PreLab 5: Using PCA for Analysis of Time Series Data

Jared Gridley

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Mouse cerebral cortex analysis

** This prelab will help you prepare for your group mini-project that will be created for you and has been posted to LMS prior. Please be sure to come to the lab sessions assigned to your team. There are 5 groups per Lab. All the coding has been done for you. You just have to read 'Prelab5.pdf' and do the quiz. Please answer the quiz using "~/MATP-4400/Lab5/Prelab5.pdf" or equivalently 'Prelab5.pdf' version uploaded to LMS. This will make sure your clusters match the clusters used to prepare the lab. **

Scott the Scientist did an analysis of RNA-Seq data from the development of the mouse cortex at days -8, -4, 0, 1, 7, 16, 21, and 26 taken from the Allen Brain Atlas Developing Mouse Portal http://developingmouse.brain-map.org/. This type of data is know as time series data since the features are taken through time. Here is his preliminary report; your job is to help Scott understand the results!

Preparation of the Mouse Homologs Data

Scott begins by reading in the dataset and preparing the data frame. The columns are days at which the samples are collected. The entries in the columns are the amount of RNA for each gene detected on that day in the mouse embryo cerebral cortex. We can use summary() and see that the column mean in not 0. This data has already been scaled, so each row has $mean\ 0$ and $sd\ 1$. This is different from prior labs where we scaled by column. Row scaling has been done so that the analysis can focus on the shape of the time series rather than specific magnitudes. Scott confirmed the scaling was successful by calculating the row means and making sure their norm was near 0.

Here is the mapping of the columns to their actual meanings:

- DayNeg8 = 8 days before birth
- DayNeg4 = 4 days before birth
- Day0 = day of birth
- DayPos1 = 1 day after birth
- DayPos7 = 7 days after birth
- DayPos16 = 16 days after birth
- DayPos21 = 21 days after birth
- DayPos28 = 28 days after birth

```
# Read in the data and create a dataframe
# We read in the csv indicating that we have row names.
Mouse.df <-read.csv("~/MATP-4400/data/MouseHomologData.csv", row.names = 1)
# Use shorter column names
colnames(Mouse.df)<-c("-8","-4","0","1","7","16","21","28")
# Create a matrix for our analysis
Mouse.matrix <- as.matrix(Mouse.df)</pre>
```

```
# Summarize; note the scaling
summary(Mouse.df)
##
          -8
                              -4
                                                 0
                                                                    1
##
    Min.
           :-2.2436
                               :-2.3882
                                                  :-2.3330
                                                                     :-2.02584
                       Min.
                                          Min.
                                                              Min.
##
    1st Qu.:-0.9138
                       1st Qu.:-1.0365
                                           1st Qu.:-0.5185
                                                              1st Qu.:-0.52867
##
    Median :-0.1530
                       Median :-0.5435
                                          Median : 0.1894
                                                              Median :-0.07383
##
    Mean
           : 0.2260
                       Mean
                               :-0.3335
                                                  : 0.3118
                                                                     : 0.13705
                                          Mean
                                                              Mean
##
    3rd Qu.: 1.4918
                       3rd Qu.: 0.3735
                                          3rd Qu.: 1.0578
                                                              3rd Qu.: 0.76547
           : 2.4749
##
                               : 2.4710
                                                  : 2.4739
                                                                     : 2.47368
    Max.
                       Max.
                                          Max.
                                                              Max.
##
          7
                               16
                                                  21
                                                                     28
##
   Min.
           :-1.58200
                        Min.
                                :-1.7886
                                           Min.
                                                   :-1.7495
                                                               Min.
                                                                       :-2.04906
##
   1st Qu.:-0.44628
                        1st Qu.:-0.7918
                                           1st Qu.:-0.7839
                                                               1st Qu.:-0.75671
   Median :-0.04786
                        Median :-0.4784
                                           Median :-0.4840
                                                               Median :-0.44015
##
##
    Mean
           : 0.18491
                                :-0.2320
                                           Mean
                                                   :-0.1980
                                                               Mean
                                                                       :-0.09629
                        Mean
##
    3rd Qu.: 0.71298
                        3rd Qu.: 0.3390
                                           3rd Qu.: 0.4180
                                                               3rd Qu.: 0.51383
   Max.
           : 2.45886
                        Max.
                                : 2.4749
                                           Max.
                                                   : 2.4749
                                                               Max.
                                                                      : 2.47487
# Demonstrate the scaling by viewing the norm
# norm(rowMeans(Mouse.matrix))
```

