AMAS_JGLAHE

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Chapter 1

AMAS

Alignment manipulation and summary statistics

If you are using this program, please cite `this publication`:
```
Borowiec, M.L. 2016. AMAS: a fast tool for alignment manipulation and computing of summary statistics. PeerJ
    4:e1660.
```

1.1 This fork: AMAS_JGLAHE

Fork of the main repo which:

- Adds a `metapartition` command -> collates discontinuous metapartitions within a superalignment and concatenates them into a new superalignment of contiguous metapartitons.

- Reduces restrictions on input partition file formatting -> accepts partition files for RAxML(-NG) and IQ-TREE2 (best_scheme, best_scheme.nex and best_model.nex).

1.2 Installation

Use `AMAS.py` in the `amas` directory as a stand-alone program or clone it if you have `git installed` on your system.

If your system doesn't have a Python version 3.4 or newer (`AMAS` will work under Python 3.0 but you may noy be able to use it with multiple cores), you will need to `download and install it`. On Linux-like systems (including Ubuntu) you can install it from the command line using

It may be possible to use this version as a module, but only through manual configuration.

1.3 Command line interface

`AMAS` can be run from the command line. Here is the general usage (you can view this in your command line with
`python3` `AMAS.py` `-h`):
```
usage: AMAS <command> [<args>]

The AMAS commands are:
  concat          Concatenate input alignments.
  convert         Convert to other file format.
  replicate       Create replicate data sets for phylogenetic jackknife.
  split           Split alignment according to a partitions file.
  summary         Write alignment summary.
  remove          Remove taxa from alignment.
  translate       Translate DNA alignment into protein alignment.
  trim            Remove columns from alignment.
  metapartitions  Runs `split' and concatenates the output.

Use AMAS <command> -h for help with arguments of the command of interest

positional arguments:
  command     Subcommand to run

optional arguments:
  -h, --help  show this help message and exit
```

To show help for individual commands, use `AMAS.py` `<command>` `-h` or `AMAS.py` `<command>` `--help`.

1.3.1 Examples

For every `AMAS.py` run on the command line you need to specify action with `concat`, `convert`, `replicate`, `split`, or `summary` for the input to be processed. Additionally, you need to provide three arguments required for all commands. The order in which the arguments are given does not matter:

1) input file name(s) with `-i` (or in long version: `--in-files`),

2) format with `-f` (`--in-format`),

3) and data type with `-d` (`--data-type`).

The options available for the format are `fasta`, `phylip`, `nexus` (sequential), `phylip-int`, and `nexus-int` (interleaved). Data types are `aa` for protein alignments and `dna` for nucleotide alignments.

For example:
```
python3 AMAS.py concat -i gene1.nex gene2.nex -f nexus -d dna
```

If you have many files that you want to input in one run, you can use multiple cores of your computer to process them in parallel. The `summary` command supports `-c` or `--cores` with which you can specify the number of cores to be used:
```
python3 AMAS.py summary -f phylip -d dna -i *phy -c 12
```

In the above, we specified 12 cores. Note that this won't improve computing time if you're working with only one or very few files. The parallel processing is only used for the file parsing step and calculating alignment summaries.

In addition to overall alignment summaries, you can also print statistics calculated on a sequence (taxon) by sequence basis. Use `-s` or `--by-taxon` flag to turn it on. `AMAS` in this mode will print out one file with overall alignment summaries and a file with taxon summaries for each input alignment.

IMPORTANT! `AMAS` is fast and powerful, but be careful: it assumes you know what you are doing and will not prevent you overwriting a file. It will, however, print out a warning if this has happened. `AMAS` was also written to work with aligned data and some of the output generated from unaligned sequences won't make sense. Because of computing efficiency `AMAS` by default does not check if input sequences are aligned. You can turn this option on with `-e` or `--check-align`.

1.3.1.1 Concatenating alignments

For example, if you want to concatenate all DNA phylip files in a directory and all of them have the `.phy` extension, you can run:

```
python3 AMAS.py concat -f phylip -d dna -i *phy
```

By default the output will be written to two files: `partitions.txt`, containing partitions from which your new alignment was constructed, and `concatenated.out` with the alignment itself in the fasta format. You can change the default names for these files with `-p` (`--concat-part`) and `-t` (`--concat-out`), respectively, followed by the desired name. The output format is specified by `-u` (`--out-format`) and can also be any of the following: `fasta`, `phylip`, `nexus` (sequential), `phylip-int`, or `nexus-int` (interleaved).

Below is a command specifying the concatenated file output format as nexus with `-u nexus`:

```
python3 AMAS.py concat -f fasta -d aa -i *fas -u nexus
```

Alignments to be concatenated need not have identical sets of taxa before processing: the concatenated alignment will be populated with missing data where a given locus is missing a taxon. However, if every file to be concatenated includes only unique names (for example species name plus sequence name: `D_melanogaster_NW↵_001845408.1` in one alignment, `D_melanogaster_NW_001848855.1` in other alignment etc.), you will first need to trim those names so that sequences from one taxon have equivalents in all files.

In addition to the name, you can also specify the format of the partitions output file. By default, the format is the following:

```
AA = 1-605
AK = 606-1200
28S = 1201-1800
```

RAxML:

```
python3 AMAS.py concat -f phylip -d dna -i *phy --part-format raxml
DNA, AA = 1-605
DNA, AK = 606-1200
DNA, 28S = 1201-1800
```

Nexus:

```
python3 AMAS.py concat -f phylip -d dna -i *phy --part-format nexus
#NEXUS
Begin sets;
    charset AA = 1-605;
    charset AK = 606-1200;
    charset 28S = 1201-1800;
End;
```

Partitions can also be written by codon positions using the `-n` or `--codons` flag, either for alignments containing first and second or all three positions. In the above example, supplying `-n 123` would result in:

```
AA_pos1 = 1-605\3
AA_pos2 = 2-605\3
AA_pos3 = 3-605\3
AK_pos1 = 606-1200\3
AK_pos2 = 607-1200\3
AK_pos3 = 608-1200\3
28S_pos1 = 1201-1800\3
28S_pos2 = 1202-1800\3
28S_pos3 = 1203-1800\3
```

1.3.1.2 Getting alignment statistics

This is an example of how you can summarize two protein fasta alignments by running:

```
python3 AMAS.py summary -f fasta -d aa -i my_aln.fasta my_aln2.fasta
```

By default `AMAS` will write a file with the summary of the alignment in `summary.txt`. You can change the name of this file with `-o` or `--summary-out`. You can also summarize a single or multiple sequence alignments at once.

The statistics calculated include the number of taxa, alignment length, total number of matrix cells, overall number of undetermined characters, percent of missing data, AT and GC contents (for DNA alignments), number and proportion of variable sites, number and proportion of parsimony informative sites, and counts of all characters present in the relevant alphabet.

1.3.1.3 Converting among formats

To convert all nucleotide fasta files with a `.fas` extension in a directory to nexus alignments, you could use:
```
python3 AMAS.py convert -d dna -f fasta -i *fas -u nexus
```

In the above, the required options are combined with `convert` command to convert the input files and `-u nexus` which indicates the output format.

`AMAS` will not overwrite over input here but will create new files instead, automatically appending appropriate extensions to the input file's name: `-out.fas`, `-out.phy`, `-out.int-phy`, `-out.nex`, or `-out.int-nex`.

1.3.1.4 Splitting alignment by partitions (updated partition parsing AMAS_JGLAHE)

If you have a partition file, you can split a concatenated alignment and write a file for each partition:
```
python3 AMAS.py split -f nexus -d dna -i concat.nex -l partitions.txt -u nexus
```

In the above one input file `concat.nex` was provided for splitting with `split` and partitions file `partitions.txt` with `-l` (same as `--split-by`). For splitting you should only use one input and one partition file at a time. This is an example partition file:
```
AApos1&2  =  1-604\3, 2-605\3
AApos3  =  3-606\3
28SAutapoInDels=7583, 7584, 7587, 7593
```

If this was the `partitions.txt` file from the example command above, `AMAS` would write three output files called `concat_AApos1&2.nex`, `concat_AApos3.nex`, and `concat_28SautapoInDels.nex`. The partitions file will be parsed correctly as long as there is are none ~~text prior to the partition name (`CHARSET AApos1&2` or `DNA, AApos1&2` will not work) and commas separate ranges or individual sites in each partition~~ of the following characters in partition names: `<>!?$^;:\/,.`; members of [\s] are also not allowed in partition names.

Sometimes after splitting you will have alignments with taxa that have only gaps `-` or missing data `?`. If you want to these to not be included in the output , add `-j` or `--remove-empty` to the command line.

Partition files are parsed with `AMAS.FileParser.partition_parse()`, which in the JGLAHE fork of AMAS looks like this:
```
def partitions_parse(self):
    # parse partitions file using regex
    # original: `matches = re.finditer(r"^(\s+)?([^ =]+)[ =]+([\\0-9, -]+)", self.in_file_lines,
      re.MULTILINE)`
    # new version: more permissive -> handles PartionFinder/RAxML/(IQ-TREE 2)best_scheme.nex format
      partition files
    matches = re.finditer(
        r"""^[ \t]*                           # start of line w/ zero-or-more (just) whitespaces/tabs
            (
             (?P<nexus>charset[ ]+)           # case 1: (IQ-TREE 2)best_scheme.nex partition
      directive; partition name
             |
             (?P<raxml>[A-Za-z0-9_+\.]+,[ \t]+)    # case 2: RAxML/RAxML-NG model(+other pars); partition
      name
            )?
            (?P<partition_name>[A-Za-z0-9_@*&()\[\]{}'"%#|-]+) # partition name (handles various
      metcharacters, excludes [\s],<>?/:;~'\^$!)
            [ ]*=[ ]*                          # whitespace-padded (or unpadded) '=': (IQ-TREE
      2)best_scheme.nex compatabiliy
            (?P<numbers>[\\0-9, -]+)           # position ranges w/ stride (multiple intervals; from
      original regex)
            (?P<nexus_term>[ ]*[;])?           # whitespace-prepended (or unprepended) ';' (nexus
      terminator)
        """,
        self.in_file_lines,
        re.MULTILINE | re.VERBOSE
    )

    # initiate list to store dictionaries with lists
    # of slice positions as values
    partitions = []
    add_to_partitions = partitions.append

    for match in matches:
        # initiate dictionary of partition name as key
```

```
        dict_of_dicts = {}
        # and list of dictionaries with slice positions
        list_of_dicts = []
        add_to_list_of_dicts = list_of_dicts.append
        # get parition name and numbers from parsed partition strings
        partition_name = match.group('partition_name')
        numbers = match.group('numbers')
        # remove any whitespace padding '-' (to be consistent with partition-writing format)
        numbers = re.sub(r"[ ]*-[ ]*", "-", numbers)
        # find all numbers that will be used to parse positions
        positions = re.findall(r"([^ ,]+)", numbers)

        for position in positions:
            # create dictionary for slicing input sequence
            # conditioning on whether positions are represented
            # by range, range with stride, or single number
            pos_dict = {}

            if "-" in position:
                m = re.search(r"([0-9]+)-([0-9]+)", position)
                pos_dict["start"] = int(m.group(1)) - 1
                pos_dict["stop"] = int(m.group(2))
            else:
                pos_dict["start"] = int(position) - 1
                pos_dict["stop"] = int(position)

            if "\\" in position:
                # Note: the value of `N` in `...\N` isn't read: the script simply assumes `N` is consistent
    with the number of
                # increments per interval when the alignment is parsed with a stride of 3 (designating each
    cpos).
                # E.g. For the partition file:
                #       ...'1-N\2'
                #       ...'2-N\2'
                #       ...'(N+1)-M\2'
                #       ...'(N+2)-M\2'
                # 3'cpos are ignored due to the absence of intervals `3-N...', `(N+3)-M...', not because the
    associated stride values are'\2'
                pos_dict["stride"] = 3
            elif "\\" not in position:
                pos_dict["stride"] = 1

            add_to_list_of_dicts(pos_dict)

        dict_of_dicts[partition_name] = list_of_dicts
        add_to_partitions(dict_of_dicts)

    return partitions
```

This version removes some of the partition formatting restrictions of the original AMAS repo version, with the updated partitions_parse() method facilitating the uses of native RAxML(-NG) and IQ-TREE2 partition files.

The following is a contrived example to demonstrate this, along with the updated `metapartions` command. Consider the following superalignment `concat.fas`:

```
>S01
CCCCCTGGCGCCGCCGCCGCCCCTCCTCCTCCTCCTCCCCCCCCC
>S02
AACAATGGAGCAGAAGAAGAAAATAATAATAATAATAAAAAAAAA
>S03
TTCTTTGGTGCTGATGTTGTTTTTTTTTTTTTTTTATTTTTTTTT
>S04
GGCGGTGGGGCGGAGGTGGGGGGTGGTGGTGTTGATGGGGGGGGG
>S05
CCCCCTGGCGCCGACGTCGGCCCTCCTCGTCTTCATCCCCCCCCC
>S06
AACAATGGCGCAGAAGTAGGAAATACTAGTATTAATAAAAAAAAA
>S07
TTCTTTGGCGCAGATGTTGGTTATTCTTGTTTTTATTTTTTTTTT
>S08
GGCGTTGGCGCAGATGTGGGGGATGCTGGTGTTGATGGGGGGGGG
>S09
CGCCTTGGCGCAGATGTGGGCCATCCTCGTCTTCATCCCCCCCCC
>S10
AGCATTGGCGCAGATGTGGGCAATACTAGTATTCATAAAAAAAAA
>S11
TGCTTTGGCGCAGATGTGGGCTATTCTTGTATTCATTTTATTTTT
>S12
GGCGTTGGCGCAGATGTGGGCGATGCTTGTATTCATGGGAGGTGG
>S13
CGCCTTGGCGCAGATGTGGGCCATGCTTGTATTCATCCCAGCTCC
>S14
AGCATTGGCGCAGATGTGGGCCATGCTTGTATTCATAAAAGATCA
>S15
TGCATTGGCGCAGATGTGGGCCATGCTTGTATTCATTTTAGATCT
```

```
>S16
TGCATGTTCTCATATTTGTGCCAGGCGTGGATGCAGGG?AGATCT
>S17
TGCATGTTCTCATATTTGTGCCAGGCGTGGATGCAGTTTAGATCT
>S18
AGCATGTTCTCATATTTGTGCCAGGCGTGGATGCAGAAAAGATCA
>S19
CGCCTGTTCTCATATTTGTGCCAGGCGTGGATGCAGCCCAGCTCC
>S20
GGCGTGTTCTCATATTTGTGCGAGGCGTGGATGCAGGGGAGGTGG
>S21
TGCTTGTTCTCATATTTGTGCTAGTCGTGGATGCAGTTTATTTTT
>S22
AGCATGTTCTCATATTTGTGCAAGACGAGGATGCAGAAAAAAAAA
>S23
CGCCTGTTCTCATATTTGTGCCAGCCGCGGCTGCAGCCCCCCCCC
>S24
GGCGTGTTCTCATATTTGTGGGAGGCGGGGGTGGAGGGGGGGGGG
>S25
TTCTTGTTCTCATATTTTTGTTAGTCGTGGTTGTAGTTTTTTTTT
>S26
AACAAGTTCTCATAATTATGAAAGACGAGGATGAAGAAAAAAAAA
>S27
CCCCCGTTCTCCTACTTCTGCCCGCCGCGGCTGCAGCCCCCCCCC
>S28
GGCGGGTTGTCGTAGTTGTGGGGGGGGGGGGGTGGAGGGGGGGGGG
>S29
TTCTTGTTTTCTTATTTTTTTTTGTTGTTGTTGTAGTTTTTTTTT
>S30
AACAAGTTATCATAATAATAAAAGAAGAAGAAGAAGAAAAAAAAA
>S31
CCCCCGTTCTCCTCCTCCTCCCCGCCGCCGCCGCCGCCCCCCCCC
```

Its partition file `partitions.txt` contain various formatting challenges to test the parser, but it includes the basic directives found in native RAxML(-NG) and IQ-TREE2 partition files:

```
charset @@(partition_A_pos1) =      7 - 21\3
  charset %s = 8  -21\3
    charset "partition_A_pos3" = 9-   21\3
Q.insect, (partition_B_pos)1 = 40-   45\3
Q.insect,   '[partition_B_pos2] = 41   -45\3
Q.insect, par&&tition_B_pos3  =„ 42-45\3
    GTR+G4,    pa&rtition_C =  , , , „38 39         37,
9.20b, #partition&|_D_pos1 = 1-6\3 22 - 36\3
      partition_D_pos2= 2-6\3 23 -36\3
p1_3_a          = 3-6\3              24 -     36\3
  charpartition mymodels =
    9.20b: partition_D_pos1;
end;
```

While most of the partitions here are not contiguous with respect to the `concat.fas`, this superalignment can be converted into one with contiguous metapartitons by running the command:

```
./AMAS.py metapartitions -i concat.fas -f fasta -d dna --no-san --no-mpan -l partitions.txt -t
     concat.out.fas`
```

This generates `concat.out.fas`:

```
>S01
GGGGGGCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCTTTTTT
>S02
GGGGGGCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAACTTTTTT
>S03
GGGGGGCATTTTTTTTTTTTTTTTTTTTTTTTTTTTTACTTTTTT
>S04
GGGGGGCATGGGGGGGGGGGGGGGGGGGGGGGGGGGGTACTTTTTT
>S05
GGGGGGCATGCCCCCCCCCCCCCCCCCCCCCCCCCGTACTTTTTT
>S06
GGGGGGCATGCAAAAAAAAAAAAAAAAAAAAAAACGTACTTTTTT
>S07
GGGGGGCATGCATTTTTTTTTTTTTTTTTTTTTACGTACTTTTTT
>S08
GGGGGGCATGCATGGGGGGGGGGGGGGGGGGGGTACGTACTTTTTT
>S09
GGGGGGCATGCATGCCCCCCCCCCCCCCCCCCGTACGTACTTTTTT
>S10
GGGGGGCATGCATGCAAAAAAAAAAAAAAAACGTACGTACTTTTTT
>S11
GGGGGGCATGCATGCATTTTTTTTTTTTTTACGTACGTACTTTTTT
>S12
GGGGGGCATGCATGCATGGGGGGGGGGGGTACGTACGTACTTTTTT
>S13
GGGGGGCATGCATGCATGCCCCCCCCCCGTACGTACGTACTTTTTT
>S14
GGGGGGCATGCATGCATGCAAAAAAAACGTACGTACGTACTTTTTT
```

```
>S15
GGGGGGCATGCATGCATGCATTTTTACGTACGTACGTACTTTTTT
>S16
TTTTTTCATGCATGCATGCATG?GTACGTACGTACGTACGGGGGG
>S17
TTTTTTCATGCATGCATGCATTTTTACGTACGTACGTACGGGGGG
>S18
TTTTTTCATGCATGCATGCAAAAAAACGTACGTACGTACGGGGGG
>S19
TTTTTTCATGCATGCATGCCCCCCCCCCGTACGTACGTACGGGGGG
>S20
TTTTTTCATGCATGCATGGGGGGGGGGGGTACGTACGTACGGGGGG
>S21
TTTTTTCATGCATGCATTTTTTTTTTTTTACGTACGTACGGGGGG
>S22
TTTTTTCATGCATGCAAAAAAAAAAAAAAACGTACGTACGGGGGG
>S23
TTTTTTCATGCATGCCCCCCCCCCCCCCCCCCGTACGTACGGGGGG
>S24
TTTTTTCATGCATGGGGGGGGGGGGGGGGGGGGGTACGTACGGGGGG
>S25
TTTTTTCATGCATTTTTTTTTTTTTTTTTTTTTTACGTACGGGGGG
>S26
TTTTTTCATGCAAAAAAAAAAAAAAAAAAAAAAACGTACGGGGGG
>S27
TTTTTTCATGCCCCCCCCCCCCCCCCCCCCCCCCCCGTACGGGGGG
>S28
TTTTTTCATGGGGGGGGGGGGGGGGGGGGGGGGGGGGTACGGGGGG
>S29
TTTTTTCATTTTTTTTTTTTTTTTTTTTTTTTTTTTTACGGGGGG
>S30
TTTTTTCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAACGGGGGG
>S31
TTTTTTCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCGGGGGG
```

The figure below more clearly demonstrates this transformation: `concat.fas` on the left is converted to `concat.out.fas` on the right using the `metapartitions` command above; visualized in Aliview v1.28

1.3.1.5 Translating a DNA alignment into aligned protein sequences

You can translate a nucleotide alignment to amino acids with AMAS using one of the NCBI translation tables. For example, to correctly translate an insect mitochondrial gene alignment that begins at a second codon position:

```
python3 AMAS.py translate -f nexus -d dna -i concat.nex --code 5 --reading-frame 2 --out-format phylip
```

`--code` and `--reading-frame` are the same as `-b` and `-k` and are both set to 1 (the standard genetic code and the first character of the alignment corresponds to the first codon position) by default. When translating, AMAS will contract gaps `-` and missing `?`, such that `---` becomes `-` in the translated alignment. A warning will be printed if stop codons are found and these are indicated as asterisks `*` in the output. See `AMAS.py translate -h` for more info.

1.3.1.6 Creating replicate data sets

With AMAS you can create concatenated alignments from a proportion of randomly chosen alignments that can be used for, for example, a phylogenetic jackknife analysis. Say you have 1000 phylip files, each containing a single aligned locus, and you want to create 200 replicate phylip alignments, each built from 100 loci randomly chosen from all the input files. You can do this by specifying `replicate` command and following it with `-r` or `--rep-aln` followed by the number of replicates (in this case `200`) and number of alignments (`100`). Remember to supply the output format with `-u` if you want it to be other than `fasta`:

```
python3 AMAS.py replicate -r 200 100 -d dna -f phylip -i *phy -u phylip
```

1.3.1.7 Removing taxa/sequences from alignment

It is possible to remove taxa from alignments:
```
python3 AMAS.py remove -x species1 species2 -d dna -f nexus -i *nex -u nexus-int -g no_species12_
```

The above will process all `nexus` files in the directory and remove taxa called `species1` and `species2`. The argument `-x` (the same as `--taxa-to-remove`) is followed by the names of sequences to be removed. Note that `AMAS` converts spaces into underscores and strips any quotes present in input sequence names before processing, so you may need to modify your names to remove accordingly. The argument `-g` (the same as `--out-prefix`) specifies a prefix to be added to output file names. The default prefix is 'reduced_'. You may want to realign your files after taxon removal.

1.3.1.8 Checking if input is aligned

By specifying optional argument `-e` (`--check-align`), you can make `AMAS` check if your input files contain only aligned sequences. This option is disabled by default because it can substantially increase computation times in files with many taxa. Enabling this option also provides an additional check against misspecified input file format.

1.3.1.9 TODO Metapartitions

1.4 TODO AMAS as a Python module

Using `AMAS` inside your Python pipeline gives you much more flexibility in how the input and output are being processed. All the major functions of the command line interface can recreated using `AMAS` as a module. Following installation from `pip` use:
```
pydoc amas.AMAS
```

To access detailed documentation for the classes and functions available.

You can import `AMAS` to your script with:
```
from amas import AMAS
```

The class used to manipulate alignments in `AMAS` is `MetaAlignment`. This class has to be instantiated with the same, named arguments as on the command line: `in_files`, `data_type`, `in_format`. You also need to supply the number of cores to be used with `cores`. MetaAlignment holds one or multiple alignments and its `in_files` option must be a list, even if only one file is being read.
```
meta_aln = AMAS.MetaAlignment(in_files=["gene1.phy"], data_type="dna",in_format="phylip", cores=1)
```

Creating MetaAlignment with multiple files is easy:
```
multi_meta_aln = AMAS.MetaAlignment(in_files=["gene1.phy", "gene1.phy"], data_type="dna",
    in_format="phylip", cores=2)
```

Now you can call the various methods on your alignments. `.get_summaries()` method will compute summaries for your alignments and produce headers for them as atuple with first element being the header and the second element a list of lists with the statistics:
```
summaries = meta_aln.get_summaries()
```

The header is different for nucleotide and amino acid data. You may choose to skip it and print only the second element of the tuple, that is a list of summary statistics:
```
= summaries[1]
```

`.get_parsed_alignments()` returns a list of dictionaries where each dictionary is an alignment and where taxa are the keys and sequences are the values. This allows you to, for example, print only taxa names in each alignment or do other manipulation of the sequence data:
```
# get parsed dictionaries
```

```
aln_dicts = multi_meta_aln.get_parsed_alignments()

# print only taxa names in the alignments:
for alignment in aln_dicts:
    for taxon_name in alignment.keys():
        print(taxon_name)
```

Similar to the above example, it is also easy to get translated amino acid alignment as a list of dictionaries (one per input alignment):

```
# get parsed dictionaties
aln_dicts = multi_meta_aln.get_translated(2, 1) # 2: vertebrate mitochondrial genetic code and 1: reading
    frame starting at first character
```

To split alignment use `.get_partitioned("your_partitions_file")` on a `MetaAlignment` with a single input file. `.get_partitioned()` returns a list of dictionaries of dictionaries, with `{ partition_name : { taxon : sequence } }` structure for each partition:

```
partitions = meta_aln.get_partitioned("partitions.txt")
```

AMAS uses `.get_partitions("your_partitions_file")` to parse the partition file:

```
parsed_parts = meta_aln.get_partitions("partitions.txt")
print(parsed_parts)
```

`.get_replicate(no_replicates, no_loci)` gives a list of parsed alignments (dictionaries), each a replicate constructed from the specified number of loci:

```
replicate_sets = multi_meta_aln.get_replicate(2, 2)
```

To concatenate multiple alignments first parse them with `.get_parsed_alignments()`, then pass to `.get←_concatenated(your_parsed_alignments)`. This will return a tuple where the first element is the `{ taxon : sequence }` dictionary of concatenated alignment and the second element is the partitions dict with `{ name : range }`.

```
parsed_alns = multi_meta_aln.get_parsed_alignments()
concat_tuple = multi_meta_aln.get_concatenated(parsed_alns)
concatenated_alignments = concat_tuple[0]
concatenated_partitions = concat_tuple[1]
```

Removing taxa from alignments is very easy:

```
spp_to_remove = ["taxon1", "taxon2", "taxon3"]
reduced_alns = multi_meta_aln.remove_taxa(spp_to_remove)
```

To print to file or convert among file formats use one of the `.print_format(parsed_alignment)` methods called with a parsed dictionary as an argument. These methods include `.print_fasta()`, `.print_←nexus()`, `.print_nexus_int()`, `print_phylip()`, and `.print_phylip_int()`. They return an appropriately formatted string.

```
for alignment in concatenated_alignments:
    nex_int_string = meta_aln.print_nexus_int(alignment)
    print(nex_int_string)
```

Chapter 2

Namespace Index

2.1 Namespace List

Here is a list of all namespaces with brief descriptions:

Chapter 3

Hierarchical Index

3.1 Class Hierarchy

This inheritance list is sorted roughly, but not completely, alphabetically:

Chapter 4

Class Index

4.1 Class List

Here are the classes, structs, unions and interfaces with brief descriptions:

Chapter 5

File Index

5.1 File List

Here is a list of all files with brief descriptions:

Chapter 6

Namespace Documentation

6.1 amas Namespace Reference

Namespaces

- namespace AMAS

Variables

- str __author__ = 'Marek Borowiec'
- str __email__ = 'petiolus@gmail.com'
- str __version__ = '1.02'
- __all__ = dir()

6.1.1 Variable Documentation

6.1.1.1 __all__

```
amas.__all__ = dir()  [private]
```

Definition at line 6 of file __init__.py.

6.1.1.2 __author__

```
str amas.__author__ = 'Marek Borowiec'  [private]
```

Definition at line 3 of file __init__.py.

6.1.1.3 __email__

```
str amas.__email__ = 'petiolus@gmail.com'  [private]
```

Definition at line 4 of file __init__.py.

6.1.1.4 __version__

```
str amas.__version__ = '1.02'  [private]
```

Definition at line 5 of file __init__.py.

6.2 amas.AMAS Namespace Reference

Classes

- class Alignment
- class AminoAcidAlignment
- class DNAAlignment
- class FileHandler
- class FileParser
- class MetaAlignment
- class ParsedArgs

Functions

- proportion (x)
- main ()
- run ()

6.2.1 Detailed Description

```
This stand-alone program allows manipulations of multiple sequence
alignments. It supports sequential FASTA, PHYLIP, NEXUS, and interleaved PHYLIP
and NEXUS formats for DNA and aino acid sequences. It can print summary statistics,
convert among formats, and concatenate alignments.

Current statistics include the number of taxa, alignment length, total number
of matrix cells, overall number of undetermined characters, percent of missing
data, AT and GC contents (for DNA alignments), number and proportion of
variable sites, number and proportion of parsimony informative sites,
and counts of all characters present in the relevant (nucleotide or amino acid) alphabet.
```

6.2.2 Function Documentation

6.2.2.1 main()

```
amas.AMAS.main ( )
```

Definition at line 2361 of file AMAS.py.

```
02361 def main():
02362
02363     # initialize parsed arguments and meta alignment objects
02364     kwargs = run()
02365     meta_aln = MetaAlignment(**kwargs)
02366
02367     if meta_aln.command == "summary":
02368         meta_aln.write_summaries(kwargs["summary_out"])
02369     if meta_aln.by_taxon_summary:
02370         print("Printing taxon summaries")
02371         meta_aln.write_taxa_summaries()
```

```
02372      if meta_aln.command == "convert":
02373          meta_aln.write_out("convert", kwargs["out_format"])
02374      if meta_aln.command == "concat":
02375          meta_aln.write_out("concat", kwargs["out_format"])
02376          meta_aln.write_partitions(kwargs["concat_part"], kwargs["part_format"], kwargs["data_type"],
      kwargs["codons"])
02377      if meta_aln.command == "replicate":
02378          meta_aln.write_out("replicate", kwargs["out_format"])
02379      if meta_aln.command == "split":
02380          meta_aln.write_out("split", kwargs["out_format"])
02381      if meta_aln.command == "remove":
02382          meta_aln.write_out("remove", kwargs["out_format"])
02383      if meta_aln.command == "translate":
02384          meta_aln.write_out("translate", kwargs["out_format"])
02385      if meta_aln.command == "trim":
02386          meta_aln.write_out("trim", kwargs["out_format"])
02387
02388      if meta_aln.command == "metapartitions":
02389          # `metapartitions` is essentially `split` + `concat`. Currently you can't set an out_format:
02390          # it's automatically set to match the in_format because the intermediate `split` outputs
      become
02391          # the 'new' in_files for the `concat` operation, and then calling either:
02392          #  -> AminoAcidAlignment(Alignment.__init__(self, in_file, in_format, data_type))
02393          #  -> DNAAlignment(Alignment.__init__(self, in_file, in_format, data_type))
02394          # through MetaAlignment.get_alignment_object(alignment, self.in_format, self.data_type)
02395          meta_aln.write_out("metapartitions", kwargs["in_format"])
02396          meta_aln.write_partitions(kwargs["concat_part"], kwargs["part_format"], kwargs["data_type"],
      "none")
02397
02398          # meta_aln.write_out("translate", kwargs["out_format"])
02399
```

References amas.AMAS.run().

Referenced by amas.AMAS.run().

Here is the call graph for this function:

Here is the caller graph for this function:

6.2.2.2 proportion()

```
amas.AMAS.proportion (
              x )
```

Definition at line 43 of file AMAS.py.

```
00043 def proportion(x):
00044     # needed to prevent input of invalid floats in trim mode
00045     x = float(x)
00046     if x < 0.0 or x > 1.0:
00047         raise argparse.ArgumentTypeError("%r not in range [0.0, 1.0]" % (x,))
00048     return x
00049
```

6.2.2.3 run()

```
amas.AMAS.run ( )
```

Definition at line 2400 of file AMAS.py.

```
02400 def run():
02401
02402     # initialize parsed arguments
02403     config = ParsedArgs()
02404     # get arguments
02405     config_dict = config.get_args_dict()
02406     return config_dict
02407
```

References amas.AMAS.main().

Referenced by amas.AMAS.main().

Here is the call graph for this function:

Here is the caller graph for this function:

Chapter 7

Class Documentation

7.1 amas.AMAS.Alignment Class Reference

Inheritance diagram for amas.AMAS.Alignment:
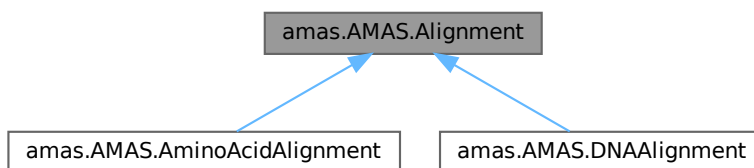
Public Member Functions

- __init__ (self, in_file, in_format, data_type)
- __str__ (self)
- get_aln_input (self)
- get_parsed_aln (self)
- summarize_alignment (self)
- summarize_alignment_by_taxa (self)
- get_char_summary (self)
- get_taxon_char_summary (self)
- append_count (self, char_dict)
- matrix_creator (self)
- get_column (self, i)
- all_same (self, site)
- get_sites_no_missing_ambiguous (self)
- get_site_no_missing_ambiguous (self, column)
- replace_missing (self, column)
- get_trim_selection (self, trim_fraction, parsimony_check)
- get_variable (self)
- get_parsimony_informative (self)

- get_prop_variable (self)
- get_prop_parsimony (self)
- get_name (self)
- get_taxa_no (self)
- get_alignment_length (self)
- get_matrix_cells (self)
- get_missing (self)
- get_missing_percent (self)
- get_missing_from_parsed (self)
- get_missing_from_seq (self, seq)
- get_missing_percent_from_seq (self, seq)
- get_counts (self)
- get_counts_from_parsed (self)
- get_counts_from_seq (self, seq)
- check_data_type (self)

Public Attributes

- in_file
- in_format
- data_type
- parsed_aln
- length
- matrix
- no_missing_ambiguous
- variable_sites
- prop_variable
- parsimony_informative
- prop_parsimony
- missing_records
- missing
- check

Static Public Attributes

- all_matrix_cells

7.1.1 Detailed Description

```
Base class: Gets in parsed sequences as input and summarizes their stats.
Based on the data type, the subclasses AminoAcidAlignment & DNAAlignment define the attributes:
'alphabet', 'missing_ambiguous_chars', 'missing_chars', 'non_alphabet'
```

Definition at line 805 of file AMAS.py.

7.1.2 Constructor & Destructor Documentation

7.1.2.1 __init__()

```
amas.AMAS.Alignment.__init__ (
            self,
            in_file,
            in_format,
            data_type )
```

Definition at line 811 of file AMAS.py.

```
00811    def __init__(self, in_file, in_format, data_type):
00812        # initialize alignment class with parsed records and alignment name as arguments,
00813        # create empty lists for list of sequences, sites without
00814        # ambiguous or missing characters, and initialize variable for the number
00815        # of parsimony informative sites
00816        self.in_file = in_file
00817        self.in_format = in_format
00818        self.data_type = data_type
00819
00820        self.parsed_aln = self.get_parsed_aln()
00821
```

7.1.3 Member Function Documentation

7.1.3.1 __str__()

```
amas.AMAS.Alignment.__str__ (
            self )
```

Definition at line 822 of file AMAS.py.

```
00822    def __str__(self):
00823        #  purpose of override? (originally returned method object)
00824        return self.get_name()
00825
```

References amas.AMAS.Alignment.get_name().

Here is the call graph for this function:

7.1.3.2 all_same()

```
amas.AMAS.Alignment.all_same (
            self,
            site )
```

Definition at line 937 of file AMAS.py.

```
00937    def all_same(self, site):
00938        # check if all elements of a site are the same
00939        return not site or site.count(site[0]) == len(site)
```

```
00940
```

Referenced by amas.AMAS.Alignment.get_variable().

Here is the caller graph for this function:
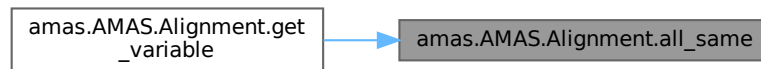
7.1.3.3 append_count()

```
amas.AMAS.Alignment.append_count (
              self,
              char_dict )
```

Definition at line 919 of file AMAS.py.

```
00919    def append_count(self, char_dict):
00920        count_list = []
00921        for char in self.alphabet:
00922            if char in char_dict.keys():
00923                count_list.append(char_dict[char])
00924            else:
00925                count_list.append(0)
00926        return count_list
00927
```

References amas.AMAS.AminoAcidAlignment.alphabet, and amas.AMAS.DNAAlignment.alphabet.

Referenced by amas.AMAS.Alignment.get_taxon_char_summary().

Here is the caller graph for this function:

7.1.3.4 check_data_type()

```
amas.AMAS.Alignment.check_data_type (
              self )
```

Definition at line 1074 of file AMAS.py.

```
01074    def check_data_type(self):
01075        # check if the data type is correct; only one seq to save on computation
01076        seq = next(iter(self.parsed_aln.values()))
01077        self.check = any(char in self.non_alphabet for char in seq)
01078        if self.check is True:
```

```
01079              print(
01080                  "WARNING: found non-" + self.data_type + " characters. "
01081                  "Are you sure you specified the right data type?"
01082              )
01083
01084
```

References amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the caller graph for this function:
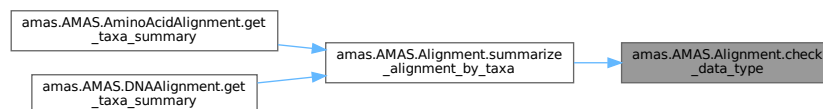
7.1.3.5 get_alignment_length()

```
amas.AMAS.Alignment.get_alignment_length (
              self )
```

Definition at line 1011 of file AMAS.py.
```
01011      def get_alignment_length(self):
01012          # get alignment length by just checking the first seq length
01013          # this assumes that all sequences are of equal length
01014          return len(next(iter(self.parsed_aln.values())))
01015
```

References amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.Alignment.get_missing_percent_from_seq(), amas.AMAS.Alignment.get_sites_no_missing_ambiguous(), amas.AMAS.Alignment.get_trim_selection(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the caller graph for this function:
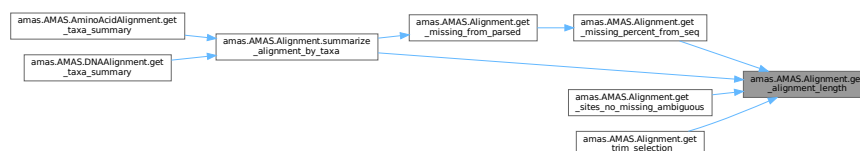
7.1.3.6 get_aln_input()

```
amas.AMAS.Alignment.get_aln_input (
            self )
```

Definition at line 826 of file AMAS.py.

```
00826    def get_aln_input(self):
00827        # open and parse input file
00828        aln_input = FileParser(self.in_file)
00829        return aln_input
00830
```

References amas.AMAS.FileHandler.in_file, amas.AMAS.FileParser.in_file, and amas.AMAS.Alignment.in_file.

Referenced by amas.AMAS.Alignment.get_parsed_aln().

Here is the caller graph for this function:

7.1.3.7 get_char_summary()

```
amas.AMAS.Alignment.get_char_summary (
            self )
```

Definition at line 899 of file AMAS.py.

```
00899    def get_char_summary(self):
00900        # get summary of frequencies for all characters
00901        characters = []
00902        counts = []
00903        add_to_chars = characters.append
00904        add_to_counts = counts.append
00905        char_count_dicts = self.get_counts()
00906        for char in self.alphabet:
00907            add_to_chars(char)
00908            if char in char_count_dicts.keys():
00909                add_to_counts(str(char_count_dicts[char]))
00910            else:
00911                add_to_counts("0")
00912        return characters, counts
00913
```

References amas.AMAS.AminoAcidAlignment.alphabet, amas.AMAS.DNAAlignment.alphabet, and amas.AMAS.Alignment.get_count

Referenced by amas.AMAS.AminoAcidAlignment.get_summary(), and amas.AMAS.DNAAlignment.get_summary().

Here is the call graph for this function:

Here is the caller graph for this function:
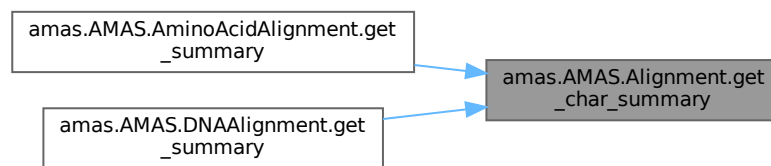
7.1.3.8 get_column()

```
amas.AMAS.Alignment.get_column (
            self,
            i )
```

Definition at line 933 of file AMAS.py.

```
00933    def get_column(self, i):
00934        # get site from the character matrix
00935        return [row[i] for row in self.matrix]
00936
```

References amas.AMAS.Alignment.matrix.

Referenced by amas.AMAS.Alignment.get_site_no_missing_ambiguous(), and amas.AMAS.Alignment.replace_missing().

Here is the caller graph for this function:

7.1.3.9 get_counts()

```
amas.AMAS.Alignment.get_counts (
            self )
```

Definition at line 1052 of file AMAS.py.

```
01052    def get_counts(self):
01053        # get counts of each character in the used alphabet for all sequences
01054        counters = [Counter(chars) for taxon, chars in self.get_counts_from_parsed()]
01055        all_counts = sum(counters, Counter())
01056        counts_dict = dict(all_counts)
01057        return counts_dict
01058
```

References amas.AMAS.Alignment.get_counts_from_parsed().

Referenced by amas.AMAS.Alignment.get_char_summary().

Here is the call graph for this function:

Here is the caller graph for this function:
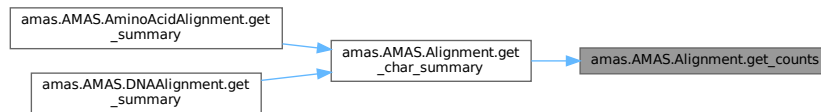
7.1.3.10 get_counts_from_parsed()

```
amas.AMAS.Alignment.get_counts_from_parsed (
            self )
```

Definition at line 1059 of file AMAS.py.

```
01059    def get_counts_from_parsed(self):
01060        # get counts of all characters from parsed alignment
01061        # return a list of tuples with taxon name and counts
01062        return sorted(
01063            [
01064                (taxon, self.get_counts_from_seq(seq))
01065                for taxon, seq in self.parsed_aln.items()
01066            ]
01067        )
01068
```

References amas.AMAS.Alignment.get_counts_from_seq(), and amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.Alignment.get_counts(), and amas.AMAS.Alignment.get_taxon_char_summary().

