

# [Tutorial] Gene expression profiles in cancer patients

## 1. Introduction

Lung cancer one of major cancers, accounting for 2.09 million deaths out of the 9.6 million total cancer deaths in 2018. There are two main types of lung cancer: small cell lung carcinoma (SCLC) and non-small cell lung carcinoma (NSCLC). NSCLC is responsible for 85–90% of lung cancer cases and its two largest subtypes are lung adenocarcinoma (LUAD) and lung squamous cell carcinoma (LUSC). LUAD develops in the periphery of the lungs and may be associated with smoking, but is the most common lung cancer type among non-smokers. In contrast, LUSC accounts for 25–30% of all total lung cancer cases while LUAD accounts for 40% of all total lung cancer cases. LUSC are likely to be found in the middle of the lungs and is associated with smoking. In this tutorial, we will explore the sample dataset of LUSC and LUAD from the The Cancer Genome Atlas (TCGA), a public resource for the genomic dataset. Here we use two types of lung cancers - LUAD and LUSC and will examine which information we can use for genomic analyses of lung cancers.

## 2. Obtain the gene expression profile dataset

To access the gene expression data, there are two ways

1. Formal way (but slow): you can download the data as described in 2.1
2. Simple ways: get the file from my dropbox link where I downloaded and save to R object as described in 2.2

### 2.1. Download the data from GDC portal

You will need to install the TCGAbiolinks package to obtain the lung cancer gene expression profiles from TCGA.

```
library(tidyverse)
```

```
## -- Attaching packages ----- tidyverse 1.3.1 --
```

```
## v ggplot2 3.3.5      v purrr   0.3.4
## v tibble  3.1.4      v dplyr  1.0.7
## v tidyr   1.1.3      v stringr 1.4.0
## v readr   2.0.1      v forcats 0.5.1
```

```
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
```

```
# Install the TCGAbiolinks package if necessary
#if (!requireNamespace("BiocManager", quietly = TRUE))
#  install.packages("BiocManager")
#BiocManager::install("TCGAbiolinks")

library(TCGAbiolinks)
```

Then, Obtain the TCGA dataset from the GDC portal. You first download the gene expression dataset for Lung Adenocarcinoma (LUAD).

```
#query <-
#  GDCquery(project = "TCGA-LUAD",
#           data.category = "Transcriptome #Profiling",
#           data.type = "Gene Expression #Quantification",
#           workflow.type = "HTSeq - FPKM-UQ")
#GDCdownload(query)
#d_luad0 <- GDCprepare(query)
#d_luad = as.data.frame(d_luad0@colData)
```

Now we are downloading the dataset for Lung Squamous Cell Carcinoma (LUSC).

```
#query <- GDCquery(project = "TCGA-LUSC",
#                  data.category = "Transcriptome #Profiling",
#                  data.type = "Gene Expression #Quantification",
#                  workflow.type = "HTSeq - FPKM-UQ")
#GDCdownload(query)
#d_lusc0 <- GDCprepare(query)
#d_lusc = as.data.frame(d_lusc0@colData)
```

For simplicity, we are choosing only genes on the chromosome 1. Please save to the Rdata into your working folder.

```
library(SummarizedExperiment)
```

```
##           : MatrixGenerics
```

```
##           : matrixStats
```

```
##
##           : 'matrixStats'
```

```
## The following object is masked from 'package:dplyr':
```

```
##
```

```
##      count
```

```
##
```

```
##           : 'MatrixGenerics'
```

```
## The following objects are masked from 'package:matrixStats':
```

```
##
```

```

##      colAlls, colAnyNAs, colAnys, colAvgPerRowSet, colCollapse,
##      colCounts, colCummaxs, colCummins, colCumprods, colCumsums,
##      colDiffs, colIQRDiffs, colIQRs, colLogSumExps, colMadDiffs,
##      colMads, colMaxs, colMeans2, colMedians, colMins, colOrderStats,
##      colProds, colQuantiles, colRanges, colRanks, colSdDiffs, colSds,
##      colSums2, colTabulates, colVarDiffs, colVars, colWeightedMads,
##      colWeightedMeans, colWeightedMedians, colWeightedSds,
##      colWeightedVars, rowAlls, rowAnyNAs, rowAnys, rowAvgPerColSet,
##      rowCollapse, rowCounts, rowCummaxs, rowCummins, rowCumprods,
##      rowCumsums, rowDiffs, rowIQRDiffs, rowIQRs, rowLogSumExps,
##      rowMadDiffs, rowMads, rowMaxs, rowMeans2, rowMedians, rowMins,
##      rowOrderStats, rowProds, rowQuantiles, rowRanges, rowRanks,
##      rowSdDiffs, rowSds, rowSums2, rowTabulates, rowVarDiffs, rowVars,
##      rowWeightedMads, rowWeightedMeans, rowWeightedMedians,
##      rowWeightedSds, rowWeightedVars

##      : GenomicRanges

##      : stats4

##      : BiocGenerics

##      : parallel

##
##      : 'BiocGenerics'

## The following objects are masked from 'package:parallel':
##
##      clusterApply, clusterApplyLB, clusterCall, clusterEvalQ,
##      clusterExport, clusterMap, parApply, parCapply, parLapply,
##      parLapplyLB, parRapply, parSapply, parSapplyLB

## The following objects are masked from 'package:dplyr':
##
##      combine, intersect, setdiff, union

## The following objects are masked from 'package:stats':
##
##      IQR, mad, sd, var, xtabs

## The following objects are masked from 'package:base':
##
##      anyDuplicated, append, as.data.frame, basename, cbind, colnames,
##      dirname, do.call, duplicated, eval, evalq, Filter, Find, get, grep,
##      grepl, intersect, is.unsorted, lapply, Map, mapply, match, mget,
##      order, paste, pmax, pmax.int, pmin, pmin.int, Position, rank,
##      rbind, Reduce, rownames, sapply, setdiff, sort, table, tapply,
##      union, unique, unsplit, which.max, which.min

##      : S4Vectors

