTM-FABM* using:

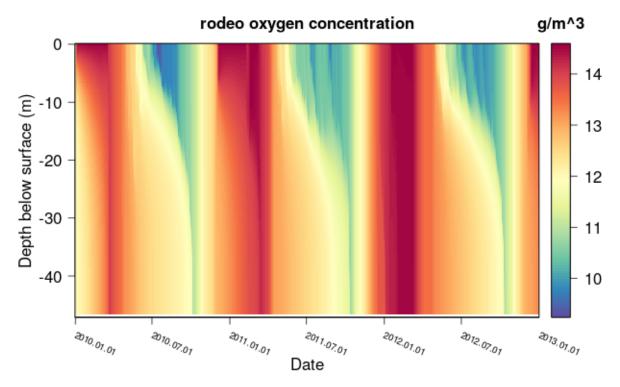
```
# run gotm
system2("./gotm")
```

After successfully running there are two new files: output.nc and restart.nc, both are netcdf files. output.nc is the output of the model run and restart.nc is a netcdf file that can be used to initialize a simulation with states stored from the previous run. We can plot the results e.g. by using the plot_var() function:

```
# plot temperature
plot_var("output.nc", "temp")

# plot oxygen
plot_var("output.nc", "rodeo_C_02")
```





These are the essential steps used to build and run a *GOTM-FABM* model. In the next section we will go a little bit more into the details of building a own model using the **rodeoFABM** package and explain them step by step.

3.2 Create a model step by step

In order to demonstrate the necessary steps and functionalities we will create a simple phytoplankton-nutrients model and progressively add more processes and state variables.

All used Libre Office spread sheets containing the model information, the GOTM controll file, and the forcing data are contained in the rodeoFABM package. You can copy all necessary files to run GOTM using the same method as described in the first example. We will use the same meteorological forcing (meteo_file.dat) and hypsographic curve (hypsograph.dat) as in the first example. Additionally we now add one inflow and one outflow (files inflow_m.dat, outflow.dat, and inflow_wq_m.dat containing the nutrient concentrations of the inflow)

```
## copy necessary files
# GOTM controll fille
yaml <- system.file("extdata/examples/gotm.yaml",</pre>
                   package = "rodeoFABM")
file.copy(from = yaml, to = ".", recursive = TRUE)
# inflow hydrological data
infl <- system.file("extdata/examples/inflow_m.dat",</pre>
                   package = "rodeoFABM")
file.copy(from = infl, to = ".", recursive = TRUE)
# inflow nutient data
nut <- system.file("extdata/examples/inflow_wq_m.dat",</pre>
                   package = "rodeoFABM")
file.copy(from = nut, to = ".", recursive = TRUE)
# outflow data
out <- system.file("extdata/examples/outflow.dat",</pre>
                   package = "rodeoFABM")
file.copy(from = out, to = ".", recursive = TRUE)
```

The inflows and especially the inflow of state variables to a *FABM* model are defined in the **streams** section of the *GOTM* control file (*gotm.yaml*). The section looks like this:

```
streams:
   inflow:
                                               # stream configuration
                                                 # inflow method, default=1
      method: 4
      zu: 0.0
                                                 # upper limit m
      zl: 0.0
                                                 # lower limit m
      flow:
                                                 # water flow
                                                 # 0=constant, 2=from file, default = 0
         method: 2
                                                 # constant value( m^3/s)
         constant_value: 1.0
         file: inflow_m.dat
                                                 # path to file with time series
                                                 # index of column to read from
         column: 1
                                                 # flow temperature
      temp:
                                                 # O=constant, 2=from file; default=0
         method: 2
         constant_value: 10.0
                                                 # constant value (°C)
                                                 # path to file with time series
         file: inflow_m.dat
         column: 2
                                                 # index of column to read from
      salt:
                                                 # flow salinity
                                                 # O=constant, 2=from file; default=0
         method: 0
         constant_value: -1.0
                                                 # constant value (PSU)
         file: inflow.dat
                                                 # path to file with time series
         column: 3
                                                 # index of column to read from
      rodeo_HPO4:
                                                 # rodeo phosphprus
         method: 0
                                                 # O=constant, 2=from file; default=0
```

```
constant_value: 0.5  # constant value (gP/m^3)

file: inflow_wq_m.dat  # path to file with time series

column: 4  # index of column to read from
```

Within the streams section several in- and outflows can be defined with any desired name (here "inflow"). The inflow/outflow depth is defined by streams/method, whereas 1 means surface, 2 means bottom, 3 means a specified range of depths defined by streams/zu (upper) and streams/zl (l), and 4 means inflow to the depth with same temperature as the inflow temperature. Every in- or outflow needs the streams/flow section defining the flow rate in m³/s and can have additional entries like streams/temp for temperature or inflowing state variables of the FABM model (like streams/rodeo_HPO4). The FABM sate variables need to start with rodeo_followd by the defined state variable name. The values can either be constant (streams/rodeo_HPO4/method = 0) or a time series given by a tab separated file (streams/rodeo_HPO4/method = 2) with first column datetime (as YYYY-mm-dd HH:MM:ss). The name of the file is supplied by streams/rodeo_HPO4/file and the column the variable is in by streams/rodeo_HPO4/column, take care: the first column with datetime is not counted and if the columns have a header it needs to start with an excalamation mark "!".

We can create the source code of the phytoplankton nutients model in the same way as we created the source code in the first example:

This first model is a simple model with two state variables, which are declared in the **vars** data frame. The table needs to have at least three columns: *name* giving the identifier of the state variable, *unit* giving the used unit, and *description* giving a short description of the state variable. If additionally the column *default* is supplied the initial value will be included in the *FABM* control file (fabm.yaml), which is automatically generated by gen_fabm_code().