Here is the call graph for this function:

Here is the caller graph for this function:
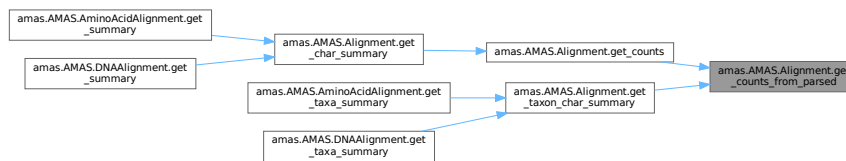
7.1.3.11 get_counts_from_seq()

```
amas.AMAS.Alignment.get_counts_from_seq (
            self,
            seq )
```

Definition at line 1069 of file AMAS.py.

```
01069     def get_counts_from_seq(self, seq):
01070         # get all alphabet chars count for individual sequence
01071         char_counts = {char : seq.count(char) for char in self.alphabet}
01072         return char_counts
01073
```

References amas.AMAS.AminoAcidAlignment.alphabet, and amas.AMAS.DNAAlignment.alphabet.

Referenced by amas.AMAS.Alignment.get_counts_from_parsed().

Here is the caller graph for this function:
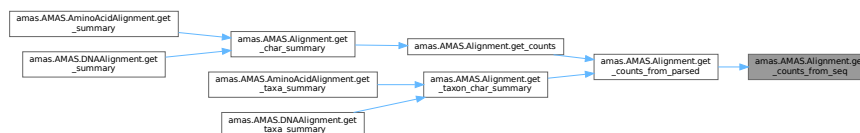
7.1.3.12 get_matrix_cells()

```
amas.AMAS.Alignment.get_matrix_cells (
            self )
```

Definition at line 1016 of file AMAS.py.

```
01016     def get_matrix_cells(self):
```

7.1.3.13 get_missing()

```
amas.AMAS.Alignment.get_missing (
            self )
```

Definition at line 1021 of file AMAS.py.

```
01021     def get_missing(self):
01022         # count missing characters from the list of missing for all sequences
01023         self.missing = sum(count for taxon, count, percent in self.missing_records)
01024         return self.missing
01025
```

7.1.3.14 get_missing_from_parsed()

amas.AMAS.Alignment.get_missing_from_parsed (
        self)

Definition at line 1031 of file AMAS.py.

```
01031    def get_missing_from_parsed(self):
01032        # get missing count and percent from parsed alignment
01033        # return a list of tuples with taxon name, count, and percent missing
01034        self.missing_records = sorted(
01035            [
01036                (taxon, self.get_missing_from_seq(seq), self.get_missing_percent_from_seq(seq))
01037                for taxon, seq in self.parsed_aln.items()
01038            ]
01039        )
01040        return self.missing_records
01041
```

References amas.AMAS.Alignment.get_missing_from_seq(), amas.AMAS.Alignment.get_missing_percent_from_seq(), amas.AMAS.Alignment.missing_records, and amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the call graph for this function:
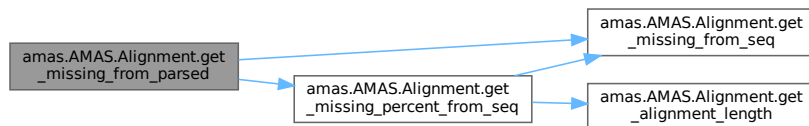
Here is the caller graph for this function:
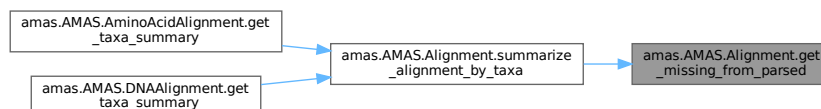
7.1.3.15 get_missing_from_seq()

amas.AMAS.Alignment.get_missing_from_seq (
        self,
        seq)

Definition at line 1042 of file AMAS.py.

```
01042    def get_missing_from_seq(self, seq):
01043        # count missing characters for individual sequence
01044        missing_count = sum(seq.count(char) for char in self.missing_chars)
01045        return missing_count
01046
```

References amas.AMAS.AminoAcidAlignment.missing_chars, and amas.AMAS.DNAAlignment.missing_chars.

Referenced by amas.AMAS.Alignment.get_missing_from_parsed(), and amas.AMAS.Alignment.get_missing_percent_from_seq().

Here is the caller graph for this function:

7.1.3.16 get_missing_percent()

```
amas.AMAS.Alignment.get_missing_percent (
            self )
```

Definition at line 1026 of file AMAS.py.

```
01026    def get_missing_percent(self):
01027        # get missing percent
01028        missing_percent = round((self.missing / self.all_matrix_cells * 100), 3)
01029        return missing_percent
01030
```

References amas.AMAS.Alignment.all_matrix_cells, and amas.AMAS.Alignment.missing.

7.1.3.17 get_missing_percent_from_seq()

```
amas.AMAS.Alignment.get_missing_percent_from_seq (
            self,
            seq )
```

Definition at line 1047 of file AMAS.py.

```
01047    def get_missing_percent_from_seq(self, seq):
01048        # get missing percent from individual sequence
01049        missing_seq_percent = round((self.get_missing_from_seq(seq) / self.get_alignment_length() *
    100), 3)
01050        return missing_seq_percent
01051
```

References amas.AMAS.Alignment.get_alignment_length(), and amas.AMAS.Alignment.get_missing_from_seq().

Referenced by amas.AMAS.Alignment.get_missing_from_parsed().

Here is the call graph for this function:
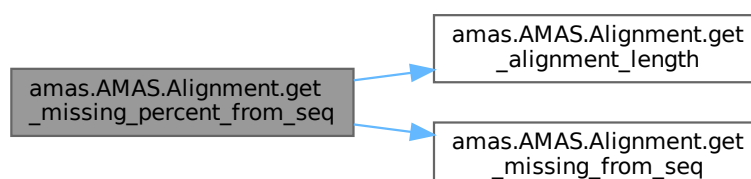
Here is the caller graph for this function:
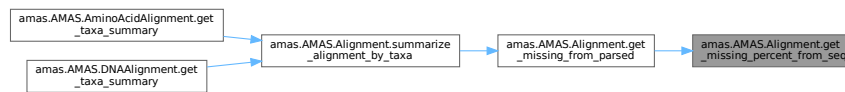
7.1.3.18 get_name()

```
amas.AMAS.Alignment.get_name (
                self )
```
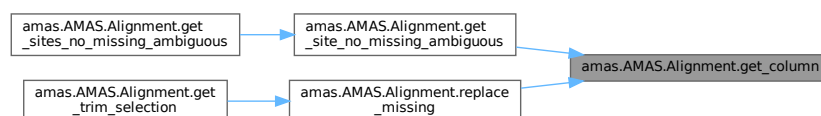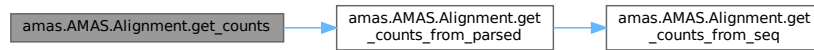
Definition at line 1002 of file AMAS.py.

```
01002      def get_name(self):
01003          # get input file name
01004          in_filename = path.basename(self.in_file)
01005          return in_filename
01006
```

References amas.AMAS.FileHandler.in_file, amas.AMAS.FileParser.in_file, and amas.AMAS.Alignment.in_file.

Referenced by amas.AMAS.Alignment.__str__(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the caller graph for this function:

7.1.3.19 get_parsed_aln()

```
amas.AMAS.Alignment.get_parsed_aln (
                self )
```

Definition at line 831 of file AMAS.py.

```
00831      def get_parsed_aln(self):
00832          # parse according to the given format
00833          aln_input = self.get_aln_input()
00834          if self.in_format == "fasta":
00835              parsed_aln = aln_input.fasta_parse()
00836          elif self.in_format == "phylip":
00837              parsed_aln = aln_input.phylip_parse()
00838          elif self.in_format == "phylip-int":
00839              parsed_aln = aln_input.phylip_interleaved_parse()
00840          elif self.in_format == "nexus":
00841              parsed_aln = aln_input.nexus_parse()
00842          elif self.in_format == "nexus-int":
00843              parsed_aln = aln_input.nexus_interleaved_parse()
00844
00845          return parsed_aln
00846
```

References amas.AMAS.Alignment.get_aln_input(), amas.AMAS.Alignment.in_format, and amas.AMAS.MetaAlignment.in_format.

Here is the call graph for this function:
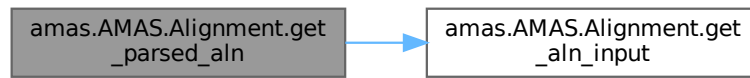
7.1.3.20 get_parsimony_informative()

```
amas.AMAS.Alignment.get_parsimony_informative (
                self )
```

Definition at line 978 of file AMAS.py.

```
00978     def get_parsimony_informative(self):
00979         # if the count for a unique character in a site is at least two,
00980         # and there are at least two such characters in a site without missing
00981         # or ambiguous characters, consider it parsimony informative
00982         parsimony_informative = 0
00983         for site in self.no_missing_ambiguous:
00984             unique_chars = set(site)
00985             pattern = [base for base in unique_chars if site.count(base) >= 2]
00986             no_patterns = len(pattern)
00987
00988             if no_patterns >= 2:
00989                 parsimony_informative += 1
00990         return parsimony_informative
00991
```

References amas.AMAS.Alignment.no_missing_ambiguous.

7.1.3.21 get_prop_parsimony()

```
amas.AMAS.Alignment.get_prop_parsimony (
                self )
```

Definition at line 997 of file AMAS.py.

```
00997     def get_prop_parsimony(self):
00998         # get proportion of parsimony informative sites to all sites
00999         prop_parsimony = self.parsimony_informative / int(self.length)
01000         return round(prop_parsimony, 3)
01001
```

References amas.AMAS.Alignment.length, and amas.AMAS.Alignment.parsimony_informative.

7.1.3.22 get_prop_variable()

```
amas.AMAS.Alignment.get_prop_variable (
                self )
```

Definition at line 992 of file AMAS.py.

```
00992     def get_prop_variable(self):
00993         # get proportion of variable sites to all sites
00994         prop_variable = self.variable_sites / int(self.length)
00995         return round(prop_variable, 3)
00996
```

References amas.AMAS.Alignment.length, and amas.AMAS.Alignment.variable_sites.

7.1.3.23 get_site_no_missing_ambiguous()

amas.AMAS.Alignment.get_site_no_missing_ambiguous (

       self,

       column)

Definition at line 946 of file AMAS.py.

```
00946    def get_site_no_missing_ambiguous(self, column):
00947        site = self.get_column(column)
00948        return [char for char in site if char not in self.missing_ambiguous_chars]
00949
```

References amas.AMAS.Alignment.get_column(), amas.AMAS.AminoAcidAlignment.missing_ambiguous_chars, and amas.AMAS.DNAAlignment.missing_ambiguous_chars.

Referenced by amas.AMAS.Alignment.get_sites_no_missing_ambiguous().

Here is the call graph for this function:
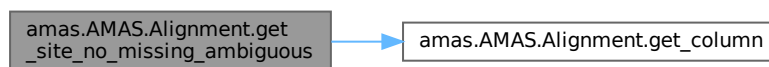
Here is the caller graph for this function:

7.1.3.24 get_sites_no_missing_ambiguous()

amas.AMAS.Alignment.get_sites_no_missing_ambiguous (

       self)

Definition at line 941 of file AMAS.py.

```
00941    def get_sites_no_missing_ambiguous(self):
00942        # get each site without missing or ambiguous characters
00943        no_missing_ambiguous_sites = [self.get_site_no_missing_ambiguous(column) for column in
        range(self.get_alignment_length())]
00944        return no_missing_ambiguous_sites
00945
```

References amas.AMAS.Alignment.get_alignment_length(), and amas.AMAS.Alignment.get_site_no_missing_ambiguous().

Here is the call graph for this function:
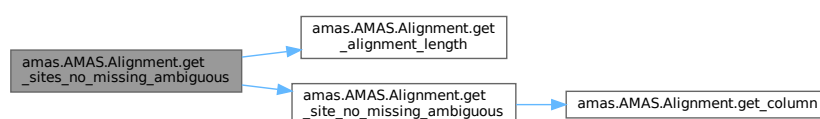
7.1.3.25 get_taxa_no()

```
amas.AMAS.Alignment.get_taxa_no (
            self )
```

Definition at line 1007 of file AMAS.py.

```
01007     def get_taxa_no(self):
01008         # get number of taxa
01009         return len(self.parsed_aln.values())
01010
```

References amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.DNAAlignment.get_atgc_content(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the caller graph for this function:
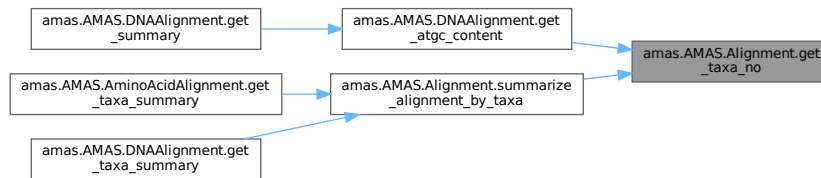
7.1.3.26 get_taxon_char_summary()

```
amas.AMAS.Alignment.get_taxon_char_summary (
            self )
```

Definition at line 914 of file AMAS.py.

```
00914     def get_taxon_char_summary(self):
00915         # get summary of frequencies for all characters
00916         records = (self.append_count(char_dict) for taxon, char_dict in self.get_counts_from_parsed())
00917         return records
00918
```

References amas.AMAS.Alignment.append_count(), and amas.AMAS.Alignment.get_counts_from_parsed().

Referenced by amas.AMAS.AminoAcidAlignment.get_taxa_summary(), and amas.AMAS.DNAAlignment.get_taxa_summary().

Here is the call graph for this function:
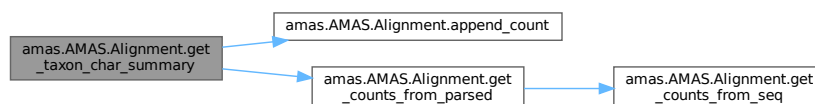
Here is the caller graph for this function:
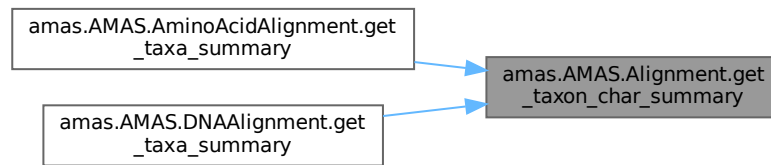
7.1.3.27 get_trim_selection()

```
amas.AMAS.Alignment.get_trim_selection (
                self,
                trim_fraction,
                parsimony_check )
```

Definition at line 953 of file AMAS.py.

```
00953     def get_trim_selection(self, trim_fraction, parsimony_check):
00954         # this checks each column of alignment for minimum occupancy
00955         self.matrix = self.matrix_creator()
00956         trim_vector = []
00957         for column in range(self.get_alignment_length()):
00958             site = self.replace_missing(column)
00959             occ = (len(site) - site.count("-")) / len(site)
00960             if parsimony_check:
00961                 unique_chars = set(site)
00962                 try:
00963                     unique_chars.remove("-")
00964                 except KeyError:
00965                     pass # this occurs if we have no missing data
00966                 pattern = [base for base in unique_chars if site.count(base) >= 2]
00967                 trim_vector.append(len(pattern) >= 2 and occ >= trim_fraction)
00968             else:
00969                 trim_vector.append(occ >= trim_fraction)
00970         return trim_vector
00971
```

References amas.AMAS.Alignment.get_alignment_length(), amas.AMAS.Alignment.matrix, amas.AMAS.Alignment.matrix_creator(), and amas.AMAS.Alignment.replace_missing().

Here is the call graph for this function:
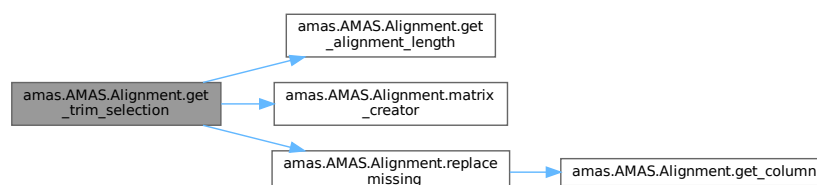
7.1.3.28 get_variable()

```
amas.AMAS.Alignment.get_variable (
          self )
```
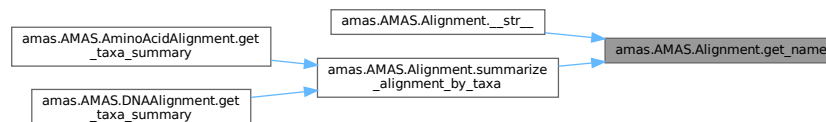
Definition at line 972 of file AMAS.py.

```
00972     def get_variable(self):
00973         # if all elements of a site without missing or ambiguous characters
00974         # are not the same, consider it variable
00975         variable = len([site for site in self.no_missing_ambiguous if not self.all_same(site)])
00976         return variable
00977
```

References amas.AMAS.Alignment.all_same(), and amas.AMAS.Alignment.no_missing_ambiguous.

Here is the call graph for this function:
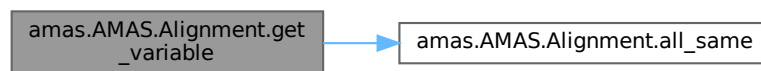
7.1.3.29 matrix_creator()

```
amas.AMAS.Alignment.matrix_creator (
          self )
```

Definition at line 928 of file AMAS.py.

```
00928     def matrix_creator(self):
00929         # decompose character matrix into a two-dimensional list
00930         matrix = [list(sequence) for sequence in self.parsed_aln.values()]
00931         return matrix
00932
```

References amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.Alignment.get_trim_selection().

Here is the caller graph for this function:

7.1.3.30 replace_missing()

```
amas.AMAS.Alignment.replace_missing (
            self,
            column )
```

Definition at line 950 of file AMAS.py.

```
00950     def replace_missing(self, column):
00951         return ["-" if x in self.missing_chars else x for x in self.get_column(column)]
00952
```

References amas.AMAS.Alignment.get_column(), amas.AMAS.AminoAcidAlignment.missing_chars, and amas.AMAS.DNAAlignment.missing_chars.

Referenced by amas.AMAS.Alignment.get_trim_selection().

Here is the call graph for this function:
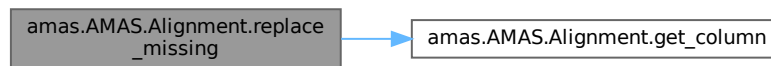
Here is the caller graph for this function:

7.1.3.31 summarize_alignment()

```
amas.AMAS.Alignment.summarize_alignment (
            self )
```

Definition at line 847 of file AMAS.py.

```
00847     def summarize_alignment(self):
00848         # call methods to create sequences list, matrix, sites without ambiguous or
00849         # missing characters; get and summarize alignment statistics
00850         summary = []
00851         self.length = str(self.get_alignment_length())
00852         self.matrix = self.matrix_creator()
00853         self.no_missing_ambiguous = self.get_sites_no_missing_ambiguous()
00854         self.variable_sites = self.get_variable()
00855         self.prop_variable = self.get_prop_variable()
00856         self.parsimony_informative = self.get_parsimony_informative()
00857         self.prop_parsimony = self.get_prop_parsimony()
00858         self.missing_records = self.get_missing_from_parsed()
00859         name = str(self.get_name())
00860         taxa_no = str(self.get_taxa_no())
00861         cells = str(self.get_matrix_cells())
00862         missing = str(self.get_missing())
00863         missing_percent = str(self.get_missing_percent())
```

```
00864          self.check_data_type()
00865          summary = [
00866              name,
00867              taxa_no,
00868              self.length,
00869              cells,
00870              missing,
00871              missing_percent,
00872              str(self.variable_sites),
00873              str(self.prop_variable),
00874              str(self.parsimony_informative),
00875              str(self.prop_parsimony)
00876          ]
00877          return summary
00878
```

Referenced by amas.AMAS.AminoAcidAlignment.get_summary(), and amas.AMAS.DNAAlignment.get_summary().

Here is the caller graph for this function:
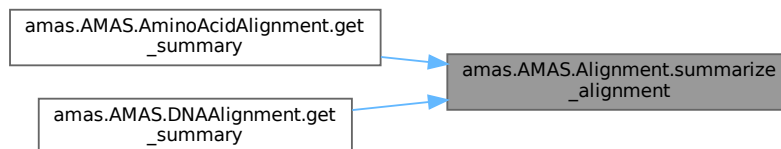
7.1.3.32 summarize_alignment_by_taxa()

```
amas.AMAS.Alignment.summarize_alignment_by_taxa (
              self )
```

Definition at line 879 of file AMAS.py.

```
00879      def summarize_alignment_by_taxa(self):
00880          # get summary for all taxa/sequences in alignment
00881          per_taxon_summary = []
00882          taxa_no = self.get_taxa_no()
00883          self.missing_records = self.get_missing_from_parsed()
00884          self.length = self.get_alignment_length()
00885          lengths = (self.length for i in range(taxa_no))
00886          name = self.get_name()
00887          names = (name for i in range(taxa_no))
00888          taxa_names = (
00889              taxon.replace(" ", "_").replace(".", "_").replace("'", "")
00890              for taxon, missing_count, missing_percent in self.missing_records
00891          )
00892          missing = (missing_count for taxon, missing_count, missing_percent in self.missing_records)
00893          missing_percent = (missing_percent for taxon, missing_count, missing_percent in
      self.missing_records)
00894          self.check_data_type()
00895          per_taxon_summary = (names, taxa_names, lengths, missing, missing_percent)
00896          zipped = list(zip(*per_taxon_summary))
00897          return zipped
00898
```

References amas.AMAS.Alignment.check_data_type(), amas.AMAS.Alignment.get_alignment_length(), amas.AMAS.Alignment.get_ amas.AMAS.Alignment.get_name(), amas.AMAS.Alignment.get_taxa_no(), amas.AMAS.Alignment.length, and amas.AMAS.Alignment.missing_records.

Referenced by amas.AMAS.AminoAcidAlignment.get_taxa_summary(), and amas.AMAS.DNAAlignment.get_taxa_summary().

Here is the call graph for this function:
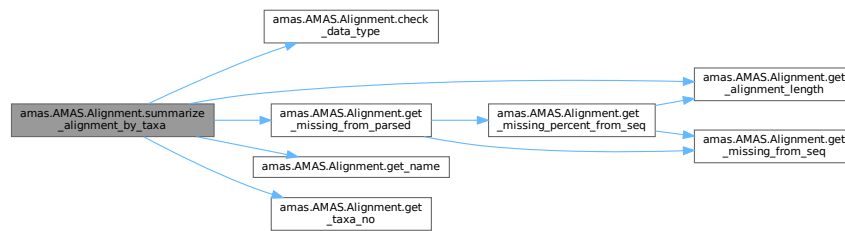
Here is the caller graph for this function:
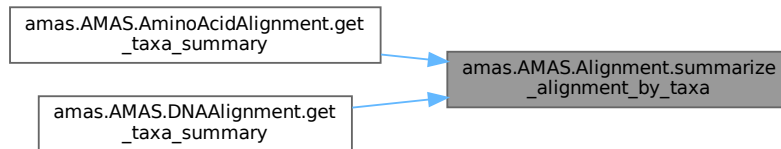
7.1.4 Member Data Documentation

7.1.4.1 all_matrix_cells

```
amas.AMAS.Alignment.all_matrix_cells  [static]
```

Definition at line 1018 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_missing_percent().

7.1.4.2 check

```
amas.AMAS.Alignment.check
```

Definition at line 1077 of file AMAS.py.

7.1.4.3 data_type

```
amas.AMAS.Alignment.data_type
```

Definition at line 818 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_object(), amas.AMAS.MetaAlignment.get_summaries(), amas.AMAS.MetaAlignment.get_taxon_summaries(), amas.AMAS.MetaAlignment.print_nexus(), amas.AMAS.MetaAlignment.print_r and amas.AMAS.MetaAlignment.write_out().

7.1.4.4 in_file

`amas.AMAS.Alignment.in_file`

Definition at line 816 of file AMAS.py.

Referenced by amas.AMAS.FileHandler.__exit__(), amas.AMAS.Alignment.get_aln_input(), and amas.AMAS.Alignment.get_name().

7.1.4.5 in_format

`amas.AMAS.Alignment.in_format`

Definition at line 817 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_object(), amas.AMAS.Alignment.get_parsed_aln(), and amas.AMAS.MetaAlignment.write_out().

7.1.4.6 length

`amas.AMAS.Alignment.length`

Definition at line 851 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_prop_parsimony(), amas.AMAS.Alignment.get_prop_variable(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

7.1.4.7 matrix

`amas.AMAS.Alignment.matrix`

Definition at line 852 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_column(), and amas.AMAS.Alignment.get_trim_selection().

7.1.4.8 missing

`amas.AMAS.Alignment.missing`

Definition at line 1023 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_missing_percent().

7.1.4.9 missing_records

`amas.AMAS.Alignment.missing_records`

Definition at line 858 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_missing_from_parsed(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

7.1.4.10 no_missing_ambiguous

`amas.AMAS.Alignment.no_missing_ambiguous`

Definition at line 853 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_parsimony_informative(), and amas.AMAS.Alignment.get_variable().

7.1.4.11 parsed_aln

`amas.AMAS.Alignment.parsed_aln`

Definition at line 820 of file AMAS.py.

Referenced by amas.AMAS.Alignment.check_data_type(), amas.AMAS.Alignment.get_alignment_length(), amas.AMAS.DNAAlignment.get_atgc_from_parsed(), amas.AMAS.Alignment.get_counts_from_parsed(), amas.AMAS.Alignment.get amas.AMAS.Alignment.get_taxa_no(), and amas.AMAS.Alignment.matrix_creator().

7.1.4.12 parsimony_informative

`amas.AMAS.Alignment.parsimony_informative`

Definition at line 856 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_prop_parsimony().

7.1.4.13 prop_parsimony

`amas.AMAS.Alignment.prop_parsimony`

Definition at line 857 of file AMAS.py.

7.1.4.14 prop_variable

`amas.AMAS.Alignment.prop_variable`

Definition at line 855 of file AMAS.py.

7.1.4.15 variable_sites

`amas.AMAS.Alignment.variable_sites`

Definition at line 854 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_prop_variable().

The documentation for this class was generated from the following file:

- amas/AMAS.py

7.2 amas.AMAS.AminoAcidAlignment Class Reference

Inheritance diagram for amas.AMAS.AminoAcidAlignment:

```
          ┌─────────────────────────────┐
          │     amas.AMAS.Alignment     │
          └─────────────────────────────┘
                         ▲
                         │
          ┌─────────────────────────────┐
          │ amas.AMAS.AminoAcidAlignment │
          └─────────────────────────────┘
```

Collaboration diagram for amas.AMAS.AminoAcidAlignment:
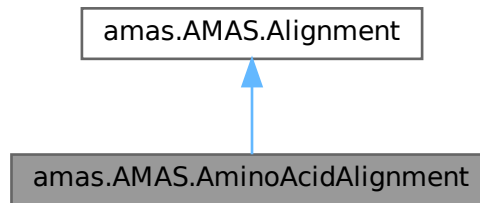
```
          ┌─────────────────────────────┐
          │     amas.AMAS.Alignment     │
          └─────────────────────────────┘
                         ▲
                         │
          ┌─────────────────────────────┐
          │ amas.AMAS.AminoAcidAlignment │
          └─────────────────────────────┘
```
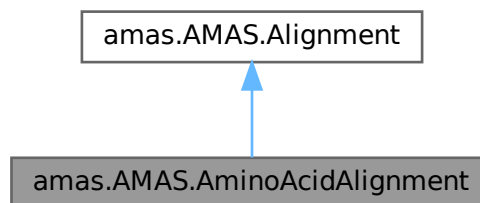
Public Member Functions

- get_summary (self)
- get_taxa_summary (self)

Static Public Attributes

- list alphabet = ["A", "C", "D", "E", "F", "G", "H", "I", "K", "L", "M", "N", "P", "Q", "R", "S", "T", "V", "W", "Y", "B", "J", "Z", "X", ".", "∗", "-", "?"]
- list missing_ambiguous_chars = ["B", "J", "Z", "X", ".", "∗", "-", "?"]
- list missing_chars = ["X", ".", "∗", "-", "?"]
- list non_alphabet = ["O"]

7.2.1 Detailed Description

```
Alphabets specific to amino acid alignments
```

Definition at line 1085 of file AMAS.py.

7.2.2 Member Function Documentation

7.2.2.1 get_summary()

```
amas.AMAS.AminoAcidAlignment.get_summary (
            self )
```

Definition at line 1093 of file AMAS.py.
```
01093    def get_summary(self):
01094        # get alignment summary specific to amino acids
01095        data = self.summarize_alignment()
01096        new_data = data + list(self.get_char_summary()[1])
01097        return new_data
01098
```

References amas.AMAS.Alignment.get_char_summary(), and amas.AMAS.Alignment.summarize_alignment().

Here is the call graph for this function:

7.2.2.2 get_taxa_summary()

```
amas.AMAS.AminoAcidAlignment.get_taxa_summary (
            self )
```

Definition at line 1099 of file AMAS.py.
```
01099    def get_taxa_summary(self):
01100        # get per-taxon/sequence alignment summary specific to amino acids
01101        data = self.summarize_alignment_by_taxa()
01102        aa_summary = (data, self.get_taxon_char_summary())
01103        zipped_list = list(zip(*aa_summary))
01104        new_data = [list(data_tupl) + chars for data_tupl, chars in zipped_list]
01105        return new_data
01106
```

References amas.AMAS.Alignment.get_taxon_char_summary(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the call graph for this function:
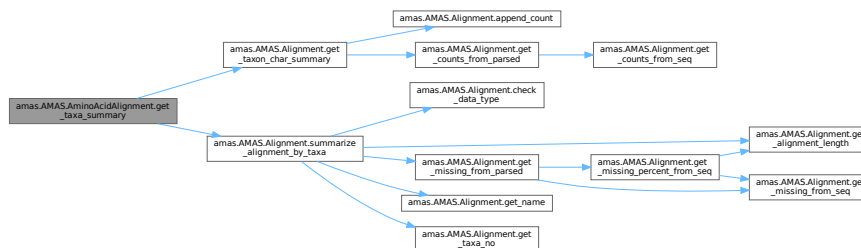
7.2.3 Member Data Documentation

7.2.3.1 alphabet

```
list amas.AMAS.AminoAcidAlignment.alphabet = ["A", "C", "D", "E", "F", "G", "H", "I", "K",
"L", "M", "N", "P", "Q", "R", "S", "T", "V", "W", "Y", "B", "J", "Z", "X", ".", "*", "-", "?"]
[static]
```

Definition at line 1088 of file AMAS.py.

Referenced by amas.AMAS.Alignment.append_count(), amas.AMAS.Alignment.get_char_summary(), and amas.AMAS.Alignment.get_counts_from_seq().

7.2.3.2 missing_ambiguous_chars

```
list amas.AMAS.AminoAcidAlignment.missing_ambiguous_chars = ["B", "J", "Z", "X", ".", "*",
"-", "?"] [static]
```

Definition at line 1089 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_site_no_missing_ambiguous().

7.2.3.3 missing_chars

```
list amas.AMAS.AminoAcidAlignment.missing_chars = ["X", ".", "*", "-", "?"] [static]
```

Definition at line 1090 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_missing_from_seq(), and amas.AMAS.Alignment.replace_missing().

7.2.3.4 non_alphabet

```
list amas.AMAS.AminoAcidAlignment.non_alphabet = ["O"] [static]
```

Definition at line 1091 of file AMAS.py.

The documentation for this class was generated from the following file:

- amas/AMAS.py

7.3 amas.AMAS.DNAAlignment Class Reference

Inheritance diagram for amas.AMAS.DNAAlignment:

```
amas.AMAS.Alignment
        ↑
amas.AMAS.DNAAlignment
```

Collaboration diagram for amas.AMAS.DNAAlignment:
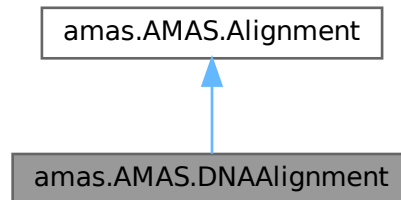
```
amas.AMAS.Alignment
        ↑
amas.AMAS.DNAAlignment
```
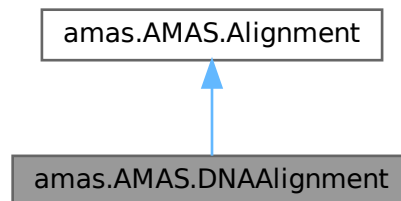
Public Member Functions

- get_summary (self)
- get_taxa_summary (self)
- get_atgc_content (self)
- get_list_from_atgc (self)
- get_atgc_from_parsed (self)
- get_atgc_from_seq (self, seq)

Static Public Attributes

- list alphabet = ["A", "C", "G", "T", "K", "M", "R", "Y", "S", "W", "B", "V", "H", "D", "X", "N", "O", "-", "?"]
- list missing_ambiguous_chars = ["K", "M", "R", "Y", "S", "W", "B", "V", "H", "D", "X", "N", "O", "-", "?"]
- list missing_chars = ["X", "N", "O", "-", "?"]
- list non_alphabet = ["E", "F", "I", "L", "P", "Q", "J", "Z", ".", "∗"]

7.3.1 Detailed Description

```
Alphabets specific to DNA alignments
```

Definition at line 1107 of file AMAS.py.

7.3.2 Member Function Documentation

7.3.2.1 get_atgc_content()

```
amas.AMAS.DNAAlignment.get_atgc_content (
              self )
```

Definition at line 1129 of file AMAS.py.

```
01129      def get_atgc_content(self):
01130          # get AC and GC contents for all sequences
01131          # AT content is the first element of AT, GC content tuple
01132          # returned by get_atgc_from_seq()
01133          atgc_records = self.get_atgc_from_parsed()
01134          at_content = round(sum(atgc[0] for taxon, atgc in atgc_records) / self.get_taxa_no(), 3)
01135          gc_content = round(1 - float(at_content), 3)
01136
01137          atgc_content = [str(at_content), str(gc_content)]
01138          return atgc_content
01139
```

References amas.AMAS.DNAAlignment.get_atgc_from_parsed(), and amas.AMAS.Alignment.get_taxa_no().

Referenced by amas.AMAS.DNAAlignment.get_summary().

Here is the call graph for this function:
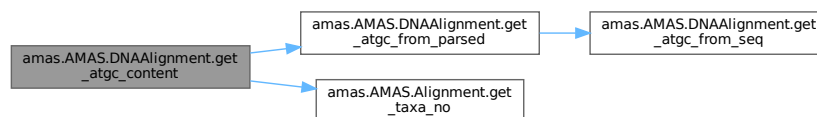
Here is the caller graph for this function:

7.3.2.2 get_atgc_from_parsed()

```
amas.AMAS.DNAAlignment.get_atgc_from_parsed (
            self )
```

Definition at line 1144 of file AMAS.py.

```
01144     def get_atgc_from_parsed(self):
01145         # get AT and GC contents from parsed alignment dictionary
01146         # return a list of tuples with taxon name, AT content, and GC content
01147         return sorted([(taxon, self.get_atgc_from_seq(seq)) for taxon, seq in
    self.parsed_aln.items()])
01148
```

References amas.AMAS.DNAAlignment.get_atgc_from_seq(), and amas.AMAS.Alignment.parsed_aln.

Referenced by amas.AMAS.DNAAlignment.get_atgc_content(), and amas.AMAS.DNAAlignment.get_list_from_atgc().

Here is the call graph for this function:

Here is the caller graph for this function:
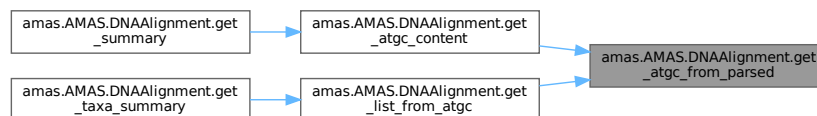
7.3.2.3 get_atgc_from_seq()

```
amas.AMAS.DNAAlignment.get_atgc_from_seq (
            self,
            seq )
```

Definition at line 1149 of file AMAS.py.

```
01149     def get_atgc_from_seq(self, seq):
01150         # get AT and GC contents from individual sequences
01151
01152         at_count = seq.count("A") + seq.count("T") + seq.count("W")
01153         gc_count = seq.count("G") + seq.count("C") + seq.count("S")
01154
01155         try:
01156             at_content = round(at_count / (at_count + gc_count), 3)
01157             gc_content = round(1 - float(at_content), 3)
01158
01159         except ZeroDivisionError:
01160             at_content = 0
01161             gc_content = 0
01162
01163         return at_content, gc_content
```

01164

Referenced by amas.AMAS.DNAAlignment.get_atgc_from_parsed().

Here is the caller graph for this function:
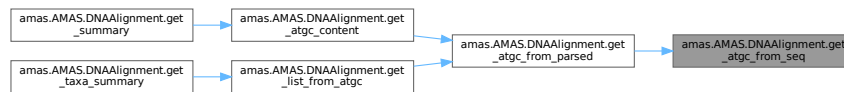
7.3.2.4 get_list_from_atgc()

```
amas.AMAS.DNAAlignment.get_list_from_atgc (
              self )
```

Definition at line 1140 of file AMAS.py.

```
01140    def get_list_from_atgc(self):
01141        records = (atgc for taxon, atgc in self.get_atgc_from_parsed())
01142        return records
01143
```

References amas.AMAS.DNAAlignment.get_atgc_from_parsed().

Referenced by amas.AMAS.DNAAlignment.get_taxa_summary().

Here is the call graph for this function:
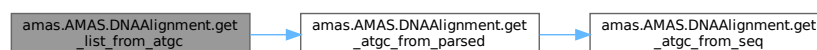
Here is the caller graph for this function:

7.3.2.5 get_summary()

```
amas.AMAS.DNAAlignment.get_summary (
            self )
```

Definition at line 1115 of file AMAS.py.

```
01115    def get_summary(self):
01116        # get alignment summarry specific to nucleotide
01117        data = self.summarize_alignment()
01118        new_data = data + self.get_atgc_content() + list(self.get_char_summary()[1])
01119        return new_data
01120
```

References amas.AMAS.DNAAlignment.get_atgc_content(), amas.AMAS.Alignment.get_char_summary(), and amas.AMAS.Alignment.summarize_alignment().

Here is the call graph for this function:
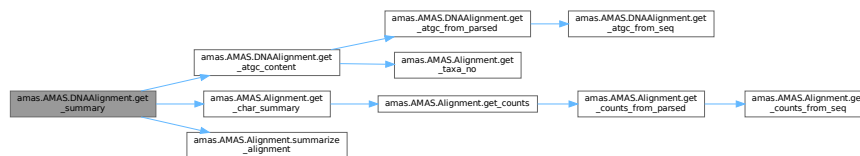
7.3.2.6 get_taxa_summary()

```
amas.AMAS.DNAAlignment.get_taxa_summary (
            self )
```

Definition at line 1121 of file AMAS.py.

```
01121    def get_taxa_summary(self):
01122        # get per-taxon/sequence alignment summary specific to nucleotides
01123        data = self.summarize_alignment_by_taxa()
01124        dna_summary = (data, self.get_list_from_atgc(), self.get_taxon_char_summary())
01125        zipped_list = list(zip(*dna_summary))
01126        new_data = [list(data_tupl) + list(atgc) + chars for data_tupl, atgc, chars in zipped_list]
01127        return new_data
01128
```

References amas.AMAS.DNAAlignment.get_list_from_atgc(), amas.AMAS.Alignment.get_taxon_char_summary(), and amas.AMAS.Alignment.summarize_alignment_by_taxa().

Here is the call graph for this function:
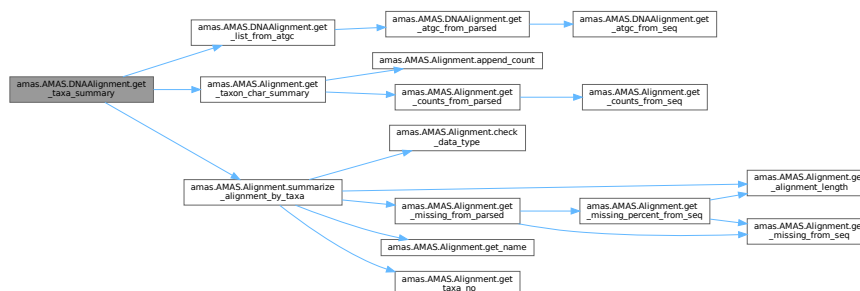
7.3.3 Member Data Documentation

7.3.3.1 alphabet

```
list amas.AMAS.DNAAlignment.alphabet = ["A", "C", "G", "T", "K", "M", "R", "Y", "S", "W", "B",
"V", "H", "D", "X", "N", "O", "-", "?"]  [static]
```

Definition at line 1110 of file AMAS.py.

Referenced by amas.AMAS.Alignment.append_count(), amas.AMAS.Alignment.get_char_summary(), and amas.AMAS.Alignment.get_counts_from_seq().

7.3.3.2 missing_ambiguous_chars

```
list amas.AMAS.DNAAlignment.missing_ambiguous_chars = ["K", "M", "R", "Y", "S", "W", "B", "V",
"H", "D", "X", "N", "O", "-", "?"]  [static]
```

Definition at line 1111 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_site_no_missing_ambiguous().

7.3.3.3 missing_chars

```
list amas.AMAS.DNAAlignment.missing_chars = ["X", "N", "O", "-", "?"]  [static]
```

Definition at line 1112 of file AMAS.py.

Referenced by amas.AMAS.Alignment.get_missing_from_seq(), and amas.AMAS.Alignment.replace_missing().

7.3.3.4 non_alphabet

```
list amas.AMAS.DNAAlignment.non_alphabet = ["E", "F", "I", "L", "P", "Q", "J", "Z", ".", "*"]
[static]
```

Definition at line 1113 of file AMAS.py.

The documentation for this class was generated from the following file:

- amas/AMAS.py

7.4 amas.AMAS.FileHandler Class Reference

Public Member Functions

- __init__ (self, file_name)
- __enter__ (self)
- __exit__ (self, ∗args)
- get_file_name (self)

Public Attributes

- file_name
- in_file

7.4.1 Detailed Description

```
Define file handle that closes when out of scope
```

Definition at line 518 of file AMAS.py.