```

```

##
##           : 'S4Vectors'

## The following objects are masked from 'package:dplyr':
##
##     first, rename

## The following object is masked from 'package:tidyr':
##
##     expand

## The following objects are masked from 'package:base':
##
##     expand.grid, I, unname

##           : IRanges

##
##           : 'IRanges'

## The following objects are masked from 'package:dplyr':
##
##     collapse, desc, slice

## The following object is masked from 'package:purrr':
##
##     reduce

## The following object is masked from 'package:grDevices':
##
##     windows

##           : GenomeInfoDb

##           : Biobase

## Welcome to Bioconductor
##
##     Vignettes contain introductory material; view with
##     'browseVignettes()'. To cite Bioconductor, see
##     'citation("Biobase")', and for packages 'citation("pkgname)".

##
##           : 'Biobase'

## The following object is masked from 'package:MatrixGenerics':
##
##     rowMedians

## The following objects are masked from 'package:matrixStats':
##
##     anyMissing, rowMedians

```

```
#e_luad = assay(d_luad0)
#e_lusc = assay(d_lusc0)

#g_luad = d_luad0[rowRanges %>% as.data.frame() %>% filter(seqnames=='chr1') %>% pull(ensembl_gene_id)
#g_lusc = d_lusc0[rowRanges %>% as.data.frame() %>% filter(seqnames=='chr1') %>% pull(ensembl_gene_id)
#e_luad = e_luad[g_luad,]
#e_lusc = e_lusc[g_lusc,]

#save(d_luad, d_lusc, e_luad, e_lusc, file='data.TCGA_LUAD_LUSC.gene_expression.Rdata')
```

## 2.2. Get the file from the link

You can download the data from this link. Please save this file to the working folder. Then, load the objects to your workspace.

```
load('data.TCGA_LUAD_LUSC.gene_expression.Rdata')
```

## 3. Explore the input dataset

Let's explore which columns are provided in the LUAD dataset.

```
colnames(d_luad)
```

```
## [1] "barcode"
## [2] "patient"
## [3] "sample"
## [4] "shortLetterCode"
## [5] "definition"
## [6] "sample_submitter_id"
## [7] "sample_type_id"
## [8] "sample_id"
## [9] "sample_type"
## [10] "days_to_collection"
## [11] "state"
## [12] "initial_weight"
## [13] "intermediate_dimension"
## [14] "pathology_report_uuid"
## [15] "submitter_id"
## [16] "shortest_dimension"
## [17] "oct_embedded"
## [18] "longest_dimension"
## [19] "is_ffpe"
## [20] "tissue_type"
## [21] "synchronous_malignancy"
## [22] "ajcc_pathologic_stage"
## [23] "tumor_stage"
## [24] "days_to_diagnosis"
## [25] "treatments"
## [26] "last_known_disease_status"
## [27] "tissue_or_organ_of_origin"
## [28] "days_to_last_follow_up"
```

```

## [29] "age_at_diagnosis"
## [30] "primary_diagnosis"
## [31] "prior_malignancy"
## [32] "year_of_diagnosis"
## [33] "prior_treatment"
## [34] "ajcc_staging_system_edition"
## [35] "ajcc_pathologic_t"
## [36] "morphology"
## [37] "ajcc_pathologic_n"
## [38] "ajcc_pathologic_m"
## [39] "classification_of_tumor"
## [40] "diagnosis_id"
## [41] "icd_10_code"
## [42] "site_of_resection_or_biopsy"
## [43] "tumor_grade"
## [44] "progression_or_recurrence"
## [45] "cigarettes_per_day"
## [46] "alcohol_history"
## [47] "exposure_id"
## [48] "years_smoked"
## [49] "pack_years_smoked"
## [50] "gender"
## [51] "ethnicity"
## [52] "race"
## [53] "vital_status"
## [54] "age_at_index"
## [55] "days_to_birth"
## [56] "year_of_birth"
## [57] "demographic_id"
## [58] "days_to_death"
## [59] "year_of_death"
## [60] "bcr_patient_barcode"
## [61] "primary_site"
## [62] "disease_type"
## [63] "project_id"
## [64] "releasable"
## [65] "name"
## [66] "released"
## [67] "paper_patient"
## [68] "paper_Sex"
## [69] "paper_Age.at.diagnosis"
## [70] "paper_T.stage"
## [71] "paper_N.stage"
## [72] "paper_Tumor.stage"
## [73] "paper_Smoking.Status"
## [74] "paper_Survival"
## [75] "paper_Transversion.High.Low"
## [76] "paper_Nonsilent.Mutations"
## [77] "paper_Nonsilent.Mutations.per.Mb"
## [78] "paper_Oncogene.Negative.or.Positive.Groups"
## [79] "paper_Fusions"
## [80] "paper_expression_subtype"
## [81] "paper_chromosome.affected.by.chromothripsis"
## [82] "paper_iCluster.Group"