Table 1: Data frame vars: Declaration of state variables.

name	unit	description	default
\overline{C}	gDM/m ³	algae concentration	0.0
HPO4	gP/m^3	phosphorus concentration	0.1

The models parameters are defined in the pars data frame in a similar fashion. They need the same three columns *name*, *unit*, and *description* and can have the aditional column *default* as well. Take care that *FABM* requires all parameters with relation to time to be in units of second.

Table 2: Data frame pars: Declaration of model parameters.

name	unit	description	default
$\overline{mu_max}$	1/s	maximum growth rate	1e-05
K_P	W/m^2	half saturation concentration of HPO4 limitation	2e-02
k_death	1/s	death rate	2e-06

name	unit	description	default
a_P	$\mathrm{gP/gDM}$	phosphorus content of phytoplankton	5e-02

External functions, or forcing data that needs to be obtained from the physical host model (e.g. water temperature) are defined in funs. As this first model has no such functions or dependencies this is explaine in the later steps. As in this example the data frame is not needed it hast to be set to NULL.

The declaration of the processes and process rates is done in the pros data frame. It has four required columns: name giving the name of the process, unit giving the unit of the process rate (again in seconds), description giving a short description of the process, and expression giving the mathematical expression of the process. There can be additional columns to define the saptial domain of the process, or to declare sinking processes and they will be explained later.

Table 3: Data frame pros: Declaration of processes.

name	unit	description	expression
growth death		growth of algae death of algae	$\begin{array}{c} C \cdot mu_max \cdot HPO4/(HPO4 + K_P) \\ C \cdot k_death \end{array}$

In this model the phytoplankton have a simple linear growth term with a Monod like limitation for the limiting nutrient Phosphorus and a linear decay/death term.

The last data frame stoi gives the stoichiometry table (in long format) connecting the process rates with the state variables. It has three required columns: *variable* giving the variable affected by the *process*, and *expression* giving a factor to multiply the process rate by:

Table 4: Data frame stoi: Declaration of stoichiometry matrix in long format.

variable	process	expression
\overline{C}	growth	1
C	death	-1
HPO4	growth	$-1 \cdot a_P$

The growth of phytoplankton is increasing its concentration C and decreasing the nutrient HPO4 by the fraction of a_P , which is the Phosphorus conten of the phytoplankton. Decay/death is decreasing phytoplankton concentration C.

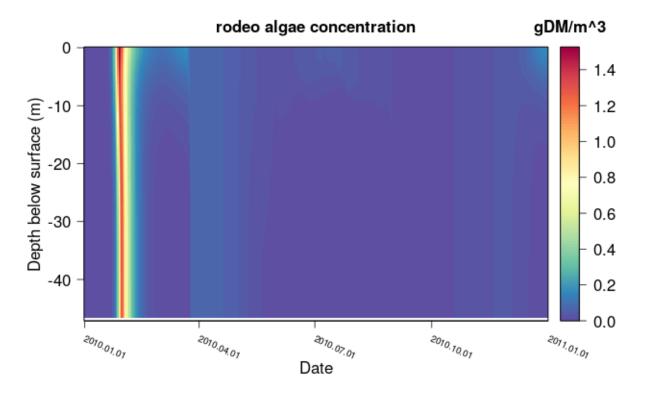
Having declared all five data frames we can now generate the fortran code using <code>gen_fabm_code()</code>. This will also perfome some automated checks e.g. if all used parameters and state variables are also declared, and will issue a warning if the used units are not using seconds for time. It will also create the <code>FABM</code> control file <code>fabm.yaml</code> and insert the default values for parameters and initial values (if declared). If the argument <code>diags</code> is set to <code>TRUE</code> the process rates are stored as diagnostic variables in the output netcdf file.

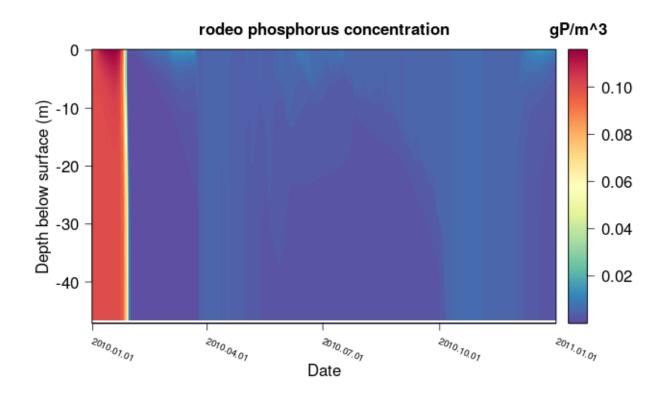
```
# create the fabm code
gen_fabm_code(vars, pars, funs, pros, stoi, "model_1.f90", diags = TRUE)
```

After creating the fortran source code, *GOTM-FABM* can be automatically compiled unsing the function build_GOTM() (assuming the source code was allready fetched and prepared for compilation using clone_GOTM()), this will also copy the compiled executable to the current working directory, which then can be ran using e.g. system2("./gotm").