7.4.2 Constructor & Destructor Documentation

7.4.2.1 __init__()

```
amas.AMAS.FileHandler.__init__ (
             self,
             file_name )
```

Definition at line 521 of file AMAS.py.
```
00521     def __init__(self, file_name):
00522         self.file_name = file_name
00523
```

7.4.3 Member Function Documentation

7.4.3.1 __enter__()

```
amas.AMAS.FileHandler.__enter__ (
             self )
```

Definition at line 524 of file AMAS.py.
```
00524     def __enter__(self):
00525         try:
00526             self.in_file = open(self.file_name, "r", encoding="utf-8")
00527         except FileNotFoundError:
00528             print("ERROR: File '" + self.file_name + "' not found.")
00529             sys.exit()
00530         return self.in_file
00531
```

7.4.3.2 __exit__()

```
amas.AMAS.FileHandler.__exit__ (
             self,
             * args )
```

Definition at line 532 of file AMAS.py.
```
00532     def __exit__(self, *args):
00533         self.in_file.close()
00534
```

References amas.AMAS.FileHandler.in_file, amas.AMAS.FileParser.in_file, and amas.AMAS.Alignment.in_file.

7.4.3.3 get_file_name()

```
amas.AMAS.FileHandler.get_file_name (
            self )
```

Definition at line 535 of file AMAS.py.
```
00535     def get_file_name(self):
00536         return self.file_name
00537
```

References amas.AMAS.FileHandler.file_name.

7.4.4 Member Data Documentation

7.4.4.1 file_name

```
amas.AMAS.FileHandler.file_name
```

Definition at line 522 of file AMAS.py.

Referenced by amas.AMAS.FileHandler.get_file_name().

7.4.4.2 in_file

```
amas.AMAS.FileHandler.in_file
```

Definition at line 526 of file AMAS.py.

Referenced by amas.AMAS.FileHandler.__exit__(), amas.AMAS.Alignment.get_aln_input(), and amas.AMAS.Alignment.get_name().

The documentation for this class was generated from the following file:

- amas/AMAS.py

7.5 amas.AMAS.FileParser Class Reference

Public Member Functions

- __init__ (self, in_file)
- fasta_parse (self)
- phylip_parse (self)
- phylip_interleaved_parse (self)
- nexus_parse (self)
- nexus_interleaved_parse (self)
- translate_ambiguous (self, seq)
- partitions_parse (self)

Public Attributes

- in_file
- in_file_lines

Static Public Attributes

- tax_chars_matches
- name_matches
- seq_matches
- tax_match = match.group(2)
- chars_match = match.group(3)
- list taxa = [ ]
- list sequences = [ ]
- dict records = {}
- int counter = 0
- name_match = match.group(2).replace("\n", "")
- seq_match = match.group(3).replace("\n", "").upper()
- matches
- str seq_match = "".join(seq_match.split())
- str sequence = ""

7.5.1 Detailed Description

```
Parse file contents and return sequences and sequence names
```

Definition at line 538 of file AMAS.py.

7.5.2 Constructor & Destructor Documentation

7.5.2.1 __init__()

```
amas.AMAS.FileParser.__init__ (
            self,
            in_file )
```

Definition at line 541 of file AMAS.py.
```
00541    def __init__(self, in_file):
00542        self.in_file = in_file
00543        with FileHandler(in_file) as handle:
00544            self.in_file_lines = handle.read().rstrip("\r\n")
00545
```

7.5.3 Member Function Documentation

7.5.3.1 fasta_parse()

```
amas.AMAS.FileParser.fasta_parse (
            self )
```

Definition at line 546 of file AMAS.py.
```
00546    def fasta_parse(self):
00547        # use regex to parse names and sequences in sequential fasta files
00548        matches = re.finditer(
00549            r"^>(.*[^$])([^>]*)",
00550            self.in_file_lines, re.MULTILINE
00551        )
00552        records = {}
00553
00554        for match in matches:
```

```
00555            name_match = match.group(1).replace("\n", "")
00556            seq_match = match.group(2).replace("\n", "").upper()
00557            seq_match = self.translate_ambiguous(seq_match)
00558            records[name_match] = seq_match
00559
00560        return records
00561
```

References amas.AMAS.FileParser.in_file_lines, and amas.AMAS.FileParser.translate_ambiguous().

Here is the call graph for this function:

7.5.3.2 nexus_interleaved_parse()

```
amas.AMAS.FileParser.nexus_interleaved_parse (
            self )
```

Definition at line 671 of file AMAS.py.
```
00671    def nexus_interleaved_parse(self):
00672        # use regex to parse names and sequences in sequential nexus files
00673        # find the matrix block
00674        matches = re.finditer(
00675            r"(\s+)?(MATRIX\n|matrix\n|MATRIX\r\n|matrix\r\n)(.*?;)",
00676            self.in_file_lines, re.DOTALL
00677        )
00678        # initiate lists for taxa names and sequence strings on separate lines
00679        taxa = []
00680        sequences = []
00681        # initiate a dictionary for the name:sequence records
00682        records = {}
00683
00684        for match in matches:
00685            matrix_match = match.group(3)
00686            # get names and sequences from the matrix block
00687            seq_matches = re.finditer(
00688                r"^(\s+)?['']?(\S+\s\S+|\S+)['']?\s+([A-Za-z*?.{}-]+)($|\s+\[[0-9]+\]$)",
00689                matrix_match, re.MULTILINE
00690            )
00691
00692            for match in seq_matches:
00693                name_match = match.group(2)
00694                if name_match not in taxa:
00695                    taxa.append(name_match)
00696                seq_match = match.group(3)
00697
00698                sequences.append(seq_match)
00699
00700        # initiate a counter to keep track of sequences strung together
00701        # from separate lines
00702        counter = 0
00703
00704        for taxon_no in range(len(taxa)):
00705
00706            full_length_sequence = "".join([sequences[index] for index in
    range(counter,len(sequences),len(taxa))])
00707            records[taxa[taxon_no]] = self.translate_ambiguous(full_length_sequence).replace("\n",
    "").upper()
00708            counter += 1
00709
00710        return records
00711
```

References amas.AMAS.FileParser.in_file_lines, and amas.AMAS.FileParser.translate_ambiguous().

Here is the call graph for this function:

7.5.3.3 nexus_parse()

```
amas.AMAS.FileParser.nexus_parse (
            self )
```

Definition at line 645 of file AMAS.py.

```
00645    def nexus_parse(self):
00646        # use regex to parse names and sequences in sequential nexus files
00647        # find the matrix block
00648        matches = re.finditer(
00649            r"(\s+)?(MATRIX\n|matrix\n|MATRIX\r\n|matrix\r\n)(.*?;)",
00650            self.in_file_lines, re.DOTALL
00651        )
00652
00653        records = {}
00654        # get names and sequences from the matrix block
00655
00656        for match in matches:
00657            matrix_match = match.group(3)
00658            seq_matches = re.finditer(
00659                r"^(\s+)?['']?(\S+\s\S+|\S+)['']?\s+([A-Za-z*?.{}-]+)($|\s+\[[0-9]+\]$)",
00660                matrix_match, re.MULTILINE
00661            )
00662
00663            for match in seq_matches:
00664                name_match = match.group(2).replace("\n", "")
00665                seq_match = match.group(3).replace("\n", "").upper()
00666                seq_match = self.translate_ambiguous(seq_match)
00667                records[name_match] = seq_match
00668
00669        return records
00670
```

References amas.AMAS.FileParser.in_file_lines, and amas.AMAS.FileParser.translate_ambiguous().

Here is the call graph for this function:

7.5.3.4 partitions_parse()

```
amas.AMAS.FileParser.partitions_parse (
            self )
```

Definition at line 731 of file AMAS.py.

```
00731     def partitions_parse(self):
00732         # parse partitions file using regex
00733         # original: `matches = re.finditer(r"^(\s+)?([^ =]+)[ =]+([\\0-9, -]+)", self.in_file_lines,
      re.MULTILINE)`
00734         # new version: more permissive -> handles PartionFinder/RAxML/(IQ-TREE 2)best_scheme.nex
      format partition files
00735         matches = re.finditer(
00736             r"""^[ \t]*                               # start of line w/ zero-or-more (just)
      whitespaces/tabs
00737                 (
00738                     (?P<nexus>charset[ ]+)            # case 1: (IQ-TREE 2)best_scheme.nex partition
      directive; partition name
00739                     |
00740                     (?P<raxml>[A-Za-z0-9_+\.]+,[ \t]+)    # case 2: RAxML/RAxML-NG model(+other pars);
      partition name
00741                 )?
00742                 (?P<partition_name>[A-Za-z0-9_@*&()\[\]{}'"%#|-]+) # partition name (handles various
      metcharacters, excludes [\s],<>?/:;~`\^$!)
00743                 [ ]*=[ ]*                             # whitespace-padded (or unpadded) '=':
      (IQ-TREE 2)best_scheme.nex compatabiliy
00744                 (?P<numbers>[\\0-9, -]+)              # position ranges w/ stride (multiple
      intervals; from original regex)
00745                 (?P<nexus_term>[ ]*[;])?              # whitespace-prepended (or unprepended) ';'
      (nexus terminator)
00746             """,
00747             self.in_file_lines,
00748             re.MULTILINE | re.VERBOSE
00749         )
00750
00751         # initiate list to store dictionaries with lists
00752         # of slice positions as values
00753         partitions = []
00754         add_to_partitions = partitions.append
00755
00756         for match in matches:
00757             # initiate dictionary of partition name as key
00758             dict_of_dicts = {}
00759             # and list of dictionaries with slice positions
00760             list_of_dicts = []
00761             add_to_list_of_dicts = list_of_dicts.append
00762             # get parition name and numbers from parsed partition strings
00763             partition_name = match.group('partition_name')
00764             numbers = match.group('numbers')
00765             # remove any whitespace padding '-' (to be consistent with partition-writing format)
00766             numbers = re.sub(r"[ ]*-[ ]*", "-", numbers)
00767             # find all numbers that will be used to parse positions
00768             positions = re.findall(r"([^ ,]+)", numbers)
00769
00770             for position in positions:
00771                 # create dictionary for slicing input sequence
00772                 # conditioning on whether positions are represented
00773                 # by range, range with stride, or single number
00774                 pos_dict = {}
00775
00776                 if "-" in position:
00777                     m = re.search(r"([0-9]+)-([0-9]+)", position)
00778                     pos_dict["start"] = int(m.group(1)) - 1
00779                     pos_dict["stop"] = int(m.group(2))
00780                 else:
00781                     pos_dict["start"] = int(position) - 1
00782                     pos_dict["stop"] = int(position)
00783
00784                 if "\\" in position:
00785                     # Note: the value of `N` in `...\N` isn't read: the script simply assumes `N` is
      consistent with the number of
00786                     # increments per interval when the alignment is parsed with a stride of 3
      (designating each cpos).
00787                     # E.g. For the partition file:
00788                     #      ...'1-N\2'
00789                     #      ...'2-N\2'
00790                     #      ...'(N+1)-M\2'
00791                     #      ...'(N+2)-M\2'
00792                     # 3'cpos are ignored due to the absence of intervals `3-N...`, `(N+3)-M...`, not
      because the associated stride values are'\2'
00793                     pos_dict["stride"] = 3
00794                 elif "\\" not in position:
00795                     pos_dict["stride"] = 1
00796
00797                 add_to_list_of_dicts(pos_dict)
00798
00799             dict_of_dicts[partition_name] = list_of_dicts
00800             add_to_partitions(dict_of_dicts)
00801
00802         return partitions
00803
00804
```

References amas.AMAS.FileParser.in_file_lines.

7.5.3.5 phylip_interleaved_parse()

```
amas.AMAS.FileParser.phylip_interleaved_parse (
                self )
```

Definition at line 579 of file AMAS.py.
```
00579     def phylip_interleaved_parse(self):
```

7.5.3.6 phylip_parse()

```
amas.AMAS.FileParser.phylip_parse (
                self )
```
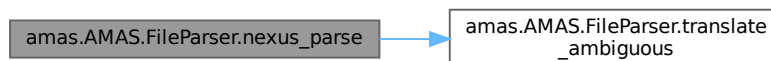
Definition at line 562 of file AMAS.py.
```
00562     def phylip_parse(self):
00563         # use regex to parse names and sequences in sequential phylip files
00564         matches = re.finditer(
00565             r"^(\s+)?(\S+)\s+([A-Za-z*?.{}-]+)",
00566             self.in_file_lines, re.MULTILINE
00567         )
00568
00569         records = {}
00570
00571         for match in matches:
00572             name_match = match.group(2).replace("\n", "")
00573             seq_match = match.group(3).replace("\n", "").upper()
00574             seq_match = self.translate_ambiguous(seq_match)
00575             records[name_match] = seq_match
00576
00577         return records
00578
```

References amas.AMAS.FileParser.in_file_lines, and amas.AMAS.FileParser.translate_ambiguous().

Here is the call graph for this function:
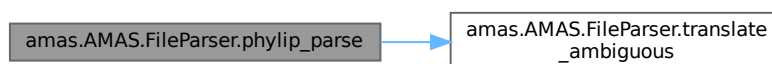
7.5.3.7 translate_ambiguous()

```
amas.AMAS.FileParser.translate_ambiguous (
                self,
                seq )
```

Definition at line 712 of file AMAS.py.
```
00712     def translate_ambiguous(self, seq):
00713         # translate ambiguous characters from curly bracket format
00714         # to single letter format
00715         # also remove spaces from sequences
00716         seq = seq.replace("{GT}", "K")
00717         seq = seq.replace("{AC}", "M")
00718         seq = seq.replace("{AG}", "R")
```

```
00719            seq = seq.replace("{CT}", "Y")
00720            seq = seq.replace("{CG}", "S")
00721            seq = seq.replace("{AT}", "W")
00722            seq = seq.replace("{CGT}", "B")
00723            seq = seq.replace("{ACG}", "V")
00724            seq = seq.replace("{ACT}", "H")
00725            seq = seq.replace("{AGT}", "D")
00726            seq = seq.replace("{GATC}", "N")
00727            seq = seq.replace(" ", "")
00728
00729            return seq
00730
```

Referenced by amas.AMAS.FileParser.fasta_parse(), amas.AMAS.FileParser.nexus_interleaved_parse(), amas.AMAS.FileParser.nexus_parse(), and amas.AMAS.FileParser.phylip_parse().

Here is the caller graph for this function:
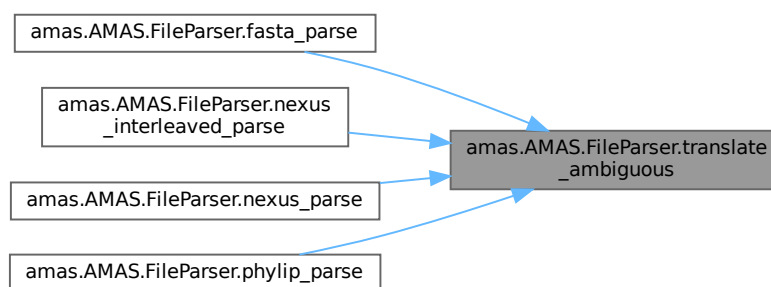
7.5.4 Member Data Documentation

7.5.4.1 chars_match

```
amas.AMAS.FileParser.chars_match = match.group(3)  [static]
```

Definition at line 596 of file AMAS.py.

7.5.4.2 counter

```
int amas.AMAS.FileParser.counter = 0  [static]
```

Definition at line 605 of file AMAS.py.

7.5.4.3 in_file

```
amas.AMAS.FileParser.in_file
```

Definition at line 542 of file AMAS.py.

Referenced by amas.AMAS.FileHandler.__exit__(), amas.AMAS.Alignment.get_aln_input(), and amas.AMAS.Alignment.get_name().

7.5.4.4 in_file_lines

```
amas.AMAS.FileParser.in_file_lines
```

Definition at line 544 of file AMAS.py.

Referenced by amas.AMAS.FileParser.fasta_parse(), amas.AMAS.FileParser.nexus_interleaved_parse(), amas.AMAS.FileParser.nexus_parse(), amas.AMAS.FileParser.partitions_parse(), and amas.AMAS.FileParser.phylip_parse().

7.5.4.5 matches

```
amas.AMAS.FileParser.matches  [static]
```

Initial value:
```
=  re.finditer(
                r"(^(\s+)?(\S+)( ){2,}|^\s+)([ A-Za-z*?.{}-]+)",
                self.in_file_lines, re.MULTILINE
            )
```

Definition at line 619 of file AMAS.py.

7.5.4.6 name_match

```
amas.AMAS.FileParser.name_match = match.group(2).replace("\n", "")  [static]
```

Definition at line 608 of file AMAS.py.

7.5.4.7 name_matches

```
amas.AMAS.FileParser.name_matches  [static]
```

Initial value:
```
=  re.finditer(
            r"^(\s+)?(\S+)[ \t]+[A-Za-z*?.{}-]+",
            self.in_file_lines, re.MULTILINE
        )
```

Definition at line 585 of file AMAS.py.

7.5.4.8 records

```
dict amas.AMAS.FileParser.records = {}  [static]
```

Definition at line 602 of file AMAS.py.

7.5.4.9 seq_match [1/2]

```
amas.AMAS.FileParser.seq_match = match.group(3).replace("\n", "").upper()  [static]
```

Definition at line 612 of file AMAS.py.

7.5.4.10 seq_match `[2/2]`

```
str amas.AMAS.FileParser.seq_match = "".join(seq_match.split())  [static]
```

Definition at line 631 of file AMAS.py.

7.5.4.11 seq_matches

```
amas.AMAS.FileParser.seq_matches  [static]
```

Initial value:
```
= re.finditer(
        r"(^(\s+)?\S+[ \t]+|^)([A-Za-z*?.{}-]+)$",
        self.in_file_lines, re.MULTILINE
    )
```

Definition at line 589 of file AMAS.py.

7.5.4.12 sequence

```
str amas.AMAS.FileParser.sequence = ""  [static]
```

Definition at line 636 of file AMAS.py.

7.5.4.13 sequences

```
list amas.AMAS.FileParser.sequences = []  [static]
```

Definition at line 600 of file AMAS.py.

7.5.4.14 tax_chars_matches

```
amas.AMAS.FileParser.tax_chars_matches  [static]
```

Initial value:
```
= re.finditer(
        r"^(\s+)?([0-9]+)[ \t]+([0-9]+)",
        self.in_file_lines, re.MULTILINE
    )
```

Definition at line 581 of file AMAS.py.

7.5.4.15 tax_match

```
amas.AMAS.FileParser.tax_match = match.group(2)  [static]
```

Definition at line 595 of file AMAS.py.

7.5.4.16 taxa

```
list amas.AMAS.FileParser.taxa = []  [static]
```

Definition at line 599 of file AMAS.py.

The documentation for this class was generated from the following file:

- amas/AMAS.py

7.6 amas.AMAS.MetaAlignment Class Reference

Public Member Functions

- __init__ (self, ∗∗kwargs)
- translate_dna_to_aa (self, seq, translation_table, frame)
- translate_dict (self, source_dict)
- get_translated (self, translation_table, reading_frame)
- trim_dict (self, alignment)
- get_trimmed (self, trim_fraction, parsimony_check)
- remove_unknown_chars (self, seq)
- remove_empty_sequences (self, split_alignment)
- get_partitions (self, partitions_file)
- get_alignment_object (self, alignment)
- get_alignment_objects (self)
- get_parsed_alignments (self)
- get_partitioned (self, partitions_file)
- get_summaries (self)
- summarize_alignments (self, alignment)
- get_taxon_summaries (self)
- summarize_alignments_taxa (self, alignment)
- write_summaries (self, file_name)
- write_taxa_summaries (self)
- get_replicate (self, no_replicates, no_loci)
- get_concatenated (self, alignments)
- remove_from_alignment (self, alignment, species_to_remove_set, index)
- remove_taxa (self, species_to_remove_set)
- print_fasta (self, source_dict)
- print_phylip (self, source_dict)
- print_phylip_int (self, source_dict)
- print_nexus (self, source_dict)
- print_nexus_int (self, source_dict)
- natural_sort (self, a_list)
- print_unspecified_partitions (self, data_type, codons)
- print_nexus_partitions (self, data_type, codons)
- print_iqtree_nexus_partitions (self, data_type, codons)
- print_raxml_partitions (self, data_type, codons)
- replace_string_in_file (self, file_name, old_string, new_string)
- write_partitions (self, file_name, part_format, data_type, codons)
- get_extension (self, file_format)
- get_metapartition_extension (self, file_format)
- file_overwrite_error (self, file_name)

- write_formatted_file (self, file_format, file_name, alignment)
- get_alignment_name (self, i, extension)
- get_alignment_name_no_ext (self, i)
- write_concat (self, file_format)
- write_convert (self, index, alignment, file_format, extension)
- write_replicate (self, index, alignment, file_format, extension)
- write_split (self, item, file_format, extension)
- write_reduced (self, file_format, extension)
- write_translated (self, index, alignment, file_format, extension)
- write_trimmed (self, index, alignment, file_format, extension)
- write_metapartitions (self, file_format)
- write_out (self, action, file_format)

Public Attributes

- in_files
- in_format
- data_type
- command
- concat_out
- using_metapartitions
- check_align
- cores
- by_taxon_summary
- no_sup_aln_name
- no_mpan
- codons
- no_replicates
- no_loci
- split
- remove_empty
- prepend_label
- species_to_remove
- species_to_remove_set
- reduced_file_prefix
- check_taxa
- reading_frame
- genetic_code
- trim_fraction
- trim_out
- parsimony_check
- alignment_objects
- parsed_alignments
- codes_list
- gencode_NCBI_1
- gencode_NCBI_2
- gencode_NCBI_3
- gencode_NCBI_4
- gencode_NCBI_5
- gencode_NCBI_6
- gencode_NCBI_9
- gencode_NCBI_10
- gencode_NCBI_11
- gencode_NCBI_12

- gencode_NCBI_13
- gencode_NCBI_14
- gencode_NCBI_16
- gencode_NCBI_21
- gencode_NCBI_22
- gencode_NCBI_23
- gencode_NCBI_24
- gencode_NCBI_25
- gencode_NCBI_26
- codes

7.6.1 Detailed Description

```
Class of multiple sequence alignments
```

Definition at line 1165 of file AMAS.py.

7.6.2 Constructor & Destructor Documentation

7.6.2.1 __init__()

```
amas.AMAS.MetaAlignment.__init__ (
            self,
         ** kwargs )
```

Definition at line 1168 of file AMAS.py.
```
01168      def __init__(self, **kwargs):
01169          # set defaults and get values from kwargs
01170          self.in_files = kwargs.get("in_files")
01171          self.in_format = kwargs.get("in_format")
01172          self.data_type = kwargs.get("data_type")
01173          self.command = kwargs.get("command")
01174          self.concat_out = kwargs.get("concat_out", "concatenated.out")
01175          self.using_metapartitions = False
01176          self.check_align = kwargs.get("check_align", False)
01177          self.cores = kwargs.get("cores")
01178          self.by_taxon_summary = kwargs.get("by_taxon_summary")
01179          self.no_sup_aln_name = False
01180          self.no_mpan = False
01181
01182          if self.command == "concat":
01183              self.codons = kwargs.get("codons", "none")
01184              if self.data_type == "aa" and self.codons != "none":
01185                  print("ERROR: when option -d|--data-type is set to 'aa', option -n|--codons must be
      set to 'none'.")
01186                  sys.exit(1)
01187
01188          if self.command == "replicate":
01189              self.no_replicates = kwargs.get("replicate_args")[0]
01190              self.no_loci = kwargs.get("replicate_args")[1]
01191
01192          if self.command == "split":
01193              self.split = kwargs.get("split_by")
01194              self.remove_empty = kwargs.get("remove_empty", False)
01195              self.no_sup_aln_name = kwargs.get("no_sup_aln_name", False)
01196
01197          if self.command == "metapartitions":
01198              self.using_metapartitions = True
01199              self.split = kwargs.get("split_by")
01200              self.remove_empty = kwargs.get("remove_empty", False)
01201              self.no_sup_aln_name = kwargs.get("no_sup_aln_name", False)
01202              self.no_mpan = kwargs.get("no_mpan", False)
01203              self.prepend_label = kwargs.get("prepend_label")
01204              if self.prepend_label is not None and isinstance(self.prepend_label, str):
01205                  self.prepend_label = self.prepend_label + "_"
01206              else:
```

```
01207                    self.prepend_label = ""
01208
01209          if self.command == "remove":
01210              self.species_to_remove = kwargs.get("taxa_to_remove")
01211              self.species_to_remove_set = set(self.species_to_remove)
01212              self.reduced_file_prefix = kwargs.get("out_prefix")
01213              self.check_taxa = kwargs.get("check_taxa", False)
01214
01215          if self.command == "translate":
01216              self.reading_frame = kwargs.get("reading_frame")
01217              self.genetic_code = kwargs.get("genetic_code")
01218
01219          if self.command == "trim":
01220              self.trim_fraction = kwargs.get("trim_fraction")
01221              self.trim_out = kwargs.get("trim_out")
01222              self.parsimony_check = kwargs.get("parsimony_check", False)
01223
01224          self.alignment_objects = self.get_alignment_objects()
01225          self.parsed_alignments = self.get_parsed_alignments()
01226
01227          # The code list:
01228          self.codes_list = """
01229           1. The Standard Code
01230           2. The Vertebrate Mitochondrial Code
01231           3. The Yeast Mitochondrial Code
01232           4. The Mold, Protozoan, and Coelenterate Mitochondrial Code and the Mycoplasma/Spiroplasma
      Code
01233           5. The Invertebrate Mitochondrial Code
01234           6. The Ciliate, Dasycladacean and Hexamita Nuclear Code
01235           9. The Echinoderm and Flatworm Mitochondrial Code
01236          10. The Euplotid Nuclear Code
01237          11. The Bacterial, Archaeal and Plant Plastid Code
01238          12. The Alternative Yeast Nuclear Code
01239          13. The Ascidian Mitochondrial Code
01240          14. The Alternative Flatworm Mitochondrial Code
01241          16. Chlorophycean Mitochondrial Code
01242          21. Trematode Mitochondrial Code
01243          22. Scenedesmus obliquus Mitochondrial Code
01244          23. Thraustochytrium Mitochondrial Code
01245          24. Pterobranchia Mitochondrial Code
01246          25. Candidate Division SR1 and Gracilibacteria Code
01247          26. Pachysolen tannophilus Nuclear Code
01248          """
01249
01250          # 1: The Standard Code
01251          self.gencode_NCBI_1 = {
01252          "TTT" : "F", # Phe
01253          "TCT" : "S", # Ser
01254          "TAT" : "Y", # Tyr
01255          "TGT" : "C", # Cys
01256          "TTC" : "F", # Phe
01257          "TCC" : "S", # Ser
01258          "TAC" : "Y", # Tyr
01259          "TGC" : "C", # Cys
01260          "TTA" : "L", # Leu
01261          "TCA" : "S", # Ser
01262          "TAA" : "*", # Ter
01263          "TGA" : "*", # Ter
01264          "TTG" : "L", # Leu i
01265          "TCG" : "S", # Ser
01266          "TAG" : "*", # Ter
01267          "TGG" : "W", # Trp
01268          "CTT" : "L", # Leu
01269          "CCT" : "P", # Pro
01270          "CAT" : "H", # His
01271          "CGT" : "R", # Arg
01272          "CTC" : "L", # Leu
01273          "CCC" : "P", # Pro
01274          "CAC" : "H", # His
01275          "CGC" : "R", # Arg
01276          "CTA" : "L", # Leu
01277          "CCA" : "P", # Pro
01278          "CAA" : "Q", # Gln
01279          "CGA" : "R", # Arg
01280          "CTG" : "L", # Leu i
01281          "CCG" : "P", # Pro
01282          "CAG" : "Q", # Gln
01283          "CGG" : "R", # Arg
01284          "ATT" : "I", # Ile
01285          "ACT" : "T", # Thr
01286          "AAT" : "N", # Asn
01287          "AGT" : "S", # Ser
01288          "ATC" : "I", # Ile
01289          "ACC" : "T", # Thr
01290          "AAC" : "N", # Asn
01291          "AGC" : "S", # Ser
01292          "ATA" : "I", # Ile
```

```
01293            "ACA" : "T",  # Thr
01294            "AAA" : "K",  # Lys
01295            "AGA" : "R",  # Arg
01296            "ATG" : "M",  # Met i
01297            "ACG" : "T",  # Thr
01298            "AAG" : "K",  # Lys
01299            "AGG" : "R",  # Arg
01300            "GTT" : "V",  # Val
01301            "GCT" : "A",  # Ala
01302            "GAT" : "D",  # Asp
01303            "GGT" : "G",  # Gly
01304            "GTC" : "V",  # Val
01305            "GCC" : "A",  # Ala
01306            "GAC" : "D",  # Asp
01307            "GGC" : "G",  # Gly
01308            "GTA" : "V",  # Val
01309            "GCA" : "A",  # Ala
01310            "GAA" : "E",  # Glu
01311            "GGA" : "G",  # Gly
01312            "GTG" : "V",  # Val
01313            "GCG" : "A",  # Ala
01314            "GAG" : "E",  # Glu
01315            "GGG" : "G",  # Gly
01316            "---" : "-",  # Gap
01317            "???" : "?",  # Unk
01318            "NNN" : "X",  # Unk
01319            }
01320
01321            # 2: The Vertebrate Mitochondrial Code
01322            self.gencode_NCBI_2 = self.gencode_NCBI_1.copy()
01323            self.gencode_NCBI_2["AGA"] = "*" # Ter
01324            self.gencode_NCBI_2["AGG"] = "*" # Ter
01325            self.gencode_NCBI_2["ATA"] = "M" # Met
01326            self.gencode_NCBI_2["TGA"] = "W" # Trp
01327
01328            # 3: The Yeast Mitochondrial Code
01329            self.gencode_NCBI_3 = self.gencode_NCBI_1.copy()
01330            self.gencode_NCBI_3["ATA"] = "M" # Met
01331            self.gencode_NCBI_3["CTT"] = "T" # Thr
01332            self.gencode_NCBI_3["CTC"] = "T" # Thr
01333            self.gencode_NCBI_3["CTA"] = "T" # Thr
01334            self.gencode_NCBI_3["CTG"] = "T" # Thr
01335            self.gencode_NCBI_3["TGA"] = "W" # Trp
01336
01337            del self.gencode_NCBI_3["CGA"]
01338            del self.gencode_NCBI_3["CGC"]
01339
01340            # 4: The Mold, Protozoan, and Coelenterate Mitochondrial Code and the Mycoplasma/Spiroplasma
       Code
01341            self.gencode_NCBI_4 = self.gencode_NCBI_1.copy()
01342            self.gencode_NCBI_4["TGA"] = "W" # Trp
01343
01344            # 5: The Invertebrate Mitochondrial Code
01345            self.gencode_NCBI_5 = self.gencode_NCBI_1.copy()
01346            self.gencode_NCBI_5["AGA"] = "S" # Ser
01347            self.gencode_NCBI_5["AGG"] = "S" # Ser
01348            self.gencode_NCBI_5["ATA"] = "M" # Met
01349            self.gencode_NCBI_5["TGA"] = "W" # Trp
01350
01351            # 6: The Ciliate, Dasycladacean and Hexamita Nuclear Code
01352            self.gencode_NCBI_6 = self.gencode_NCBI_1.copy()
01353            self.gencode_NCBI_6["TAA"] = "Q" # Gln
01354            self.gencode_NCBI_6["TAG"] = "Q" # Gln
01355
01356            # 9: The Echinoderm and Flatworm Mitochondrial Code
01357            self.gencode_NCBI_9 = self.gencode_NCBI_1.copy()
01358            self.gencode_NCBI_9["AAA"] = "N" # Asn
01359            self.gencode_NCBI_9["AGA"] = "S" # Ser
01360            self.gencode_NCBI_9["AGG"] = "S" # Ser
01361            self.gencode_NCBI_9["TGA"] = "W" # Trp
01362
01363            # 10: The Euplotid Nuclear Code
01364            self.gencode_NCBI_10 = self.gencode_NCBI_1.copy()
01365            self.gencode_NCBI_10["TGA"] = "C" # Cys
01366
01367            # 11: The Bacterial, Archaeal and Plant Plastid Code
01368            self.gencode_NCBI_11 = self.gencode_NCBI_1.copy()
01369
01370            # 12: The Alternative Yeast Nuclear Code
01371            self.gencode_NCBI_12 = self.gencode_NCBI_1.copy()
01372            self.gencode_NCBI_12["CTG"] = "S" # Ser
01373
01374            # 13: The Ascidian Mitochondrial Code
01375            self.gencode_NCBI_13 = self.gencode_NCBI_1.copy()
01376            self.gencode_NCBI_13["AGA"] = "G" # Gly
01377            self.gencode_NCBI_13["AGG"] = "G" # Gly
01378            self.gencode_NCBI_13["ATA"] = "M" # Met
```

```
01379            self.gencode_NCBI_13["TGA"] = "W" # Trp
01380
01381            # 14: The Alternative Flatworm Mitochondrial Code
01382            self.gencode_NCBI_14 = self.gencode_NCBI_1.copy()
01383            self.gencode_NCBI_14["AAA"] = "N" # Asn
01384            self.gencode_NCBI_14["AGA"] = "S" # Ser
01385            self.gencode_NCBI_14["AGG"] = "S" # Ser
01386            self.gencode_NCBI_14["TAA"] = "Y" # Tyr
01387            self.gencode_NCBI_14["TGA"] = "W" # Trp
01388
01389            # 16: Chlorophycean Mitochondrial Code
01390            self.gencode_NCBI_16 = self.gencode_NCBI_1.copy()
01391            self.gencode_NCBI_16["TAG"] = "L" # Leu
01392
01393            # 21: Trematode Mitochondrial Code
01394            self.gencode_NCBI_21 = self.gencode_NCBI_1.copy()
01395            self.gencode_NCBI_21["TGA"] = "W" # Trp
01396            self.gencode_NCBI_21["ATA"] = "M" # Met
01397            self.gencode_NCBI_21["AGA"] = "S" # Ser
01398            self.gencode_NCBI_21["AGG"] = "S" # Ser
01399            self.gencode_NCBI_21["AAA"] = "N" # Asn
01400
01401            # 22: Scenedesmus obliquus Mitochondrial Code
01402            self.gencode_NCBI_22 = self.gencode_NCBI_1.copy()
01403            self.gencode_NCBI_22["TCA"] = "*" # Ter
01404            self.gencode_NCBI_22["TAG"] = "L" # Leu
01405
01406            # 23: Thraustochytrium Mitochondrial Code
01407            self.gencode_NCBI_23 = self.gencode_NCBI_1.copy()
01408            self.gencode_NCBI_23["TTA"] = "*" # Ter
01409
01410            # 24: Pterobranchia Mitochondrial Code
01411            self.gencode_NCBI_24 = self.gencode_NCBI_1.copy()
01412            self.gencode_NCBI_24["AGA"] = "S" # Ser
01413            self.gencode_NCBI_24["AGG"] = "K" # Lys
01414            self.gencode_NCBI_24["TGA"] = "W" # Trp
01415
01416            # 25: Candidate Division SR1 and Gracilibacteria Code
01417            self.gencode_NCBI_25 = self.gencode_NCBI_1.copy()
01418            self.gencode_NCBI_25["TGA"] = "G" # Gly
01419
01420            # 26: Pachysolen tannophilus Nuclear Code
01421            self.gencode_NCBI_26 = self.gencode_NCBI_1.copy()
01422            self.gencode_NCBI_26["CTG"] = "A" # Ala
01423
01424            self.codes = {
01425            1 : self.gencode_NCBI_1,
01426            2 : self.gencode_NCBI_2,
01427            3 : self.gencode_NCBI_3,
01428            4 : self.gencode_NCBI_4,
01429            5 : self.gencode_NCBI_5,
01430            6 : self.gencode_NCBI_6,
01431            9 : self.gencode_NCBI_9,
01432            10 : self.gencode_NCBI_10,
01433            11 : self.gencode_NCBI_11,
01434            12 : self.gencode_NCBI_12,
01435            13 : self.gencode_NCBI_13,
01436            14 : self.gencode_NCBI_14,
01437            16 : self.gencode_NCBI_16,
01438            21 : self.gencode_NCBI_21,
01439            22 : self.gencode_NCBI_22,
01440            23 : self.gencode_NCBI_23,
01441            24 : self.gencode_NCBI_24,
01442            25 : self.gencode_NCBI_25,
01443            26 : self.gencode_NCBI_26
01444            }
01445
```

7.6.3 Member Function Documentation

7.6.3.1 file_overwrite_error()

```
amas.AMAS.MetaAlignment.file_overwrite_error (
            self,
            file_name )
```

Definition at line 2157 of file AMAS.py.
```
02157     def file_overwrite_error(self, file_name):
```

```
02158          # print warning when overwriting a file
02159          if path.exists(file_name):
02160              print("WARNING: You are overwriting '" + file_name + "'")
02161
```

Referenced by amas.AMAS.MetaAlignment.write_concat(), amas.AMAS.MetaAlignment.write_convert(), amas.AMAS.MetaAlignment amas.AMAS.MetaAlignment.write_reduced(), amas.AMAS.MetaAlignment.write_replicate(), amas.AMAS.MetaAlignment.write_split() amas.AMAS.MetaAlignment.write_summaries(), amas.AMAS.MetaAlignment.write_taxa_summaries(), amas.AMAS.MetaAlignment.v and amas.AMAS.MetaAlignment.write_trimmed().

Here is the caller graph for this function:

7.6.3.2 get_alignment_name()

```
amas.AMAS.MetaAlignment.get_alignment_name (
            self,
            i,
            extension )
```

Definition at line 2176 of file AMAS.py.

```
02176      def get_alignment_name(self, i, extension):
02177          # get file name
02178          file_name = self.alignment_objects[i].get_name() + extension
02179
02180          return file_name
02181
```

References amas.AMAS.MetaAlignment.alignment_objects.

Referenced by amas.AMAS.MetaAlignment.write_convert(), amas.AMAS.MetaAlignment.write_translated(), and amas.AMAS.MetaAlignment.write_trimmed().

Here is the caller graph for this function:
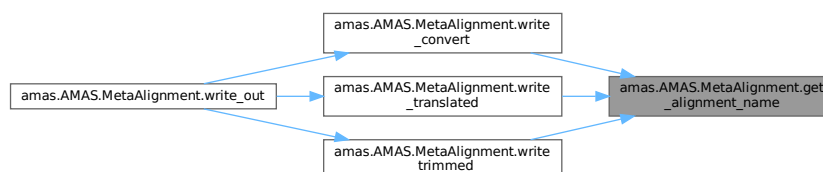
7.6.3.3 get_alignment_name_no_ext()

```
amas.AMAS.MetaAlignment.get_alignment_name_no_ext (
            self,
            i )
```

Definition at line 2182 of file AMAS.py.

```
02182     def get_alignment_name_no_ext(self, i):
02183         # get file name without extension
02184         file_name = self.alignment_objects[i].get_name()
02185
02186         return file_name
02187
```

References amas.AMAS.MetaAlignment.alignment_objects.

Referenced by amas.AMAS.MetaAlignment.remove_from_alignment().

Here is the caller graph for this function:

7.6.3.4 get_alignment_object()

```
amas.AMAS.MetaAlignment.get_alignment_object (
            self,
            alignment )
```

Definition at line 1523 of file AMAS.py.

```
01523     def get_alignment_object(self, alignment):
01524         # parse according to the given alphabet;
01525         # Note:('alignment') <=> `in_file' outside MetaAlignment, e.g.
01526         #
      AminoAcidAlignment(Alignment<-.get_parsed_aln<-.get_aln_input)<-FileParser.__init__(in_file)<-FileHandler(...open(self.i
01527         if self.data_type == "aa":
01528             aln = AminoAcidAlignment(alignment, self.in_format, self.data_type)
01529         elif self.data_type == "dna":
01530             aln = DNAAlignment(alignment, self.in_format, self.data_type)
01531         return aln
01532
```

References amas.AMAS.Alignment.data_type, amas.AMAS.MetaAlignment.data_type, amas.AMAS.Alignment.in_format, and amas.AMAS.MetaAlignment.in_format.

Referenced by amas.AMAS.MetaAlignment.get_alignment_objects().

Here is the caller graph for this function:

7.6.3.5 get_alignment_objects()

```
amas.AMAS.MetaAlignment.get_alignment_objects (
              self )
```

Definition at line 1533 of file AMAS.py.

```
01533      def get_alignment_objects(self):
01534          # get alignment objects on which statistics can be computed
01535          # use multiprocessing if more than one core specified
01536          if int(self.cores) == 1:
01537              alignments = [self.get_alignment_object(alignment) for alignment in self.in_files]
01538          elif int(self.cores) > 1:
01539              pool = mp.Pool(int(self.cores))
01540              alignments = pool.map(self.get_alignment_object, self.in_files)
01541          return alignments
01542
```

References   amas.AMAS.MetaAlignment.cores,   amas.AMAS.MetaAlignment.get_alignment_object(),   and amas.AMAS.MetaAlignment.in_files.

Referenced by amas.AMAS.MetaAlignment.write_metapartitions().