```

```
## [83] "paper_CIMP.methylation.signature."
## [84] "paper_MTOR.mechanism.of.mTOR.pathway.activation"
## [85] "paper_Ploidy.ABSOLUTE.calls"
## [86] "paper_Purity.ABSOLUTE.calls"
```

Do we see the same columns for the LUSC dataset?

```
colnames(d_lusc)
```

```
## [1] "barcode" "patient"
## [3] "sample" "shortLetterCode"
## [5] "definition" "sample_submitter_id"
## [7] "sample_type_id" "sample_id"
## [9] "sample_type" "days_to_collection"
## [11] "state" "initial_weight"
## [13] "intermediate_dimension" "pathology_report_uuid"
## [15] "submitter_id" "shortest_dimension"
## [17] "oct_embedded" "longest_dimension"
## [19] "is_ffpe" "tissue_type"
## [21] "synchronous_malignancy" "ajcc_pathologic_stage"
## [23] "tumor_stage" "days_to_diagnosis"
## [25] "treatments" "last_known_disease_status"
## [27] "tissue_or_organ_of_origin" "days_to_last_follow_up"
## [29] "primary_diagnosis" "age_at_diagnosis"
## [31] "prior_malignancy" "year_of_diagnosis"
## [33] "prior_treatment" "ajcc_staging_system_edition"
## [35] "ajcc_pathologic_t" "morphology"
## [37] "ajcc_pathologic_n" "ajcc_pathologic_m"
## [39] "classification_of_tumor" "diagnosis_id"
## [41] "icd_10_code" "site_of_resection_or_biopsy"
## [43] "tumor_grade" "progression_or_recurrence"
## [45] "pack_years_smoked" "cigarettes_per_day"
## [47] "alcohol_history" "exposure_id"
## [49] "years_smoked" "race"
## [51] "ethnicity" "gender"
## [53] "vital_status" "age_at_index"
## [55] "days_to_birth" "year_of_birth"
## [57] "demographic_id" "days_to_death"
## [59] "year_of_death" "bcr_patient_barcode"
## [61] "primary_site" "project_id"
## [63] "disease_type" "name"
## [65] "releasable" "released"
## [67] "paper_patient" "paper_Sex"
## [69] "paper_Age.at.diagnosis" "paper_T.stage"
## [71] "paper_N.stage" "paper_M.stage"
## [73] "paper_Smoking.Status" "paper_Pack.years"
## [75] "paper_Nonsilent.Mutations" "paper_Nonsilent.Mutations.per.Mb"
## [77] "paper_Selected.Mutation.Summary" "paper_High.Level.Amplifications"
## [79] "paper_Homozygous.Deletions" "paper_Expression.Subtype"
```

### 3.1. Which tissues or samples are available for your analysis?

Lung cancer samples in the dataset are provided in tumor or normal tissues. The column shortLetterCode contains Sample Type Codes to describe the type of tissues collected in the dataset.

- TP: Primary solid Tumor
- TR: Recurrent solid Tumor
- NT: Solid Tissue Normal

Check which samples are included in the LUAD dataset.

```
d_luad %>% dplyr::count(shortLetterCode)
```

```
##   shortLetterCode    n
## 1                NT  59
## 2                TP 533
## 3                TR   2
```

Check which samples are included in the LUSC dataset.