Cluster and PCA Analysis

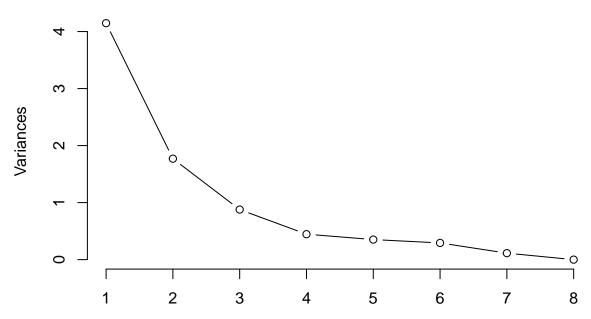
We used Kmeans to create five clusters based on domain knowledge: biologists believe there are five stages of brain development, so we select five clusters. Examining the plot of the kmean objective by cluster size using the "elbow test" suggests that a smaller number of clusters *could* be used; you can verify this on your own. But as we will see, five clusters proves to be an appropriate number for this analysis.

```
set.seed(300)
km <-kmeans(Mouse.matrix, 5)</pre>
```

We visualize the cluster using a biplot of two components generated by PCA which explain 74% of the variance. The scree plot suggests that PC3 might also contain significant variance.

```
# Calculate the PCA
my.pca <- prcomp(Mouse.matrix, retx=TRUE, center=TRUE, scale=TRUE)
# Summarize, to see the complete PCA result
summary(my.pca)
## Importance of components:
##
                             PC1
                                    PC2
                                           PC3
                                                    PC4
                                                            PC5
                                                                    PC6
                                                                            PC7
                          2.0364 1.3306 0.9370 0.66801 0.59296 0.54132 0.33712
## Standard deviation
## Proportion of Variance 0.5184 0.2213 0.1098 0.05578 0.04395 0.03663 0.01421
## Cumulative Proportion 0.5184 0.7397 0.8494 0.90521 0.94917 0.98579 1.00000
                                PC8
                          3.306e-11
## Standard deviation
## Proportion of Variance 0.000e+00
## Cumulative Proportion
                          1.000e+00
# Generate a scree plot
screeplot(my.pca, type = "lines",
          main = 'Explained Variance of Mouse Genes')
```

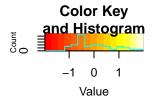
Explained Variance of Mouse Genes



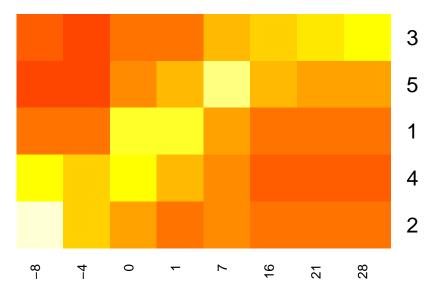
Examining the heatmap of the Kmeans cluster centers, we can see that each cluster corresponds to different average peaks of gene expressions. A line plot is also a very effective way to view the means as time-series.

```
## Warning in plot.window(...): "alpha" is not a graphical parameter
## Warning in plot.xy(xy, type, ...): "alpha" is not a graphical parameter
```

Warning in title(...): "alpha" is not a graphical parameter



Kmeans Cluster Centers



We can also see the time trends in the clusters means by plotting each cluster mean as a line. The cluster means have to be reformatted into a data frame with columns 'Cluster', 'Day' and 'Mean'. This is done using the 'dplyr' package 'gather' command. The factors are also recoded to look nice on the plot.