Here is the call graph for this function:

Here is the caller graph for this function:
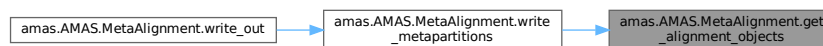
7.6.3.6 get_concatenated()

```
amas.AMAS.MetaAlignment.get_concatenated (
              self,
              alignments )
```

Definition at line 1746 of file AMAS.py.

```
01746      def get_concatenated(self, alignments):
01747          # concatenate muntiple input alignments
01748          # create empty dictionary of lists
01749          concatenated = defaultdict(list)
01750
01751          # first create list of taxa in all alignments
01752          # you need this to insert empty seqs in
01753          # the concatenated alignment
01754          all_taxa = []
01755          for alignment in alignments:
01756              for taxon in alignment.keys():
01757                  if taxon not in all_taxa:
01758                      all_taxa.append(taxon)
01759
```

```
01760            # start counters to keep track of partitions
01761            partition_counter = 1
01762            position_counter = 1
01763            # get dict for alignment name and partition
01764            partitions = {}
01765            digits_to_pad = len(str(len(alignments)))
01766
01767            for alignment in alignments:
01768                # get alignment length from a random taxon
01769                partition_length = len(alignment[list(alignment.keys())[0]])
01770                # get base name of each alignment for use when writing partitions file
01771                # NOTE: the base name here is whatever comes before fist period in the file name
01772                alignment_name = self.alignment_objects[partition_counter - 1].get_name().split('.')[0]
01773
01774                if self.using_metapartitions:
01775                    # implementation of option --no-mpan; option --prepend(-label) will assign a string or
       "" (see class definition)
01776                    if self.no_mpan:
01777                        # omit original alignment names from the printed partition file
01778                        partition_name = self.prepend_label + "p" +
       str(partition_counter).zfill(digits_to_pad)
01779                    else:
01780                        # keep original alignment names in the printed partition file
01781                        partition_name = self.prepend_label + "p" +
       str(partition_counter).zfill(digits_to_pad) + "_" + alignment_name
01782                else:
01783                    partition_name = "p" + str(partition_counter) + "_" + alignment_name
01784
01785                start = position_counter
01786                position_counter += partition_length
01787                end = position_counter - 1
01788                partitions[partition_name] = str(start) + "-" + str(end)
01789                partition_counter += 1
01790
01791                # get empty sequence if there is missing taxon
01792                # getting length from first element of list of keys
01793                # created from the original dict for this alignment
01794                empty_seq = '?' * partition_length
01795
01796                for taxon in all_taxa:
01797
01798                    if taxon not in alignment.keys():
01799                        concatenated[taxon].append(empty_seq)
01800                    else:
01801                        concatenated[taxon].append(alignment[taxon])
01802
01803            concatenated = {taxon:".join(seqs) for taxon, seqs in concatenated.items()}
01804
01805            return concatenated, partitions
01806
```

References amas.AMAS.MetaAlignment.alignment_objects, amas.AMAS.MetaAlignment.no_mpan, amas.AMAS.MetaAlignment.prep
amas.AMAS.MetaAlignment.split, and amas.AMAS.MetaAlignment.using_metapartitions.

Referenced by amas.AMAS.MetaAlignment.get_replicate(), amas.AMAS.MetaAlignment.print_iqtree_nexus_partitions(), amas.AMAS.MetaAlignment.print_nexus_partitions(), amas.AMAS.MetaAlignment.print_raxml_partitions(), amas.AMAS.MetaAlignment.print_unspecified_partitions(), and amas.AMAS.MetaAlignment.write_concat().

Here is the caller graph for this function:
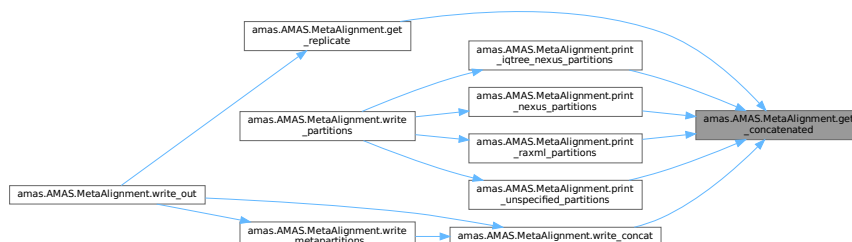
7.6.3.7 get_extension()

```
amas.AMAS.MetaAlignment.get_extension (
            self,
            file_format )
```

Definition at line 2127 of file AMAS.py.

```
02127      def get_extension(self, file_format):
02128          # get proper extension string
02129          if file_format == "phylip":
02130              extension = "-out.phy"
02131          elif file_format == "phylip-int":
02132              extension = "-out.int-phy"
02133          elif file_format == "fasta":
02134              extension = "-out.fas"
02135          elif file_format == "nexus":
02136              extension = "-out.nex"
02137          elif file_format == "nexus-int":
02138              extension = "-out.int-nex"
02139
02140          return extension
02141
```

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the caller graph for this function:
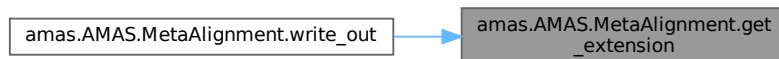
7.6.3.8 get_metapartition_extension()

```
amas.AMAS.MetaAlignment.get_metapartition_extension (
            self,
            file_format )
```

Definition at line 2142 of file AMAS.py.

```
02142      def get_metapartition_extension(self, file_format):
02143          # get proper metapartition_extension string
02144          if file_format == "phylip":
02145              metapartition_extension = "-meta.phy"
02146          elif file_format == "phylip-int":
02147              metapartition_extension = "-meta.int-phy"
02148          elif file_format == "fasta":
02149              metapartition_extension = "-meta.fas"
02150          elif file_format == "nexus":
02151              metapartition_extension = "-meta.nex"
02152          elif file_format == "nexus-int":
02153              metapartition_extension = "-meta.int-nex"
02154
02155          return metapartition_extension
02156
```

Referenced by amas.AMAS.MetaAlignment.write_metapartitions().

Here is the caller graph for this function:
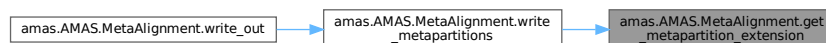
7.6.3.9 get_parsed_alignments()

```
amas.AMAS.MetaAlignment.get_parsed_alignments (
                self )
```

Definition at line 1543 of file AMAS.py.

```
01543    def get_parsed_alignments(self):
01544        # get parsed dictionaries with taxa and sequences
01545        parsed_alignments = []
01546        add_to_parsed_alignments = parsed_alignments.append
01547        for alignment in self.alignment_objects:
01548            parsed = alignment.parsed_aln
01549            add_to_parsed_alignments(parsed)
01550            # checking if every seq has the same length or if parsed is not empty; exit if false
01551            if self.check_align:
01552                equal = all(
01553                    x == [len(list(parsed.values())[i]) for i in
        range(0,len(list(parsed.values())))][0]
01554                    for x in [len(list(parsed.values())[i]) for i in
        range(0,len(list(parsed.values())))]
01555                )
01556                if equal is False:
01557                    print("ERROR: Sequences in input are of varying lengths. Be sure to align them
        first.")
01558                    sys.exit()
01559
01560            if not parsed.keys() or not any(parsed.values()):
01561                print(
01562                    "ERROR: Parsed sequences of " + alignment.in_file + " are empty. "
01563                    "Are you sure you specified the right input format and/or that input is a valid
        alignment?"
01564                )
01565                sys.exit()
01566
01567        return parsed_alignments
01568
```

References amas.AMAS.MetaAlignment.alignment_objects, and amas.AMAS.MetaAlignment.check_align.

Referenced by amas.AMAS.MetaAlignment.write_metapartitions().

Here is the caller graph for this function:
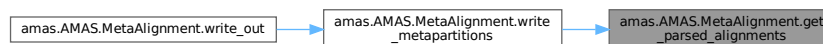
7.6.3.10 get_partitioned()

```
amas.AMAS.MetaAlignment.get_partitioned (
                self,
                partitions_file )
```

Definition at line 1569 of file AMAS.py.

```
01569    def get_partitioned(self, partitions_file):
01570        # partition alignment according to a partitions file
01571        partitions = self.get_partitions(partitions_file)
01572        alignment = self.parsed_alignments[0]
01573
01574        # initiate list of newly partitioned alignments
01575        list_of_parts = []
01576        add_to_list_of_parts = list_of_parts.append
01577        for partition in partitions:
01578            # loop over all parsed partitions, adding taxa and sliced sequences
01579            for name, elements in partition.items():
01580                new_dict = {}
01581
```

```
01582                     for taxon, seq in alignment.items():
01583                         new_seq = ""
01584
01585                         for dictionary in elements:
01586                             new_seq = new_seq +
       seq[dictionary["start"]:dictionary["stop"]:dictionary["stride"]]
01587                         new_dict[taxon] = new_seq
01588
01589                     if self.remove_empty:
01590                         # check if remove empty sequences
01591                         no_empty_dict = self.remove_empty_sequences(new_dict)
01592                         add_to_list_of_parts({name : no_empty_dict})
01593                     else:
01594                         # add partition name : dict of taxa and sequences to the list
01595                         add_to_list_of_parts({name : new_dict})
01596
01597             return list_of_parts
01598
```

References   amas.AMAS.MetaAlignment.get_partitions(),   amas.AMAS.MetaAlignment.parsed_alignments, amas.AMAS.MetaAlignment.remove_empty, and amas.AMAS.MetaAlignment.remove_empty_sequences().

Referenced by amas.AMAS.MetaAlignment.write_metapartitions(), and amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
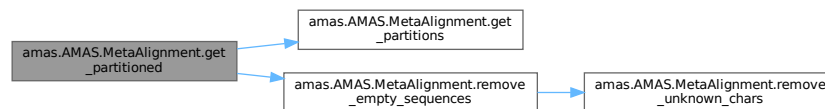
Here is the caller graph for this function:
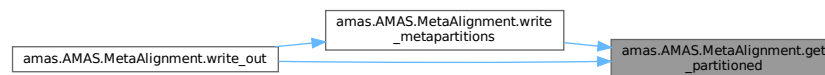
7.6.3.11   get_partitions()

```
amas.AMAS.MetaAlignment.get_partitions (
              self,
              partitions_file )
```

Definition at line 1516 of file AMAS.py.

```
01516     def get_partitions(self, partitions_file):
01517         # parse and get partitions from partitions file
01518         partitions = FileParser(partitions_file)
01519         parsed_partitions = partitions.partitions_parse()
01520
01521         return parsed_partitions
01522
```

Referenced by amas.AMAS.MetaAlignment.get_partitioned().

Here is the caller graph for this function:

7.6.3.12 get_replicate()

```
amas.AMAS.MetaAlignment.get_replicate (
            self,
            no_replicates,
            no_loci )
```

Definition at line 1726 of file AMAS.py.

```
01726     def get_replicate(self, no_replicates, no_loci):
01727         # construct replicate data sets for phylogenetic jackknife
01728         replicates = []
01729         add_to_replicates = replicates.append
01730         counter = 1
01731         for replicate in range(no_replicates):
01732
01733             try:
01734                 random_alignments = sample(self.parsed_alignments, no_loci)
01735             except ValueError:
01736                 print("ERROR: You specified more loci per replicate than there are in your input.")
01737                 sys.exit()
01738
01739             random_alignments = sample(self.parsed_alignments, no_loci)
01740             concat_replicate = self.get_concatenated(random_alignments)[0]
01741             add_to_replicates(concat_replicate)
01742             counter += 1
01743
01744         return replicates
01745
```

References amas.AMAS.MetaAlignment.get_concatenated(), and amas.AMAS.MetaAlignment.parsed_alignments.

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:

Here is the caller graph for this function:
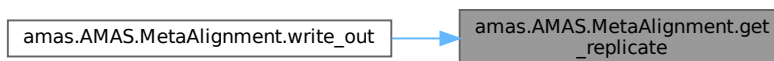
7.6.3.13 get_summaries()

```
amas.AMAS.MetaAlignment.get_summaries (
            self )
```

Definition at line 1599 of file AMAS.py.

```
01599     def get_summaries(self):
```

```
01600            # get summaries for all alignment objects
01601
01602            # define different headers for aa and dna alignments
01603            aa_header = [
01604                "Alignment_name",
01605                "No_of_taxa",
01606                "Alignment_length",
01607                "Total_matrix_cells",
01608                "Undetermined_characters",
01609                "Missing_percent",
01610                "No_variable_sites",
01611                "Proportion_variable_sites",
01612                "Parsimony_informative_sites",
01613                "Proportion_parsimony_informative"
01614            ]
01615
01616            dna_header = [
01617                "Alignment_name",
01618                "No_of_taxa",
01619                "Alignment_length",
01620                "Total_matrix_cells",
01621                "Undetermined_characters",
01622                "Missing_percent",
01623                "No_variable_sites",
01624                "Proportion_variable_sites",
01625                "Parsimony_informative_sites",
01626                "Proportion_parsimony_informative",
01627                "AT_content",
01628                "GC_content"
01629            ]
01630
01631            alignments = self.alignment_objects
01632            parsed_alignments = self.parsed_alignments
01633            freq_header = [char for char in alignments[0].alphabet]
01634
01635            if self.data_type == "aa":
01636                header = aa_header + freq_header
01637            elif self.data_type == "dna":
01638                header = dna_header + freq_header
01639
01640            # use multiprocessing if more than one core specified
01641            if int(self.cores) == 1:
01642                summaries = [alignment.get_summary() for alignment in alignments]
01643            elif int(self.cores) > 1:
01644                pool = mp.Pool(int(self.cores))
01645                summaries = pool.map(self.summarize_alignments, alignments)
01646            return header, summaries
01647
```

References amas.AMAS.MetaAlignment.alignment_objects, amas.AMAS.MetaAlignment.cores, amas.AMAS.Alignment.data_type, amas.AMAS.MetaAlignment.data_type, amas.AMAS.MetaAlignment.parsed_alignments, and amas.AMAS.MetaAlignment.summarize

Referenced by amas.AMAS.MetaAlignment.write_summaries().

Here is the call graph for this function:

Here is the caller graph for this function:

7.6.3.14 get_taxon_summaries()

```
amas.AMAS.MetaAlignment.get_taxon_summaries (
            self )
```

Definition at line 1653 of file AMAS.py.

```
01653      def get_taxon_summaries(self):
01654          # get per-sequence summaries for all alignment objects
01655
01656          # define different headers for aa and dna alignments
01657          aa_header = [
01658              "Alignment_name",
01659              "Taxon_name",
01660              "Sequence_length",
01661              "Undetermined_characters",
01662              "Missing_percent"
01663          ]
01664
01665          dna_header = [
01666              "Alignment_name",
01667              "Taxon_name",
01668              "Sequence_length",
01669              "Undetermined_characters",
01670              "Missing_percent",
01671              "AT_content",
01672              "GC_content"
01673          ]
01674
01675          alignments = self.alignment_objects
01676          parsed_alignments = self.parsed_alignments
01677          freq_header = alignments[0].alphabet
01678
01679          if self.data_type == "aa":
01680              header = aa_header + freq_header
01681          elif self.data_type == "dna":
01682              header = dna_header + freq_header
01683
01684          # use multiprocessing if more than one core specified
01685          if int(self.cores) == 1:
01686              summaries = [alignment.get_taxa_summary() for alignment in alignments]
01687          elif int(self.cores) > 1:
01688              pool = mp.Pool(int(self.cores))
01689              summaries = pool.map(self.summarize_alignments_taxa, alignments)
01690
01691          return header, summaries
01692
```

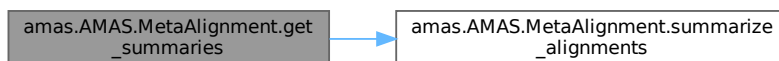References amas.AMAS.MetaAlignment.alignment_objects, amas.AMAS.MetaAlignment.cores, amas.AMAS.Alignment.data_type, amas.AMAS.MetaAlignment.data_type, amas.AMAS.MetaAlignment.parsed_alignments, and amas.AMAS.MetaAlignment.summarize

Referenced by amas.AMAS.MetaAlignment.write_taxa_summaries().

Here is the call graph for this function:
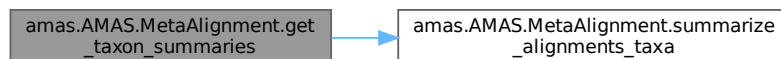
Here is the caller graph for this function:

7.6.3.15 get_translated()

```
amas.AMAS.MetaAlignment.get_translated (
            self,
            translation_table,
            reading_frame )
```

Definition at line 1478 of file AMAS.py.

```
01478      def get_translated(self, translation_table, reading_frame):
01479          if int(self.cores) == 1:
01480              translated_alignments = [self.translate_dict(alignment) for alignment in
    self.parsed_alignments]
01481          elif int(self.cores) > 1:
01482              pool = mp.Pool(int(self.cores))
01483              translated_alignments = pool.map(self.translate_dict, self.parsed_alignments)
01484
01485          return translated_alignments
01486
```

References amas.AMAS.MetaAlignment.cores, amas.AMAS.MetaAlignment.parsed_alignments, and amas.AMAS.MetaAlignment.tra

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:

Here is the caller graph for this function:
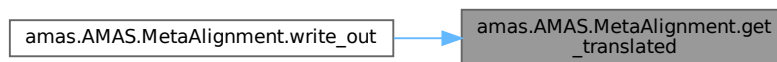
7.6.3.16 get_trimmed()

```
amas.AMAS.MetaAlignment.get_trimmed (
            self,
            trim_fraction,
            parsimony_check )
```

Definition at line 1495 of file AMAS.py.

```
01495      def get_trimmed(self, trim_fraction, parsimony_check):
01496          if int(self.cores) == 1:
01497              trimmed_alignments = [self.trim_dict(alignment) for alignment in self.alignment_objects]
01498          elif int(self.cores) > 1:
01499              pool = mp.Pool(int(self.cores))
01500              trimmed_alignments = pool.map(self.trim_dict, self.alignment_objects)
01501
01502          return trimmed_alignments
01503
```

References amas.AMAS.MetaAlignment.alignment_objects, amas.AMAS.MetaAlignment.cores, and amas.AMAS.MetaAlignment.trim

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
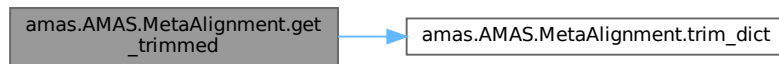
Here is the caller graph for this function:
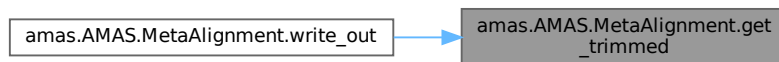
7.6.3.17 natural_sort()

```
amas.AMAS.MetaAlignment.natural_sort (
            self,
            a_list )
```

Definition at line 1960 of file AMAS.py.
```
01960    def natural_sort(self, a_list):
01961        # create a function that does 'human sort' on a list
01962        convert = lambda text: int(text) if text.isdigit() else text.lower()
01963        alphanum_key = lambda key: [convert(c) for c in re.split('([0-9]+)', key)]
01964        return sorted(a_list, key = alphanum_key)
01965
```

Referenced by amas.AMAS.MetaAlignment.print_iqtree_nexus_partitions(), amas.AMAS.MetaAlignment.print_nexus_partitions(), amas.AMAS.MetaAlignment.print_raxml_partitions(), and amas.AMAS.MetaAlignment.print_unspecified_partitions().

Here is the caller graph for this function:
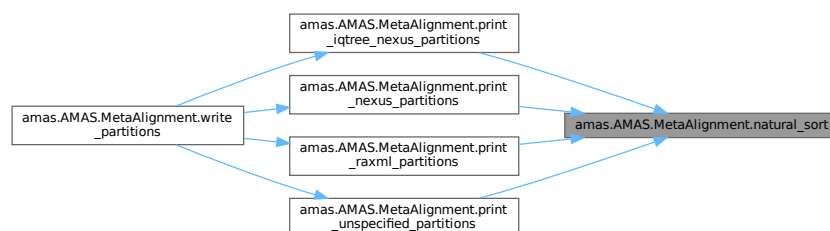
7.6.3.18 print_fasta()

```
amas.AMAS.MetaAlignment.print_fasta (
             self,
             source_dict )
```

Definition at line 1833 of file AMAS.py.

```
01833    def print_fasta(self, source_dict):
01834        # print fasta-formatted string from a dictionary
01835        fasta_string = ""
01836        # each sequence line will have 80 characters
01837        n = 80
01838
01839        for taxon, seq in sorted(source_dict.items()):
01840            # split dictionary values to a list of string, each n chars long
01841            seq = [seq[i:i+n] for i in range(0, len(seq), n)]
01842            # in case there are unwanted spaces in taxon names
01843            taxon = taxon.replace(" ", "_").strip("'")
01844            fasta_string += ">" + taxon + "\n"
01845            for element in seq:
01846                fasta_string += element + "\n"
01847
01848        return fasta_string
01849
```

Referenced by amas.AMAS.MetaAlignment.write_formatted_file().

Here is the caller graph for this function:

7.6.3.19 print_iqtree_nexus_partitions()

```
amas.AMAS.MetaAlignment.print_iqtree_nexus_partitions (
             self,
             data_type,
             codons )
```

Definition at line 2027 of file AMAS.py.

```
02027    def print_iqtree_nexus_partitions(self, data_type, codons):
02028        # print partitions for concatenated alignment
02029        part_string = ""
02030        part_dict = self.get_concatenated(self.parsed_alignments)[1]
02031        part_list = self.natural_sort(part_dict.keys())
02032        # write beginning of nexus sets
02033        part_string += "#nexus\n"
02034        part_string += "begin sets;\n"
02035
02036        if data_type == "dna":
02037            if codons == "none":
02038                for key in part_list:
02039                    part_string += "  charset " + key + " = " + str(part_dict[key]) + ";\n"
02040            elif codons == "12":
02041                for key in part_list:
02042                    start, end = str(part_dict[key]).split("-")
02043                    part_string += "  charset " + key + "_pos1" + " = " + start + " - " + end + "\\2"
     + ";\n"
```

```
02044                         part_string += "  charset " + key + "_pos2" + " = " + str(int(start) + 1) + " - "
      + end + "\\2" + ";\n"
02045               elif codons == "123":
02046                   for key in part_list:
02047                       start, end = str(part_dict[key]).split("-")
02048                       part_string += "  charset " + key + "_pos1" + " = " + start + " - " + end + "\\3"
      + ";\n"
02049                       part_string += "  charset " + key + "_pos2" + " = " + str(int(start) + 1) + " - "
      + end + "\\3" + ";\n"
02050                       part_string += "  charset " + key + "_pos3" + " = " + str(int(start) + 2) + " - "
      + end + "\\3" + ";\n"
02051               part_string += "end;\n"
02052
02053           elif data_type == "aa":
02054               for key in part_list:
02055                   part_string += "  charset " + key + " = " + str(part_dict[key]) + ";\n"
02056               part_string += "end;\n"
02057
02058           return part_string
02059
```

References amas.AMAS.MetaAlignment.get_concatenated(), amas.AMAS.MetaAlignment.natural_sort(), amas.AMAS.MetaAlignmen and amas.AMAS.MetaAlignment.split.

Referenced by amas.AMAS.MetaAlignment.write_partitions().

Here is the call graph for this function:
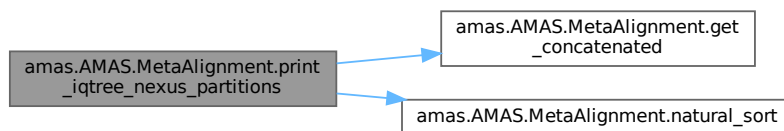
Here is the caller graph for this function:

7.6.3.20   print_nexus()

```
amas.AMAS.MetaAlignment.print_nexus (
                self,
                source_dict )
```
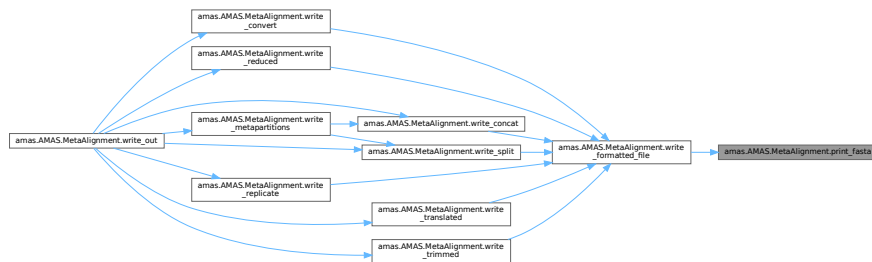
Definition at line 1897 of file AMAS.py.

```
01897      def print_nexus(self, source_dict):
01898          # print nexus-formatted string from a dictionary
01899          if self.data_type == "aa" or self.command == "translate":
01900              data_type = "PROTEIN"
01901          elif self.data_type == "dna":
01902              data_type = "DNA"
01903
```

```
01904          taxa_list = list(source_dict.keys())
01905          no_taxa = len(taxa_list)
01906          pad_longest_name = len(max(taxa_list, key=len)) + 3
01907          seq_length = len(next(iter(source_dict.values())))
01908          header = str(len(source_dict)) + " " + str(seq_length)
01909          nexus_string = (
01910              "#NEXUS\n\nBEGIN DATA;\n\tDIMENSIONS  NTAX=" + str(no_taxa) + " NCHAR=" + str(seq_length)
01911              + ";\n\tFORMAT DATATYPE=" + data_type + "  GAP = - MISSING = ?;\n\tMATRIX\n"
01912          )
01913
01914          for taxon, seq in sorted(source_dict.items()):
01915              taxon = taxon.replace(" ", "_").strip("'")
01916              nexus_string += "\t" + taxon.ljust(pad_longest_name, ' ') + seq + "\n"
01917          nexus_string += "\n;\n\nEND;"
01918
01919          return nexus_string
01920
```

References [amas.AMAS.MetaAlignment.command](#), [amas.AMAS.Alignment.data_type](#), and [amas.AMAS.MetaAlignment.data_type](#).

Referenced by [amas.AMAS.MetaAlignment.write_formatted_file()](#).

Here is the caller graph for this function:
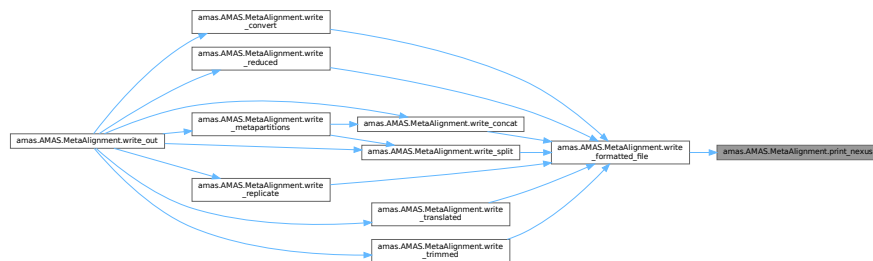
7.6.3.21   print_nexus_int()

```
amas.AMAS.MetaAlignment.print_nexus_int (
              self,
              source_dict )
```

Definition at line [1921](#) of file [AMAS.py](#).

```
01921      def print_nexus_int(self, source_dict):
01922          # print nexus interleaved-formatted string from a dictionary
01923          if self.data_type == "aa":
01924              data_type = "PROTEIN"
01925          elif self.data_type == "dna":
01926              data_type = "DNA"
01927
01928          taxa_list = list(source_dict.keys())
01929          no_taxa = len(taxa_list)
01930          pad_longest_name = len(max(taxa_list, key=len)) + 3
01931          seq_length = len(next(iter(source_dict.values())))
01932          header = str(len(source_dict)) + " " + str(seq_length)
01933          # this will be a list of tuples to hold taxa names and sequences
01934          seq_matrix = []
01935          nexus_int_string = (
01936              "#NEXUS\n\nBEGIN DATA;\n\tDIMENSIONS  NTAX=" + str(no_taxa) + " NCHAR=" + str(seq_length)
01937              + ";\n\tFORMAT   INTERLEAVE" + "  DATATYPE=" + data_type + "  GAP = - MISSING =
       ?;\n\tMATRIX\n"
01938          )
01939          # each sequence line will have 500 characters
01940          n = 500
01941
01942          # recreate sequence matrix
01943          add_to_matrix = seq_matrix.append
01944          for taxon, seq in sorted(source_dict.items()):
01945              add_to_matrix((taxon, [seq[i:i+n] for i in range(0, len(seq), n)]))
01946
```

```
01947            first_seq = seq_matrix[0][1]
01948            for index, item in enumerate(first_seq):
01949                for taxon, sequence in seq_matrix:
01950                    if index == 0:
01951                        nexus_int_string += taxon.ljust(pad_longest_name, ' ') + sequence[index] + "\n"
01952                    else:
01953                        nexus_int_string += sequence[index] + "\n"
01954                nexus_int_string += "\n"
01955
01956        nexus_int_string += "\n;\n\nEND;"
01957
01958        return nexus_int_string
01959
```

References amas.AMAS.Alignment.data_type, and amas.AMAS.MetaAlignment.data_type.

Referenced by amas.AMAS.MetaAlignment.write_formatted_file().

Here is the caller graph for this function:

7.6.3.22 print_nexus_partitions()

```
amas.AMAS.MetaAlignment.print_nexus_partitions (
            self,
            data_type,
            codons )
```

Definition at line 1994 of file AMAS.py.

```
01994        def print_nexus_partitions(self, data_type, codons):
01995            # print partitions for concatenated alignment
01996            part_string = ""
01997            part_dict = self.get_concatenated(self.parsed_alignments)[1]
01998            part_list = self.natural_sort(part_dict.keys())
01999            # write beginning of nexus sets
02000            part_string += "#NEXUS\n\n"
02001            part_string += "BEGIN SETS;\n"
02002
02003            if data_type == "dna":
02004                if codons == "none":
02005                    for key in part_list:
02006                        part_string += "\tcharset " + key + " = " + str(part_dict[key]) + ";\n"
02007                elif codons == "12":
02008                    for key in part_list:
02009                        start, end = str(part_dict[key]).split("-")
02010                        part_string += "\tcharset " + key + "_pos1" + " = " + start + "-" + end + "\\2" +
";\n"
02011                        part_string += "\tcharset " + key + "_pos2" + " = " + str(int(start) + 1) + "-" +
    end + "\\2" + ";\n"
02012                elif codons == "123":
02013                    for key in part_list:
02014                        start, end = str(part_dict[key]).split("-")
02015                        part_string += "\tcharset " + key + "_pos1" + " = " + start + "-" + end + "\\3" +
";\n"
02016                        part_string += "\tcharset " + key + "_pos2" + " = " + str(int(start) + 1) + "-" +
    end + "\\3" + ";\n"
02017                        part_string += "\tcharset " + key + "_pos3" + " = " + str(int(start) + 2) + "-" +
    end + "\\3" + ";\n"
02018            part_string += "END;"
```

```
02019
02020            elif data_type == "aa":
02021                for key in part_list:
02022                    part_string += "\tcharset " + key + " = " + str(part_dict[key]) + ";\n"
02023                part_string += "END;"
02024
02025            return part_string
02026
```

References amas.AMAS.MetaAlignment.get_concatenated(), amas.AMAS.MetaAlignment.natural_sort(), amas.AMAS.MetaAlignmen and amas.AMAS.MetaAlignment.split.

Referenced by amas.AMAS.MetaAlignment.write_partitions().

Here is the call graph for this function:
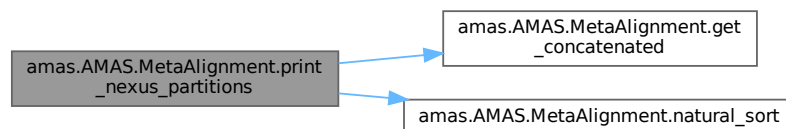
Here is the caller graph for this function:

7.6.3.23 print_phylip()

```
amas.AMAS.MetaAlignment.print_phylip (
            self,
            source_dict )
```

Definition at line 1850 of file AMAS.py.

```
01850    def print_phylip(self, source_dict):
01851        # print phylip-formatted string from a dictionary
01852        taxa_list = list(source_dict.keys())
01853        no_taxa = len(taxa_list)
01854        # figure out the max length of a taxon for nice padding of sequences
01855        pad_longest_name = len(max(taxa_list, key=len)) + 3
01856        # get sequence length from a random value
01857        seq_length = len(next(iter(source_dict.values())))
01858        header = str(len(source_dict)) + " " + str(seq_length)
01859        phylip_string = header + "\n"
01860        for taxon, seq in sorted(source_dict.items()):
01861            taxon = taxon.replace(" ", "_").strip("'")
01862            # left-justify taxon names relative to sequences
01863            phylip_string += taxon.ljust(pad_longest_name, ' ') + seq + "\n"
01864
01865        return phylip_string
01866
```

Referenced by amas.AMAS.MetaAlignment.write_formatted_file().

Here is the caller graph for this function:

7.6.3.24 print_phylip_int()

```
amas.AMAS.MetaAlignment.print_phylip_int (
            self,
            source_dict )
```

Definition at line 1867 of file AMAS.py.

```
01867     def print_phylip_int(self, source_dict):
01868         # print phylip interleaved-formatted string from a dictionary
01869         taxa_list = list(source_dict.keys())
01870         no_taxa = len(taxa_list)
01871         pad_longest_name = len(max(taxa_list, key=len)) + 3
01872         seq_length = len(next(iter(source_dict.values())))
01873         header = str(len(source_dict)) + " " + str(seq_length)
01874         phylip_int_string = header + "\n\n"
01875         # this will be a list of tuples to hold taxa names and sequences
01876         seq_matrix = []
01877
01878         # each sequence line will have 500 characters
01879         n = 500
01880
01881         # recreate sequence matrix
01882         add_to_matrix = seq_matrix.append
01883         for taxon, seq in sorted(source_dict.items()):
01884             add_to_matrix((taxon, [seq[i:i+n] for i in range(0, len(seq), n)]))
01885
01886         first_seq = seq_matrix[0][1]
01887         for index, item in enumerate(first_seq):
01888             for taxon, sequence in seq_matrix:
01889                 if index == 0:
01890                     phylip_int_string += taxon.ljust(pad_longest_name, ' ') + sequence[index] + "\n"
01891                 else:
01892                     phylip_int_string += sequence[index] + "\n"
01893             phylip_int_string += "\n"
01894
01895         return phylip_int_string
01896
```

Referenced by amas.AMAS.MetaAlignment.write_formatted_file().

Here is the caller graph for this function:

7.6.3.25 print_raxml_partitions()

amas.AMAS.MetaAlignment.print_raxml_partitions (

       self,

       data_type,

       codons)

Definition at line 2060 of file AMAS.py.

```
02060        def print_raxml_partitions(self, data_type, codons):
02061            # print partitions for concatenated alignment
02062            part_string = ""
02063            part_dict = self.get_concatenated(self.parsed_alignments)[1]
02064            part_list = self.natural_sort(part_dict.keys())
02065
02066            if data_type == "dna":
02067                if codons == "none":
02068                    for key in part_list:
02069                        part_string += "DNA, " + key + " = " + str(part_dict[key]) + "\n"
02070                elif codons == "12":
02071                    for key in part_list:
02072                        start, end = str(part_dict[key]).split("-")
02073                        part_string += "DNA, " + key + "_pos1" + " = " + start + "-" + end + "\\2" + "\n"
02074                        part_string += "DNA, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end +
      "\\2" + "\n"
02075                elif codons == "123":
02076                    for key in part_list:
02077                        start, end = str(part_dict[key]).split("-")
02078                        part_string += "DNA, " + key + "_pos1" + " = " + start + "-" + end + "\\3" + "\n"
02079                        part_string += "DNA, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end +
      "\\3" + "\n"
02080                        part_string += "DNA, " + key + "_pos3" + " = " + str(int(start) + 2) + "-" + end +
      "\\3" + "\n"
02081
02082            elif data_type == "aa":
02083                for key in part_list:
02084                    part_string += "WAG, " + key + " = " + str(part_dict[key]) + "\n"
02085
02086                #  aa-partition files with strides are probably not useful? (original below)
02087 #            elif codons == "12":
02088 #                for key in part_list:
02089 #                    start, end = str(part_dict[key]).split("-")
02090 #                    part_string += "WAG, " + key + "_pos1" + " = " + start + "-" + end + "\\2" + "\n"
02091 #                    part_string += "WAG, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end
      + "\\2" + "\n"
02092 #            elif codons == "123":
02093 #                for key in part_list:
02094 #                    start, end = str(part_dict[key]).split("-")
02095 #                    part_string += "WAG, " + key + "_pos1" + " = " + start + "-" + end + "\\3" + "\n"
02096 #                    part_string += "WAG, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end
      + "\\3" + "\n"
02097 #                    part_string += "WAG, " + key + "_pos3" + " = " + str(int(start) + 2) + "-" + end
      + "\\3" + "\n"
02098            return part_string
02099
```

References amas.AMAS.MetaAlignment.get_concatenated(), amas.AMAS.MetaAlignment.natural_sort(), amas.AMAS.MetaAlignment and amas.AMAS.MetaAlignment.split.

Referenced by amas.AMAS.MetaAlignment.write_partitions().

Here is the call graph for this function:

Here is the caller graph for this function:

7.6.3.26 print_unspecified_partitions()

amas.AMAS.MetaAlignment.print_unspecified_partitions (
        self,
        data_type,
        codons)

Definition at line 1966 of file AMAS.py.
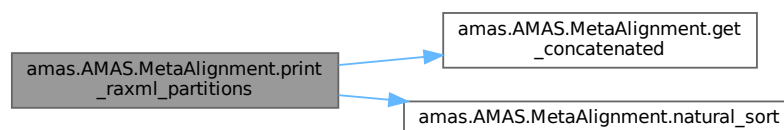
```
01966    def print_unspecified_partitions(self, data_type, codons):
01967        # print partitions for concatenated alignment
01968        part_string = ""
01969        part_dict = self.get_concatenated(self.parsed_alignments)[1]
01970        part_list = self.natural_sort(part_dict.keys())
01971
01972        if data_type == "dna":
01973            if codons == "none":
01974                for key in part_list:
01975                    part_string += key + " = " + str(part_dict[key]) + "\n"
01976            elif codons == "12":
01977                for key in part_list:
01978                    start, end = str(part_dict[key]).split("-")
01979                    part_string += key + "_pos1" + " = " + start + "-" + end + "\\2" + "\n"
01980                    part_string += key + "_pos2" + " = " + str(int(start) + 1) + "-" + end + "\\2" +
        "\n"
01981            elif codons == "123":
01982                for key in part_list:
01983                    start, end = str(part_dict[key]).split("-")
01984                    part_string += key + "_pos1" + " = " + start + "-" + end + "\\3" + "\n"
01985                    part_string += key + "_pos2" + " = " + str(int(start) + 1) + "-" + end + "\\3" +
        "\n"
01986                    part_string += key + "_pos3" + " = " + str(int(start) + 2) + "-" + end + "\\3" +
        "\n"
01987
01988        elif data_type == "aa":
01989            for key in part_list:
01990                part_string += key + " = " + str(part_dict[key]) + "\n"
01991
01992        return part_string
01993
```

References amas.AMAS.MetaAlignment.get_concatenated(), amas.AMAS.MetaAlignment.natural_sort(), amas.AMAS.MetaAlignmen
and amas.AMAS.MetaAlignment.split.

Referenced by amas.AMAS.MetaAlignment.write_partitions().

Here is the call graph for this function:

Here is the caller graph for this function:

7.6.3.27 remove_empty_sequences()

```
amas.AMAS.MetaAlignment.remove_empty_sequences (
            self,
            split_alignment )
```

Definition at line 1510 of file AMAS.py.

```
01510    def remove_empty_sequences(self, split_alignment):
01511        # remove taxa from alignment if they are composed of only empty sequences
01512        new_alignment = {taxon : seq for taxon, seq in split_alignment.items() if
        self.remove_unknown_chars(seq)}
01513
01514        return new_alignment
01515
```

References amas.AMAS.MetaAlignment.remove_unknown_chars().

Referenced by amas.AMAS.MetaAlignment.get_partitioned().

Here is the call graph for this function:

Here is the caller graph for this function:

7.6.3.28 remove_from_alignment()

```
amas.AMAS.MetaAlignment.remove_from_alignment (
            self,
            alignment,
            species_to_remove_set,
            index )
```

Definition at line 1807 of file AMAS.py.

```
01807     def remove_from_alignment(self, alignment, species_to_remove_set, index):
01808         # remove taxa from alignment
01809         aln_name = self.get_alignment_name_no_ext(index)
01810         for taxon in species_to_remove_set:
01811             if taxon not in alignment.keys():
01812                 print(
01813                     "WARNING: Taxon '" + taxon + "' not found in '" + aln_name + "'.\nIf you expected
      it to be there, "
01814                     "make sure to replace all taxon name spaces with underscores and that you are not
      using quotes."
01815                 )
01816         # originally within for-loop scope (redundancy)
01817         new_alignment = {species: seq for species, seq in alignment.items() if species not in
      species_to_remove_set}
01818
01819         return aln_name, new_alignment
01820
```

References amas.AMAS.MetaAlignment.get_alignment_name_no_ext().

Referenced by amas.AMAS.MetaAlignment.remove_taxa().

Here is the call graph for this function:
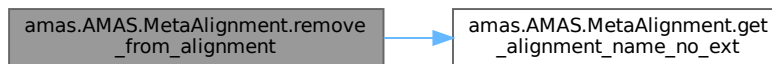
Here is the caller graph for this function:

7.6.3.29 remove_taxa()

```
amas.AMAS.MetaAlignment.remove_taxa (
            self,
            species_to_remove_set )
```

Definition at line 1821 of file AMAS.py.

```
01821     def remove_taxa(self, species_to_remove_set):
01822         new_alns = {}
01823         for index, alignment in enumerate(self.parsed_alignments):
01824             aln_name, aln_dict = self.remove_from_alignment(alignment, species_to_remove_set, index)
01825             # check if alignment is not empty:
01826             if aln_dict:
```

```
01827                new_alns[aln_name] = aln_dict
01828           else:
01829                print("ERROR: You asked to remove all taxa from the alignment " + aln_name + ". No
     output file will be written.")
01830
01831        return new_alns
01832
```

References amas.AMAS.MetaAlignment.parsed_alignments, and amas.AMAS.MetaAlignment.remove_from_alignment().

Referenced by amas.AMAS.MetaAlignment.write_reduced().

Here is the call graph for this function:
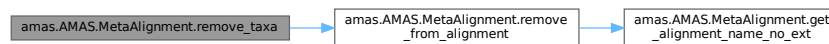
Here is the caller graph for this function:

7.6.3.30 remove_unknown_chars()

amas.AMAS.MetaAlignment.remove_unknown_chars (
            self,
            seq)

Definition at line 1504 of file AMAS.py.
```
01504    def remove_unknown_chars(self, seq):
01505        # remove unknown characters from sequence
01506        new_seq = seq.replace("?", "").replace("-", "")
01507
01508        return new_seq
01509
```

Referenced by amas.AMAS.MetaAlignment.remove_empty_sequences().

Here is the caller graph for this function:

7.6.3.31 replace_string_in_file()

```
amas.AMAS.MetaAlignment.replace_string_in_file (
            self,
            file_name,
            old_string,
            new_string )
```

Definition at line 2100 of file AMAS.py.

```
02100     def replace_string_in_file(self, file_name, old_string, new_string):
02101         # global string replacement in file
02102         with open(file_name, "r", encoding="utf-8") as file:
02103             file_content = file.read()
02104         # write globally replaced content back to file
02105         glb_replaced_content = file_content.replace(old_string, new_string)
02106         with open(file_name, "w", encoding="utf-8") as file:
02107             file.write(glb_replaced_content)
02108
```

Referenced by amas.AMAS.MetaAlignment.write_partitions().