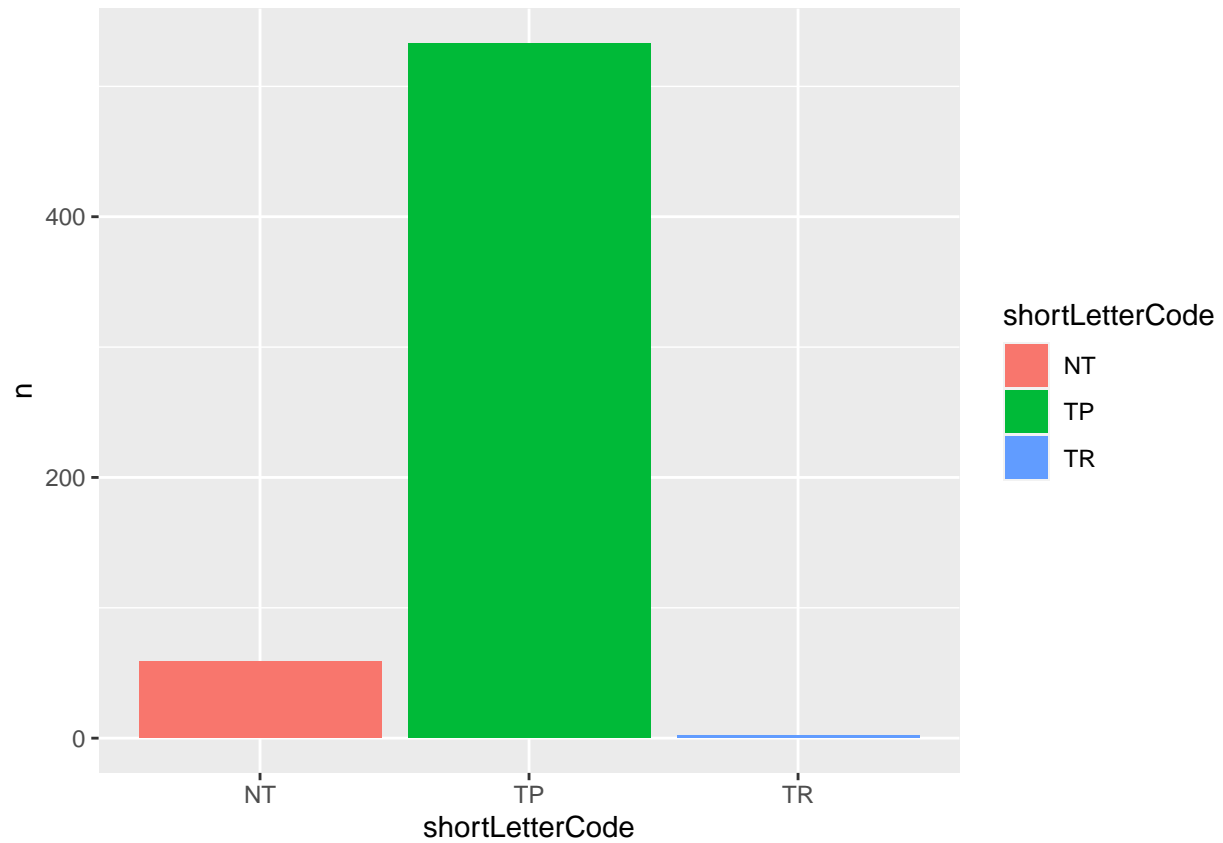
```
d_lusc %>% dplyr::count(shortLetterCode)
```

```
##   shortLetterCode    n
## 1                NT  49
## 2                TP 502
```

Let's make a simple bar plot to compare the number of tissues between two tissue types. We will look at the LUAD samples first.