We can now plot the model results e.g. using the plot_var() function:

```
# plot the variables
plot_var("output.nc", "rodeo_C")
plot_var("output.nc", "rodeo_HPO4")
```





3.2.1 Getting dependencies from the host model

As many biogeochemical processes depend on external forcing, such as temperature or available iradiation, these values can be obtained from the physical host model. In the next step we want to add the dependency of phytoplankton growth on available iradiation. We first copy the prepared spread sheet and declare the data frames:

We need to get values for the photosynthetic active radiation (PAR) from GOTM. FABM has so called "standard-variables" with defined names (stored in the std_names_FABM data). If you want to access these variables you need to defind them as a function in the funs data frame and add the additional column dependency wich contains the full standard-variable name. The data frame funs has three required columns that are the same as in vars, and pars: name, unit, and description, additionally the column dependency. If you declare several functions of whome some are not dependencies the corresponding entry in column dependency needs to be empty (NA) for these and the corresponding standard-name for the ones that are dependencies.

Table 5: Data frame funs: Declaration of model functions and dependencies from the host model.

name	unit	description	dependency
\overline{par}	W/m^2	Downwelling photosynthetic radiative flux	$downwelling_photosynthetic_radiative_flux$

The declared functions/dependencies can now be used in the process expression, same as parameters and state variables. We added a Monod Term for light limitation in the *growth* process:

Table 6: Data frame pros: Declaration of processes.

name	unit	description	expression
		growth of algae death of algae	$\begin{array}{c} C \cdot mu_max \cdot HPO4/(HPO4 + K_P) \cdot par/(par + K_par) \\ C \cdot k_death \end{array}$

For this we need to declare the additional parameter K_par for the half-saturation irradiation in the pars data frame:

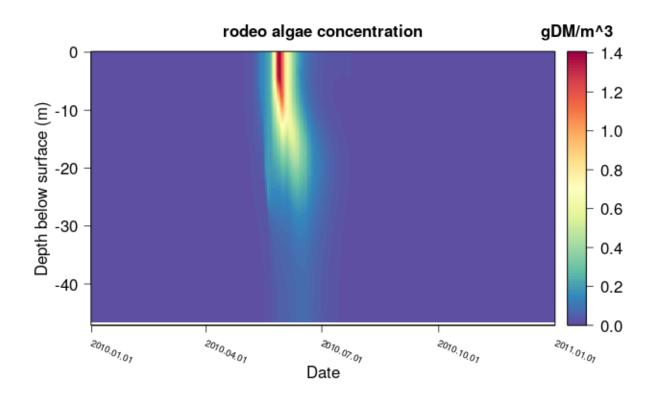
Table 7: Data frame pars: Declaration of model parameters.

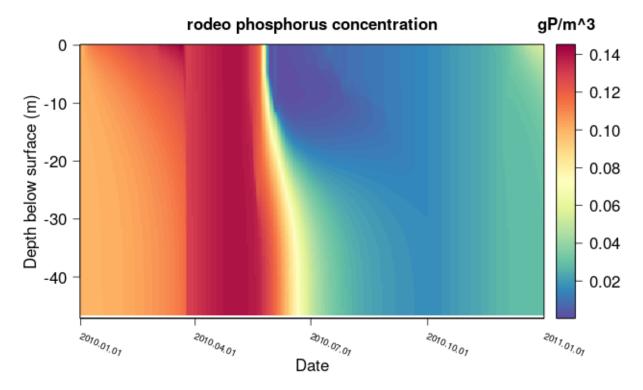
name	unit	description	default
$\overline{mu_max}$	1/s	maximum growth rate	1.0e-05
K_P	W/m^2	half saturation of photosyntetic flux	2.0e-02
k_death	1/s	death rate	2.0e-06
a_P	gP/gDM	phosphorus content of phytoplankton	5.0e-02
K_par	W/m^2	half saturation of photosyntetic flux	2.7e + 01

Now we can generate the fortran code, compile GOTM-FABM, and run the adapted model.

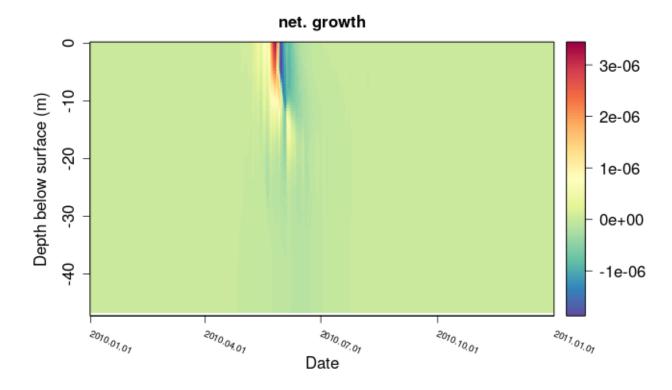
And plot some of the simulated state variables:

```
# plot the variables
plot_var("output.nc", "rodeo_C")
plot_var("output.nc", "rodeo_HPO4")
```





From the saved process rates (the diagnostic variables) we can e.g. plot the net growth rate. We can acces the values stored in the netcdf file e.g. by using the function get_var() and plot them using plot3D::image2D(): library(plot3D)



3.2.2 Sedimentation

Often in biogeochemical models some state variables are sinking in the water body (e.g. phytoplankton or particulated organic matter). In the next adaptation of the model we want to include a constant sinking velocity for the phytoplankton. Therefore, we again copy the spread sheets from the package data in order to declare the data frames:

```
stoi <- read_ods("simple_alg_par_sed.ods", sheet = "stoi")</pre>
```

FABM allows for time varying sinking of state variables. This in implemented in *rodeoFABM* as a process declared in the **pros** data frame that has a logical flag set in an additional column called *sedi*. The expression for this can also be a function of external dependencies (e.g. water density) or internal state variables (e.g. nutrient concentration), in this simple case we choose a constant sinking velocity:

Table 8: Data frame pros: Declaration of processes.

name	unit description	expression	sedi
growth	g/m ³ /d growth of	$C \cdot mu_max \cdot HPO4/(HPO4 + K_P) \cdot par/(par + K_par)$	NA
death sed	algae $g/m^3/d$ death of algae $g/m^3/s$ sinking	$C \cdot k_death \\ v_sed$	NA TRUE

For this to work we need to declare the additional parameter for the sinking velocity:

Table 9: Data frame pars: Declaration of model parameters.