Here is the caller graph for this function:

7.6.3.32 summarize_alignments()

```
amas.AMAS.MetaAlignment.summarize_alignments (
            self,
            alignment )
```

Definition at line 1648 of file AMAS.py.

```
01648     def summarize_alignments(self, alignment):
01649         # helper function to summarize alignments
01650         summary = alignment.get_summary()
01651         return summary
01652
```

Referenced by amas.AMAS.MetaAlignment.get_summaries().

Here is the caller graph for this function:

7.6.3.33 summarize_alignments_taxa()

```
amas.AMAS.MetaAlignment.summarize_alignments_taxa (
            self,
            alignment )
```

Definition at line 1693 of file AMAS.py.

```
01693    def summarize_alignments_taxa(self, alignment):
01694        # helper function to summarize alignments by taxon
01695        summary = alignment.get_taxa_summary()
01696        return summary
01697
```

Referenced by amas.AMAS.MetaAlignment.get_taxon_summaries().

Here is the caller graph for this function:
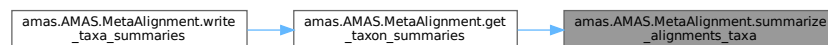
7.6.3.34 translate_dict()

```
amas.AMAS.MetaAlignment.translate_dict (
            self,
            source_dict )
```

Definition at line 1467 of file AMAS.py.

```
01467    def translate_dict(self, source_dict):
01468        translation_table = self.codes.get(self.genetic_code)
01469        translated_dict = {}
01470        for taxon, seq in sorted(source_dict.items()):
01471            translated_seq = self.translate_dna_to_aa(seq, translation_table, self.reading_frame)
01472            if "*" in translated_seq:
01473                print("WARNING: stop codon(s), indicated as *, found in {} sequence".format(taxon))
01474            translated_dict[taxon] = translated_seq
01475
01476        return translated_dict
01477
```

References amas.AMAS.MetaAlignment.codes, amas.AMAS.MetaAlignment.genetic_code, amas.AMAS.MetaAlignment.reading_fram and amas.AMAS.MetaAlignment.translate_dna_to_aa().

Referenced by amas.AMAS.MetaAlignment.get_translated().

Here is the call graph for this function:
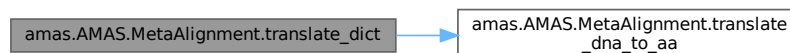
Here is the caller graph for this function:
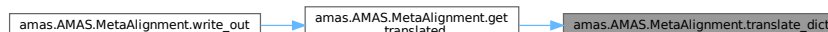
7.6.3.35 translate_dna_to_aa()

```
amas.AMAS.MetaAlignment.translate_dna_to_aa (
        self,
        seq,
        translation_table,
        frame )
```

Definition at line 1446 of file AMAS.py.

```
01446    def translate_dna_to_aa(self, seq, translation_table, frame):
01447        # translate DNA string into amino acids
01448        # where the last codon starts
01449        last_codon_start = len(seq) - 2
01450        # where the first codon starts
01451        if frame == 1:
01452            first = 0
01453        elif frame == 2:
01454            first = 1
01455        elif frame == 3:
01456            first = 2
01457        # create protein sequence by growing list
01458        protein = []
01459        add_to_protein = protein.append
01460        for start in range(first, last_codon_start, 3):
01461            codon = seq[start : start + 3]
01462            aa = translation_table.get(codon.upper(), 'X')
01463            add_to_protein(aa)
01464
01465        return "".join(protein)
01466
```

Referenced by amas.AMAS.MetaAlignment.translate_dict().

Here is the caller graph for this function:

```
amas.AMAS.MetaAlignment.write_out → amas.AMAS.MetaAlignment.get_translated → amas.AMAS.MetaAlignment.translate_dict → amas.AMAS.MetaAlignment.translate_dna_to_aa
```

7.6.3.36 trim_dict()

```
amas.AMAS.MetaAlignment.trim_dict (
        self,
        alignment )
```

Definition at line 1487 of file AMAS.py.

```
01487    def trim_dict(self, alignment):
01488        trim_vector = alignment.get_trim_selection(self.trim_fraction, self.parsimony_check)
01489        aln_dict = alignment.parsed_aln
01490        for key in aln_dict:
01491            aln_dict[key] = "".join(list(compress(aln_dict[key], trim_vector)))
01492
01493        return aln_dict
01494
```

References amas.AMAS.MetaAlignment.parsimony_check, and amas.AMAS.MetaAlignment.trim_fraction.

Referenced by amas.AMAS.MetaAlignment.get_trimmed().

Here is the caller graph for this function:

```
amas.AMAS.MetaAlignment.write_out → amas.AMAS.MetaAlignment.get_trimmed → amas.AMAS.MetaAlignment.trim_dict
```
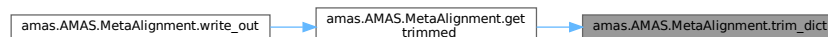
7.6.3.37 write_concat()

```
amas.AMAS.MetaAlignment.write_concat (
            self,
            file_format )
```

Definition at line 2188 of file AMAS.py.

```
02188    def write_concat(self, file_format):
02189        # write concatenated alignment into a file
02190        concatenated_alignment = self.get_concatenated(self.parsed_alignments)[0]
02191        file_name = self.concat_out
02192        self.file_overwrite_error(file_name)
02193        self.write_formatted_file(file_format, file_name, concatenated_alignment)
02194
02195        print("Wrote concatenated sequences to " + file_format + " file '" + file_name + "'")
02196
```

References amas.AMAS.MetaAlignment.concat_out, amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.
amas.AMAS.MetaAlignment.parsed_alignments, and amas.AMAS.MetaAlignment.write_formatted_file().

Referenced by amas.AMAS.MetaAlignment.write_metapartitions(), and amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
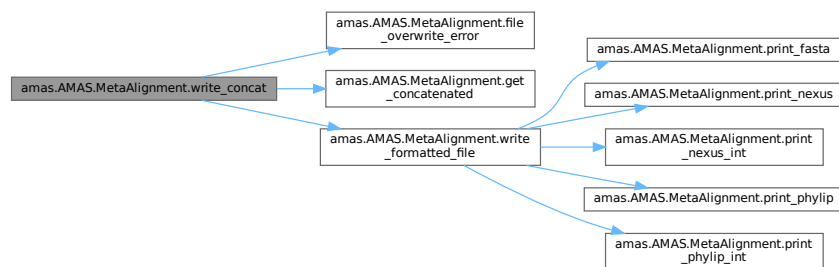
Here is the caller graph for this function:
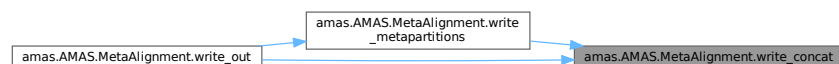
7.6.3.38 write_convert()

```
amas.AMAS.MetaAlignment.write_convert (
            self,
            index,
            alignment,
            file_format,
            extension )
```

Definition at line 2197 of file AMAS.py.

```
02197    def write_convert(self, index, alignment, file_format, extension):
02198        # write converted alignment into a file
```

```
02199            file_name = self.get_alignment_name(index, extension)
02200            self.file_overwrite_error(file_name)
02201            self.write_formatted_file(file_format, file_name, alignment)
02202
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.get_alignment_name(), and amas.AMAS.MetaAlignment.write_formatted_file().

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
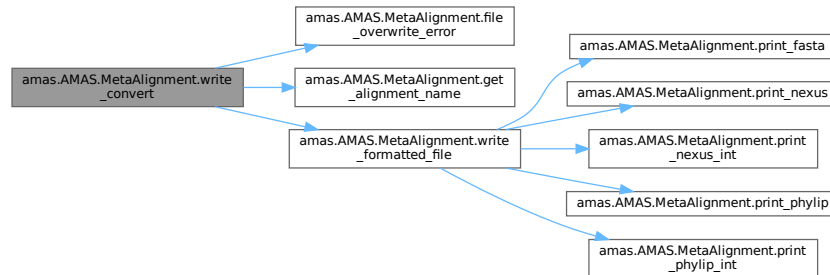
Here is the caller graph for this function:
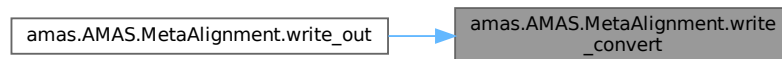
7.6.3.39 write_formatted_file()

```
amas.AMAS.MetaAlignment.write_formatted_file (
            self,
            file_format,
            file_name,
            alignment )
```

Definition at line 2162 of file AMAS.py.

```
02162    def write_formatted_file(self, file_format, file_name, alignment):
02163        # write the correct format string into a file
02164        with open(file_name, "w", encoding="utf-8") as out_file:
02165            if file_format == "phylip":
02166                out_file.write(self.print_phylip(alignment))
02167            elif file_format == "fasta":
02168                out_file.write(self.print_fasta(alignment))
02169            elif file_format == "phylip-int":
02170                out_file.write(self.print_phylip_int(alignment))
02171            elif file_format == "nexus":
02172                out_file.write(self.print_nexus(alignment))
02173            elif file_format == "nexus-int":
02174                out_file.write(self.print_nexus_int(alignment))
02175
```

References amas.AMAS.MetaAlignment.print_fasta(), amas.AMAS.MetaAlignment.print_nexus(), amas.AMAS.MetaAlignment.print_n
amas.AMAS.MetaAlignment.print_phylip(), and amas.AMAS.MetaAlignment.print_phylip_int().

Referenced by amas.AMAS.MetaAlignment.write_concat(), amas.AMAS.MetaAlignment.write_convert(), amas.AMAS.MetaAlignment
amas.AMAS.MetaAlignment.write_replicate(), amas.AMAS.MetaAlignment.write_split(), amas.AMAS.MetaAlignment.write_translated
and amas.AMAS.MetaAlignment.write_trimmed().

Here is the call graph for this function:
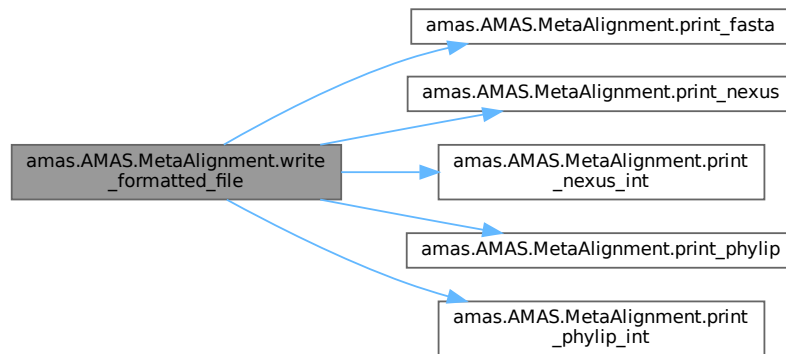
Here is the caller graph for this function:
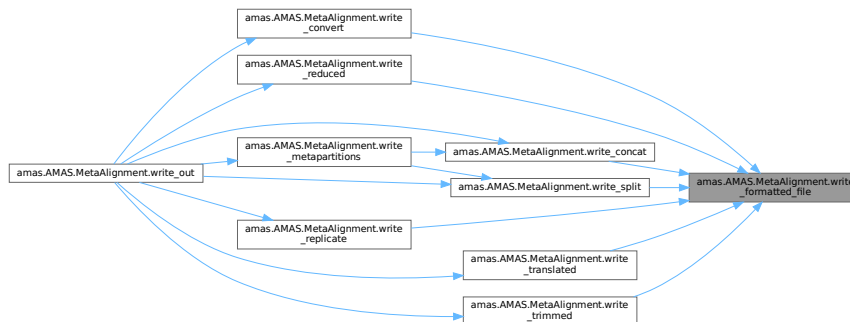
7.6.3.40 write_metapartitions()

amas.AMAS.MetaAlignment.write_metapartitions (
        self,
        file_format)

Definition at line 2263 of file AMAS.py.

```
02263        def write_metapartitions(self, file_format):
02264            # write metapartitions - combines split and concat
02265            print("write_out elif action == metapartitions")
02266            metapartition_extension = self.get_metapartition_extension(file_format)
02267            list_of_alignments = self.get_partitioned(self.split)
02268            written_split_files = []
02269            err_indx = 0
```

```
02270
02271         for item in list_of_alignments:
02272             try:
02273                 for split_file in self.write_split(item, file_format, metapartition_extension):
02274                     written_split_files.append(split_file)
02275             except ValueError as e:
02276                 print("WARNING: ", e)
02277                 err_indx += 1
02278         if len(written_split_files) > 0:
02279             print("Wrote %d %s metapartition files from partitions provided" %
        (len(written_split_files), file_format))
02280         if err_indx > 0:
02281             print("WARNING: %d file(s) raised an error while writing (see above)." % err_indx)
02282
02283         # now set inputs to be the collated metapartition alignment files
02284         self.in_files = written_split_files
02285         self.alignment_objects = self.get_alignment_objects()
02286         self.parsed_alignments = self.get_parsed_alignments()
02287
02288         # concat metapartition alignment files
02289         self.write_concat(file_format)
02290
```

References amas.AMAS.MetaAlignment.alignment_objects, amas.AMAS.MetaAlignment.get_alignment_objects(), amas.AMAS.MetaAlignment.get_metapartition_extension(), amas.AMAS.MetaAlignment.get_parsed_alignments(), amas.AMAS.MetaAlignment.get_partitioned(), amas.AMAS.MetaAlignment.in_files, amas.AMAS.MetaAlignment.parsed_alignments, amas.AMAS.ParsedArgs.split(), amas.AMAS.MetaAlignment.split, amas.AMAS.MetaAlignment.write_concat(), and amas.AMAS.MetaAlignment.write_split().

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:

Here is the caller graph for this function:
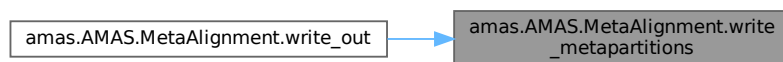
7.6.3.41 write_out()

```
amas.AMAS.MetaAlignment.write_out (
            self,
```

```
            action,
            file_format )
```

Definition at line 2291 of file AMAS.py.

```
02291      def write_out(self, action, file_format):
02292          # write other output files depending on command (action)
02293          extension = self.get_extension(file_format)
02294
02295          if action == "concat":
02296              self.write_concat(file_format)
02297
02298          elif action == "convert":
02299              length = len(self.alignment_objects)
02300              [
02301                  self.write_convert(i, alignment, file_format, extension)
02302                  for i, alignment in enumerate(self.parsed_alignments)
02303              ]
02304              print("Converted " + str(length) + " files from " + self.in_format + " to " + file_format)
02305
02306          elif action == "replicate":
02307              [
02308                  self.write_replicate(i, alignment, file_format, extension)
02309                  for i, alignment in enumerate(self.get_replicate(self.no_replicates, self.no_loci))
02310              ]
02311
02312              print("Constructed " + str(self.no_replicates) + " replicate data sets, each from " +
       str(self.no_loci) + " alignments")
02313
02314          elif action == "split":
02315              list_of_alignments = self.get_partitioned(self.split)
02316              written_split_files = []
02317              err_indx = 0
02318
02319              for item in list_of_alignments:
02320                  try:
02321                      for split_file in self.write_split(item, file_format, extension):
02322                          written_split_files.append(split_file)
02323                  except ValueError as e:
02324                      print("WARNING: ", e)
02325                      err_indx += 1
02326              if len(written_split_files) > 0:
02327                  print("Wrote %d %s files from partitions provided" % (len(written_split_files),
       file_format))
02328              if err_indx > 0:
02329                  print("WARNING: %d file(s) raised an error while writing (see above)." % err_indx)
02330
02331          elif action == "metapartitions":
02332              self.write_metapartitions(file_format)
02333
02334          elif action == "remove":
02335              aln_no = self.write_reduced(file_format, extension)
02336              if aln_no:
02337                  print("Wrote " + str(aln_no) + " " + str(file_format) + " files with reduced taxon
       set")
02338
02339          elif action == "translate":
02340              if self.data_type == "aa":
02341                  print("ERROR: cannot translate; you said your alignment already contains amino acids")
02342                  sys.exit()
02343              translated_alignment_dicts = self.get_translated(self.genetic_code, self.reading_frame)
02344              length = len(self.alignment_objects)
02345              [
02346                  self.write_translated(i, alignment, file_format, extension)
02347                  for i, alignment in enumerate(translated_alignment_dicts)
02348              ]
02349              print("Translated " + str(length) + " files to amino acid sequences")
02350
02351          elif action == "trim": # self.trim_fraction, self.parsimony_check
02352              trimmed_alignment_dicts = self.get_trimmed(self.trim_fraction, self.parsimony_check)
02353              length = len(self.alignment_objects)
02354              [
02355                  self.write_trimmed(i, alignment, file_format, extension)
02356                  for i, alignment in enumerate(trimmed_alignment_dicts)
02357              ]
02358              print("Trimmed", str(length), "file(s) to have", self.trim_fraction, "minimum occupancy
       per alignment column")
02359
02360
```
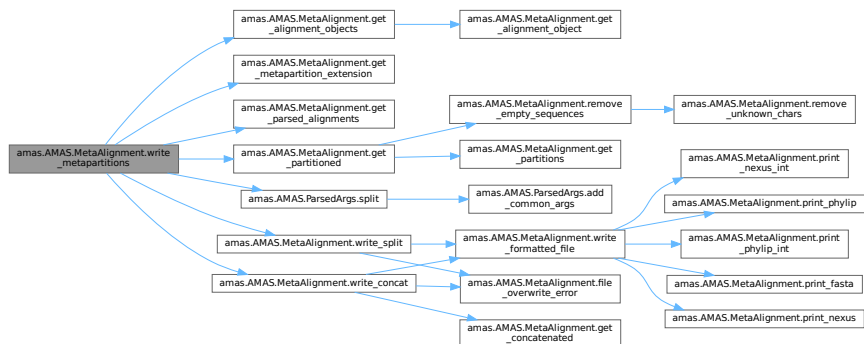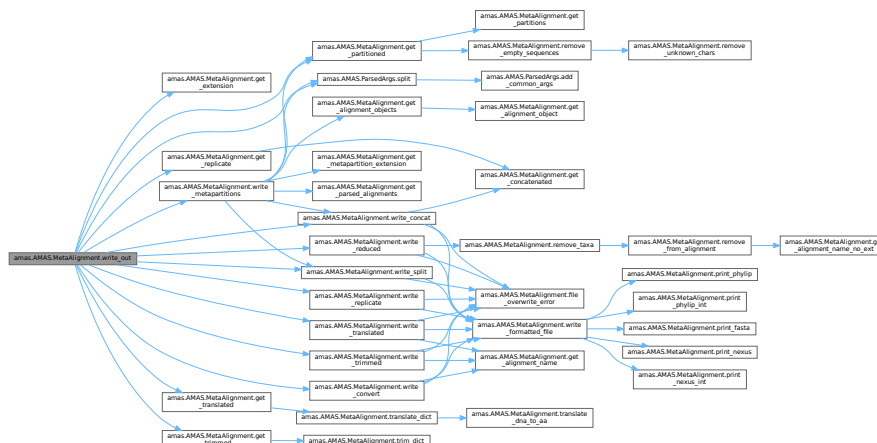
References amas.AMAS.MetaAlignment.alignment_objects, amas.AMAS.Alignment.data_type, amas.AMAS.MetaAlignment.data_typ
amas.AMAS.MetaAlignment.genetic_code, amas.AMAS.MetaAlignment.get_extension(), amas.AMAS.MetaAlignment.get_partitioned
amas.AMAS.MetaAlignment.get_replicate(), amas.AMAS.MetaAlignment.get_translated(), amas.AMAS.MetaAlignment.get_trimmed(
amas.AMAS.Alignment.in_format, amas.AMAS.MetaAlignment.in_format, amas.AMAS.MetaAlignment.no_loci,

amas.AMAS.MetaAlignment.no_replicates, amas.AMAS.MetaAlignment.parsed_alignments, amas.AMAS.MetaAlignment.parsimony_
amas.AMAS.MetaAlignment.reading_frame, amas.AMAS.ParsedArgs.split(), amas.AMAS.MetaAlignment.split,
amas.AMAS.MetaAlignment.trim_fraction, amas.AMAS.MetaAlignment.write_concat(), amas.AMAS.MetaAlignment.write_convert(),
amas.AMAS.MetaAlignment.write_metapartitions(), amas.AMAS.MetaAlignment.write_reduced(), amas.AMAS.MetaAlignment.write_
amas.AMAS.MetaAlignment.write_split(), amas.AMAS.MetaAlignment.write_translated(), and amas.AMAS.MetaAlignment.write_trim

Here is the call graph for this function:

7.6.3.42 write_partitions()

```
amas.AMAS.MetaAlignment.write_partitions (
            self,
            file_name,
            part_format,
            data_type,
            codons )
```

Definition at line 2109 of file AMAS.py.

```
02109      def write_partitions(self, file_name, part_format, data_type, codons):
02110          # write partitions file for concatenated alignment
02111          self.file_overwrite_error(file_name)
02112          with open(file_name, "w", encoding="utf-8") as part_file:
02113              if part_format == "nexus":
02114                  part_file.write(self.print_nexus_partitions(data_type, codons))
02115              if part_format == "iqtree-nexus":
02116                  part_file.write(self.print_iqtree_nexus_partitions(data_type, codons))
02117              if part_format == "raxml":
02118                  part_file.write(self.print_raxml_partitions(data_type, codons))
02119              if part_format == "unspecified":
02120                  part_file.write(self.print_unspecified_partitions(data_type, codons))
02121
02122              if self.using_metapartitions:
02123                  self.replace_string_in_file(file_name, '-meta =', ' =')
02124
02125          print("Wrote partitions for the concatenated file to '" + file_name + "'")
02126
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.print_iqtree_nexus_partitions(),
amas.AMAS.MetaAlignment.print_nexus_partitions(), amas.AMAS.MetaAlignment.print_raxml_partitions(),
amas.AMAS.MetaAlignment.print_unspecified_partitions(), amas.AMAS.MetaAlignment.replace_string_in_file(),
and amas.AMAS.MetaAlignment.using_metapartitions.

Here is the call graph for this function:
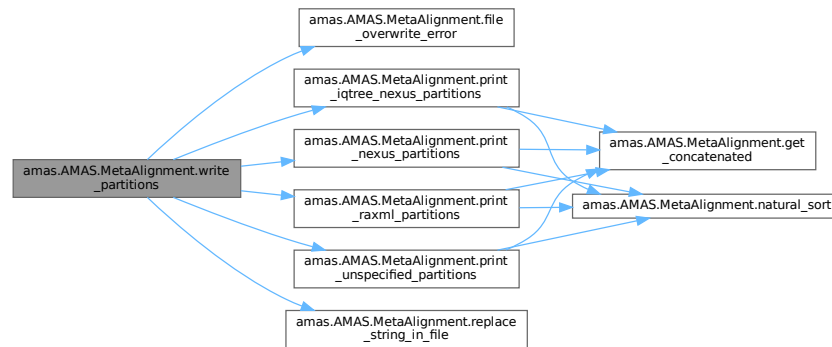
7.6.3.43 write_reduced()

```
amas.AMAS.MetaAlignment.write_reduced (
            self,
            file_format,
            extension )
```

Definition at line 2234 of file AMAS.py.

```
02234    def write_reduced(self, file_format, extension):
02235        # write alignment with taxa removed into a file
02236        prefix = self.reduced_file_prefix
02237        alns = self.remove_taxa(self.species_to_remove)
02238        for file_name, aln_dict in alns.items():
02239            out_file_name = prefix + file_name + extension
02240            self.file_overwrite_error(out_file_name)
02241            self.write_formatted_file(file_format, out_file_name, aln_dict)
02242        return len(alns)
02243
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.reduced_file_prefix, amas.AMAS.MetaAlignment.remove_taxa(), amas.AMAS.MetaAlignment.species_to_remove, and amas.AMAS.MetaAlignment.write_

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:

Here is the caller graph for this function:

7.6.3.44 write_replicate()

```
amas.AMAS.MetaAlignment.write_replicate (
            self,
            index,
            alignment,
            file_format,
            extension )
```

Definition at line 2203 of file AMAS.py.

```
02203    def write_replicate(self, index, alignment, file_format, extension):
02204        # write replicate alignment into a file
02205        file_name = "replicate" + str(index + 1) + "_" + str(self.no_loci) + "-loci" + extension
02206        self.file_overwrite_error(file_name)
02207        self.write_formatted_file(file_format, file_name, alignment)
02208
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.no_loci, and amas.AMAS.MetaAlignment

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:

Here is the caller graph for this function:
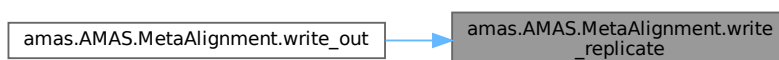
7.6.3.45 write_split()

```
amas.AMAS.MetaAlignment.write_split (
              self,
              item,
              file_format,
              extension )
```
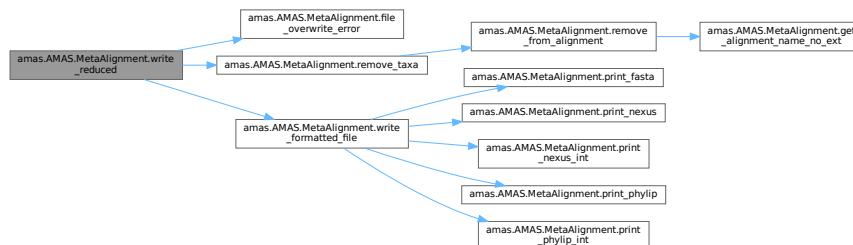
Definition at line 2209 of file AMAS.py.

```
02209      def write_split(self, item, file_format, extension):
02210          # write split alignments from partitions file
02211          # bad practice with the dicts; figure out better solution
02212          partition_name = list(item.keys())[0]
02213          alignment = item[partition_name]
02214
02215          if not alignment:
02216              # If the alignment dict is empty, i.e. no alignment associated with partition name, raise
     error
02217              raise ValueError("Partition '%s' is empty. No sequences to write." % partition_name)
02218
02219          # implementation of option --no-san (don't prepend input superalignment filename to the
     'split' outputs)
02220          if self.no_sup_aln_name:
02221              file_name = partition_name + extension
02222          else:
02223              file_name = str(self.in_files[0].split('.')[0]) + "_" + partition_name + extension
02224
02225          try:
02226              self.file_overwrite_error(file_name)
02227              self.write_formatted_file(file_format, file_name, alignment)
02228              yield file_name
02229          except ValueError as e:
02230              print("There was an issue writing file '%s': %s" % (file_name, str(e)))
02231              remove(file_name)
02232              raise
02233
```
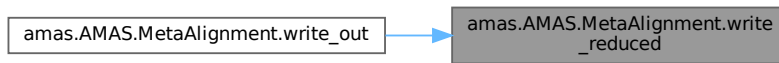
References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.in_files, amas.AMAS.MetaAlignment.no_
amas.AMAS.MetaAlignment.split, and amas.AMAS.MetaAlignment.write_formatted_file().

Referenced by amas.AMAS.MetaAlignment.write_metapartitions(), and amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:

Here is the caller graph for this function:

7.6.3.46 write_summaries()

```
amas.AMAS.MetaAlignment.write_summaries (
            self,
            file_name )
```

Definition at line 1698 of file AMAS.py.

```
01698      def write_summaries(self, file_name):
01699          # write summaries to file
01700
01701          self.file_overwrite_error(file_name)
01702
01703          with open(file_name, "w", encoding="utf-8") as summary_file:
01704              summary_out = self.get_summaries()
01705              header = '\t'.join(summary_out[0])
01706              new_summ = ['\t'.join(summary) for summary in summary_out[1]]
01707              summary_file.write(header + '\n')
01708              summary_file.write('\n'.join(new_summ))
01709              summary_file.write('\n')
01710              print("Wrote summaries to file '" + file_name + "'")
01711
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), and amas.AMAS.MetaAlignment.get_summaries().

Here is the call graph for this function:
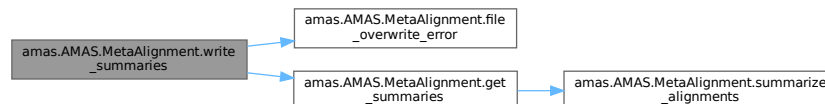
7.6.3.47 write_taxa_summaries()

```
amas.AMAS.MetaAlignment.write_taxa_summaries (
            self )
```

Definition at line 1712 of file AMAS.py.

```
01712      def write_taxa_summaries(self):
01713          # write by-taxon summaries to file
01714          for index, in_file_name in enumerate(self.in_files):
01715              out_file_name = in_file_name + "-seq-summary.txt"
01716              self.file_overwrite_error(out_file_name)
01717              with open(out_file_name, "w", encoding="utf-8") as summary_file:
01718                  summary_out = self.get_taxon_summaries()
01719                  header = '\t'.join(summary_out[0])
01720                  summ = [[str(col) for col in element] for element in summary_out[1][index]]
01721                  new_summ = ['\t'.join(row) for row in summ]
01722                  summary_file.write(header + '\n')
01723                  summary_file.write('\n'.join(new_summ))
01724                  summary_file.write('\n')
01725
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.get_taxon_summaries(), and amas.AMAS.MetaAlignment.in_files.

Here is the call graph for this function:
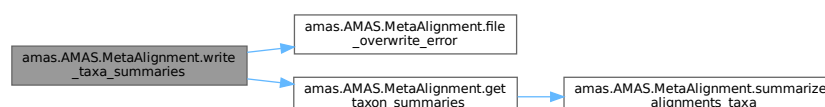
7.6.3.48   write_translated()

```
amas.AMAS.MetaAlignment.write_translated (
            self,
            index,
            alignment,
            file_format,
            extension )
```
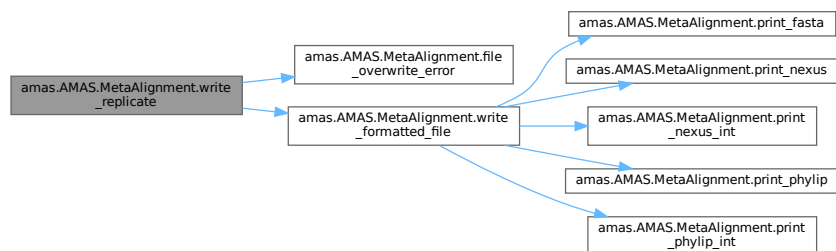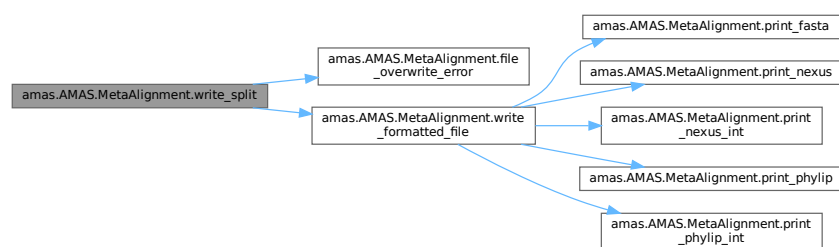
Definition at line 2244 of file AMAS.py.

```
02244      def write_translated(self, index, alignment, file_format, extension):
02245          # write alignments translated into amino acids
02246          prefix = "translated_"
02247          file_name = self.get_alignment_name(index, extension)
02248          out_file_name = prefix + file_name + extension
02249          self.file_overwrite_error(out_file_name)
02250          self.write_formatted_file(file_format, out_file_name, alignment)
02251
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.get_alignment_name(), and amas.AMAS.MetaAlignment.write_formatted_file().

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
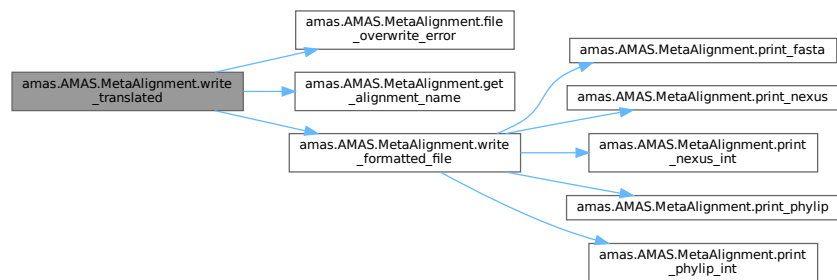
Here is the caller graph for this function:
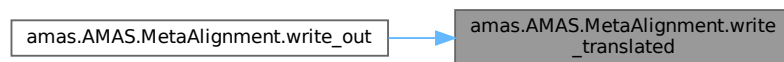
7.6.3.49   write_trimmed()

```
amas.AMAS.MetaAlignment.write_trimmed (
            self,
            index,
            alignment,
```

```
                    file_format,
                    extension )
```

Definition at line 2252 of file AMAS.py.

```
02252      def write_trimmed(self, index, alignment, file_format, extension):
02253          # write trimmed alignments
02254          if self.trim_out:
02255              out_file_name = self.trim_out
02256          else:
02257              prefix = "trimmed_"
02258              file_name = self.get_alignment_name(index, extension)
02259              out_file_name = prefix + file_name
02260          self.file_overwrite_error(out_file_name)
02261          self.write_formatted_file(file_format, out_file_name, alignment)
02262
```

References amas.AMAS.MetaAlignment.file_overwrite_error(), amas.AMAS.MetaAlignment.get_alignment_name(), amas.AMAS.MetaAlignment.trim_out, and amas.AMAS.MetaAlignment.write_formatted_file().

Referenced by amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
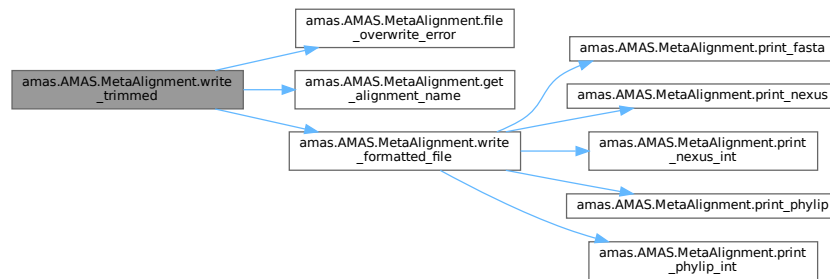
Here is the caller graph for this function:
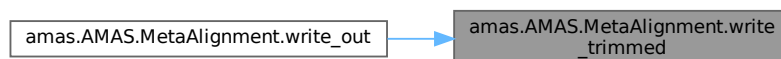
7.6.4 Member Data Documentation

7.6.4.1 alignment_objects

```
amas.AMAS.MetaAlignment.alignment_objects
```

Definition at line 1224 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_name(), amas.AMAS.MetaAlignment.get_alignment_name_no_ext(), amas.AMAS.MetaAlignment.get_concatenated(), amas.AMAS.MetaAlignment.get_parsed_alignments(), amas.AMAS.MetaAlignment, amas.AMAS.MetaAlignment.get_taxon_summaries(), amas.AMAS.MetaAlignment.get_trimmed(), amas.AMAS.MetaAlignment.write_, and amas.AMAS.MetaAlignment.write_out().

7.6.4.2 by_taxon_summary

`amas.AMAS.MetaAlignment.by_taxon_summary`

Definition at line 1178 of file AMAS.py.

7.6.4.3 check_align

`amas.AMAS.MetaAlignment.check_align`

Definition at line 1176 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_parsed_alignments().

7.6.4.4 check_taxa

`amas.AMAS.MetaAlignment.check_taxa`

Definition at line 1213 of file AMAS.py.

7.6.4.5 codes

`amas.AMAS.MetaAlignment.codes`

Definition at line 1424 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.translate_dict().

7.6.4.6 codes_list

`amas.AMAS.MetaAlignment.codes_list`

Definition at line 1228 of file AMAS.py.

7.6.4.7 codons

`amas.AMAS.MetaAlignment.codons`

Definition at line 1183 of file AMAS.py.

7.6.4.8 command

`amas.AMAS.MetaAlignment.command`

Definition at line 1173 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.print_nexus().

7.6.4.9 concat_out

`amas.AMAS.MetaAlignment.concat_out`

Definition at line 1174 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_concat().

7.6.4.10 cores

`amas.AMAS.MetaAlignment.cores`

Definition at line 1177 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_objects(), amas.AMAS.MetaAlignment.get_summaries(), amas.AMAS.MetaAlignment.get_taxon_summaries(), amas.AMAS.MetaAlignment.get_translated(), and amas.AMAS.MetaAlignment.

7.6.4.11 data_type

`amas.AMAS.MetaAlignment.data_type`

Definition at line 1172 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_object(), amas.AMAS.MetaAlignment.get_summaries(), amas.AMAS.MetaAlignment.get_taxon_summaries(), amas.AMAS.MetaAlignment.print_nexus(), amas.AMAS.MetaAlignment.print_n and amas.AMAS.MetaAlignment.write_out().

7.6.4.12 gencode_NCBI_1

`amas.AMAS.MetaAlignment.gencode_NCBI_1`

Definition at line 1251 of file AMAS.py.

7.6.4.13 gencode_NCBI_10

`amas.AMAS.MetaAlignment.gencode_NCBI_10`

Definition at line 1364 of file AMAS.py.

7.6.4.14 gencode_NCBI_11

`amas.AMAS.MetaAlignment.gencode_NCBI_11`

Definition at line 1368 of file AMAS.py.

7.6.4.15 gencode_NCBI_12

`amas.AMAS.MetaAlignment.gencode_NCBI_12`

Definition at line 1371 of file AMAS.py.

7.6.4.16 gencode_NCBI_13

`amas.AMAS.MetaAlignment.gencode_NCBI_13`

Definition at line 1375 of file AMAS.py.

7.6.4.17 gencode_NCBI_14

`amas.AMAS.MetaAlignment.gencode_NCBI_14`

Definition at line 1382 of file AMAS.py.

7.6.4.18 gencode_NCBI_16

`amas.AMAS.MetaAlignment.gencode_NCBI_16`

Definition at line 1390 of file AMAS.py.

7.6.4.19 gencode_NCBI_2

`amas.AMAS.MetaAlignment.gencode_NCBI_2`

Definition at line 1322 of file AMAS.py.

7.6.4.20 gencode_NCBI_21

`amas.AMAS.MetaAlignment.gencode_NCBI_21`

Definition at line 1394 of file AMAS.py.

7.6.4.21 gencode_NCBI_22

`amas.AMAS.MetaAlignment.gencode_NCBI_22`

Definition at line 1402 of file AMAS.py.

7.6.4.22 gencode_NCBI_23

`amas.AMAS.MetaAlignment.gencode_NCBI_23`

Definition at line 1407 of file AMAS.py.

7.6.4.23 gencode_NCBI_24

`amas.AMAS.MetaAlignment.gencode_NCBI_24`

Definition at line 1411 of file AMAS.py.

7.6.4.24 gencode_NCBI_25

`amas.AMAS.MetaAlignment.gencode_NCBI_25`

Definition at line 1417 of file AMAS.py.

7.6.4.25 gencode_NCBI_26

`amas.AMAS.MetaAlignment.gencode_NCBI_26`

Definition at line 1421 of file AMAS.py.

7.6.4.26 gencode_NCBI_3

`amas.AMAS.MetaAlignment.gencode_NCBI_3`

Definition at line 1329 of file AMAS.py.

7.6.4.27 gencode_NCBI_4

`amas.AMAS.MetaAlignment.gencode_NCBI_4`

Definition at line 1341 of file AMAS.py.

7.6.4.28 gencode_NCBI_5

`amas.AMAS.MetaAlignment.gencode_NCBI_5`

Definition at line 1345 of file AMAS.py.

7.6.4.29 gencode_NCBI_6

`amas.AMAS.MetaAlignment.gencode_NCBI_6`

Definition at line 1352 of file AMAS.py.

7.6.4.30 gencode_NCBI_9

`amas.AMAS.MetaAlignment.gencode_NCBI_9`

Definition at line 1357 of file AMAS.py.

7.6.4.31 genetic_code

`amas.AMAS.MetaAlignment.genetic_code`

Definition at line 1217 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.translate_dict(), and amas.AMAS.MetaAlignment.write_out().

7.6.4.32 in_files

`amas.AMAS.MetaAlignment.in_files`

Definition at line 1170 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_objects(), amas.AMAS.MetaAlignment.write_metapartitions(), amas.AMAS.MetaAlignment.write_split(), and amas.AMAS.MetaAlignment.write_taxa_summaries().

7.6.4.33 in_format

`amas.AMAS.MetaAlignment.in_format`

Definition at line 1171 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_alignment_object(), amas.AMAS.Alignment.get_parsed_aln(), and amas.AMAS.MetaAlignment.write_out().

7.6.4.34 no_loci

`amas.AMAS.MetaAlignment.no_loci`

Definition at line 1190 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_out(), and amas.AMAS.MetaAlignment.write_replicate().

7.6.4.35 no_mpan

`amas.AMAS.MetaAlignment.no_mpan`

Definition at line 1180 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_concatenated().

7.6.4.36 no_replicates

`amas.AMAS.MetaAlignment.no_replicates`

Definition at line 1189 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_out().

7.6.4.37 no_sup_aln_name

`amas.AMAS.MetaAlignment.no_sup_aln_name`

Definition at line 1179 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_split().

7.6.4.38 parsed_alignments

`amas.AMAS.MetaAlignment.parsed_alignments`

Definition at line 1225 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_partitioned(), amas.AMAS.MetaAlignment.get_replicate(), amas.AMAS.MetaAlignment.get_summaries(), amas.AMAS.MetaAlignment.get_taxon_summaries(), amas.AMAS.MetaAlignment.get amas.AMAS.MetaAlignment.print_iqtree_nexus_partitions(), amas.AMAS.MetaAlignment.print_nexus_partitions(), amas.AMAS.MetaAlignment.print_raxml_partitions(), amas.AMAS.MetaAlignment.print_unspecified_partitions(), amas.AMAS.MetaAlignment.remove_taxa(), amas.AMAS.MetaAlignment.write_concat(), amas.AMAS.MetaAlignment.write_metapart and amas.AMAS.MetaAlignment.write_out().

7.6.4.39 parsimony_check

`amas.AMAS.MetaAlignment.parsimony_check`

Definition at line 1222 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.trim_dict(), and amas.AMAS.MetaAlignment.write_out().

7.6.4.40 prepend_label

`amas.AMAS.MetaAlignment.prepend_label`

Definition at line 1203 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_concatenated().

7.6.4.41 reading_frame

`amas.AMAS.MetaAlignment.reading_frame`

Definition at line 1216 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.translate_dict(), and amas.AMAS.MetaAlignment.write_out().

7.6.4.42 reduced_file_prefix

`amas.AMAS.MetaAlignment.reduced_file_prefix`

Definition at line 1212 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_reduced().

7.6.4.43 remove_empty

`amas.AMAS.MetaAlignment.remove_empty`

Definition at line 1194 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_partitioned().

7.6.4.44 species_to_remove

`amas.AMAS.MetaAlignment.species_to_remove`

Definition at line 1210 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_reduced().

7.6.4.45 species_to_remove_set

`amas.AMAS.MetaAlignment.species_to_remove_set`

Definition at line 1211 of file AMAS.py.

7.6.4.46 split

`amas.AMAS.MetaAlignment.split`

Definition at line 1193 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_concatenated(), amas.AMAS.MetaAlignment.print_iqtree_nexus_partitions(), amas.AMAS.MetaAlignment.print_nexus_partitions(), amas.AMAS.MetaAlignment.print_raxml_partitions(), amas.AMAS.MetaAlignment.print_unspecified_partitions(), amas.AMAS.MetaAlignment.write_metapartitions(), amas.AMAS.MetaAlignment.write_out(), and amas.AMAS.MetaAlignment.write_split().

7.6.4.47 trim_fraction

`amas.AMAS.MetaAlignment.trim_fraction`

Definition at line 1220 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.trim_dict(), and amas.AMAS.MetaAlignment.write_out().

7.6.4.48 trim_out

`amas.AMAS.MetaAlignment.trim_out`

Definition at line 1221 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.write_trimmed().

7.6.4.49 using_metapartitions

`amas.AMAS.MetaAlignment.using_metapartitions`

Definition at line 1175 of file AMAS.py.

Referenced by amas.AMAS.MetaAlignment.get_concatenated(), and amas.AMAS.MetaAlignment.write_partitions().

The documentation for this class was generated from the following file:

- amas/AMAS.py

7.7 amas.AMAS.ParsedArgs Class Reference

Public Member Functions

- __init__ (self)
- add_common_args (self, parser)
- trim (self)
- summary (self)
- concat (self)
- convert (self)
- replicate (self)
- split (self)
- metapartitions (self)
- translate (self)
- remove (self)
- get_args_dict (self)

Public Attributes

- args

7.7.1 Detailed Description

Definition at line 50 of file AMAS.py.

7.7.2 Constructor & Destructor Documentation

7.7.2.1 __init__()

```
amas.AMAS.ParsedArgs.__init__ (
                self )
```

Definition at line 52 of file AMAS.py.