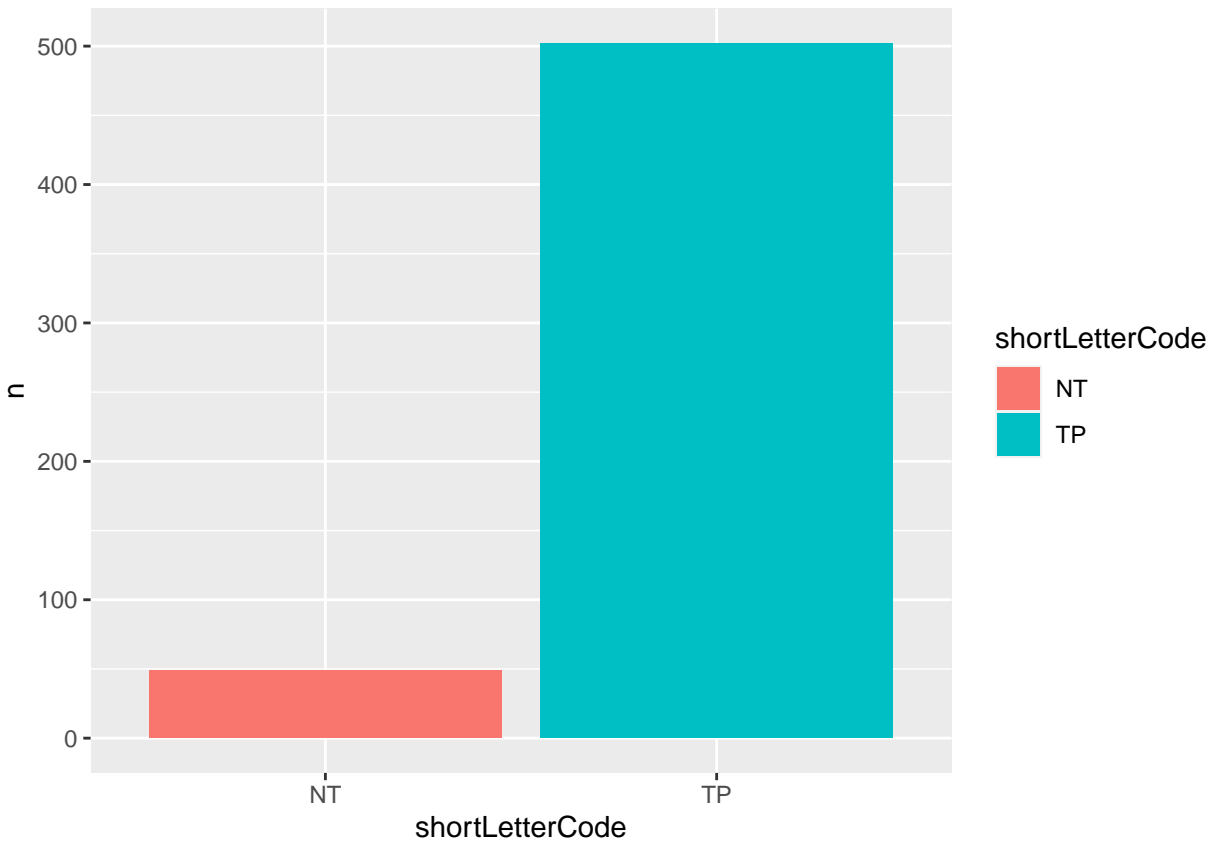
```
d_luad %>%
  dplyr::count(shortLetterCode) %>%
  ggplot(aes(shortLetterCode, n, fill = shortLetterCode)) +
  geom_bar(stat = "identity", position = position_dodge())
```





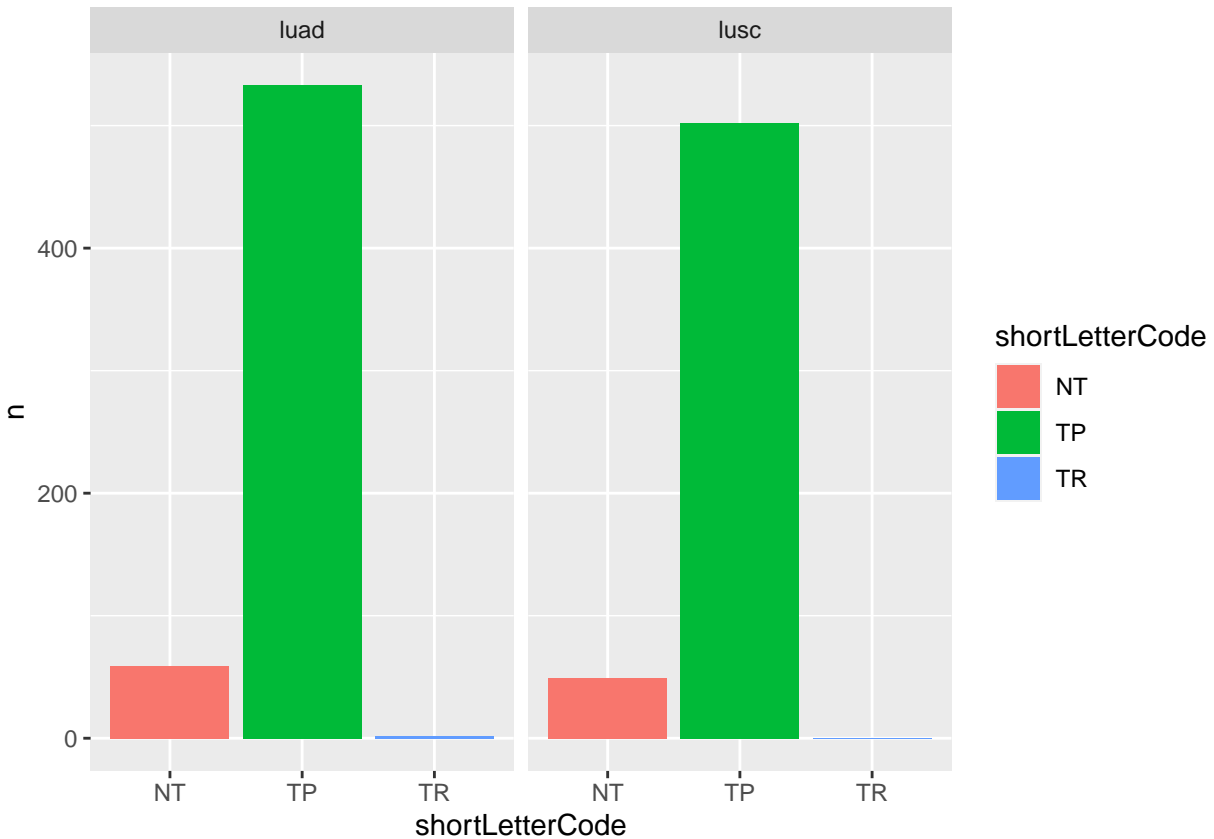
Now we can plot the LUSC samples.

```
d_lusc %>%  
  dplyr::count(shortLetterCode) %>%  
  ggplot(aes(shortLetterCode, n, fill = shortLetterCode)) +  
  geom_bar(stat = "identity", position = position_dodge())
```