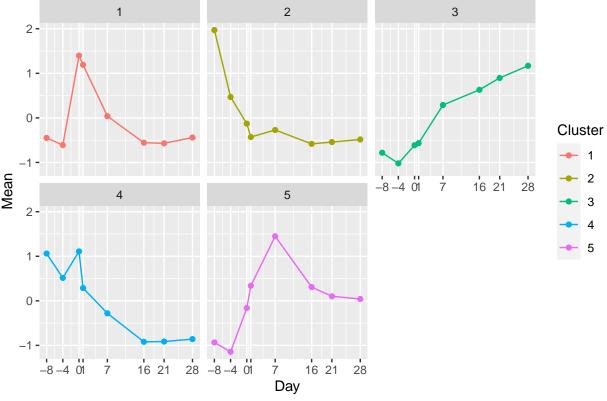
We use ggplot with geom_line to make the plots. Note how the 'facet_grid()

```
tics<-c(-8,-4,0,1,7,16,21,28) # x-axis "tics" (for the plot)
clustermean<-km$centers # Extract the cluster means from km
# Set up qqplot-based line plot
# We must "pivot" at dataframe version of clustermean (ie convert from "wide" to "long")
# Rows are our groups
clustermean.df <- as.data.frame(clustermean, row.names = c("1","2","3","4","5"))</pre>
# Tidyverse pipeline
# This is making a data frame of the form
# Cluster Day Mean
clustermeanlong.df <- clustermean.df %>%
  rownames_to_column("Cluster") %>% # Make a new column called Cluster
  gather(key="Day", value="Mean", -Cluster) %>% # Make a skinny data frame
  #Recode the factors to have short names
 # mutate(Day=recode(Day, "DayNeg8"="-8", "DayNeg4"="-4", "DayO"="0", "DayPos1"= "1",
         "DayPos7"= "7", "DayPos16"= "16", "DayPos21"= "21", "DayPos28"= "28"))%>%
  # convert Day to an integer
 convert(int(Day))
# see what data frame looks like.,
kable(head(clustermeanlong.df))
```

Cluster	Day	Mean
1	-8	-0.4496261
2	-8	1.9712955
3	-8	-0.7822509
4	-8	1.0614302
5	-8	-0.9336692
1	-4	-0.6106791

```
#Plot the mean of each cluster in a separate graph":
ggplot(clustermeanlong.df,aes(x=Day, y=Mean, col=Cluster)) +
geom_line() +
geom_point() +
scale_x_continuous(breaks=tics) +
labs(title="Cluster Centers") +
# Use facet_wrap to make a separate plot for each cluster
facet_wrap(Cluster ~.)
```

Cluster Centers



Take a look, what patterns do you see in the clusters?

Exercise 1

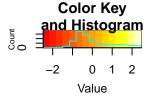
For each cluster mean, identify the peak, i.e. the day with the highest average value. Cluster 5 has been completed for you.

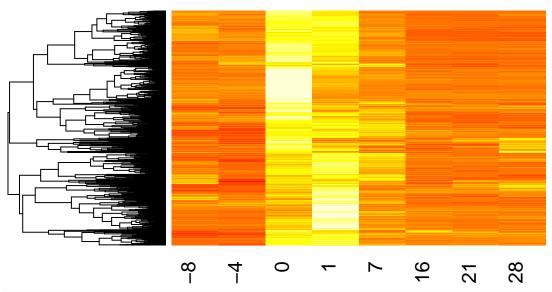
- Cluster 1 = Day 0
- Cluster 2 = Day -8
- Cluster 3 = Day 28

```
• Cluster 4 = Day 0
```

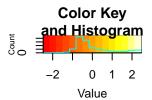
Exercise 2

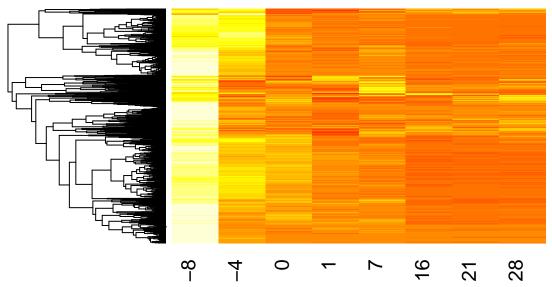
Make a separate heat map for each of the five clusters. Plot the cluster heatmaps in the order they occur in development. **Do not scale the heatmaps.** Cluster the genes but not the days. An example of how to plot Cluster 5 using heatmap is provided. How do the heatmaps match your results for exercise 1?