```
00052    def __init__(self):
00053        parser = argparse.ArgumentParser(
00054            usage='''AMAS <command> [<args>]
00055
00056 The AMAS commands are:
00057   concat          Concatenate input alignments.
00058   convert         Convert to other file format.
00059   replicate       Create replicate data sets for phylogenetic jackknife.
00060   split           Split alignment according to a partitions file.
00061   summary         Write alignment summary.
00062   remove          Remove taxa from alignment.
00063   translate       Translate DNA alignment into protein alignment.
00064   trim            Remove columns from alignment.
00065   metapartitions  Runs `split` and concatenates the output.
00066
00067
00068 Use AMAS <command> -h for help with arguments of the command of interest
00069 '''
00070        )
00071
00072        parser.add_argument(
00073            "command",
00074            help="Subcommand to run"
00075        )
00076
00077        # parse_args defaults to [1:] for args, but you need to
00078        # exclude the rest of the args too, or validation will fail
00079        self.args = parser.parse_args(sys.argv[1:2])
00080        if not hasattr(self, self.args.command):
00081            print('Unrecognized command')
00082            parser.print_help()
00083            exit(1)
00084        # use dispatch pattern to invoke method with same name
00085        getattr(self, self.args.command)()
00086
```

7.7.3 Member Function Documentation

7.7.3.1 add_common_args()

```
amas.AMAS.ParsedArgs.add_common_args (
                self,
                parser )
```

Definition at line 87 of file AMAS.py.

```
00087    def add_common_args(self, parser):
00088        # define required arguments for every command
00089        requiredNamed = parser.add_argument_group('required arguments')
00090        parser.add_argument(
00091            "-e",
00092            "--check-align",
00093            dest = "check_align",
00094            action = "store_true",
00095            default = False,
00096            help = "Check if input sequences are aligned. Default: no check"
00097        )
00098        parser.add_argument(
00099            # parallelization is used for file parsing and calculating summary stats
00100            "-c",
00101            "--cores",
00102            dest = "cores",
00103            default = 1,
00104            help = "Number of cores used. Default: 1"
00105        )
00106
```

```
00107            requiredNamed.add_argument(
00108                "-i",
00109                "--in-files",
00110                nargs = "+",
00111                type = str,
00112                dest = "in_files",
00113                required = True,
00114                help = """Alignment files to be taken as input.
00115                You can specify multiple files using wildcards (e.g. --in-files *fasta)"""
00116            )
00117            requiredNamed.add_argument(
00118                "-f",
00119                "--in-format",
00120                dest = "in_format",
00121                required = True,
00122                choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00123                help = "The format of input alignment"
00124            )
00125            requiredNamed.add_argument(
00126                "-d",
00127                "--data-type",
00128                dest = "data_type",
00129                required = True,
00130                choices = ["aa", "dna"],
00131                help = "Type of data"
00132            )
00133
```

Referenced by amas.AMAS.ParsedArgs.concat(), amas.AMAS.ParsedArgs.convert(), amas.AMAS.ParsedArgs.metapartitions(), amas.AMAS.ParsedArgs.remove(), amas.AMAS.ParsedArgs.replicate(), amas.AMAS.ParsedArgs.split(), amas.AMAS.ParsedArgs.su amas.AMAS.ParsedArgs.translate(), and amas.AMAS.ParsedArgs.trim().

Here is the caller graph for this function:
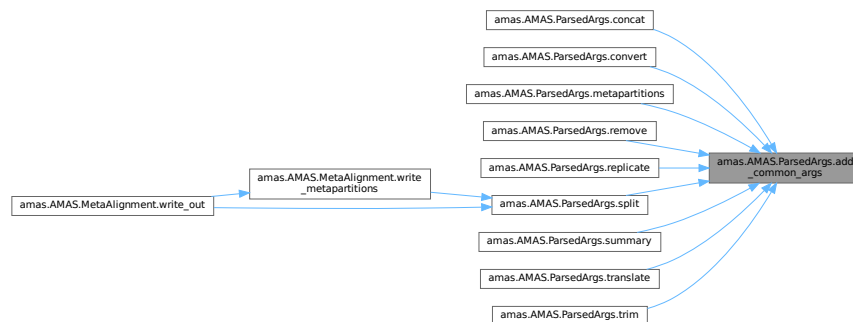
7.7.3.2 concat()

```
amas.AMAS.ParsedArgs.concat (
                self )
```

Definition at line 201 of file AMAS.py.

```
00201        def concat(self):
00202            # concat command
00203            parser = argparse.ArgumentParser(
00204                description="Concatenate input alignments"
00205            )
00206            parser.add_argument(
00207                "-p",
00208                "--concat-part",
00209                dest = "concat_part",
00210                default = "partitions.txt",
00211                help = "File name for the concatenated alignment partitions. Default: 'partitions.txt'"
00212            )
00213            parser.add_argument(
00214                "-t",
00215                "--concat-out",
```

```
00216                    dest = "concat_out",
00217                    default = "concatenated.out",
00218                    help = "File name for the concatenated alignment. Default: 'concatenated.out'"
00219               )
00220          parser.add_argument(
00221                    "-u",
00222                    "--out-format",
00223                    dest = "out_format",
00224                    choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00225                    default = "fasta",
00226                    help = "File format for the output alignment. Default: fasta"
00227               )
00228          parser.add_argument(
00229                    "-y",
00230                    "--part-format",
00231                    dest = "part_format",
00232                    choices = ["nexus", "iqtree-nexus", "raxml", "unspecified"],
00233                    default = "unspecified",
00234                    help = "Format of the partitions file. Default: 'unspecified'"
00235               )
00236          parser.add_argument(
00237                    "-n",
00238                    "--codons",
00239                    dest = "codons",
00240                    choices = ["none", "12", "123"],
00241                    default = "none",
00242                    help = "Use codon partitioning for 1st and 2nd or all three positions. Default: Don't use"
00243               )
00244          # add shared arguments
00245          self.add_common_args(parser)
00246          args = parser.parse_args(sys.argv[2:])
00247          return args
00248
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:
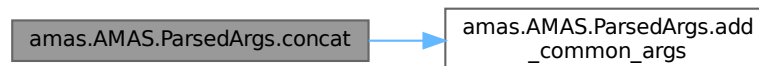
7.7.3.3 convert()

```
amas.AMAS.ParsedArgs.convert (
              self )
```

Definition at line 249 of file AMAS.py.

```
00249     def convert(self):
00250          # convert command
00251          parser = argparse.ArgumentParser(
00252               description="Convert to other file format",
00253          )
00254          parser.add_argument(
00255                    "-u",
00256                    "--out-format",
00257                    dest = "out_format",
00258                    choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00259                    default = "fasta",
00260                    help = "File format for the output alignment. Default: fasta"
00261               )
00262          # add shared arguments
00263          self.add_common_args(parser)
00264          args = parser.parse_args(sys.argv[2:])
00265          return args
00266
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:
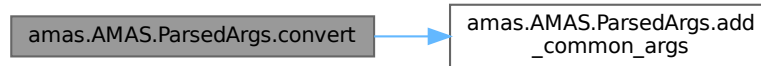
7.7.3.4 get_args_dict()

```
amas.AMAS.ParsedArgs.get_args_dict (
            self )
```

Definition at line 508 of file AMAS.py.

```
00508    def get_args_dict(self):
00509        # store arguments in a dictionary
00510        command = self.args.__dict__
00511        arguments = getattr(self, self.args.command)().__dict__
00512        argument_dictionary = command.copy()
00513        argument_dictionary.update(arguments)
00514
00515        return argument_dictionary
00516
00517
```

References amas.AMAS.ParsedArgs.args.

7.7.3.5 metapartitions()

```
amas.AMAS.ParsedArgs.metapartitions (
            self )
```

Definition at line 334 of file AMAS.py.

```
00334    def metapartitions(self):
00335        # metapartitions command
00336        parser = argparse.ArgumentParser(
00337            formatter_class=argparse.RawTextHelpFormatter,
00338            description="'Split alignment according to a partition file, then concatenate the
    output."'
00339            "'\n\nuse case:\n"'
00340            "'    Some utilities cannot parse partition definitions containing strides (\\) and/or
    discontinuous ranges.\n"'
00341            "'    In such case, running `split` + `concat` in separate passes can convert a
    corresponding (super)alignment it into an\n"'
00342            "'    equivalent compatible form with contiguous (meta)partitions; this may also require
    renaming metapartiton alignments\n"'
00343            "'    and partition file entries in order to remove tags applied by each respective
    operation.\n\n"'
00344            "'    `metapartitions` combines these steps into one command, with the options `--prepend`
    and `--no-mpan`\n"'
00345            "'    providing additional control over the collated (meta)partition names (see their
    respective help entries).\n\n"'
00346            "'    Note: in this mode, the format of the input (super)alignment file determines that of
    all outputs (-u|--out-format is disabled)!\n\n"'
00347        )
00348        parser.add_argument(
00349            "-p",
00350            "--concat-part",
00351            dest = "concat_part",
00352            default = "metapartitions.txt",
00353            help = "Partition file(name) for the final concatenated alignment of metapartitions.
    Default: 'metapartitions.txt'"
```

```
00354            )
00355        parser.add_argument(
00356            "-t",
00357            "--concat-out",
00358            dest = "concat_out",
00359            default = "concatenated-meta.out",
00360            help = "File name for the concatenated alignment of metapartitions. Default:
    'concatenated-meta.out'"
00361            )
00362        parser.add_argument(
00363            "-y",
00364            "--part-format",
00365            dest = "part_format",
00366            choices = ["nexus", "iqtree-nexus", "raxml", "unspecified"],
00367            default = "unspecified",
00368            help = "Partitions file format for the final concatenated alignment of metapartitions.
    Default: 'unspecified'"
00369            )
00370        parser.add_argument(
00371            "-l",
00372            "--split-by",
00373            dest = "split_by",
00374            help = "Partition file(name) to be used for splitting the initial concatenated
    alignment.",
00375            required = True
00376            )
00377        parser.add_argument(
00378            "-j",
00379            "--remove-empty",
00380            dest = "remove_empty",
00381            action = "store_true",
00382            default = False,
00383            help = "Remove taxa with sequences composed of only undetermined characters? Default:
    Don't remove"
00384            )
00385        parser.add_argument(
00386            "--no-san",
00387            dest = "no_sup_aln_name",
00388            action = "store_true",
00389            default = False,
00390            help = "'Don't prepend the input (super)alignment filename to the
    (meta)partition-alignment filenames output by `split`'"
00391            )
00392        parser.add_argument(
00393            "--prepend",
00394            dest = "prepend_label",
00395            default = None,
00396            help = "'Prepend <string> to the partition counter in partition file, e.g.'"
00397            "'\n          --prepend <string>:  <string>_p001_metapartition_alignment_name = 1-1200
    ...'"
00398            "'\n          Default (None):       p001_metapartition_alignment_name = 1-1200
    ...'"
00399            "'\n--no-mpan + --prepend <string>:  <string>_p001 = 1-1200 ...'"
00400            )
00401        parser.add_argument(
00402            "--no-mpan",
00403            dest = "no_mpan",
00404            action = "store_true",
00405            default = False,
00406            help = "'Omits (meta)partition alignment names when printing partition file, e.g.'"
00407            "'\n                --no-mpan:       p001 = 1-1200 ...'"
00408            "'\n          Default (False):      p001_metapartition_alignment_name = 1-1200
    ...'"
00409            "'\n--prepend <string> + --no-mpan: <string>_p001 = 1-1200 ...'"
00410            )
00411        # add shared arguments
00412        self.add_common_args(parser)
00413        args = parser.parse_args(sys.argv[2:])
00414        return args
00415
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:

7.7.3.6 remove()

```
amas.AMAS.ParsedArgs.remove (
                self )
```

Definition at line 474 of file AMAS.py.
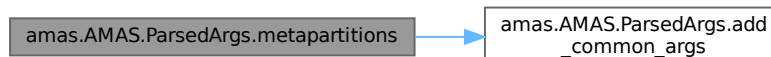
```
00474     def remove(self):
00475         # remove taxa command
00476         parser = argparse.ArgumentParser(
00477             description="Remove taxa from alignment",
00478         )
00479         parser.add_argument(
00480             "-x",
00481             "--taxa-to-remove",
00482             nargs = "+",
00483             type = str,
00484             dest = "taxa_to_remove",
00485             help = "Taxon/sequence names to be removed.",
00486             required = True
00487         )
00488         parser.add_argument(
00489             "-u",
00490             "--out-format",
00491             dest = "out_format",
00492             choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00493             default = "fasta",
00494             help = "File format for the output alignment. Default: fasta"
00495         )
00496         parser.add_argument(
00497             "-g",
00498             "--out-prefix",
00499             dest = "out_prefix",
00500             default = "reduced_",
00501             help = "File name prefix for the concatenated alignment. Default: 'reduced_'"
00502         )
00503         # add shared arguments
00504         self.add_common_args(parser)
00505         args = parser.parse_args(sys.argv[2:])
00506         return args
00507
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:
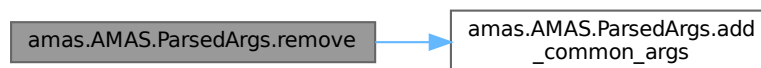
7.7.3.7 replicate()

```
amas.AMAS.ParsedArgs.replicate (
                self )
```

Definition at line 267 of file AMAS.py.

```
00267     def replicate(self):
00268         # replicate command
00269         parser = argparse.ArgumentParser(
00270             description="Create replicate datasets for phylogenetic jackknife",
00271         )
00272         parser.add_argument(
00273             "-r",
00274             "--rep-aln",
00275             nargs = 2,
00276             type = int,
```

```
00277                dest = "replicate_args",
00278                help = "Create replicate data sets for phylogenetic jackknife [replicates, no alignments
     for each replicate]",
00279                required = True
00280            )
00281            parser.add_argument(
00282                "-u",
00283                "--out-format",
00284                dest = "out_format",
00285                choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00286                default = "fasta",
00287                help = "File format for the output alignment. Default: fasta"
00288            )
00289            # add shared arguments
00290            self.add_common_args(parser)
00291            args = parser.parse_args(sys.argv[2:])
00292            return args
00293
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:
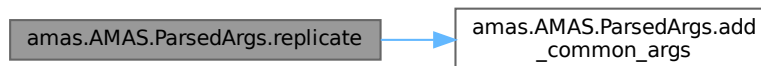
7.7.3.8 split()

```
amas.AMAS.ParsedArgs.split (
                self )
```

Definition at line 294 of file AMAS.py.

```
00294    def split(self):
00295        # split command
00296        parser = argparse.ArgumentParser(
00297            description="Split alignment according to a partitions file",
00298        )
00299        parser.add_argument(
00300            "-l",
00301            "--split-by",
00302            dest = "split_by",
00303            help = "File name for partitions to be used for alignment splitting.",
00304            required = True
00305        )
00306        parser.add_argument(
00307            "-j",
00308            "--remove-empty",
00309            dest = "remove_empty",
00310            action = "store_true",
00311            default = False,
00312            help = "Remove taxa with sequences composed of only undetermined characters? Default:
     Don't remove"
00313        )
00314        parser.add_argument(
00315            "-u",
00316            "--out-format",
00317            dest = "out_format",
00318            choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00319            default = "fasta",
00320            help = "File format for the output alignment. Default: fasta"
00321        )
00322        parser.add_argument(
00323            "--no-san",
00324            dest = "no_sup_aln_name",
00325            action = "store_true",
00326            default = False,
```

```
00327            help = "'Don't prepend the input (super)alignment filename to the partition-alignment
      filenames output by `split`'"
00328        )
00329        # add shared arguments
00330        self.add_common_args(parser)
00331        args = parser.parse_args(sys.argv[2:])
00332        return args
00333
```

References amas.AMAS.ParsedArgs.add_common_args().

Referenced by amas.AMAS.MetaAlignment.write_metapartitions(), and amas.AMAS.MetaAlignment.write_out().

Here is the call graph for this function:
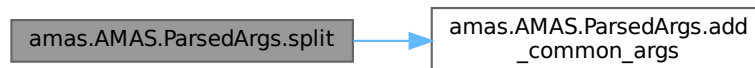
Here is the caller graph for this function:
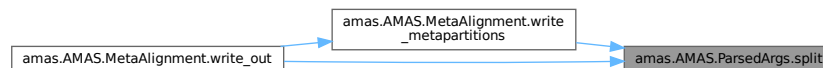
7.7.3.9 summary()

```
amas.AMAS.ParsedArgs.summary (
            self )
```

Definition at line 176 of file AMAS.py.
```
00176    def summary(self):
00177        # summary command
00178        parser = argparse.ArgumentParser(
00179            description="Write alignment summary",
00180        )
00181        parser.add_argument(
00182            "-o",
00183            "--summary-out",
00184            dest = "summary_out",
00185            default = "summary.txt",
00186            help = "File name for the alignment summary. Default: 'summary.txt'"
00187        )
00188        parser.add_argument(
00189            "-s",
00190            "--by-taxon",
00191            dest = "by_taxon_summary",
00192            action = "store_true",
00193            default = False,
00194            help = "In addition to alignment summary, write by sequence/taxon summaries. Default:
      Don't write"
00195        )
00196        # add shared arguments
00197        self.add_common_args(parser)
00198        args = parser.parse_args(sys.argv[2:])
00199        return args
00200
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:

7.7.3.10 translate()

```
amas.AMAS.ParsedArgs.translate (
              self )
```

Definition at line 416 of file AMAS.py.

```
00416    def translate(self):
00417        # translate command
00418        parser = argparse.ArgumentParser(
00419            formatter_class=argparse.RawTextHelpFormatter,
00420            description="Translate a protein-coding DNA alignment into amino acids"
00421        )
00422        parser.add_argument(
00423            "-b",
00424            "--code",
00425            type = int,
00426            dest = "genetic_code",
00427            choices = [1, 2, 3, 4, 5, 6, 9, 10, 11, 12, 13, 14, 16, 21, 22, 23, 24, 25, 26],
00428            default = 1,
00429            help = "'\nNCBI genetic code to use (Default: 1):"'
00430 "'
00431   1. The Standard Code
00432   2. The Vertebrate Mitochondrial Code
00433   3. The Yeast Mitochondrial Code
00434   4. The Mold, Protozoan, and Coelenterate Mitochondrial Code and the Mycoplasma/Spiroplasma Code
00435   5. The Invertebrate Mitochondrial Code
00436   6. The Ciliate, Dasycladacean and Hexamita Nuclear Code
00437   9. The Echinoderm and Flatworm Mitochondrial Code
00438  10. The Euplotid Nuclear Code
00439  11. The Bacterial, Archaeal and Plant Plastid Code
00440  12. The Alternative Yeast Nuclear Code
00441  13. The Ascidian Mitochondrial Code
00442  14. The Alternative Flatworm Mitochondrial Code
00443  16. Chlorophycean Mitochondrial Code
00444  21. Trematode Mitochondrial Code
00445  22. Scenedesmus obliquus Mitochondrial Code
00446  23. Thraustochytrium Mitochondrial Code
00447  24. Pterobranchia Mitochondrial Code
00448  25. Candidate Division SR1 and Gracilibacteria Code
00449  26. Pachysolen tannophilus Nuclear Code\n
00450 "'
00451        )
00452        parser.add_argument(
00453            "-k",
00454            "--reading-frame",
00455            type = int,
00456            dest = "reading_frame",
00457            choices = [1, 2, 3],
00458            default = 1,
00459            help = "Number specifying reading frame; i.e. '2' means codons start at the second
       character of the alignment. Default: 1",
00460        )
00461        parser.add_argument(
00462            "-u",
00463            "--out-format",
00464            dest = "out_format",
00465            choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00466            default = "fasta",
00467            help = "File format for the output alignment. Default: fasta"
00468        )
00469        # add shared arguments
```

```
00470          self.add_common_args(parser)
00471          args = parser.parse_args(sys.argv[2:])
00472          return args
00473
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:
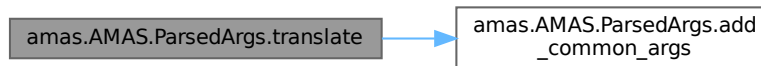
7.7.3.11 trim()

```
amas.AMAS.ParsedArgs.trim (
              self )
```
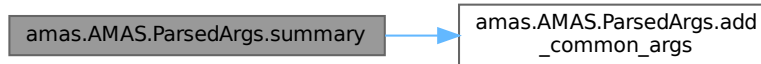
Definition at line 134 of file AMAS.py.

```
00134      def trim(self):
00135          # trim command
00136          parser = argparse.ArgumentParser(
00137              formatter_class=argparse.RawDescriptionHelpFormatter,
00138              description="'Trim alignment by occupancy. Optionally removes sites that are not parsimony
     informative."'
00139              "'\nCAUTION: when running on amino acids stop codons marked with * will be treated as
     missing data!"'
00140          )
00141          parser.add_argument(
00142              "-u",
00143              "--out-format",
00144              dest = "out_format",
00145              choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00146              default = "fasta",
00147              help = "File format for the output alignment. Default: fasta"
00148          )
00149          parser.add_argument(
00150              "-o",
00151              "--trim-out",
00152              dest = "trim_out",
00153              help = "File name for the trimmed alignment when providing a single file as input."
00154          )
00155          parser.add_argument(
00156              "-t",
00157              "--trim-fraction",
00158              type = proportion,
00159              dest = "trim_fraction",
00160              default = 0.6,
00161              help = "Columns in the alignments with occupancy lower than this value will be removed.
     Default: 0.6"
00162          )
00163          parser.add_argument(
00164              "-p",
00165              "--retain-only-parsimony-sites",
00166              dest = "parsimony_check",
00167              action = "store_true",
00168              default = False,
00169              help = "Only write parsimony informative columns in trimmed alignment Default: write all
     columns"
00170          )
00171          # add shared arguments
00172          self.add_common_args(parser)
00173          args = parser.parse_args(sys.argv[2:])
00174          return args
00175
```

References amas.AMAS.ParsedArgs.add_common_args().

Here is the call graph for this function:
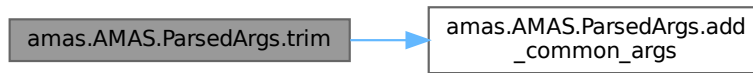
7.7.4 Member Data Documentation

7.7.4.1 args

`amas.AMAS.ParsedArgs.args`

Definition at line 79 of file AMAS.py.

Referenced by amas.AMAS.ParsedArgs.get_args_dict().

The documentation for this class was generated from the following file:

- amas/AMAS.py

Chapter 8

File Documentation

8.1 amas/__init__.py File Reference

Namespaces

- namespace amas

Variables

- str amas.__author__ = 'Marek Borowiec'
- str amas.__email__ = 'petiolus@gmail.com'
- str amas.__version__ = '1.02'
- amas.__all__ = dir()

8.2 __init__.py

Go to the documentation of this file.
```
00001 # -*- coding: utf-8 -*-
00002
00003 __author__ = 'Marek Borowiec'
00004 __email__ = 'petiolus@gmail.com'
00005 __version__ = '1.02'
00006 __all__ = dir()
```

8.3 amas/AMAS.py File Reference

Classes

- class amas.AMAS.ParsedArgs
- class amas.AMAS.FileHandler
- class amas.AMAS.FileParser
- class amas.AMAS.Alignment
- class amas.AMAS.AminoAcidAlignment
- class amas.AMAS.DNAAlignment
- class amas.AMAS.MetaAlignment

Namespaces

- namespace amas
- namespace amas.AMAS

Functions

- amas.AMAS.proportion (x)
- amas.AMAS.main ()
- amas.AMAS.run ()