We made two separate plots for the LUAD and LUSC dataset. It would be great if we can merge them into one plot. Here I put the code for this. It would be good practice if you comment or delete the line that you don't understand fully, you will compare the difference between outcomes. Also, please figure out what Qs do in this plot.

```
bind_rows(d_luad %>%
  mutate(type='luad') %>%
  select(type, shortLetterCode, tumor_stage),
d_lusc %>%
  mutate(type='lusc') %>%
  select(type, shortLetterCode, tumor_stage)) %>%
dplyr::count(shortLetterCode, type) %>%
complete(type, shortLetterCode, fill = list(n = 0)) %>% # Q1: What is this?
ggplot(., aes(shortLetterCode, n, fill=shortLetterCode)) +
geom_bar(stat="identity", position=position_dodge()) +
facet_wrap(~type, scales = 'free_x', ncol=5) # Q2: What is this?
```



Q1. `complete(type, shortLetterCode, fill = list(n = 0))` -> What is this?

It fills missing values(NA) with 0.

Q2. `facet_wrap(~type, scales = 'free_x', ncol=5)` -> What is this?

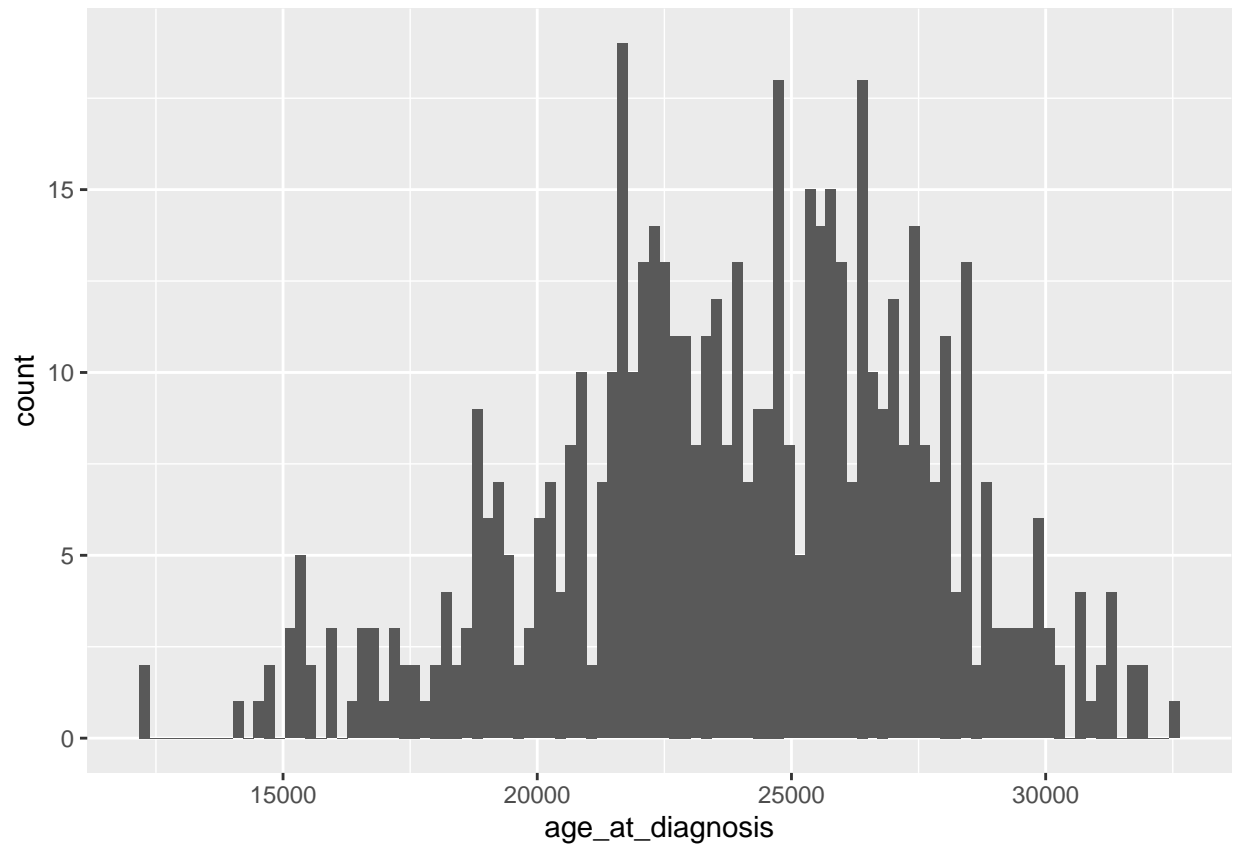
It wraps a 1d sequence of panels into 2d. -scale = 'free\_x' : Allow plots to have different x-axis scales -ncol : number of plots to be on the same row