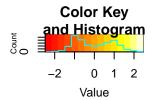


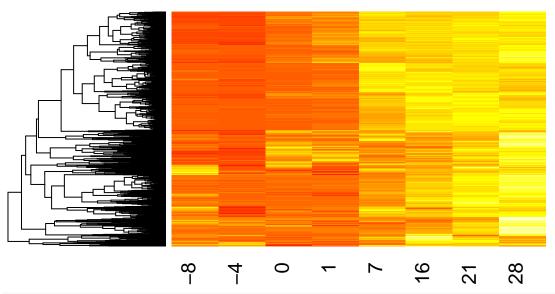


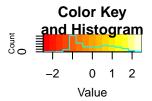
[•] Cluster 5 = Day 7

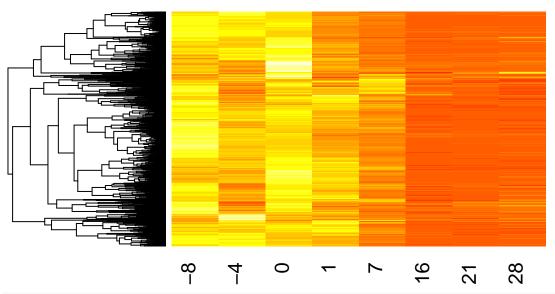


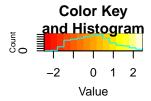


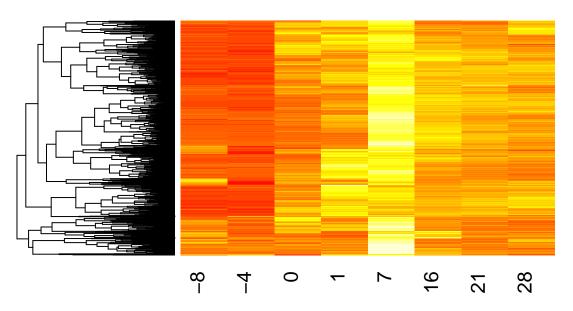








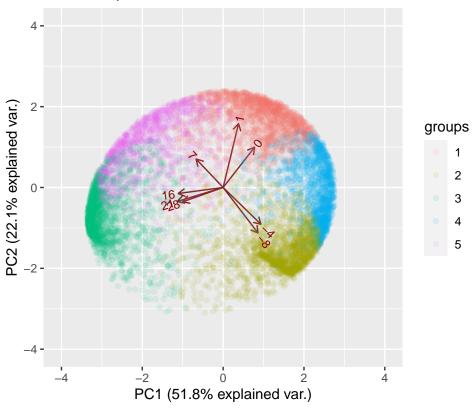




Visualization by biplot

We display the points in a biplot. The projection of the points makes an interesting disc-type shape which is less dense in the middle, like a donut. We can see that the clusters are arranged in time order around the disc. Why do we see this?

Mouse Biplot for PC1 and PC2



Exercise 3

We hypothesize that each cluster of genes represents one of the five "stages" of brain development labeled A,B,C,D,E. Assign each cluster to its corresponding stage using the biplot. For example, Cluster 5 represents Stage D of development since it peaks at the first at the point Day7.

- Stage A = Cluster 2
- Stage B = Cluster 1 (Can also be 4 if not allowing for duplicates, but matches 1 better)
- Stage C = Cluster 1
- Stage D = Cluster 5
- Stage E = Cluster 3

Exercise 4

Examine the scalar projections of the coordinate axes in the biplot, for Days -8, -4, 0, 1, 7, 16, 21 and 28. Notice that the coordinate vectors act as hours on a "developmental time clock" that starts at Day -8. Does time on this development clock run clockwise or counterclockwise? Counterclockwise

You've now completed in class Prelab5! Go to LMS and complete the online quiz.

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