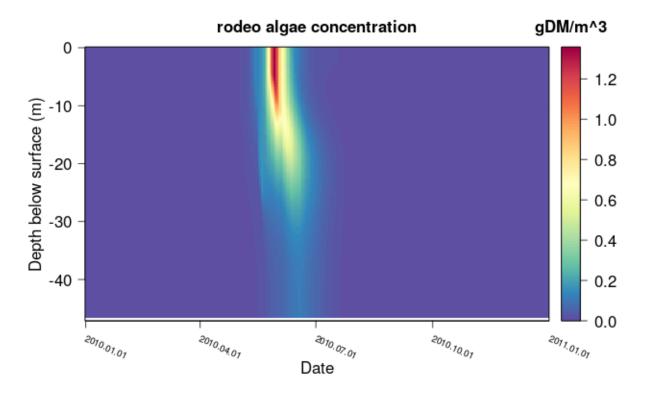
name	unit	description	default
$\overline{mu_max}$	1/s	maximum growth rate	1.0e-05
K_P	W/m^2	half saturation of photosyntetic flux	2.0e-02
k_death	1/s	death rate	2.0e-06
a_P	gP/gDM	phosphorus content of phytoplankton	5.0e-02
K_par	W/m^2	half saturation of photosyntetic flux	2.7e + 01
v_sed	m/s	sedimentation velocity	1.0e-06

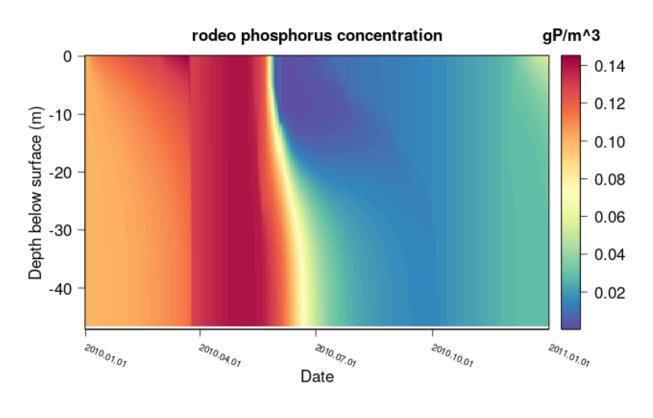
And add the process to the stopichiometry table:

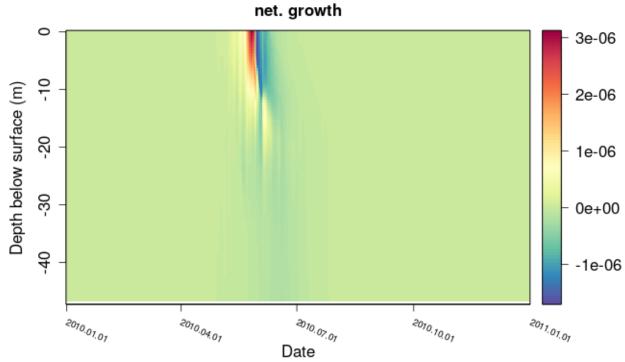
Table 10: Data frame stoi: Declaration of stoichiometry matrix in long format.

variable	process	expression
\overline{C}	growth	1
C	death	-1
C	sed	-1
HPO4	growth	$-1 \cdot a_P$

Now we can create the FORTRAN source code file, compile GOTM-FABM, run the model, and plot some of the results:







3.2.3 Processes at the surface and sediment

There are some processes that only take place at the surface or bottom (sediment) of lakes. FABM knows three spatial domains: open water (pelagial), surface, and bottom (sediment) and processes can be declared

to only take place at one of these domaines. To demonstrate this we add the new state variable Oxygen along with the processes of surface exchange and a constant oxygen consumption in the sediment to the model. We again start by copying the spread sheet from the package data:

Here we added the new state variable O2 to the vars data frame:

Table 11: Data frame vars: Declaration of state variables.

name	unit	description	default
\overline{C}	gDM/m^3	algae concentration	0.0
HPO4	gP/m^3	phosphorus concentration	0.1
O2	gO/m^3	oxygen concentration	10.0

If processes occure only at the surface or bottom interface we can declare this by setting a logical flag in additional columns in the pros data frame called *bot* and *surf*. We have two new processes $O2_exch$, and $O2_cons$ and the flags in the corresponding columns are set to TRUE:

Table 12: Data frame 'pros': Declaration of processes.

name	unit	description	expression	surf	bot	sedi
growth death sed O2_exch O2_cons	g/m^3/d g/m^3/d g/m^3/s g/m^3/d g/m^3/d	growth of algae death of algae sinking exchange of Oxygen at the surface consumption of Oxygen in the pelagial	$\begin{array}{l} C \cdot mu_max \cdot HPO4/(HPO4 + K_P) \cdot par/(par + K_par) \\ C \cdot k_death \\ v_sed \\ v_02 \cdot (exp(7.7117 - 1.31403 \cdot log(Temp + 45.93)) \cdot (p/101325) - O2) \\ O2/(O2 + K_O2) \cdot k_O2_cons \end{array}$	TRUE	TRUE	TRUE