### 3.2. Distribution of clinical variables

We will plot the distribution of age at diagnosis using histogram. Few things you can check: - Is the distribution continuous? - Is it following the normal distribution? - What is the scale on the x-axis? - If the distribution is stratified, which makes it?

```
# Plot histogram
ggplot(d_luad, aes(age_at_diagnosis)) + geom_histogram(bins=100)
```

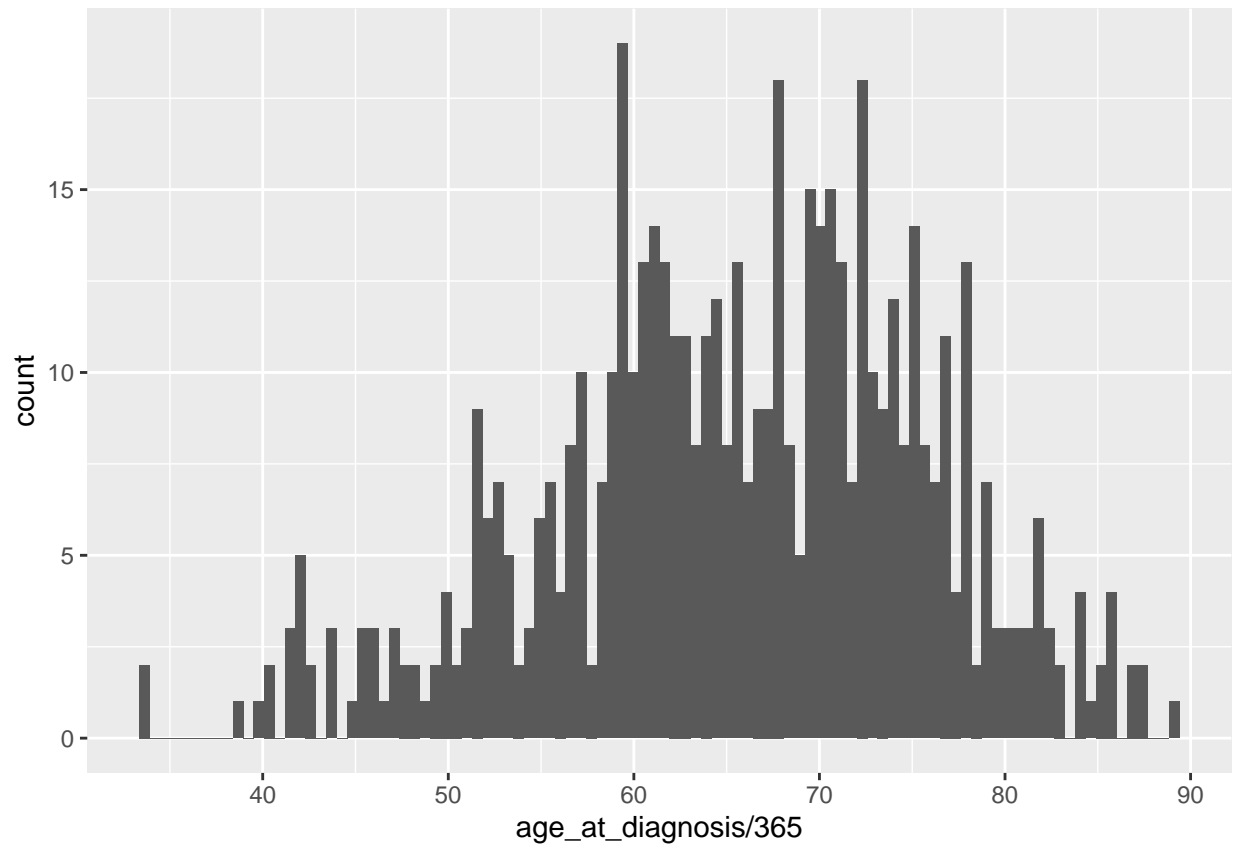
```
## Warning: Removed 37 rows containing non-finite values (stat_bin).
```