8.4 AMAS.py

Go to the documentation of this file.
```
00001 #! /usr/bin/env python3
00002 # -*- coding: utf-8 -*-
00003 # vim:fileencoding=utf-8
00004
00005 #   Program to calculate various statistics on a multiple sequence alignment
00006 #   and allow efficient manipulation of phylogenomic data sets
00007
00008 #   Copyright (C) 2015 Marek Borowiec
00009
00010 #   This program is free software: you can redistribute it and/or modify
00011 #   it under the terms of the GNU General Public License as published by
00012 #   the Free Software Foundation, either version 3 of the License, or
00013 #   (at your option) any later version.
00014
00015 #   This program is distributed in the hope that it will be useful,
00016 #   but WITHOUT ANY WARRANTY; without even the implied warranty of
00017 #   MERCHANTABILITY or FITNESS FOR A PARTICULAR PURPOSE.  See the
00018 #   GNU General Public License for more details.
00019
00020 #   You should have received a copy of the GNU General Public License
00021 #   along with this program.  If not, see <http://www.gnu.org/licenses/>.
00022
00023 """
00024 This stand-alone program allows manipulations of multiple sequence
00025 alignments. It supports sequential FASTA, PHYLIP, NEXUS, and interleaved PHYLIP
00026 and NEXUS formats for DNA and aino acid sequences. It can print summary statistics,
00027 convert among formats, and concatenate alignments.
00028
00029 Current statistics include the number of taxa, alignment length, total number
00030 of matrix cells, overall number of undetermined characters, percent of missing
00031 data, AT and GC contents (for DNA alignments), number and proportion of
00032 variable sites, number and proportion of parsimony informative sites,
00033 and counts of all characters present in the relevant (nucleotide or amino acid) alphabet.
00034 """
00035
00036
00037 import argparse, multiprocessing as mp, re, sys
00038 from random import sample
00039 from os import path, remove
00040 from collections import defaultdict, Counter
00041 from itertools import compress
00042
00043 def proportion(x):
00044     # needed to prevent input of invalid floats in trim mode
00045     x = float(x)
00046     if x < 0.0 or x > 1.0:
00047         raise argparse.ArgumentTypeError("%r not in range [0.0, 1.0]" % (x,))
00048     return x
00049
00050 class ParsedArgs:
00051
00052     def __init__(self):
00053         parser = argparse.ArgumentParser(
00054             usage="'AMAS <command> [<args>]
00055
00056 The AMAS commands are:
00057   concat            Concatenate input alignments.
00058   convert           Convert to other file format.
00059   replicate         Create replicate data sets for phylogenetic jackknife.
00060   split             Split alignment according to a partitions file.
00061   summary           Write alignment summary.
```

```
00062   remove            Remove taxa from alignment.
00063   translate         Translate DNA alignment into protein alignment.
00064   trim              Remove columns from alignment.
00065   metapartitions    Runs `split` and concatenates the output.
00066
00067
00068 Use AMAS <command> -h for help with arguments of the command of interest
00069 ”’
00070           )
00071
00072           parser.add_argument(
00073               "command",
00074               help="Subcommand to run"
00075           )
00076
00077           # parse_args defaults to [1:] for args, but you need to
00078           # exclude the rest of the args too, or validation will fail
00079           self.args = parser.parse_args(sys.argv[1:2])
00080           if not hasattr(self, self.args.command):
00081               print(‘Unrecognized command’)
00082               parser.print_help()
00083               exit(1)
00084           # use dispatch pattern to invoke method with same name
00085           getattr(self, self.args.command)()
00086
00087       def add_common_args(self, parser):
00088           # define required arguments for every command
00089           requiredNamed = parser.add_argument_group(‘required arguments’)
00090           parser.add_argument(
00091               "-e",
00092               "--check-align",
00093               dest = "check_align",
00094               action = "store_true",
00095               default = False,
00096               help = "Check if input sequences are aligned. Default: no check"
00097           )
00098           parser.add_argument(
00099               # parallelization is used for file parsing and calculating summary stats
00100               "-c",
00101               "--cores",
00102               dest = "cores",
00103               default = 1,
00104               help = "Number of cores used. Default: 1"
00105           )
00106
00107           requiredNamed.add_argument(
00108               "-i",
00109               "--in-files",
00110               nargs = "+",
00111               type = str,
00112               dest = "in_files",
00113               required = True,
00114               help = """Alignment files to be taken as input.
00115               You can specify multiple files using wildcards (e.g. --in-files *fasta)"""
00116           )
00117           requiredNamed.add_argument(
00118               "-f",
00119               "--in-format",
00120               dest = "in_format",
00121               required = True,
00122               choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00123               help = "The format of input alignment"
00124           )
00125           requiredNamed.add_argument(
00126               "-d",
00127               "--data-type",
00128               dest = "data_type",
00129               required = True,
00130               choices = ["aa", "dna"],
00131               help = "Type of data"
00132           )
00133
00134       def trim(self):
00135           # trim command
00136           parser = argparse.ArgumentParser(
00137               formatter_class=argparse.RawDescriptionHelpFormatter,
00138               description=”’Trim alignment by occupancy. Optionally removes sites that are not parsimony
    informative.”’
00139               ”’\nCAUTION: when running on amino acids stop codons marked with * will be treated as
    missing data!”’
00140           )
00141           parser.add_argument(
00142               "-u",
00143               "--out-format",
00144               dest = "out_format",
00145               choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00146               default = "fasta",
```

```
00147                  help = "File format for the output alignment. Default: fasta"
00148              )
00149          parser.add_argument(
00150              "-o",
00151              "--trim-out",
00152              dest = "trim_out",
00153              help = "File name for the trimmed alignment when providing a single file as input."
00154          )
00155          parser.add_argument(
00156              "-t",
00157              "--trim-fraction",
00158              type = proportion,
00159              dest = "trim_fraction",
00160              default = 0.6,
00161              help = "Columns in the alignments with occupancy lower than this value will be removed.
       Default: 0.6"
00162          )
00163          parser.add_argument(
00164              "-p",
00165              "--retain-only-parsimony-sites",
00166              dest = "parsimony_check",
00167              action = "store_true",
00168              default = False,
00169              help = "Only write parsimony informative columns in trimmed alignment Default: write all
       columns"
00170          )
00171          # add shared arguments
00172          self.add_common_args(parser)
00173          args = parser.parse_args(sys.argv[2:])
00174          return args
00175
00176      def summary(self):
00177          # summary command
00178          parser = argparse.ArgumentParser(
00179              description="Write alignment summary",
00180          )
00181          parser.add_argument(
00182              "-o",
00183              "--summary-out",
00184              dest = "summary_out",
00185              default = "summary.txt",
00186              help = "File name for the alignment summary. Default: 'summary.txt'"
00187          )
00188          parser.add_argument(
00189              "-s",
00190              "--by-taxon",
00191              dest = "by_taxon_summary",
00192              action = "store_true",
00193              default = False,
00194              help = "In addition to alignment summary, write by sequence/taxon summaries. Default:
       Don't write"
00195          )
00196          # add shared arguments
00197          self.add_common_args(parser)
00198          args = parser.parse_args(sys.argv[2:])
00199          return args
00200
00201      def concat(self):
00202          # concat command
00203          parser = argparse.ArgumentParser(
00204              description="Concatenate input alignments"
00205          )
00206          parser.add_argument(
00207              "-p",
00208              "--concat-part",
00209              dest = "concat_part",
00210              default = "partitions.txt",
00211              help = "File name for the concatenated alignment partitions. Default: 'partitions.txt'"
00212          )
00213          parser.add_argument(
00214              "-t",
00215              "--concat-out",
00216              dest = "concat_out",
00217              default = "concatenated.out",
00218              help = "File name for the concatenated alignment. Default: 'concatenated.out'"
00219          )
00220          parser.add_argument(
00221              "-u",
00222              "--out-format",
00223              dest = "out_format",
00224              choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00225              default = "fasta",
00226              help = "File format for the output alignment. Default: fasta"
00227          )
00228          parser.add_argument(
00229              "-y",
00230              "--part-format",
```

```
00231                    dest = "part_format",
00232                    choices = ["nexus", "iqtree-nexus", "raxml", "unspecified"],
00233                    default = "unspecified",
00234                    help = "Format of the partitions file. Default: 'unspecified'"
00235                )
00236            parser.add_argument(
00237                "-n",
00238                "--codons",
00239                dest = "codons",
00240                choices = ["none", "12", "123"],
00241                default = "none",
00242                help = "Use codon partitioning for 1st and 2nd or all three positions. Default: Don't use"
00243            )
00244            # add shared arguments
00245            self.add_common_args(parser)
00246            args = parser.parse_args(sys.argv[2:])
00247            return args
00248
00249    def convert(self):
00250            # convert command
00251            parser = argparse.ArgumentParser(
00252                description="Convert to other file format",
00253            )
00254            parser.add_argument(
00255                "-u",
00256                "--out-format",
00257                dest = "out_format",
00258                choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00259                default = "fasta",
00260                help = "File format for the output alignment. Default: fasta"
00261            )
00262            # add shared arguments
00263            self.add_common_args(parser)
00264            args = parser.parse_args(sys.argv[2:])
00265            return args
00266
00267    def replicate(self):
00268            # replicate command
00269            parser = argparse.ArgumentParser(
00270                description="Create replicate datasets for phylogenetic jackknife",
00271            )
00272            parser.add_argument(
00273                "-r",
00274                "--rep-aln",
00275                nargs = 2,
00276                type = int,
00277                dest = "replicate_args",
00278                help = "Create replicate data sets for phylogenetic jackknife [replicates, no alignments
    for each replicate]",
00279                required = True
00280            )
00281            parser.add_argument(
00282                "-u",
00283                "--out-format",
00284                dest = "out_format",
00285                choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00286                default = "fasta",
00287                help = "File format for the output alignment. Default: fasta"
00288            )
00289            # add shared arguments
00290            self.add_common_args(parser)
00291            args = parser.parse_args(sys.argv[2:])
00292            return args
00293
00294    def split(self):
00295            # split command
00296            parser = argparse.ArgumentParser(
00297                description="Split alignment according to a partitions file",
00298            )
00299            parser.add_argument(
00300                "-l",
00301                "--split-by",
00302                dest = "split_by",
00303                help = "File name for partitions to be used for alignment splitting.",
00304                required = True
00305            )
00306            parser.add_argument(
00307                "-j",
00308                "--remove-empty",
00309                dest = "remove_empty",
00310                action = "store_true",
00311                default = False,
00312                help = "Remove taxa with sequences composed of only undetermined characters? Default:
    Don't remove"
00313            )
00314            parser.add_argument(
00315                "-u",
```

```
00316                "--out-format",
00317                dest = "out_format",
00318                choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00319                default = "fasta",
00320                help = "File format for the output alignment. Default: fasta"
00321            )
00322        parser.add_argument(
00323                "--no-san",
00324                dest = "no_sup_aln_name",
00325                action = "store_true",
00326                default = False,
00327                help = "'Don't prepend the input (super)alignment filename to the partition-alignment
       filenames output by `split`'"
00328            )
00329        # add shared arguments
00330        self.add_common_args(parser)
00331        args = parser.parse_args(sys.argv[2:])
00332        return args
00333
00334    def metapartitions(self):
00335        # metapartitions command
00336        parser = argparse.ArgumentParser(
00337            formatter_class=argparse.RawTextHelpFormatter,
00338            description="'Split alignment according to a partition file, then concatenate the
       output.'"
00339            "'\n\nuse case:\n'"
00340            "'    Some utilities cannot parse partition definitions containing strides (\\) and/or
       discontinuous ranges.\n'"
00341            "'    In such case, running `split` + `concat` in separate passes can convert a
       corresponding (super)alignment it into an\n'"
00342            "'    equivalent compatible form with contiguous (meta)partitions; this may also require
       renaming metapartiton alignments\n'"
00343            "'    and partition file entries in order to remove tags applied by each respective
       operation.\n\n'"
00344            "'    `metapartitions` combines these steps into one command, with the options `--prepend`
       and `--no-mpan`\n'"
00345            "'    providing additional control over the collated (meta)partition names (see their
       respective help entries).\n\n'"
00346            "'    Note: in this mode, the format of the input (super)alignment file determines that of
       all outputs (-u|--out-format is disabled)!\n\n'"
00347            )
00348        parser.add_argument(
00349            "-p",
00350            "--concat-part",
00351            dest = "concat_part",
00352            default = "metapartitions.txt",
00353            help = "Partition file(name) for the final concatenated alignment of metapartitions.
       Default: 'metapartitions.txt'"
00354            )
00355        parser.add_argument(
00356            "-t",
00357            "--concat-out",
00358            dest = "concat_out",
00359            default = "concatenated-meta.out",
00360            help = "File name for the concatenated alignment of metapartitions. Default:
       'concatenated-meta.out'"
00361            )
00362        parser.add_argument(
00363            "-y",
00364            "--part-format",
00365            dest = "part_format",
00366            choices = ["nexus", "iqtree-nexus", "raxml", "unspecified"],
00367            default = "unspecified",
00368            help = "Partitions file format for the final concatenated alignment of metapartitions.
       Default: 'unspecified'"
00369            )
00370        parser.add_argument(
00371            "-l",
00372            "--split-by",
00373            dest = "split_by",
00374            help = "Partition file(name) to be used for splitting the initial concatenated
       alignment.",
00375            required = True
00376            )
00377        parser.add_argument(
00378            "-j",
00379            "--remove-empty",
00380            dest = "remove_empty",
00381            action = "store_true",
00382            default = False,
00383            help = "Remove taxa with sequences composed of only undetermined characters? Default:
       Don't remove"
00384            )
00385        parser.add_argument(
00386            "--no-san",
00387            dest = "no_sup_aln_name",
00388            action = "store_true",
```

```
00389                 default = False,
00390                 help = "'Don't prepend the input (super)alignment filename to the
      (meta)partition-alignment filenames output by `split`"'
00391             )
00392         parser.add_argument(
00393             "--prepend",
00394             dest = "prepend_label",
00395             default = None,
00396             help = "'Prepend <string> to the partition counter in partition file, e.g."'
00397             "'\n             --prepend <string>:  <string>_p001_metapartition_alignment_name = 1-1200
      ..."'
00398             "'\n                  Default (None):          p001_metapartition_alignment_name = 1-1200
      ..."'
00399             "'\n--no-mpan + --prepend <string>:  <string>_p001 = 1-1200 ..."'
00400             )
00401         parser.add_argument(
00402             "--no-mpan",
00403             dest = "no_mpan",
00404             action = "store_true",
00405             default = False,
00406             help = "'Omits (meta)partition alignment names when printing partition file, e.g."'
00407             "'\n                     --no-mpan:        p001 = 1-1200 ..."'
00408             "'\n                  Default (False):        p001_metapartition_alignment_name = 1-1200
      ..."'
00409             "'\n--prepend <string> + --no-mpan: <string>_p001 = 1-1200 ..."'
00410             )
00411         # add shared arguments
00412         self.add_common_args(parser)
00413         args = parser.parse_args(sys.argv[2:])
00414         return args
00415
00416     def translate(self):
00417         # translate command
00418         parser = argparse.ArgumentParser(
00419             formatter_class=argparse.RawTextHelpFormatter,
00420             description="Translate a protein-coding DNA alignment into amino acids"
00421         )
00422         parser.add_argument(
00423             "-b",
00424             "--code",
00425             type = int,
00426             dest = "genetic_code",
00427             choices = [1, 2, 3, 4, 5, 6, 9, 10, 11, 12, 13, 14, 16, 21, 22, 23, 24, 25, 26],
00428             default = 1,
00429             help = "'\nNCBI genetic code to use (Default: 1):"'
00430 "'
00431     1. The Standard Code
00432     2. The Vertebrate Mitochondrial Code
00433     3. The Yeast Mitochondrial Code
00434     4. The Mold, Protozoan, and Coelenterate Mitochondrial Code and the Mycoplasma/Spiroplasma Code
00435     5. The Invertebrate Mitochondrial Code
00436     6. The Ciliate, Dasycladacean and Hexamita Nuclear Code
00437     9. The Echinoderm and Flatworm Mitochondrial Code
00438    10. The Euplotid Nuclear Code
00439    11. The Bacterial, Archaeal and Plant Plastid Code
00440    12. The Alternative Yeast Nuclear Code
00441    13. The Ascidian Mitochondrial Code
00442    14. The Alternative Flatworm Mitochondrial Code
00443    16. Chlorophycean Mitochondrial Code
00444    21. Trematode Mitochondrial Code
00445    22. Scenedesmus obliquus Mitochondrial Code
00446    23. Thraustochytrium Mitochondrial Code
00447    24. Pterobranchia Mitochondrial Code
00448    25. Candidate Division SR1 and Gracilibacteria Code
00449    26. Pachysolen tannophilus Nuclear Code\n
00450 "'
00451         )
00452         parser.add_argument(
00453             "-k",
00454             "--reading-frame",
00455             type = int,
00456             dest = "reading_frame",
00457             choices = [1, 2, 3],
00458             default = 1,
00459             help = "Number specifying reading frame; i.e. '2' means codons start at the second
      character of the alignment. Default: 1",
00460             )
00461         parser.add_argument(
00462             "-u",
00463             "--out-format",
00464             dest = "out_format",
00465             choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00466             default = "fasta",
00467             help = "File format for the output alignment. Default: fasta"
00468             )
00469         # add shared arguments
00470         self.add_common_args(parser)
```

```
00471            args = parser.parse_args(sys.argv[2:])
00472            return args
00473
00474     def remove(self):
00475            # remove taxa command
00476            parser = argparse.ArgumentParser(
00477                description="Remove taxa from alignment",
00478            )
00479            parser.add_argument(
00480                "-x",
00481                "--taxa-to-remove",
00482                nargs = "+",
00483                type = str,
00484                dest = "taxa_to_remove",
00485                help = "Taxon/sequence names to be removed.",
00486                required = True
00487            )
00488            parser.add_argument(
00489                "-u",
00490                "--out-format",
00491                dest = "out_format",
00492                choices = ["fasta", "phylip", "nexus", "phylip-int", "nexus-int"],
00493                default = "fasta",
00494                help = "File format for the output alignment. Default: fasta"
00495            )
00496            parser.add_argument(
00497                "-g",
00498                "--out-prefix",
00499                dest = "out_prefix",
00500                default = "reduced_",
00501                help = "File name prefix for the concatenated alignment. Default: 'reduced_'"
00502            )
00503            # add shared arguments
00504            self.add_common_args(parser)
00505            args = parser.parse_args(sys.argv[2:])
00506            return args
00507
00508     def get_args_dict(self):
00509            # store arguments in a dictionary
00510            command = self.args.__dict__
00511            arguments = getattr(self, self.args.command)().__dict__
00512            argument_dictionary = command.copy()
00513            argument_dictionary.update(arguments)
00514
00515            return argument_dictionary
00516
00517
00518 class FileHandler:
00519     """Define file handle that closes when out of scope"""
00520
00521     def __init__(self, file_name):
00522            self.file_name = file_name
00523
00524     def __enter__(self):
00525            try:
00526                self.in_file = open(self.file_name, "r", encoding="utf-8")
00527            except FileNotFoundError:
00528                print("ERROR: File '" + self.file_name + "' not found.")
00529                sys.exit()
00530            return self.in_file
00531
00532     def __exit__(self, *args):
00533            self.in_file.close()
00534
00535     def get_file_name(self):
00536            return self.file_name
00537
00538 class FileParser:
00539     """Parse file contents and return sequences and sequence names"""
00540
00541     def __init__(self, in_file):
00542            self.in_file = in_file
00543            with FileHandler(in_file) as handle:
00544                self.in_file_lines = handle.read().rstrip("\r\n")
00545
00546     def fasta_parse(self):
00547            # use regex to parse names and sequences in sequential fasta files
00548            matches = re.finditer(
00549                r"^>(.*[^$])([^>]*)",
00550                self.in_file_lines, re.MULTILINE
00551            )
00552            records = {}
00553
00554            for match in matches:
00555                name_match = match.group(1).replace("\n", "")
00556                seq_match = match.group(2).replace("\n", "").upper()
00557                seq_match = self.translate_ambiguous(seq_match)
```

```
00558                     records[name_match] = seq_match
00559
00560             return records
00561
00562     def phylip_parse(self):
00563         # use regex to parse names and sequences in sequential phylip files
00564         matches = re.finditer(
00565             r"^(\s+)?(\S+)\s+([A-Za-z*?.{}-]+)",
00566             self.in_file_lines, re.MULTILINE
00567         )
00568
00569         records = {}
00570
00571         for match in matches:
00572             name_match = match.group(2).replace("\n", "")
00573             seq_match = match.group(3).replace("\n", "").upper()
00574             seq_match = self.translate_ambiguous(seq_match)
00575             records[name_match] = seq_match
00576
00577         return records
00578
00579     def phylip_interleaved_parse(self):
00580         # use regex to parse names and sequences in interleaved phylip files
00581         tax_chars_matches = re.finditer(
00582             r"^(\s+)?([0-9]+)[ \t]+([0-9]+)",
00583             self.in_file_lines, re.MULTILINE
00584         )
00585         name_matches = re.finditer(
00586             r"^(\s+)?(\S+)[ \t]+[A-Za-z*?.{}-]+",
00587             self.in_file_lines, re.MULTILINE
00588         )
00589         seq_matches = re.finditer(
00590             r"(^(\s+)?\S+[ \t]+|^)([A-Za-z*?.{}-]+)$",
00591             self.in_file_lines, re.MULTILINE
00592         )
00593         # get number of taxa and chars
00594         for match in tax_chars_matches:
00595             tax_match = match.group(2)
00596             chars_match = match.group(3)
00597
00598         # initiate lists for taxa names and sequence strings on separate lines
00599         taxa = []
00600         sequences = []
00601         # initiate a dictionary for the name:sequence records
00602         records = {}
00603         # initiate a counter to keep track of sequences strung together
00604         # from separate lines
00605         counter = 0
00606
00607         for match in name_matches:
00608             name_match = match.group(2).replace("\n", "")
00609             taxa.append(name_match)
00610
00611         for match in seq_matches:
00612             seq_match = match.group(3).replace("\n", "").upper()
00613             seq_match = self.translate_ambiguous(seq_match)
00614             sequences.append(seq_match)
00615         # try parsing PHYLUCE-style interleaved phylip
00616         if len(taxa) != int(tax_match):
00617             taxa = []
00618             sequences = []
00619             matches = re.finditer(
00620                 r"(^(\s+)?(\S+)( ){2,}|^\s+)([ A-Za-z*?.{}-]+)",
00621                 self.in_file_lines, re.MULTILINE
00622             )
00623
00624             for match in matches:
00625                 try:
00626                     name_match = match.group(3).replace("\n", "")
00627                     taxa.append(name_match)
00628                 except AttributeError:
00629                     pass
00630                 seq_match = match.group(5).replace("\n", "").upper()
00631                 seq_match = "".join(seq_match.split())
00632                 seq_match = self.translate_ambiguous(seq_match)
00633                 sequences.append(seq_match)
00634
00635         for taxon_no in range(len(taxa)):
00636             sequence = ""
00637             for index in range(counter,len(sequences),len(taxa)):
00638                 sequence += sequences[index]
00639
00640             records[taxa[taxon_no]] = sequence
00641             counter += 1
00642
00643         return records
00644
```

```
00645      def nexus_parse(self):
00646          # use regex to parse names and sequences in sequential nexus files
00647          # find the matrix block
00648          matches = re.finditer(
00649              r"(\s+)?(MATRIX\n|matrix\n|MATRIX\r\n|matrix\r\n)(.*?;)",
00650              self.in_file_lines, re.DOTALL
00651          )
00652
00653          records = {}
00654          # get names and sequences from the matrix block
00655
00656          for match in matches:
00657              matrix_match = match.group(3)
00658              seq_matches = re.finditer(
00659                  r"^(\s+)?[']?(\S+\s\S+|\S+)[']?\s+([A-Za-z*?.{}-]+)($|\s+\[[0-9]+\]$)",
00660                  matrix_match, re.MULTILINE
00661              )
00662
00663              for match in seq_matches:
00664                  name_match = match.group(2).replace("\n", "")
00665                  seq_match = match.group(3).replace("\n", "").upper()
00666                  seq_match = self.translate_ambiguous(seq_match)
00667                  records[name_match] = seq_match
00668
00669          return records
00670
00671      def nexus_interleaved_parse(self):
00672          # use regex to parse names and sequences in sequential nexus files
00673          # find the matrix block
00674          matches = re.finditer(
00675              r"(\s+)?(MATRIX\n|matrix\n|MATRIX\r\n|matrix\r\n)(.*?;)",
00676              self.in_file_lines, re.DOTALL
00677          )
00678          # initiate lists for taxa names and sequence strings on separate lines
00679          taxa = []
00680          sequences = []
00681          # initiate a dictionary for the name:sequence records
00682          records = {}
00683
00684          for match in matches:
00685              matrix_match = match.group(3)
00686              # get names and sequences from the matrix block
00687              seq_matches = re.finditer(
00688                  r"^(\s+)?[']?(\S+\s\S+|\S+)[']?\s+([A-Za-z*?.{}-]+)($|\s+\[[0-9]+\]$)",
00689                  matrix_match, re.MULTILINE
00690              )
00691
00692              for match in seq_matches:
00693                  name_match = match.group(2)
00694                  if name_match not in taxa:
00695                      taxa.append(name_match)
00696                  seq_match = match.group(3)
00697
00698                  sequences.append(seq_match)
00699
00700          # initiate a counter to keep track of sequences strung together
00701          # from separate lines
00702          counter = 0
00703
00704          for taxon_no in range(len(taxa)):
00705
00706              full_length_sequence = "".join([sequences[index] for index in
    range(counter,len(sequences),len(taxa))])
00707              records[taxa[taxon_no]] = self.translate_ambiguous(full_length_sequence).replace("\n",
    "").upper()
00708              counter += 1
00709
00710          return records
00711
00712      def translate_ambiguous(self, seq):
00713          # translate ambiguous characters from curly bracket format
00714          # to single letter format
00715          # also remove spaces from sequences
00716          seq = seq.replace("{GT}", "K")
00717          seq = seq.replace("{AC}", "M")
00718          seq = seq.replace("{AG}", "R")
00719          seq = seq.replace("{CT}", "Y")
00720          seq = seq.replace("{CG}", "S")
00721          seq = seq.replace("{AT}", "W")
00722          seq = seq.replace("{CGT}", "B")
00723          seq = seq.replace("{ACG}", "V")
00724          seq = seq.replace("{ACT}", "H")
00725          seq = seq.replace("{AGT}", "D")
00726          seq = seq.replace("{GATC}", "N")
00727          seq = seq.replace(" ", "")
00728
00729          return seq
```

```
00730
00731     def partitions_parse(self):
00732         # parse partitions file using regex
00733         # original: `matches = re.finditer(r"^(\s+)?([^ =]+)[ =]+([\\0-9, -]+)", self.in_file_lines,
     re.MULTILINE)`
00734         # new version: more permissive -> handles PartionFinder/RAxML/(IQ-TREE 2)best_scheme.nex
     format partition files
00735         matches = re.finditer(
00736             r"""^[ \t]*                              # start of line w/ zero-or-more (just)
     whitespaces/tabs
00737                 (
00738                     (?P<nexus>charset[ ]+)           # case 1: (IQ-TREE 2)best_scheme.nex partition
     directive; partition name
00739                     |
00740                     (?P<raxml>[A-Za-z0-9_+\.]+,[ \t]+)    # case 2: RAxML/RAxML-NG model(+other pars);
     partition name
00741                 )?
00742                 (?P<partition_name>[A-Za-z0-9_@*&()\[\]{}'"%#|-]+) # partition name (handles various
     metcharacters, excludes [\s],<>?/:;~`\^$!)
00743                 [ ]*=[ ]*                            # whitespace-padded (or unpadded) '=':
     (IQ-TREE 2)best_scheme.nex compatabiliy
00744                 (?P<numbers>[\\0-9, -]+)             # position ranges w/ stride (multiple
     intervals; from original regex)
00745                 (?P<nexus_term>[ ]*[;])?             # whitespace-prepended (or unprepended) ';'
     (nexus terminator)
00746             """,
00747             self.in_file_lines,
00748             re.MULTILINE | re.VERBOSE
00749         )
00750
00751         # initiate list to store dictionaries with lists
00752         # of slice positions as values
00753         partitions = []
00754         add_to_partitions = partitions.append
00755
00756         for match in matches:
00757             # initiate dictionary of partition name as key
00758             dict_of_dicts = {}
00759             # and list of dictionaries with slice positions
00760             list_of_dicts = []
00761             add_to_list_of_dicts = list_of_dicts.append
00762             # get parition name and numbers from parsed partition strings
00763             partition_name = match.group('partition_name')
00764             numbers = match.group('numbers')
00765             # remove any whitespace padding '-' (to be consistent with partition-writing format)
00766             numbers = re.sub(r"[ ]*-[ ]*", "-", numbers)
00767             # find all numbers that will be used to parse positions
00768             positions = re.findall(r"([^ ,]+)", numbers)
00769
00770             for position in positions:
00771                 # create dictionary for slicing input sequence
00772                 # conditioning on whether positions are represented
00773                 # by range, range with stride, or single number
00774                 pos_dict = {}
00775
00776                 if "-" in position:
00777                     m = re.search(r"([0-9]+)-([0-9]+)", position)
00778                     pos_dict["start"] = int(m.group(1)) - 1
00779                     pos_dict["stop"] = int(m.group(2))
00780                 else:
00781                     pos_dict["start"] = int(position) - 1
00782                     pos_dict["stop"] = int(position)
00783
00784                 if "\\" in position:
00785                     # Note: the value of `N` in `...\N` isn't read: the script simply assumes `N` is
     consistent with the number of
00786                     # increments per interval when the alignment is parsed with a stride of 3
     (designating each cpos).
00787                     # E.g. For the partition file:
00788                     #    ...'1-N\2'
00789                     #    ...'2-N\2'
00790                     #    ...'(N+1)-M\2'
00791                     #    ...'(N+2)-M\2'
00792                     # 3'cpos are ignored due to the absence of intervals `3-N...`, `(N+3)-M...`, not
     because the associated stride values are'\2`
00793                     pos_dict["stride"] = 3
00794                 elif "\\" not in position:
00795                     pos_dict["stride"] = 1
00796
00797                 add_to_list_of_dicts(pos_dict)
00798
00799             dict_of_dicts[partition_name] = list_of_dicts
00800             add_to_partitions(dict_of_dicts)
00801
00802         return partitions
00803
00804
```

```
00805 class Alignment:
00806     """Base class: Gets in parsed sequences as input and summarizes their stats.
00807     Based on the data type, the subclasses AminoAcidAlignment & DNAAlignment define the attributes:
00808     `alphabet`, `missing_ambiguous_chars`, `missing_chars`, `non_alphabet`
00809     """
00810
00811     def __init__(self, in_file, in_format, data_type):
00812         # initialize alignment class with parsed records and alignment name as arguments,
00813         # create empty lists for list of sequences, sites without
00814         # ambiguous or missing characters, and initialize variable for the number
00815         # of parsimony informative sites
00816         self.in_file = in_file
00817         self.in_format = in_format
00818         self.data_type = data_type
00819
00820         self.parsed_aln = self.get_parsed_aln()
00821
00822     def __str__(self):
00823         #  purpose of override? (originally returned method object)
00824         return self.get_name()
00825
00826     def get_aln_input(self):
00827         # open and parse input file
00828         aln_input = FileParser(self.in_file)
00829         return aln_input
00830
00831     def get_parsed_aln(self):
00832         # parse according to the given format
00833         aln_input = self.get_aln_input()
00834         if self.in_format == "fasta":
00835             parsed_aln = aln_input.fasta_parse()
00836         elif self.in_format == "phylip":
00837             parsed_aln = aln_input.phylip_parse()
00838         elif self.in_format == "phylip-int":
00839             parsed_aln = aln_input.phylip_interleaved_parse()
00840         elif self.in_format == "nexus":
00841             parsed_aln = aln_input.nexus_parse()
00842         elif self.in_format == "nexus-int":
00843             parsed_aln = aln_input.nexus_interleaved_parse()
00844
00845         return parsed_aln
00846
00847     def summarize_alignment(self):
00848         # call methods to create sequences list, matrix, sites without ambiguous or
00849         # missing characters; get and summarize alignment statistics
00850         summary = []
00851         self.length = str(self.get_alignment_length())
00852         self.matrix = self.matrix_creator()
00853         self.no_missing_ambiguous = self.get_sites_no_missing_ambiguous()
00854         self.variable_sites = self.get_variable()
00855         self.prop_variable = self.get_prop_variable()
00856         self.parsimony_informative = self.get_parsimony_informative()
00857         self.prop_parsimony = self.get_prop_parsimony()
00858         self.missing_records = self.get_missing_from_parsed()
00859         name = str(self.get_name())
00860         taxa_no = str(self.get_taxa_no())
00861         cells = str(self.get_matrix_cells())
00862         missing = str(self.get_missing())
00863         missing_percent = str(self.get_missing_percent())
00864         self.check_data_type()
00865         summary = [
00866             name,
00867             taxa_no,
00868             self.length,
00869             cells,
00870             missing,
00871             missing_percent,
00872             str(self.variable_sites),
00873             str(self.prop_variable),
00874             str(self.parsimony_informative),
00875             str(self.prop_parsimony)
00876         ]
00877         return summary
00878
00879     def summarize_alignment_by_taxa(self):
00880         # get summary for all taxa/sequences in alignment
00881         per_taxon_summary = []
00882         taxa_no = self.get_taxa_no()
00883         self.missing_records = self.get_missing_from_parsed()
00884         self.length = self.get_alignment_length()
00885         lengths = (self.length for i in range(taxa_no))
00886         name = self.get_name()
00887         names = (name for i in range(taxa_no))
00888         taxa_names = (
00889             taxon.replace(" ", "_").replace(".", "_").replace("'", "")
00890             for taxon, missing_count, missing_percent in self.missing_records
00891         )
```

```
00892            missing = (missing_count for taxon, missing_count, missing_percent in self.missing_records)
00893            missing_percent = (missing_percent for taxon, missing_count, missing_percent in
      self.missing_records)
00894            self.check_data_type()
00895            per_taxon_summary = (names, taxa_names, lengths, missing, missing_percent)
00896            zipped = list(zip(*per_taxon_summary))
00897            return zipped
00898
00899        def get_char_summary(self):
00900            # get summary of frequencies for all characters
00901            characters = []
00902            counts = []
00903            add_to_chars = characters.append
00904            add_to_counts = counts.append
00905            char_count_dicts = self.get_counts()
00906            for char in self.alphabet:
00907                add_to_chars(char)
00908                if char in char_count_dicts.keys():
00909                    add_to_counts(str(char_count_dicts[char]))
00910                else:
00911                    add_to_counts("0")
00912            return characters, counts
00913
00914        def get_taxon_char_summary(self):
00915            # get summary of frequencies for all characters
00916            records = (self.append_count(char_dict) for taxon, char_dict in self.get_counts_from_parsed())
00917            return records
00918
00919        def append_count(self, char_dict):
00920            count_list = []
00921            for char in self.alphabet:
00922                if char in char_dict.keys():
00923                    count_list.append(char_dict[char])
00924                else:
00925                    count_list.append(0)
00926            return count_list
00927
00928        def matrix_creator(self):
00929            # decompose character matrix into a two-dimensional list
00930            matrix = [list(sequence) for sequence in self.parsed_aln.values()]
00931            return matrix
00932
00933        def get_column(self, i):
00934            # get site from the character matrix
00935            return [row[i] for row in self.matrix]
00936
00937        def all_same(self, site):
00938            # check if all elements of a site are the same
00939            return not site or site.count(site[0]) == len(site)
00940
00941        def get_sites_no_missing_ambiguous(self):
00942            # get each site without missing or ambiguous characters
00943            no_missing_ambiguous_sites = [self.get_site_no_missing_ambiguous(column) for column in
      range(self.get_alignment_length())]
00944            return no_missing_ambiguous_sites
00945
00946        def get_site_no_missing_ambiguous(self, column):
00947            site = self.get_column(column)
00948            return [char for char in site if char not in self.missing_ambiguous_chars]
00949
00950        def replace_missing(self, column):
00951            return ["-" if x in self.missing_chars else x for x in self.get_column(column)]
00952
00953        def get_trim_selection(self, trim_fraction, parsimony_check):
00954            # this checks each column of alignment for minimum occupancy
00955            self.matrix = self.matrix_creator()
00956            trim_vector = []
00957            for column in range(self.get_alignment_length()):
00958                site = self.replace_missing(column)
00959                occ = (len(site) - site.count("-")) / len(site)
00960                if parsimony_check:
00961                    unique_chars = set(site)
00962                    try:
00963                        unique_chars.remove("-")
00964                    except KeyError:
00965                        pass # this occurs if we have no missing data
00966                    pattern = [base for base in unique_chars if site.count(base) >= 2]
00967                    trim_vector.append(len(pattern) >= 2 and occ >= trim_fraction)
00968                else:
00969                    trim_vector.append(occ >= trim_fraction)
00970            return trim_vector
00971
00972        def get_variable(self):
00973            # if all elements of a site without missing or ambiguous characters
00974            # are not the same, consider it variable
00975            variable = len([site for site in self.no_missing_ambiguous if not self.all_same(site)])
00976            return variable
```

```
00977
00978    def get_parsimony_informative(self):
00979        # if the count for a unique character in a site is at least two,
00980        # and there are at least two such characters in a site without missing
00981        # or ambiguous characters, consider it parsimony informative
00982        parsimony_informative = 0
00983        for site in self.no_missing_ambiguous:
00984            unique_chars = set(site)
00985            pattern = [base for base in unique_chars if site.count(base) >= 2]
00986            no_patterns = len(pattern)
00987
00988            if no_patterns >= 2:
00989                parsimony_informative += 1
00990        return parsimony_informative
00991
00992    def get_prop_variable(self):
00993        # get proportion of variable sites to all sites
00994        prop_variable = self.variable_sites / int(self.length)
00995        return round(prop_variable, 3)
00996
00997    def get_prop_parsimony(self):
00998        # get proportion of parsimony informative sites to all sites
00999        prop_parsimony = self.parsimony_informative / int(self.length)
01000        return round(prop_parsimony, 3)
01001
01002    def get_name(self):
01003        # get input file name
01004        in_filename = path.basename(self.in_file)
01005        return in_filename
01006
01007    def get_taxa_no(self):
01008        # get number of taxa
01009        return len(self.parsed_aln.values())
01010
01011    def get_alignment_length(self):
01012        # get alignment length by just checking the first seq length
01013        # this assumes that all sequences are of equal length
01014        return len(next(iter(self.parsed_aln.values())))
01015
01016    def get_matrix_cells(self):
01017    # count all matrix cells
01018        self.all_matrix_cells = len(self.parsed_aln.values()) * int(self.length)
01019        return self.all_matrix_cells
01020
01021    def get_missing(self):
01022        # count missing characters from the list of missing for all sequences
01023        self.missing = sum(count for taxon, count, percent in self.missing_records)
01024        return self.missing
01025
01026    def get_missing_percent(self):
01027        # get missing percent
01028        missing_percent = round((self.missing / self.all_matrix_cells * 100), 3)
01029        return missing_percent
01030
01031    def get_missing_from_parsed(self):
01032        # get missing count and percent from parsed alignment
01033        # return a list of tuples with taxon name, count, and percent missing
01034        self.missing_records = sorted(
01035            [
01036                (taxon, self.get_missing_from_seq(seq), self.get_missing_percent_from_seq(seq))
01037                for taxon, seq in self.parsed_aln.items()
01038            ]
01039        )
01040        return self.missing_records
01041
01042    def get_missing_from_seq(self, seq):
01043        # count missing characters for individual sequence
01044        missing_count = sum(seq.count(char) for char in self.missing_chars)
01045        return missing_count
01046
01047    def get_missing_percent_from_seq(self, seq):
01048        # get missing percent from individual sequence
01049        missing_seq_percent = round((self.get_missing_from_seq(seq) / self.get_alignment_length() *
    100), 3)
01050        return missing_seq_percent
01051
01052    def get_counts(self):
01053        # get counts of each character in the used alphabet for all sequences
01054        counters = [Counter(chars) for taxon, chars in self.get_counts_from_parsed()]
01055        all_counts = sum(counters, Counter())
01056        counts_dict = dict(all_counts)
01057        return counts_dict
01058
01059    def get_counts_from_parsed(self):
01060        # get counts of all characters from parsed alignment
01061        # return a list of tuples with taxon name and counts
01062        return sorted(
```

```
01063                [
01064                    (taxon, self.get_counts_from_seq(seq))
01065                    for taxon, seq in self.parsed_aln.items()
01066                ]
01067            )
01068
01069    def get_counts_from_seq(self, seq):
01070        # get all alphabet chars count for individual sequence
01071        char_counts = {char : seq.count(char) for char in self.alphabet}
01072        return char_counts
01073
01074    def check_data_type(self):
01075        # check if the data type is correct; only one seq to save on computation
01076        seq = next(iter(self.parsed_aln.values()))
01077        self.check = any(char in self.non_alphabet for char in seq)
01078        if self.check is True:
01079            print(
01080                "WARNING: found non-" + self.data_type + " characters. "
01081                "Are you sure you specified the right data type?"
01082            )
01083
01084
01085 class AminoAcidAlignment(Alignment):
01086    """Alphabets specific to amino acid alignments"""
01087
01088    alphabet = ["A", "C", "D", "E", "F", "G", "H", "I", "K", "L", "M", "N", "P", "Q", "R", "S", "T",
    "V", "W", "Y", "B", "J", "Z", "X", ".", "*", "-", "?"]
01089    missing_ambiguous_chars = ["B", "J", "Z", "X", ".", "*", "-", "?"]
01090    missing_chars = ["X", ".", "*", "-", "?"]
01091    non_alphabet = ["O"]
01092
01093    def get_summary(self):
01094        # get alignment summary specific to amino acids
01095        data = self.summarize_alignment()
01096        new_data = data + list(self.get_char_summary()[1])
01097        return new_data
01098
01099    def get_taxa_summary(self):
01100        # get per-taxon/sequence alignment summary specific to amino acids
01101        data = self.summarize_alignment_by_taxa()
01102        aa_summary = (data, self.get_taxon_char_summary())
01103        zipped_list = list(zip(*aa_summary))
01104        new_data = [list(data_tupl) + chars for data_tupl, chars in zipped_list]
01105        return new_data
01106
01107 class DNAAlignment(Alignment):
01108    """Alphabets specific to DNA alignments"""
01109
01110    alphabet = ["A", "C", "G", "T", "K", "M", "R", "Y", "S", "W", "B", "V", "H", "D", "X", "N", "O",
    "-", "?"]
01111    missing_ambiguous_chars = ["K", "M", "R", "Y", "S", "W", "B", "V", "H", "D", "X", "N", "O", "-",
    "?"]
01112    missing_chars = ["X", "N", "O", "-", "?"]
01113    non_alphabet = ["E", "F", "I", "L", "P", "Q", "J", "Z", ".", "*"]
01114
01115    def get_summary(self):
01116        # get alignment summarry specific to nucleotide
01117        data = self.summarize_alignment()
01118        new_data = data + self.get_atgc_content() + list(self.get_char_summary()[1])
01119        return new_data
01120
01121    def get_taxa_summary(self):
01122        # get per-taxon/sequence alignment summary specific to nucleotides
01123        data = self.summarize_alignment_by_taxa()
01124        dna_summary = (data, self.get_list_from_atgc(), self.get_taxon_char_summary())
01125        zipped_list = list(zip(*dna_summary))
01126        new_data = [list(data_tupl) + list(atgc) + chars for data_tupl, atgc, chars in zipped_list]
01127        return new_data
01128
01129    def get_atgc_content(self):
01130        # get AC and GC contents for all sequences
01131        # AT content is the first element of AT, GC content tuple
01132        # returned by get_atgc_from_seq()
01133        atgc_records = self.get_atgc_from_parsed()
01134        at_content = round(sum(atgc[0] for taxon, atgc in atgc_records) / self.get_taxa_no(), 3)
01135        gc_content = round(1 - float(at_content), 3)
01136
01137        atgc_content = [str(at_content), str(gc_content)]
01138        return atgc_content
01139
01140    def get_list_from_atgc(self):
01141        records = (atgc for taxon, atgc in self.get_atgc_from_parsed())
01142        return records
01143
01144    def get_atgc_from_parsed(self):
01145        # get AT and GC contents from parsed alignment dictionary
01146        # return a list of tuples with taxon name, AT content, and GC content
```

```
01147            return sorted([(taxon, self.get_atgc_from_seq(seq)) for taxon, seq in
    self.parsed_aln.items()])
01148
01149      def get_atgc_from_seq(self, seq):
01150            # get AT and GC contents from individual sequences
01151
01152            at_count = seq.count("A") + seq.count("T") + seq.count("W")
01153            gc_count = seq.count("G") + seq.count("C") + seq.count("S")
01154
01155            try:
01156                at_content = round(at_count / (at_count + gc_count), 3)
01157                gc_content = round(1 - float(at_content), 3)
01158
01159            except ZeroDivisionError:
01160                at_content = 0
01161                gc_content = 0
01162
01163            return at_content, gc_content
01164
01165 class MetaAlignment:
01166      """Class of multiple sequence alignments"""
01167
01168      def __init__(self, **kwargs):
01169            # set defaults and get values from kwargs
01170            self.in_files = kwargs.get("in_files")
01171            self.in_format = kwargs.get("in_format")
01172            self.data_type = kwargs.get("data_type")
01173            self.command = kwargs.get("command")
01174            self.concat_out = kwargs.get("concat_out", "concatenated.out")
01175            self.using_metapartitions = False
01176            self.check_align = kwargs.get("check_align", False)
01177            self.cores = kwargs.get("cores")
01178            self.by_taxon_summary = kwargs.get("by_taxon_summary")
01179            self.no_sup_aln_name = False
01180            self.no_mpan = False
01181
01182            if self.command == "concat":
01183                self.codons = kwargs.get("codons", "none")
01184                if self.data_type == "aa" and self.codons != "none":
01185                    print("ERROR: when option -d|--data-type is set to 'aa', option -n|--codons must be
    set to 'none'.")
01186                    sys.exit(1)
01187
01188            if self.command == "replicate":
01189                self.no_replicates = kwargs.get("replicate_args")[0]
01190                self.no_loci = kwargs.get("replicate_args")[1]
01191
01192            if self.command == "split":
01193                self.split = kwargs.get("split_by")
01194                self.remove_empty = kwargs.get("remove_empty", False)
01195                self.no_sup_aln_name = kwargs.get("no_sup_aln_name", False)
01196
01197            if self.command == "metapartitions":
01198                self.using_metapartitions = True
01199                self.split = kwargs.get("split_by")
01200                self.remove_empty = kwargs.get("remove_empty", False)
01201                self.no_sup_aln_name = kwargs.get("no_sup_aln_name", False)
01202                self.no_mpan = kwargs.get("no_mpan", False)
01203                self.prepend_label = kwargs.get("prepend_label")
01204                if self.prepend_label is not None and isinstance(self.prepend_label, str):
01205                    self.prepend_label = self.prepend_label + "_"
01206                else:
01207                    self.prepend_label = ""
01208
01209            if self.command == "remove":
01210                self.species_to_remove = kwargs.get("taxa_to_remove")
01211                self.species_to_remove_set = set(self.species_to_remove)
01212                self.reduced_file_prefix = kwargs.get("out_prefix")
01213                self.check_taxa = kwargs.get("check_taxa", False)
01214
01215            if self.command == "translate":
01216                self.reading_frame = kwargs.get("reading_frame")
01217                self.genetic_code = kwargs.get("genetic_code")
01218
01219            if self.command == "trim":
01220                self.trim_fraction = kwargs.get("trim_fraction")
01221                self.trim_out = kwargs.get("trim_out")
01222                self.parsimony_check = kwargs.get("parsimony_check", False)
01223
01224            self.alignment_objects = self.get_alignment_objects()
01225            self.parsed_alignments = self.get_parsed_alignments()
01226
01227            # The code list:
01228            self.codes_list = """
01229             1. The Standard Code
01230             2. The Vertebrate Mitochondrial Code
01231             3. The Yeast Mitochondrial Code
```

```
01232              4. The Mold, Protozoan, and Coelenterate Mitochondrial Code and the Mycoplasma/Spiroplasma
      Code
01233              5. The Invertebrate Mitochondrial Code
01234              6. The Ciliate, Dasycladacean and Hexamita Nuclear Code
01235              9. The Echinoderm and Flatworm Mitochondrial Code
01236             10. The Euplotid Nuclear Code
01237             11. The Bacterial, Archaeal and Plant Plastid Code
01238             12. The Alternative Yeast Nuclear Code
01239             13. The Ascidian Mitochondrial Code
01240             14. The Alternative Flatworm Mitochondrial Code
01241             16. Chlorophycean Mitochondrial Code
01242             21. Trematode Mitochondrial Code
01243             22. Scenedesmus obliquus Mitochondrial Code
01244             23. Thraustochytrium Mitochondrial Code
01245             24. Pterobranchia Mitochondrial Code
01246             25. Candidate Division SR1 and Gracilibacteria Code
01247             26. Pachysolen tannophilus Nuclear Code
01248          """
01249
01250          # 1: The Standard Code