We declared the additional parameters for the oxygen exchange velocity, the constant consumption in the sediment, and the half-saturation concentration of oxygen limiting the oxygen consumption in the sediment:

Table 13: Data frame pars: Declaration of model parameters.

name	unit	description	default
$\overline{mu_max}$	1/s	maximum growth rate	1.0e-05
K_P	\dot{W}/m^2	half saturation of photosyntetic flux	2.0e-02
k_death	1/s	death rate	2.0e-06
a_P	gP/gDM	phosphorus content of phytoplankton	5.0e-02
K_par	W/m^2	half saturation of photosyntetic flux	2.7e + 01
v_sed	m/s	sedimentation velocity	1.0e-06
v_O2	1/s	speed of oxygen transfer	1.0e-05
k_O2_cons	$1/s/m^2$	Oxygen consumption rate in sediment	5.0e-07
K_O2	gO/m^3	half saturation concentration of oxygen consumption	5.0e + 00
<u>a_O</u>	gO/gDM	oxygen production per growth of algae	1.0e+00

We also declared the used functions log, and exp, as well as the external dependencies p (the barometric pressure at the surface), and Temp (water temperature) which are needed to calculate the oxygen saturation

concentration:

Table 14: Data frame funs: Declaration of model functions and dependencies from the host model.

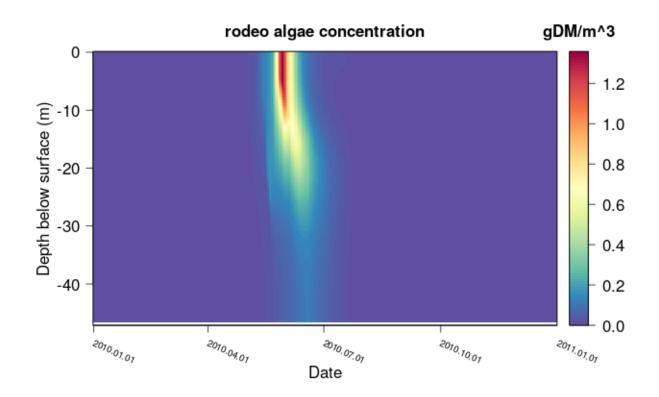
name	unit	description	dependency
\overline{par}	W/m^2	Downwelling photosynthetic radiative flux	$downwelling_photosynthetic_radiative_flux$
$p \ Temp \ exp \ log$	Pa celsius -	Atmospheric Pressure Water temperature exponential function logarithmic function	surface_air_pressure temperature NA NA

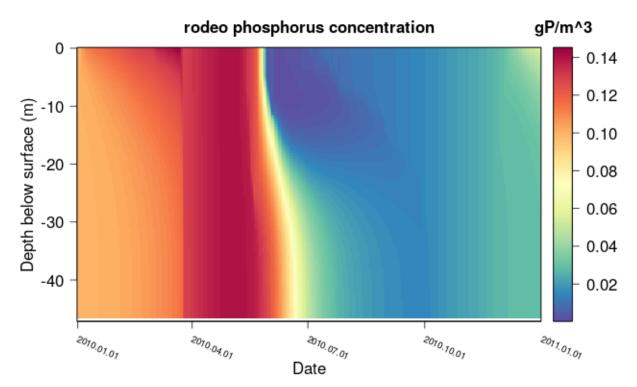
And added the new processes to the stoichiometry table:

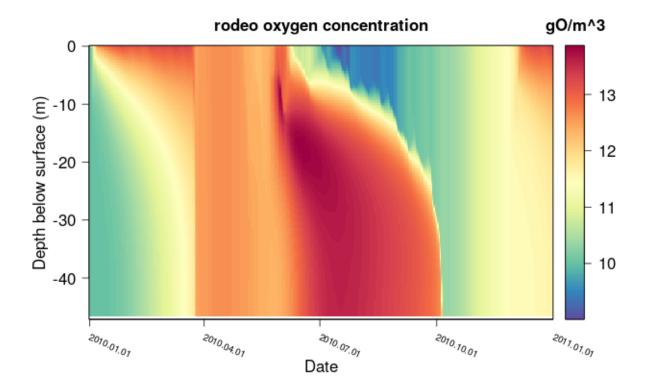
Table 15: Data frame stoi: Declaration of stoichiometry matrix in long format.

variable	process	expression
\overline{C}	growth	1
C	death	-1
C	sed	-1
HPO4	growth	$-1 \cdot a_P$
O2	$O2$ _exch	1
O2	$O2$ _cons	-1
O2	growth	a_O

We can generate the source code, compile GOTM-FABM, run the model, and plot some of the results using:







3.2.4 Sediment or surface attached state variables

As mentioned before FABM recognizes three spatial domains: open water, surface, and sediment. Like Processes, state variables can also be attached to one of these domaines (e.g. sedimented particulated organic matter). To demonstrate this feature we will include two more state variables in our model; particulated organic matter (POM) and sedimented particulated organic matter (SPOM). Again we need to copy the spread sheet from the package:

We added the two new state variables POM and SPOM and declared SPOM as bottom bound state variable by adding another column to the vars data frame called bot and set it to TRUE for all bottom bound state variables and to NA (empty) or FALSE for all others. Surface bound state variables can be declared in the same manner using a column named surf.