01251          self.gencode_NCBI_1 = {
01252          "TTT" : "F", # Phe
01253          "TCT" : "S", # Ser
01254          "TAT" : "Y", # Tyr
01255          "TGT" : "C", # Cys
01256          "TTC" : "F", # Phe
01257          "TCC" : "S", # Ser
01258          "TAC" : "Y", # Tyr
01259          "TGC" : "C", # Cys
01260          "TTA" : "L", # Leu
01261          "TCA" : "S", # Ser
01262          "TAA" : "*", # Ter
01263          "TGA" : "*", # Ter
01264          "TTG" : "L", # Leu i
01265          "TCG" : "S", # Ser
01266          "TAG" : "*", # Ter
01267          "TGG" : "W", # Trp
01268          "CTT" : "L", # Leu
01269          "CCT" : "P", # Pro
01270          "CAT" : "H", # His
01271          "CGT" : "R", # Arg
01272          "CTC" : "L", # Leu
01273          "CCC" : "P", # Pro
01274          "CAC" : "H", # His
01275          "CGC" : "R", # Arg
01276          "CTA" : "L", # Leu
01277          "CCA" : "P", # Pro
01278          "CAA" : "Q", # Gln
01279          "CGA" : "R", # Arg
01280          "CTG" : "L", # Leu i
01281          "CCG" : "P", # Pro
01282          "CAG" : "Q", # Gln
01283          "CGG" : "R", # Arg
01284          "ATT" : "I", # Ile
01285          "ACT" : "T", # Thr
01286          "AAT" : "N", # Asn
01287          "AGT" : "S", # Ser
01288          "ATC" : "I", # Ile
01289          "ACC" : "T", # Thr
01290          "AAC" : "N", # Asn
01291          "AGC" : "S", # Ser
01292          "ATA" : "I", # Ile
01293          "ACA" : "T", # Thr
01294          "AAA" : "K", # Lys
01295          "AGA" : "R", # Arg
01296          "ATG" : "M", # Met i
01297          "ACG" : "T", # Thr
01298          "AAG" : "K", # Lys
01299          "AGG" : "R", # Arg
01300          "GTT" : "V", # Val
01301          "GCT" : "A", # Ala
01302          "GAT" : "D", # Asp
01303          "GGT" : "G", # Gly
01304          "GTC" : "V", # Val
01305          "GCC" : "A", # Ala
01306          "GAC" : "D", # Asp
01307          "GGC" : "G", # Gly
01308          "GTA" : "V", # Val
01309          "GCA" : "A", # Ala
01310          "GAA" : "E", # Glu
01311          "GGA" : "G", # Gly
01312          "GTG" : "V", # Val
01313          "GCG" : "A", # Ala
01314          "GAG" : "E", # Glu
01315          "GGG" : "G", # Gly
01316          "---" : "-", # Gap
01317          "???" : "?", # Unk
```

```
01318            "NNN" : "X", # Unk
01319            }
01320
01321            # 2: The Vertebrate Mitochondrial Code
01322            self.gencode_NCBI_2 = self.gencode_NCBI_1.copy()
01323            self.gencode_NCBI_2["AGA"] = "*" # Ter
01324            self.gencode_NCBI_2["AGG"] = "*" # Ter
01325            self.gencode_NCBI_2["ATA"] = "M" # Met
01326            self.gencode_NCBI_2["TGA"] = "W" # Trp
01327
01328            # 3: The Yeast Mitochondrial Code
01329            self.gencode_NCBI_3 = self.gencode_NCBI_1.copy()
01330            self.gencode_NCBI_3["ATA"] = "M" # Met
01331            self.gencode_NCBI_3["CTT"] = "T" # Thr
01332            self.gencode_NCBI_3["CTC"] = "T" # Thr
01333            self.gencode_NCBI_3["CTA"] = "T" # Thr
01334            self.gencode_NCBI_3["CTG"] = "T" # Thr
01335            self.gencode_NCBI_3["TGA"] = "W" # Trp
01336
01337            del self.gencode_NCBI_3["CGA"]
01338            del self.gencode_NCBI_3["CGC"]
01339
01340            # 4: The Mold, Protozoan, and Coelenterate Mitochondrial Code and the Mycoplasma/Spiroplasma
       Code
01341            self.gencode_NCBI_4 = self.gencode_NCBI_1.copy()
01342            self.gencode_NCBI_4["TGA"] = "W" # Trp
01343
01344            # 5: The Invertebrate Mitochondrial Code
01345            self.gencode_NCBI_5 = self.gencode_NCBI_1.copy()
01346            self.gencode_NCBI_5["AGA"] = "S" # Ser
01347            self.gencode_NCBI_5["AGG"] = "S" # Ser
01348            self.gencode_NCBI_5["ATA"] = "M" # Met
01349            self.gencode_NCBI_5["TGA"] = "W" # Trp
01350
01351            # 6: The Ciliate, Dasycladacean and Hexamita Nuclear Code
01352            self.gencode_NCBI_6 = self.gencode_NCBI_1.copy()
01353            self.gencode_NCBI_6["TAA"] = "Q" # Gln
01354            self.gencode_NCBI_6["TAG"] = "Q" # Gln
01355
01356            # 9: The Echinoderm and Flatworm Mitochondrial Code
01357            self.gencode_NCBI_9 = self.gencode_NCBI_1.copy()
01358            self.gencode_NCBI_9["AAA"] = "N" # Asn
01359            self.gencode_NCBI_9["AGA"] = "S" # Ser
01360            self.gencode_NCBI_9["AGG"] = "S" # Ser
01361            self.gencode_NCBI_9["TGA"] = "W" # Trp
01362
01363            # 10: The Euplotid Nuclear Code
01364            self.gencode_NCBI_10 = self.gencode_NCBI_1.copy()
01365            self.gencode_NCBI_10["TGA"] = "C" # Cys
01366
01367            # 11: The Bacterial, Archaeal and Plant Plastid Code
01368            self.gencode_NCBI_11 = self.gencode_NCBI_1.copy()
01369
01370            # 12: The Alternative Yeast Nuclear Code
01371            self.gencode_NCBI_12 = self.gencode_NCBI_1.copy()
01372            self.gencode_NCBI_12["CTG"] = "S" # Ser
01373
01374            # 13: The Ascidian Mitochondrial Code
01375            self.gencode_NCBI_13 = self.gencode_NCBI_1.copy()
01376            self.gencode_NCBI_13["AGA"] = "G" # Gly
01377            self.gencode_NCBI_13["AGG"] = "G" # Gly
01378            self.gencode_NCBI_13["ATA"] = "M" # Met
01379            self.gencode_NCBI_13["TGA"] = "W" # Trp
01380
01381            # 14: The Alternative Flatworm Mitochondrial Code
01382            self.gencode_NCBI_14 = self.gencode_NCBI_1.copy()
01383            self.gencode_NCBI_14["AAA"] = "N" # Asn
01384            self.gencode_NCBI_14["AGA"] = "S" # Ser
01385            self.gencode_NCBI_14["AGG"] = "S" # Ser
01386            self.gencode_NCBI_14["TAA"] = "Y" # Tyr
01387            self.gencode_NCBI_14["TGA"] = "W" # Trp
01388
01389            # 16: Chlorophycean Mitochondrial Code
01390            self.gencode_NCBI_16 = self.gencode_NCBI_1.copy()
01391            self.gencode_NCBI_16["TAG"] = "L" # Leu
01392
01393            # 21: Trematode Mitochondrial Code
01394            self.gencode_NCBI_21 = self.gencode_NCBI_1.copy()
01395            self.gencode_NCBI_21["TGA"] = "W" # Trp
01396            self.gencode_NCBI_21["ATA"] = "M" # Met
01397            self.gencode_NCBI_21["AGA"] = "S" # Ser
01398            self.gencode_NCBI_21["AGG"] = "S" # Ser
01399            self.gencode_NCBI_21["AAA"] = "N" # Asn
01400
01401            # 22: Scenedesmus obliquus Mitochondrial Code
01402            self.gencode_NCBI_22 = self.gencode_NCBI_1.copy()
01403            self.gencode_NCBI_22["TCA"] = "*" # Ter
```

```
01404            self.gencode_NCBI_22["TAG"] = "L" # Leu
01405
01406            # 23: Thraustochytrium Mitochondrial Code
01407            self.gencode_NCBI_23 = self.gencode_NCBI_1.copy()
01408            self.gencode_NCBI_23["TTA"] = "*" # Ter
01409
01410            # 24: Pterobranchia Mitochondrial Code
01411            self.gencode_NCBI_24 = self.gencode_NCBI_1.copy()
01412            self.gencode_NCBI_24["AGA"] = "S" # Ser
01413            self.gencode_NCBI_24["AGG"] = "K" # Lys
01414            self.gencode_NCBI_24["TGA"] = "W" # Trp
01415
01416            # 25: Candidate Division SR1 and Gracilibacteria Code
01417            self.gencode_NCBI_25 = self.gencode_NCBI_1.copy()
01418            self.gencode_NCBI_25["TGA"] = "G" # Gly
01419
01420            # 26: Pachysolen tannophilus Nuclear Code
01421            self.gencode_NCBI_26 = self.gencode_NCBI_1.copy()
01422            self.gencode_NCBI_26["CTG"] = "A" # Ala
01423
01424            self.codes = {
01425            1 : self.gencode_NCBI_1,
01426            2 : self.gencode_NCBI_2,
01427            3 : self.gencode_NCBI_3,
01428            4 : self.gencode_NCBI_4,
01429            5 : self.gencode_NCBI_5,
01430            6 : self.gencode_NCBI_6,
01431            9 : self.gencode_NCBI_9,
01432            10 : self.gencode_NCBI_10,
01433            11 : self.gencode_NCBI_11,
01434            12 : self.gencode_NCBI_12,
01435            13 : self.gencode_NCBI_13,
01436            14 : self.gencode_NCBI_14,
01437            16 : self.gencode_NCBI_16,
01438            21 : self.gencode_NCBI_21,
01439            22 : self.gencode_NCBI_22,
01440            23 : self.gencode_NCBI_23,
01441            24 : self.gencode_NCBI_24,
01442            25 : self.gencode_NCBI_25,
01443            26 : self.gencode_NCBI_26
01444            }
01445
01446     def translate_dna_to_aa(self, seq, translation_table, frame):
01447            # translate DNA string into amino acids
01448            # where the last codon starts
01449            last_codon_start = len(seq) - 2
01450            # where the first codon starts
01451            if frame == 1:
01452                first = 0
01453            elif frame == 2:
01454                first = 1
01455            elif frame == 3:
01456                first = 2
01457            # create protein sequence by growing list
01458            protein = []
01459            add_to_protein = protein.append
01460            for start in range(first, last_codon_start, 3):
01461                codon = seq[start : start + 3]
01462                aa = translation_table.get(codon.upper(), 'X')
01463                add_to_protein(aa)
01464
01465            return "".join(protein)
01466
01467     def translate_dict(self, source_dict):
01468            translation_table = self.codes.get(self.genetic_code)
01469            translated_dict = {}
01470            for taxon, seq in sorted(source_dict.items()):
01471                translated_seq = self.translate_dna_to_aa(seq, translation_table, self.reading_frame)
01472                if "*" in translated_seq:
01473                    print("WARNING: stop codon(s), indicated as *, found in {} sequence".format(taxon))
01474                translated_dict[taxon] = translated_seq
01475
01476            return translated_dict
01477
01478     def get_translated(self, translation_table, reading_frame):
01479            if int(self.cores) == 1:
01480                translated_alignments = [self.translate_dict(alignment) for alignment in
    self.parsed_alignments]
01481            elif int(self.cores) > 1:
01482                pool = mp.Pool(int(self.cores))
01483                translated_alignments = pool.map(self.translate_dict, self.parsed_alignments)
01484
01485            return translated_alignments
01486
01487     def trim_dict(self, alignment):
01488            trim_vector = alignment.get_trim_selection(self.trim_fraction, self.parsimony_check)
01489            aln_dict = alignment.parsed_aln
```

```
01490            for key in aln_dict:
01491                aln_dict[key] = "".join(list(compress(aln_dict[key], trim_vector)))
01492
01493            return aln_dict
01494
01495     def get_trimmed(self, trim_fraction, parsimony_check):
01496         if int(self.cores) == 1:
01497             trimmed_alignments = [self.trim_dict(alignment) for alignment in self.alignment_objects]
01498         elif int(self.cores) > 1:
01499             pool = mp.Pool(int(self.cores))
01500             trimmed_alignments = pool.map(self.trim_dict, self.alignment_objects)
01501
01502         return trimmed_alignments
01503
01504     def remove_unknown_chars(self, seq):
01505         # remove unknown characters from sequence
01506         new_seq = seq.replace("?", "").replace("-", "")
01507
01508         return new_seq
01509
01510     def remove_empty_sequences(self, split_alignment):
01511         # remove taxa from alignment if they are composed of only empty sequences
01512         new_alignment = {taxon : seq for taxon, seq in split_alignment.items() if
     self.remove_unknown_chars(seq)}
01513
01514         return new_alignment
01515
01516     def get_partitions(self, partitions_file):
01517         # parse and get partitions from partitions file
01518         partitions = FileParser(partitions_file)
01519         parsed_partitions = partitions.partitions_parse()
01520
01521         return parsed_partitions
01522
01523     def get_alignment_object(self, alignment):
01524         # parse according to the given alphabet;
01525         # Note:('alignment') <=> `in_file` outside MetaAlignment, e.g.
01526         #
     AminoAcidAlignment(Alignment<-.get_parsed_aln<-.get_aln_input)<-FileParser.__init__(in_file)<-FileHandler(...open(self.
01527         if self.data_type == "aa":
01528             aln = AminoAcidAlignment(alignment, self.in_format, self.data_type)
01529         elif self.data_type == "dna":
01530             aln = DNAAlignment(alignment, self.in_format, self.data_type)
01531         return aln
01532
01533     def get_alignment_objects(self):
01534         # get alignment objects on which statistics can be computed
01535         # use multiprocessing if more than one core specified
01536         if int(self.cores) == 1:
01537             alignments = [self.get_alignment_object(alignment) for alignment in self.in_files]
01538         elif int(self.cores) > 1:
01539             pool = mp.Pool(int(self.cores))
01540             alignments = pool.map(self.get_alignment_object, self.in_files)
01541         return alignments
01542
01543     def get_parsed_alignments(self):
01544         # get parsed dictionaries with taxa and sequences
01545         parsed_alignments = []
01546         add_to_parsed_alignments = parsed_alignments.append
01547         for alignment in self.alignment_objects:
01548             parsed = alignment.parsed_aln
01549             add_to_parsed_alignments(parsed)
01550             # checking if every seq has the same length or if parsed is not empty; exit if false
01551             if self.check_align:
01552                 equal = all(
01553                     x == [len(list(parsed.values())[i]) for i in
     range(0,len(list(parsed.values())))][0]
01554                     for x in [len(list(parsed.values())[i]) for i in
     range(0,len(list(parsed.values())))]
01555                 )
01556                 if equal is False:
01557                     print("ERROR: Sequences in input are of varying lengths. Be sure to align them
     first.")
01558                     sys.exit()
01559
01560             if not parsed.keys() or not any(parsed.values()):
01561                 print(
01562                     "ERROR: Parsed sequences of " + alignment.in_file + " are empty. "
01563                     "Are you sure you specified the right input format and/or that input is a valid
     alignment?"
01564                 )
01565                 sys.exit()
01566
01567         return parsed_alignments
01568
01569     def get_partitioned(self, partitions_file):
01570         # partition alignment according to a partitions file
```

```
01571            partitions = self.get_partitions(partitions_file)
01572            alignment = self.parsed_alignments[0]
01573
01574            # initiate list of newly partitioned alignments
01575            list_of_parts = []
01576            add_to_list_of_parts = list_of_parts.append
01577            for partition in partitions:
01578                # loop over all parsed partitions, adding taxa and sliced sequences
01579                for name, elements in partition.items():
01580                    new_dict = {}
01581
01582                    for taxon, seq in alignment.items():
01583                        new_seq = ""
01584
01585                        for dictionary in elements:
01586                            new_seq = new_seq +
        seq[dictionary["start"]:dictionary["stop"]:dictionary["stride"]]
01587                        new_dict[taxon] = new_seq
01588
01589                    if self.remove_empty:
01590                        # check if remove empty sequences
01591                        no_empty_dict = self.remove_empty_sequences(new_dict)
01592                        add_to_list_of_parts({name : no_empty_dict})
01593                    else:
01594                        # add partition name : dict of taxa and sequences to the list
01595                        add_to_list_of_parts({name : new_dict})
01596
01597            return list_of_parts
01598
01599        def get_summaries(self):
01600            # get summaries for all alignment objects
01601
01602            # define different headers for aa and dna alignments
01603            aa_header = [
01604                "Alignment_name",
01605                "No_of_taxa",
01606                "Alignment_length",
01607                "Total_matrix_cells",
01608                "Undetermined_characters",
01609                "Missing_percent",
01610                "No_variable_sites",
01611                "Proportion_variable_sites",
01612                "Parsimony_informative_sites",
01613                "Proportion_parsimony_informative"
01614            ]
01615
01616            dna_header = [
01617                "Alignment_name",
01618                "No_of_taxa",
01619                "Alignment_length",
01620                "Total_matrix_cells",
01621                "Undetermined_characters",
01622                "Missing_percent",
01623                "No_variable_sites",
01624                "Proportion_variable_sites",
01625                "Parsimony_informative_sites",
01626                "Proportion_parsimony_informative",
01627                "AT_content",
01628                "GC_content"
01629            ]
01630
01631            alignments = self.alignment_objects
01632            parsed_alignments = self.parsed_alignments
01633            freq_header = [char for char in alignments[0].alphabet]
01634
01635            if self.data_type == "aa":
01636                header = aa_header + freq_header
01637            elif self.data_type == "dna":
01638                header = dna_header + freq_header
01639
01640            # use multiprocessing if more than one core specified
01641            if int(self.cores) == 1:
01642                summaries = [alignment.get_summary() for alignment in alignments]
01643            elif int(self.cores) > 1:
01644                pool = mp.Pool(int(self.cores))
01645                summaries = pool.map(self.summarize_alignments, alignments)
01646            return header, summaries
01647
01648        def summarize_alignments(self, alignment):
01649            # helper function to summarize alignments
01650            summary = alignment.get_summary()
01651            return summary
01652
01653        def get_taxon_summaries(self):
01654            # get per-sequence summaries for all alignment objects
01655
01656            # define different headers for aa and dna alignments
```

```
01657            aa_header = [
01658                "Alignment_name",
01659                "Taxon_name",
01660                "Sequence_length",
01661                "Undetermined_characters",
01662                "Missing_percent"
01663            ]
01664
01665            dna_header = [
01666                "Alignment_name",
01667                "Taxon_name",
01668                "Sequence_length",
01669                "Undetermined_characters",
01670                "Missing_percent",
01671                "AT_content",
01672                "GC_content"
01673            ]
01674
01675            alignments = self.alignment_objects
01676            parsed_alignments = self.parsed_alignments
01677            freq_header = alignments[0].alphabet
01678
01679            if self.data_type == "aa":
01680                header = aa_header + freq_header
01681            elif self.data_type == "dna":
01682                header = dna_header + freq_header
01683
01684            # use multiprocessing if more than one core specified
01685            if int(self.cores) == 1:
01686                summaries = [alignment.get_taxa_summary() for alignment in alignments]
01687            elif int(self.cores) > 1:
01688                pool = mp.Pool(int(self.cores))
01689                summaries = pool.map(self.summarize_alignments_taxa, alignments)
01690
01691            return header, summaries
01692
01693    def summarize_alignments_taxa(self, alignment):
01694        # helper function to summarize alignments by taxon
01695        summary = alignment.get_taxa_summary()
01696        return summary
01697
01698    def write_summaries(self, file_name):
01699        # write summaries to file
01700
01701        self.file_overwrite_error(file_name)
01702
01703        with open(file_name, "w", encoding="utf-8") as summary_file:
01704            summary_out = self.get_summaries()
01705            header = '\t'.join(summary_out[0])
01706            new_summ = ['\t'.join(summary) for summary in summary_out[1]]
01707            summary_file.write(header + '\n')
01708            summary_file.write('\n'.join(new_summ))
01709            summary_file.write('\n')
01710            print("Wrote summaries to file '" + file_name + "'")
01711
01712    def write_taxa_summaries(self):
01713        # write by-taxon summaries to file
01714        for index, in_file_name in enumerate(self.in_files):
01715            out_file_name = in_file_name + "-seq-summary.txt"
01716            self.file_overwrite_error(out_file_name)
01717            with open(out_file_name, "w", encoding="utf-8") as summary_file:
01718                summary_out = self.get_taxon_summaries()
01719                header = '\t'.join(summary_out[0])
01720                summ = [[str(col) for col in element] for element in summary_out[1][index]]
01721                new_summ = ['\t'.join(row) for row in summ]
01722                summary_file.write(header + '\n')
01723                summary_file.write('\n'.join(new_summ))
01724                summary_file.write('\n')
01725
01726    def get_replicate(self, no_replicates, no_loci):
01727        # construct replicate data sets for phylogenetic jackknife
01728        replicates = []
01729        add_to_replicates = replicates.append
01730        counter = 1
01731        for replicate in range(no_replicates):
01732
01733            try:
01734                random_alignments = sample(self.parsed_alignments, no_loci)
01735            except ValueError:
01736                print("ERROR: You specified more loci per replicate than there are in your input.")
01737                sys.exit()
01738
01739            random_alignments = sample(self.parsed_alignments, no_loci)
01740            concat_replicate = self.get_concatenated(random_alignments)[0]
01741            add_to_replicates(concat_replicate)
01742            counter += 1
01743
```

```
01744            return replicates
01745
01746       def get_concatenated(self, alignments):
01747           # concatenate muntiple input alignments
01748           # create empty dictionary of lists
01749           concatenated = defaultdict(list)
01750
01751           # first create list of taxa in all alignments
01752           # you need this to insert empty seqs in
01753           # the concatenated alignment
01754           all_taxa = []
01755           for alignment in alignments:
01756               for taxon in alignment.keys():
01757                   if taxon not in all_taxa:
01758                       all_taxa.append(taxon)
01759
01760           # start counters to keep track of partitions
01761           partition_counter = 1
01762           position_counter = 1
01763           # get dict for alignment name and partition
01764           partitions = {}
01765           digits_to_pad = len(str(len(alignments)))
01766
01767           for alignment in alignments:
01768               # get alignment length from a random taxon
01769               partition_length = len(alignment[list(alignment.keys())[0]])
01770               # get base name of each alignment for use when writing partitions file
01771               # NOTE: the base name here is whatever comes before fist period in the file name
01772               alignment_name = self.alignment_objects[partition_counter - 1].get_name().split('.')[0]
01773
01774               if self.using_metapartitions:
01775                   # implementation of option --no-mpan; option --prepend(-label) will assign a string or
       "" (see class definition)
01776                   if self.no_mpan:
01777                       # omit original alignment names from the printed partition file
01778                       partition_name = self.prepend_label + "p" +
       str(partition_counter).zfill(digits_to_pad)
01779                   else:
01780                       # keep original alignment names in the printed partition file
01781                       partition_name = self.prepend_label + "p" +
       str(partition_counter).zfill(digits_to_pad) + "_" + alignment_name
01782               else:
01783                   partition_name = "p" + str(partition_counter) + "_" + alignment_name
01784
01785               start = position_counter
01786               position_counter += partition_length
01787               end = position_counter - 1
01788               partitions[partition_name] = str(start) + "-" + str(end)
01789               partition_counter += 1
01790
01791               # get empty sequence if there is missing taxon
01792               # getting length from first element of list of keys
01793               # created from the original dict for this alignment
01794               empty_seq = '?' * partition_length
01795
01796               for taxon in all_taxa:
01797
01798                   if taxon not in alignment.keys():
01799                       concatenated[taxon].append(empty_seq)
01800                   else:
01801                       concatenated[taxon].append(alignment[taxon])
01802
01803           concatenated = {taxon:''.join(seqs) for taxon, seqs in concatenated.items()}
01804
01805           return concatenated, partitions
01806
01807       def remove_from_alignment(self, alignment, species_to_remove_set, index):
01808           # remove taxa from alignment
01809           aln_name = self.get_alignment_name_no_ext(index)
01810           for taxon in species_to_remove_set:
01811               if taxon not in alignment.keys():
01812                   print(
01813                       "WARNING: Taxon '" + taxon + "' not found in '" + aln_name + "'.\nIf you expected
       it to be there, "
01814                       "make sure to replace all taxon name spaces with underscores and that you are not
       using quotes."
01815                   )
01816           # originally within for-loop scope (redundancy)
01817           new_alignment = {species: seq for species, seq in alignment.items() if species not in
       species_to_remove_set}
01818
01819           return aln_name, new_alignment
01820
01821       def remove_taxa(self, species_to_remove_set):
01822           new_alns = {}
01823           for index, alignment in enumerate(self.parsed_alignments):
01824               aln_name, aln_dict = self.remove_from_alignment(alignment, species_to_remove_set, index)
```

```
01825                    # check if alignment is not empty:
01826                    if aln_dict:
01827                        new_alns[aln_name] = aln_dict
01828                    else:
01829                        print("ERROR: You asked to remove all taxa from the alignment " + aln_name + ". No
      output file will be written.")
01830
01831            return new_alns
01832
01833     def print_fasta(self, source_dict):
01834            # print fasta-formatted string from a dictionary
01835            fasta_string = ""
01836            # each sequence line will have 80 characters
01837            n = 80
01838
01839            for taxon, seq in sorted(source_dict.items()):
01840                # split dictionary values to a list of string, each n chars long
01841                seq = [seq[i:i+n] for i in range(0, len(seq), n)]
01842                # in case there are unwanted spaces in taxon names
01843                taxon = taxon.replace(" ", "_").strip("'")
01844                fasta_string += ">" + taxon + "\n"
01845                for element in seq:
01846                    fasta_string += element + "\n"
01847
01848            return fasta_string
01849
01850     def print_phylip(self, source_dict):
01851            # print phylip-formatted string from a dictionary
01852            taxa_list = list(source_dict.keys())
01853            no_taxa = len(taxa_list)
01854            # figure out the max length of a taxon for nice padding of sequences
01855            pad_longest_name = len(max(taxa_list, key=len)) + 3
01856            # get sequence length from a random value
01857            seq_length = len(next(iter(source_dict.values())))
01858            header = str(len(source_dict)) + " " + str(seq_length)
01859            phylip_string = header + "\n"
01860            for taxon, seq in sorted(source_dict.items()):
01861                taxon = taxon.replace(" ", "_").strip("'")
01862                # left-justify taxon names relative to sequences
01863                phylip_string += taxon.ljust(pad_longest_name, ' ') + seq + "\n"
01864
01865            return phylip_string
01866
01867     def print_phylip_int(self, source_dict):
01868            # print phylip interleaved-formatted string from a dictionary
01869            taxa_list = list(source_dict.keys())
01870            no_taxa = len(taxa_list)
01871            pad_longest_name = len(max(taxa_list, key=len)) + 3
01872            seq_length = len(next(iter(source_dict.values())))
01873            header = str(len(source_dict)) + " " + str(seq_length)
01874            phylip_int_string = header + "\n\n"
01875            # this will be a list of tuples to hold taxa names and sequences
01876            seq_matrix = []
01877
01878            # each sequence line will have 500 characters
01879            n = 500
01880
01881            # recreate sequence matrix
01882            add_to_matrix = seq_matrix.append
01883            for taxon, seq in sorted(source_dict.items()):
01884                add_to_matrix((taxon, [seq[i:i+n] for i in range(0, len(seq), n)]))
01885
01886            first_seq = seq_matrix[0][1]
01887            for index, item in enumerate(first_seq):
01888                for taxon, sequence in seq_matrix:
01889                    if index == 0:
01890                        phylip_int_string += taxon.ljust(pad_longest_name, ' ') + sequence[index] + "\n"
01891                    else:
01892                        phylip_int_string += sequence[index] + "\n"
01893                phylip_int_string += "\n"
01894
01895            return phylip_int_string
01896
01897     def print_nexus(self, source_dict):
01898            # print nexus-formatted string from a dictionary
01899            if self.data_type == "aa" or self.command == "translate":
01900                data_type = "PROTEIN"
01901            elif self.data_type == "dna":
01902                data_type = "DNA"
01903
01904            taxa_list = list(source_dict.keys())
01905            no_taxa = len(taxa_list)
01906            pad_longest_name = len(max(taxa_list, key=len)) + 3
01907            seq_length = len(next(iter(source_dict.values())))
01908            header = str(len(source_dict)) + " " + str(seq_length)
01909            nexus_string = (
01910                "#NEXUS\n\nBEGIN DATA;\n\tDIMENSIONS  NTAX=" + str(no_taxa) + " NCHAR=" + str(seq_length)
```

```
01911                    + ";\n\tFORMAT DATATYPE=" + data_type + "  GAP = - MISSING = ?;\n\tMATRIX\n"
01912                )
01913
01914            for taxon, seq in sorted(source_dict.items()):
01915                taxon = taxon.replace(" ", "_").strip("'")
01916                nexus_string += "\t" + taxon.ljust(pad_longest_name, ' ') + seq + "\n"
01917            nexus_string += "\n;\n\nEND;"
01918
01919            return nexus_string
01920
01921      def print_nexus_int(self, source_dict):
01922            # print nexus interleaved-formatted string from a dictionary
01923            if self.data_type == "aa":
01924                data_type = "PROTEIN"
01925            elif self.data_type == "dna":
01926                data_type = "DNA"
01927
01928            taxa_list = list(source_dict.keys())
01929            no_taxa = len(taxa_list)
01930            pad_longest_name = len(max(taxa_list, key=len)) + 3
01931            seq_length = len(next(iter(source_dict.values())))
01932            header = str(len(source_dict)) + " " + str(seq_length)
01933            # this will be a list of tuples to hold taxa names and sequences
01934            seq_matrix = []
01935            nexus_int_string = (
01936                "#NEXUS\n\nBEGIN DATA;\n\tDIMENSIONS  NTAX=" + str(no_taxa) + " NCHAR=" + str(seq_length)
01937                + ";\n\tFORMAT   INTERLEAVE" + "  DATATYPE=" + data_type + "  GAP = - MISSING =
       ?;\n\tMATRIX\n"
01938                )
01939            # each sequence line will have 500 characters
01940            n = 500
01941
01942            # recreate sequence matrix
01943            add_to_matrix = seq_matrix.append
01944            for taxon, seq in sorted(source_dict.items()):
01945                add_to_matrix((taxon, [seq[i:i+n] for i in range(0, len(seq), n)]))
01946
01947            first_seq = seq_matrix[0][1]
01948            for index, item in enumerate(first_seq):
01949                for taxon, sequence in seq_matrix:
01950                    if index == 0:
01951                        nexus_int_string += taxon.ljust(pad_longest_name, ' ') + sequence[index] + "\n"
01952                    else:
01953                        nexus_int_string += sequence[index] + "\n"
01954                nexus_int_string += "\n"
01955
01956            nexus_int_string += "\n;\n\nEND;"
01957
01958            return nexus_int_string
01959
01960      def natural_sort(self, a_list):
01961            # create a function that does 'human sort' on a list
01962            convert = lambda text: int(text) if text.isdigit() else text.lower()
01963            alphanum_key = lambda key: [convert(c) for c in re.split('([0-9]+)', key)]
01964            return sorted(a_list, key = alphanum_key)
01965
01966      def print_unspecified_partitions(self, data_type, codons):
01967            # print partitions for concatenated alignment
01968            part_string = ""
01969            part_dict = self.get_concatenated(self.parsed_alignments)[1]
01970            part_list = self.natural_sort(part_dict.keys())
01971
01972            if data_type == "dna":
01973                if codons == "none":
01974                    for key in part_list:
01975                        part_string += key + " = " + str(part_dict[key]) + "\n"
01976                elif codons == "12":
01977                    for key in part_list:
01978                        start, end = str(part_dict[key]).split("-")
01979                        part_string += key + "_pos1" + " = " + start + "-" + end + "\\2" + "\n"
01980                        part_string += key + "_pos2" + " = " + str(int(start) + 1) + "-" + end + "\\2" +
       "\n"
01981                elif codons == "123":
01982                    for key in part_list:
01983                        start, end = str(part_dict[key]).split("-")
01984                        part_string += key + "_pos1" + " = " + start + "-" + end + "\\3" + "\n"
01985                        part_string += key + "_pos2" + " = " + str(int(start) + 1) + "-" + end + "\\3" +
       "\n"
01986                        part_string += key + "_pos3" + " = " + str(int(start) + 2) + "-" + end + "\\3" +
       "\n"
01987
01988            elif data_type == "aa":
01989                for key in part_list:
01990                    part_string += key + " = " + str(part_dict[key]) + "\n"
01991
01992            return part_string
01993
```

```
01994    def print_nexus_partitions(self, data_type, codons):
01995        # print partitions for concatenated alignment
01996        part_string = ""
01997        part_dict = self.get_concatenated(self.parsed_alignments)[1]
01998        part_list = self.natural_sort(part_dict.keys())
01999        # write beginning of nexus sets
02000        part_string += "#NEXUS\n\n"
02001        part_string += "BEGIN SETS;\n"
02002
02003        if data_type == "dna":
02004            if codons == "none":
02005                for key in part_list:
02006                    part_string += "\tcharset " + key + " = " + str(part_dict[key]) + ";\n"
02007            elif codons == "12":
02008                for key in part_list:
02009                    start, end = str(part_dict[key]).split("-")
02010                    part_string += "\tcharset " + key + "_pos1" + " = " + start + "-" + end + "\\2" +
    ";\n"
02011                    part_string += "\tcharset " + key + "_pos2" + " = " + str(int(start) + 1) + "-" +
    end + "\\2" + ";\n"
02012            elif codons == "123":
02013                for key in part_list:
02014                    start, end = str(part_dict[key]).split("-")
02015                    part_string += "\tcharset " + key + "_pos1" + " = " + start + "-" + end + "\\3" +
    ";\n"
02016                    part_string += "\tcharset " + key + "_pos2" + " = " + str(int(start) + 1) + "-" +
    end + "\\3" + ";\n"
02017                    part_string += "\tcharset " + key + "_pos3" + " = " + str(int(start) + 2) + "-" +
    end + "\\3" + ";\n"
02018            part_string += "END;"
02019
02020        elif data_type == "aa":
02021            for key in part_list:
02022                part_string += "\tcharset " + key + " = " + str(part_dict[key]) + ";\n"
02023            part_string += "END;"
02024
02025        return part_string
02026
02027    def print_iqtree_nexus_partitions(self, data_type, codons):
02028        # print partitions for concatenated alignment
02029        part_string = ""
02030        part_dict = self.get_concatenated(self.parsed_alignments)[1]
02031        part_list = self.natural_sort(part_dict.keys())
02032        # write beginning of nexus sets
02033        part_string += "#nexus\n"
02034        part_string += "begin sets;\n"
02035
02036        if data_type == "dna":
02037            if codons == "none":
02038                for key in part_list:
02039                    part_string += "  charset " + key + " = " + str(part_dict[key]) + ";\n"
02040            elif codons == "12":
02041                for key in part_list:
02042                    start, end = str(part_dict[key]).split("-")
02043                    part_string += "  charset " + key + "_pos1" + " = " + start + " - " + end + "\\2"
    + ";\n"
02044                    part_string += "  charset " + key + "_pos2" + " = " + str(int(start) + 1) + " - "
    + end + "\\2" + ";\n"
02045            elif codons == "123":
02046                for key in part_list:
02047                    start, end = str(part_dict[key]).split("-")
02048                    part_string += "  charset " + key + "_pos1" + " = " + start + " - " + end + "\\3"
    + ";\n"
02049                    part_string += "  charset " + key + "_pos2" + " = " + str(int(start) + 1) + " - "
    + end + "\\3" + ";\n"
02050                    part_string += "  charset " + key + "_pos3" + " = " + str(int(start) + 2) + " - "
    + end + "\\3" + ";\n"
02051            part_string += "end;\n"
02052
02053        elif data_type == "aa":
02054            for key in part_list:
02055                part_string += "  charset " + key + " = " + str(part_dict[key]) + ";\n"
02056            part_string += "end;\n"
02057
02058        return part_string
02059
02060    def print_raxml_partitions(self, data_type, codons):
02061        # print partitions for concatenated alignment
02062        part_string = ""
02063        part_dict = self.get_concatenated(self.parsed_alignments)[1]
02064        part_list = self.natural_sort(part_dict.keys())
02065
02066        if data_type == "dna":
02067            if codons == "none":
02068                for key in part_list:
02069                    part_string += "DNA, " + key + " = " + str(part_dict[key]) + "\n"
02070            elif codons == "12":
```

```
02071                    for key in part_list:
02072                        start, end = str(part_dict[key]).split("-")
02073                        part_string += "DNA, " + key + "_pos1" + " = " + start + "-" + end + "\\2" + "\n"
02074                        part_string += "DNA, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end +
       "\\2" + "\n"
02075                elif codons == "123":
02076                    for key in part_list:
02077                        start, end = str(part_dict[key]).split("-")
02078                        part_string += "DNA, " + key + "_pos1" + " = " + start + "-" + end + "\\3" + "\n"
02079                        part_string += "DNA, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end +
       "\\3" + "\n"
02080                        part_string += "DNA, " + key + "_pos3" + " = " + str(int(start) + 2) + "-" + end +
       "\\3" + "\n"
02081
02082            elif data_type == "aa":
02083                for key in part_list:
02084                    part_string += "WAG, " + key + " = " + str(part_dict[key]) + "\n"
02085
02086                #  aa-partition files with strides are probably not useful? (original below)
02087 #             elif codons == "12":
02088 #                 for key in part_list:
02089 #                     start, end = str(part_dict[key]).split("-")
02090 #                     part_string += "WAG, " + key + "_pos1" + " = " + start + "-" + end + "\\2" + "\n"
02091 #                     part_string += "WAG, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end
       + "\\2" + "\n"
02092 #             elif codons == "123":
02093 #                 for key in part_list:
02094 #                     start, end = str(part_dict[key]).split("-")
02095 #                     part_string += "WAG, " + key + "_pos1" + " = " + start + "-" + end + "\\3" + "\n"
02096 #                     part_string += "WAG, " + key + "_pos2" + " = " + str(int(start) + 1) + "-" + end
       + "\\3" + "\n"
02097 #                     part_string += "WAG, " + key + "_pos3" + " = " + str(int(start) + 2) + "-" + end
       + "\\3" + "\n"
02098            return part_string
02099
02100    def replace_string_in_file(self, file_name, old_string, new_string):
02101        # global string replacement in file
02102        with open(file_name, "r", encoding="utf-8") as file:
02103            file_content = file.read()
02104        # write globally replaced content back to file
02105        glb_replaced_content = file_content.replace(old_string, new_string)
02106        with open(file_name, "w", encoding="utf-8") as file:
02107            file.write(glb_replaced_content)
02108
02109    def write_partitions(self, file_name, part_format, data_type, codons):
02110        # write partitions file for concatenated alignment
02111        self.file_overwrite_error(file_name)
02112        with open(file_name, "w", encoding="utf-8") as part_file:
02113            if part_format == "nexus":
02114                part_file.write(self.print_nexus_partitions(data_type, codons))
02115            if part_format == "iqtree-nexus":
02116                part_file.write(self.print_iqtree_nexus_partitions(data_type, codons))
02117            if part_format == "raxml":
02118                part_file.write(self.print_raxml_partitions(data_type, codons))
02119            if part_format == "unspecified":
02120                part_file.write(self.print_unspecified_partitions(data_type, codons))
02121
02122            if self.using_metapartitions:
02123                self.replace_string_in_file(file_name, '-meta =', ' =')
02124
02125        print("Wrote partitions for the concatenated file to '" + file_name + "'")
02126
02127    def get_extension(self, file_format):
02128        # get proper extension string
02129        if file_format == "phylip":
02130            extension = "-out.phy"
02131        elif file_format == "phylip-int":
02132            extension = "-out.int-phy"
02133        elif file_format == "fasta":
02134            extension = "-out.fas"
02135        elif file_format == "nexus":
02136            extension = "-out.nex"
02137        elif file_format == "nexus-int":
02138            extension = "-out.int-nex"
02139
02140        return extension
02141
02142    def get_metapartition_extension(self, file_format):
02143        # get proper metapartition_extension string
02144        if file_format == "phylip":
02145            metapartition_extension = "-meta.phy"
02146        elif file_format == "phylip-int":
02147            metapartition_extension = "-meta.int-phy"
02148        elif file_format == "fasta":
02149            metapartition_extension = "-meta.fas"
02150        elif file_format == "nexus":
02151            metapartition_extension = "-meta.nex"
```

```
02152            elif file_format == "nexus-int":
02153                metapartition_extension = "-meta.int-nex"
02154
02155            return metapartition_extension
02156
02157        def file_overwrite_error(self, file_name):
02158            # print warning when overwriting a file
02159            if path.exists(file_name):
02160                print("WARNING: You are overwriting '" + file_name + "'")
02161
02162        def write_formatted_file(self, file_format, file_name, alignment):
02163            # write the correct format string into a file
02164            with open(file_name, "w", encoding="utf-8") as out_file:
02165                if file_format == "phylip":
02166                    out_file.write(self.print_phylip(alignment))
02167                elif file_format == "fasta":
02168                    out_file.write(self.print_fasta(alignment))
02169                elif file_format == "phylip-int":
02170                    out_file.write(self.print_phylip_int(alignment))
02171                elif file_format == "nexus":
02172                    out_file.write(self.print_nexus(alignment))
02173                elif file_format == "nexus-int":
02174                    out_file.write(self.print_nexus_int(alignment))
02175
02176        def get_alignment_name(self, i, extension):
02177            # get file name
02178            file_name = self.alignment_objects[i].get_name() + extension
02179
02180            return file_name
02181
02182        def get_alignment_name_no_ext(self, i):
02183            # get file name without extension
02184            file_name = self.alignment_objects[i].get_name()
02185
02186            return file_name
02187
02188        def write_concat(self, file_format):
02189            # write concatenated alignment into a file
02190            concatenated_alignment = self.get_concatenated(self.parsed_alignments)[0]
02191            file_name = self.concat_out
02192            self.file_overwrite_error(file_name)
02193            self.write_formatted_file(file_format, file_name, concatenated_alignment)
02194
02195            print("Wrote concatenated sequences to " + file_format + " file '" + file_name + "'")
02196
02197        def write_convert(self, index, alignment, file_format, extension):
02198            # write converted alignment into a file
02199            file_name = self.get_alignment_name(index, extension)
02200            self.file_overwrite_error(file_name)
02201            self.write_formatted_file(file_format, file_name, alignment)
02202
02203        def write_replicate(self, index, alignment, file_format, extension):
02204            # write replicate alignment into a file
02205            file_name = "replicate" + str(index + 1) + "_" + str(self.no_loci) + "-loci" + extension
02206            self.file_overwrite_error(file_name)
02207            self.write_formatted_file(file_format, file_name, alignment)
02208
02209        def write_split(self, item, file_format, extension):
02210            # write split alignments from partitions file
02211            # bad practice with the dicts; figure out better solution
02212            partition_name = list(item.keys())[0]
02213            alignment = item[partition_name]
02214
02215            if not alignment:
02216                # If the alignment dict is empty, i.e. no alignment associated with partition name, raise
        error
02217                raise ValueError("Partition '%s' is empty. No sequences to write." % partition_name)
02218
02219            # implementation of option --no-san (don't prepend input superalignment filename to the
        `split` outputs)
02220            if self.no_sup_aln_name:
02221                file_name = partition_name + extension
02222            else:
02223                file_name = str(self.in_files[0].split('.')[0]) + "_" + partition_name + extension
02224
02225            try:
02226                self.file_overwrite_error(file_name)
02227                self.write_formatted_file(file_format, file_name, alignment)
02228                yield file_name
02229            except ValueError as e:
02230                print("There was an issue writing file '%s': %s" % (file_name, str(e)))
02231                remove(file_name)
02232                raise
02233
02234        def write_reduced(self, file_format, extension):
02235            # write alignment with taxa removed into a file
02236            prefix = self.reduced_file_prefix
```

```
02237            alns = self.remove_taxa(self.species_to_remove)
02238            for file_name, aln_dict in alns.items():
02239                out_file_name = prefix + file_name + extension
02240                self.file_overwrite_error(out_file_name)
02241                self.write_formatted_file(file_format, out_file_name, aln_dict)
02242            return len(alns)
02243
02244       def write_translated(self, index, alignment, file_format, extension):
02245            # write alignments translated into amino acids
02246            prefix = "translated_"
02247            file_name = self.get_alignment_name(index, extension)
02248            out_file_name = prefix + file_name + extension
02249            self.file_overwrite_error(out_file_name)
02250            self.write_formatted_file(file_format, out_file_name, alignment)
02251
02252       def write_trimmed(self, index, alignment, file_format, extension):
02253            # write trimmed alignments
02254            if self.trim_out:
02255                out_file_name = self.trim_out
02256            else:
02257                prefix = "trimmed_"
02258                file_name = self.get_alignment_name(index, extension)
02259                out_file_name = prefix + file_name
02260            self.file_overwrite_error(out_file_name)
02261            self.write_formatted_file(file_format, out_file_name, alignment)
02262
02263       def write_metapartitions(self, file_format):
02264            # write metapartitions - combines split and concat
02265            print("write_out elif action == metapartitions")
02266            metapartition_extension = self.get_metapartition_extension(file_format)
02267            list_of_alignments = self.get_partitioned(self.split)
02268            written_split_files = []
02269            err_indx = 0
02270
02271            for item in list_of_alignments:
02272                try:
02273                    for split_file in self.write_split(item, file_format, metapartition_extension):
02274                        written_split_files.append(split_file)
02275                except ValueError as e:
02276                    print("WARNING: ", e)
02277                    err_indx += 1
02278            if len(written_split_files) > 0:
02279                print("Wrote %d %s metapartition files from partitions provided" %
        (len(written_split_files), file_format))
02280            if err_indx > 0:
02281                print("WARNING: %d file(s) raised an error while writing (see above)." % err_indx)
02282
02283            # now set inputs to be the collated metapartition alignment files
02284            self.in_files = written_split_files
02285            self.alignment_objects = self.get_alignment_objects()
02286            self.parsed_alignments = self.get_parsed_alignments()
02287
02288            # concat metapartition alignment files
02289            self.write_concat(file_format)
02290
02291       def write_out(self, action, file_format):
02292            # write other output files depending on command (action)
02293            extension = self.get_extension(file_format)
02294
02295            if action == "concat":
02296                self.write_concat(file_format)
02297
02298            elif action == "convert":
02299                length = len(self.alignment_objects)
02300                [
02301                    self.write_convert(i, alignment, file_format, extension)
02302                    for i, alignment in enumerate(self.parsed_alignments)
02303                ]
02304                print("Converted " + str(length) + " files from " + self.in_format + " to " + file_format)
02305
02306            elif action == "replicate":
02307                [
02308                    self.write_replicate(i, alignment, file_format, extension)
02309                    for i, alignment in enumerate(self.get_replicate(self.no_replicates, self.no_loci))
02310                ]
02311
02312                print("Constructed " + str(self.no_replicates) + " replicate data sets, each from " +
        str(self.no_loci) + " alignments")
02313
02314            elif action == "split":
02315                list_of_alignments = self.get_partitioned(self.split)
02316                written_split_files = []
02317                err_indx = 0
02318
02319                for item in list_of_alignments:
02320                    try:
02321                        for split_file in self.write_split(item, file_format, extension):
```

```
02322                             written_split_files.append(split_file)
02323                     except ValueError as e:
02324                             print("WARNING: ", e)
02325                             err_indx += 1
02326                 if len(written_split_files) > 0:
02327                     print("Wrote %d %s files from partitions provided" % (len(written_split_files),
        file_format))
02328                 if err_indx > 0:
02329                     print("WARNING: %d file(s) raised an error while writing (see above)." % err_indx)
02330
02331         elif action == "metapartitions":
02332             self.write_metapartitions(file_format)
02333
02334         elif action == "remove":
02335             aln_no = self.write_reduced(file_format, extension)
02336             if aln_no:
02337                 print("Wrote " + str(aln_no) + " " + str(file_format) + " files with reduced taxon
        set")
02338
02339         elif action == "translate":
02340             if self.data_type == "aa":
02341                 print("ERROR: cannot translate; you said your alignment already contains amino acids")
02342                 sys.exit()
02343             translated_alignment_dicts = self.get_translated(self.genetic_code, self.reading_frame)
02344             length = len(self.alignment_objects)
02345             [
02346                 self.write_translated(i, alignment, file_format, extension)
02347                 for i, alignment in enumerate(translated_alignment_dicts)
02348             ]
02349             print("Translated " + str(length) + " files to amino acid sequences")
02350
02351         elif action == "trim": # self.trim_fraction, self.parsimony_check
02352             trimmed_alignment_dicts = self.get_trimmed(self.trim_fraction, self.parsimony_check)
02353             length = len(self.alignment_objects)
02354             [
02355                 self.write_trimmed(i, alignment, file_format, extension)
02356                 for i, alignment in enumerate(trimmed_alignment_dicts)
02357             ]
02358             print("Trimmed", str(length), "file(s) to have", self.trim_fraction, "minimum occupancy
        per alignment column")
02359
02360
02361 def main():
02362
02363     # initialize parsed arguments and meta alignment objects
02364     kwargs = run()
02365     meta_aln = MetaAlignment(**kwargs)
02366
02367     if meta_aln.command == "summary":
02368         meta_aln.write_summaries(kwargs["summary_out"])
02369     if meta_aln.by_taxon_summary:
02370         print("Printing taxon summaries")
02371         meta_aln.write_taxa_summaries()
02372     if meta_aln.command == "convert":
02373         meta_aln.write_out("convert", kwargs["out_format"])
02374     if meta_aln.command == "concat":
02375         meta_aln.write_out("concat", kwargs["out_format"])
02376         meta_aln.write_partitions(kwargs["concat_part"], kwargs["part_format"], kwargs["data_type"],
        kwargs["codons"])
02377     if meta_aln.command == "replicate":
02378         meta_aln.write_out("replicate", kwargs["out_format"])
02379     if meta_aln.command == "split":
02380         meta_aln.write_out("split", kwargs["out_format"])
02381     if meta_aln.command == "remove":
02382         meta_aln.write_out("remove", kwargs["out_format"])
02383     if meta_aln.command == "translate":
02384         meta_aln.write_out("translate", kwargs["out_format"])
02385     if meta_aln.command == "trim":
02386         meta_aln.write_out("trim", kwargs["out_format"])
02387
02388     if meta_aln.command == "metapartitions":
02389         # `metapartitions` is essentially `split` + `concat`. Currently you can't set an out_format:
02390         # it's automatically set to match the in_format because the intermediate `split` outputs
        become
02391         # the 'new' in_files for the `concat` operation, and then calling either:
02392         #   -> AminoAcidAlignment(Alignment.__init__(self, in_file, in_format, data_type))
02393         #   -> DNAAlignment(Alignment.__init__(self, in_file, in_format, data_type))
02394         # through MetaAlignment.get_alignment_object(alignment, self.in_format, self.data_type)
02395         meta_aln.write_out("metapartitions", kwargs["in_format"])
02396         meta_aln.write_partitions(kwargs["concat_part"], kwargs["part_format"], kwargs["data_type"],
        "none")
02397
02398         # meta_aln.write_out("translate", kwargs["out_format"])
02399
02400 def run():
02401
02402     # initialize parsed arguments
```

```
02403    config = ParsedArgs()
02404    # get arguments
02405    config_dict = config.get_args_dict()
02406    return config_dict
02407
02408 if __name__ == '__main__':
02409    main()
```

8.5 md/README.md File Reference

```
02408 if __name__ == '__main__':
02409    main()
```

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