Table 16: Data frame vars: Declaration of state variables.

name	unit	description	default	bot
C $HPO4$	gDM/m^3 gP/m^3	algae concentration phosphorus concentration	0.0	NA NA
O2	gC/m^3	oxygen concentration	10.0	NA

name	unit	description	default	bot
\overline{POM}	gDM/m^3	particulated organic matter	0.0	NA
SPOM	gDM/m^2	sedimented particulated organic matter	0.0	TRUE

Looking at the stoichiometry table we can see that the death of algae generates POM, which settles down, and sediments to the ground to become SPOM. Both POM and SPOM are mineralized, releasing HPO4 but the mineralization is faster in the sediment (see pars table). We added the new sinking, sedimentation, and mineralization processes to the pros data frame:

Table 17: Data frame 'pros': Declaration of processes.

name	unit	description	expression	surf	bot	sedi
growth	gDW/m^3/d	growth of algae	$C \cdot mu_max \cdot (HPO4)/(HPO4 + K_P) \cdot (par)/(par + K_par)$			
death	$gDW/m^3/d$	death of algae	$C \cdot k_death$			
sed_ALG	gDW/m^3/s	sinking of algae	v_sed_ALG			TRUE
O2 exch	gO/m^3/d	exchange of Oxygen at the surface	$v = O2 \cdot (exp(7.7117 - 1.31403 \cdot log(Temp + 45.93)) \cdot (p)/(101325) - O2)$	TRUE		
O2 cons	gO/m ³ /d	consumption of Oxygen in the pelagial	$(O2)/(O2 + K O2) \cdot k O2 \ cons$		TRUE	
sed POM	gDW/m^3/s	sinking of POM	v sed POM			TRUE
miner_POM	gDW/m^3/s	mineralization of POM	$\overrightarrow{POM} \cdot k_miner_POM \cdot (O2)/(O2 + K_miner_O2)$			
miner SPOM	gDW/m ³ /s	mineralization of SPOM	$SPOM \cdot k \ miner \ SPOM \cdot (O2)/(O2 + K \ miner \ O2)$		TRUE	
set_POM	gDW/m^3/s	settling of POM	$v_sed_POM \cdot POM$		TRUE	

And we adapted the stoichiometry table:

Table 18: Data frame stoi: Declaration of stoichiometry matrix in long format.

variable	process	expression
\overline{C}	growth	1
C	death	-1
C	sed _ALG	-1
HPO4	growth	$-1 \cdot a_P$
HPO4	miner_POM	a_P
HPO4	$miner_SPOM$	a_P
O2	O2_exch	1
O2	$O2$ _cons	-1
O2	growth	a_O
O2	miner_POM	$-1 \cdot a_miner$
O2	$miner_SPOM$	$-1 \cdot a_miner$
POM	$\operatorname{sed}\operatorname{POM}$	-1
POM	$\operatorname{set}\operatorname{POM}$	-1
POM	death	1
POM	$miner_POM$	-1
SPOM	$\operatorname{set}\operatorname{POM}$	1
SPOM	$\overline{\text{miner}}$ SPOM	-1

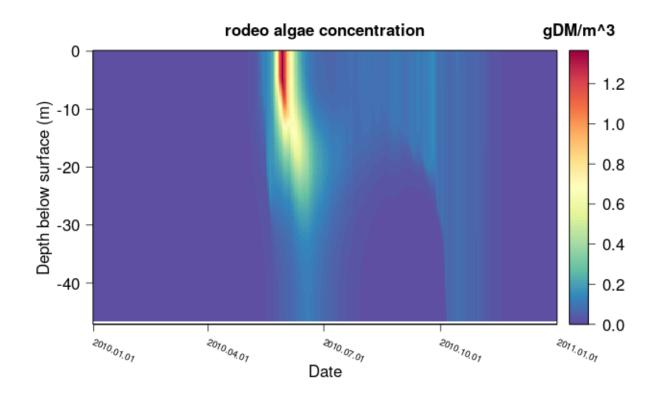
We declared the new parameters for the sinking velocity, the mineralization kinetic, and the half-saturation concentration limiting the mineralization:

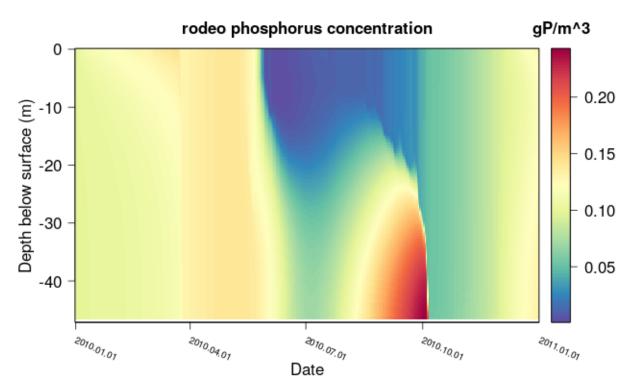
Table 19: Data frame pars: Declaration of model parameters.

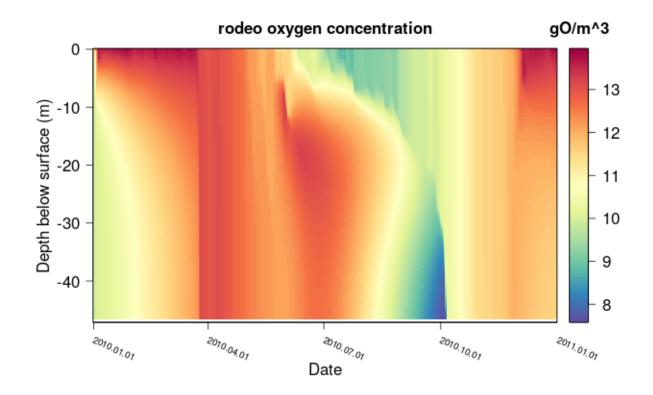
name	unit	description	default
$\overline{mu_max}$	1/s	maximum growth rate	1.0e-05
K_P	W/m^2	half saturation of photosyntetic flux	2.0e-02
k_death	1/s	death rate	2.0e-06
a_P	gP/gDM	phosphorus content of phytoplankton	5.0e-02

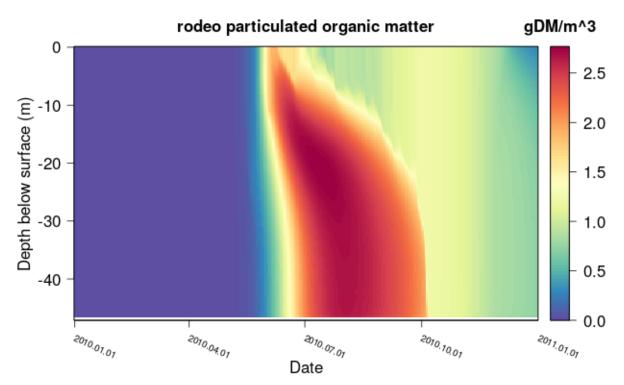
name	unit	description	default
$\overline{K_par}$	W/m^2	half saturation of photosyntetic flux	2.7e+01
v_sed_ALG	m/s	sedimentation velocity of algae	1.0e-06
v_O2	1/s	speed of oxygen transfer	1.0e-04
k_O2_cons	$1/s/m^2$	Oxygen consumption rate in sediment	5.0e-07
K_O2	gO/m^3	half saturation concentration of oxygen consumption	5.0e + 00
a_O	gO/gDM	oxygen production per growth of algae	1.0e+00
v_sed_POM	m/s	sedimentation velocity of POM	2.0e-06
K_miner_O2	gO/m^3	half saturation concentration of oxygen for mineralization	3.0e + 00
k_miner_POM	1/s	maximum mineralization rate of POM	0.0e + 00
k_miner_SPOM	I/s	maximum mineralization rate of SPOM	3.0e-07
a_miner	gO/gDM	oxygen consumption per oxygenation of $\operatorname{POM}/\operatorname{SPOM}$	1.0e+00

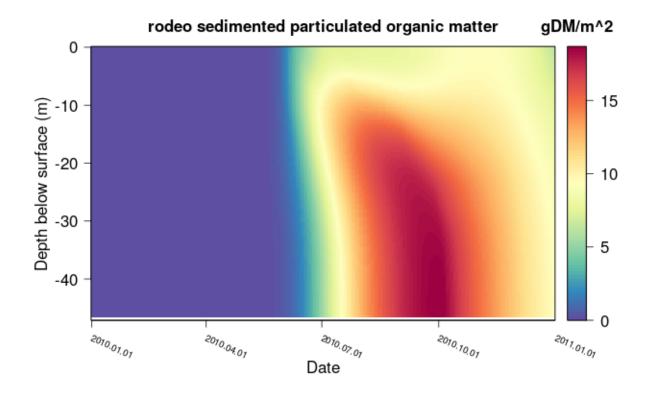
We can create fortran source code, compile GOTM-FABM, run the model, and plot the results using:











4 Additional features

4.1 State variable arguments

There are a few additional arguments for state variables that can be defined in *FABM*. In order to use them a new column in the state variable data frame needs to be added with exactly the name.

- minimum: minnimum allowed value for the state variable (used in the numerical solver)
- maximum: maximum allowed value for the state variable (used in the numerical solver)
- specific light extinction: specific light extinction coefficient of this variable
- no precipitation dilution: the variable is not diluted by precipitation (logical)
- no_river_dilution: the variable is not diluted by river inflows (logical)

4.2 Initial values for the state variables

By default the initial values for the FABM sate variables are constant throughout the whole profile. With some tinkering we can set any profide we want as the initial values. Therefore, we need to have (a) text file(s) with (the) profiles that you want to initialise and install the R packages ncdf4, data.table, gotmtools from the AEMON-J github. The approach is to first run GOTM with 0 time steps (stop date = start date). After a run with GOTM, a restart.nc file is created, that can be use to restart a simulation with the same settings that ended the previous simulation. By running with 0 time steps, this file contains the "standard" initial values, including the ones in out biogeochemical model. Then we need to replaces the homogeneous initial profiles in the restart.nc file by our specified profiles using the ncdf4 librarie. Lastly, we need to set the restart option in the gotm.yaml file to "true". If we run gotm now, it will run with the initial profiles for our biogeochemical model (note that we'll have to rerun this approach every time before we want to run GOTM, because every new GOTM run overwrites the restart.nc file). Alternatively we can save the created restart.nc file e.g. as restart_init_profiles.nc and use this file to override the restart.nc before we run GOTM.

4.3 Defining own functions

4.3.1 Defining functions in the funs data frame

It is possible to define our own functions and call them. If the function is simple i.e. can be calculated in one line of code (e.g. limiting functions for algae growth) it can be defined from the *expressions* column of the *pros* data frame. In order to define own functions two additional columns are needed in the *funs* data frame: expression and arguments. Similar to the *pros* data frame the expression column gives the mathematical expression to calculate. The arguments column gives all input arguments that the functions uses, in the same order they are supplied during function calls (e.g. in the *expression* column of the *pros* data frame), separated by commas (",").

Using this we could e.g. implement the monod function $f_{monod} = \frac{C}{C+K}$ by changing the funs data frame to:

Table 20: Data frame 'funs': Declaration of model functions and dependencies from the host model.

name	unit	description	dependency	expression	arguments
par p $Temp$ exp log f_monod	W/m ² Pa celsius - -	Downwelling photosynthetic radiative flux Atmospheric Pressure Water temperature exponential function logarithmic function Monod funciton	$downwelling_photosynthetic_radiative_flux\\ surface_air_pressure\\ temperature$	C/(C + K)	С, К

Now we can use the function in the expression column of the pros data frame e.g. by replacing:

```
C · mu_max · HP 04/(HP 04 + K_P ) · par/(par + K_par) with C · mu_max · f_monod(HP04, K_P · f_monod(par, K_par)
```

in the expression column of the growth of algae.

4.3.2 Defining functions in external fortran files

More complex functions can be supplied as external fortran code. Two additional columns are needed in the funs data frame: file and module. They give the name of the source code file and the name of the module, which is then loaded in the main source code. This feature is still experimental and might lead to errors!

4.4 Automatic model documentation

If wanted rodeoFABM can automatically generate LaTeX documentation of the state variables, parameter, processes and stoichometry. To do so the function $document_model()$ can be used. Lets create a documentation of the final phytoplankton nutrients model from our example. In order to work we need an additional column named tex (you can also use another name for this column and supply the name to $document_model()$ using the tex argument) in the data frames vars, pars, funs, and pros giving the corresponding LaTeX symbols to be used. The documentation function automatically generates LaTeX fraction, but in order for this to work all used fractions in the expression column of the pros data frame need to be in a specified format. The numerator and denominator need to be in brakets, even if they are just one single variable, number, or parameter: e.g. $(02)/(02 + K_02)$. In the example spread sheet file they are allready added:

```
# see column "tex"
head(vars)
##
     name
              unit.
                                                 description default
                                                                         bot
                                                                                tex
## 1
        C gDM/m<sup>3</sup>
                                         algae concentration
                                                                   0.0
                                                                          NA
                                                                                  C
## 2 HPO4 gP/m<sup>3</sup>
                                   phosphorus concentration
                                                                          NA HPO 4
                                                                   0.1
## 3
       02 g0/m<sup>3</sup>
                                        oxygen concentration
                                                                  10.0
                                                                          NA
                                                                                0 2
                                particulated organic matter
## 4 POM gDM/m^3
                                                                   0.0
                                                                          NA
                                                                                POM
## 5 SPOM gDM/m^2 sedimented particulated organic matter
                                                                   0.0 TRUE
                                                                              SPOM
# create LaTeX documentation for our model
document_model(vars, pars, pros, funs, stoi, landscape = FALSE)
```

```
## ## finished ## [1] TRUE
```

We can see that now there are seven additional file in our worling directory:

They are LaTeX tables of the models state variables $(tab_vars.tex)$, used model parameters $(tab_pars.tex)$, used functions $(tab_funs.tex)$, description of the models processes $(tab_pros.tex)$, description of the process equations $(pros_expr.tex)$, the stoichiometry table $(tab_stoi.tex)$, and a simple latex document that can be used to compile all of the before $(document_model.tex)$.

The created expressions of the processes now look like this:

```
## [1] "\begin{align}"
## [2] "\rho_{grouth} =&- C \\cdot \\mu_{max} \\cdot \\frac{HPO_4 }{ HPO_4 + K_P} \\cdot \\frac{par }{ par + K_{par}}\\\"
## [3] "\rho_{death} =&- C \\cdot k_{death}\\\\"
## [4] "\rho_{geath} =&- V_{sed,ALG}\\\\"
## [4] "\rho_{geath} =&- V_{sed,ALG}\\\\"
## [5] "\rho_{geath} =&- V_{sed,ALG}\\\\\"
## [6] "\rho_{geath} =&- V_{sed,ALG} \\cdot \\frac{p} { 101325} - O_2 \\right)\\\"
## [6] "\rho_{geath} =&- V_{sed,ALG} \\cdot \\frac{p} { 101325} - O_2 \\right)\\\"
```

and compiled they look like this:

head(readLines("pros_expr.tex"))

$$\rho_{growth} = C \cdot \mu_{max} \cdot \frac{HPO_4}{HPO_4 + K_P} \cdot \frac{par}{par + K_{par}}$$

$$\tag{1}$$

$$\rho_{death} = C \cdot k_{death} \tag{2}$$

$$\rho_{Sed,ALG} = v_{sed,ALG} \tag{3}$$

$$\rho_{O2,exch} = v_{exch,O2} \cdot \left(\exp\left(7.7117 - 1.31403 \cdot \log\left(\vartheta_z + 45.93\right)\right) \cdot \frac{p}{101325} - O_2 \right) \tag{4}$$

$$\rho_{O2,cons} = \frac{O_2}{O_2 + K_{O2}} \cdot k_{O2,cons} \tag{5}$$

$$\rho_{Sed,POM} = v_{sed,POM} \tag{6}$$

$$\rho_{Miner,POM} = POM \cdot k_{miner,POM} \cdot \frac{O_2}{O_2 + K_{miner,O2}} \tag{7}$$

$$\rho_{Miner,SPOM} = SPOM \cdot k_{miner,SPOM} \cdot \frac{O_2}{O_2 + K_{miner,O2}}$$
(8)

$$\rho_{Set,POM} = v_{sed,POM} \cdot POM$$
 (9)

References

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