The x-axis is a day scale. Let's convert it to year.

```
# Change the axis  
ggplot(d_luad, aes(age_at_diagnosis/365)) + geom_histogram(bins=100)
```

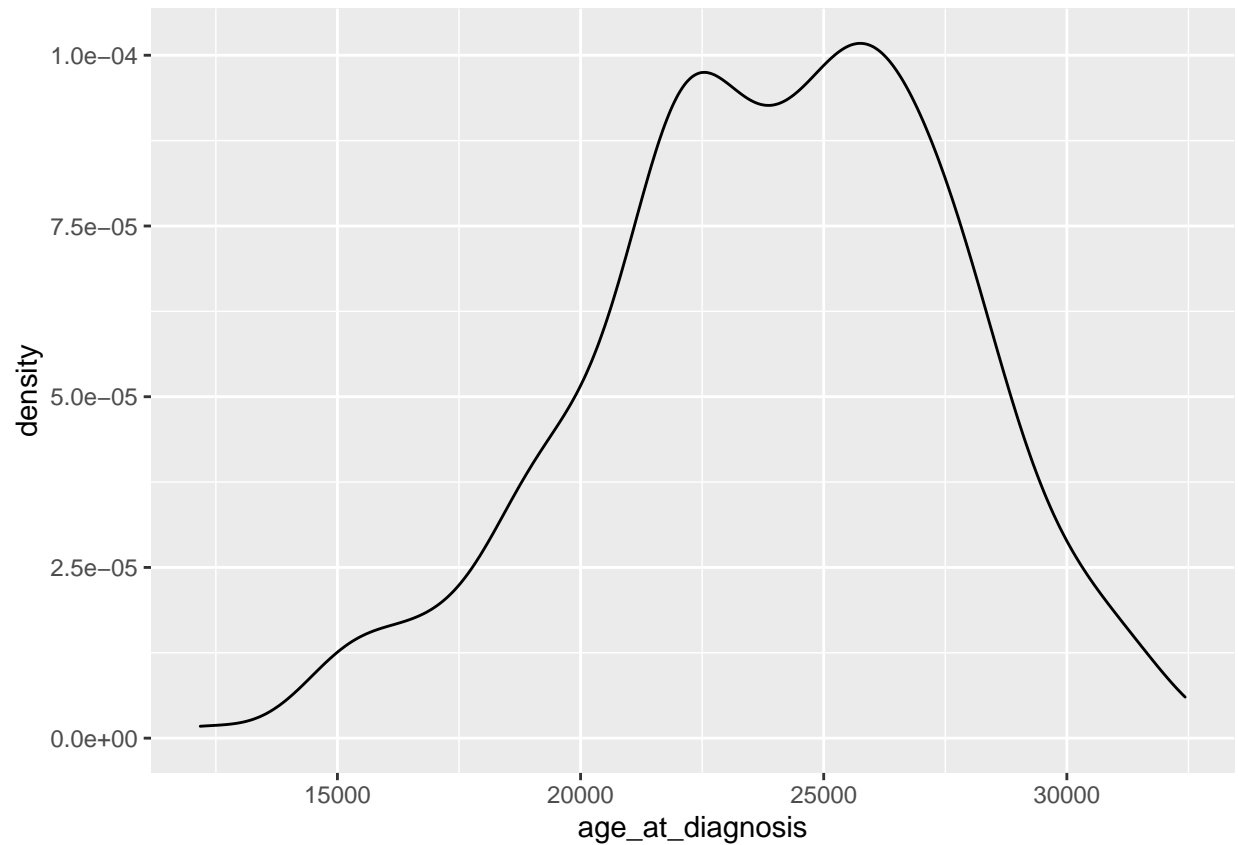
```
## Warning: Removed 37 rows containing non-finite values (stat_bin).
```



To create a smooth density, we use the `geom_density`.

```
ggplot(d_luad, aes(age_at_diagnosis)) + geom_density()
```

```
## Warning: Removed 37 rows containing non-finite values (stat_density).
```

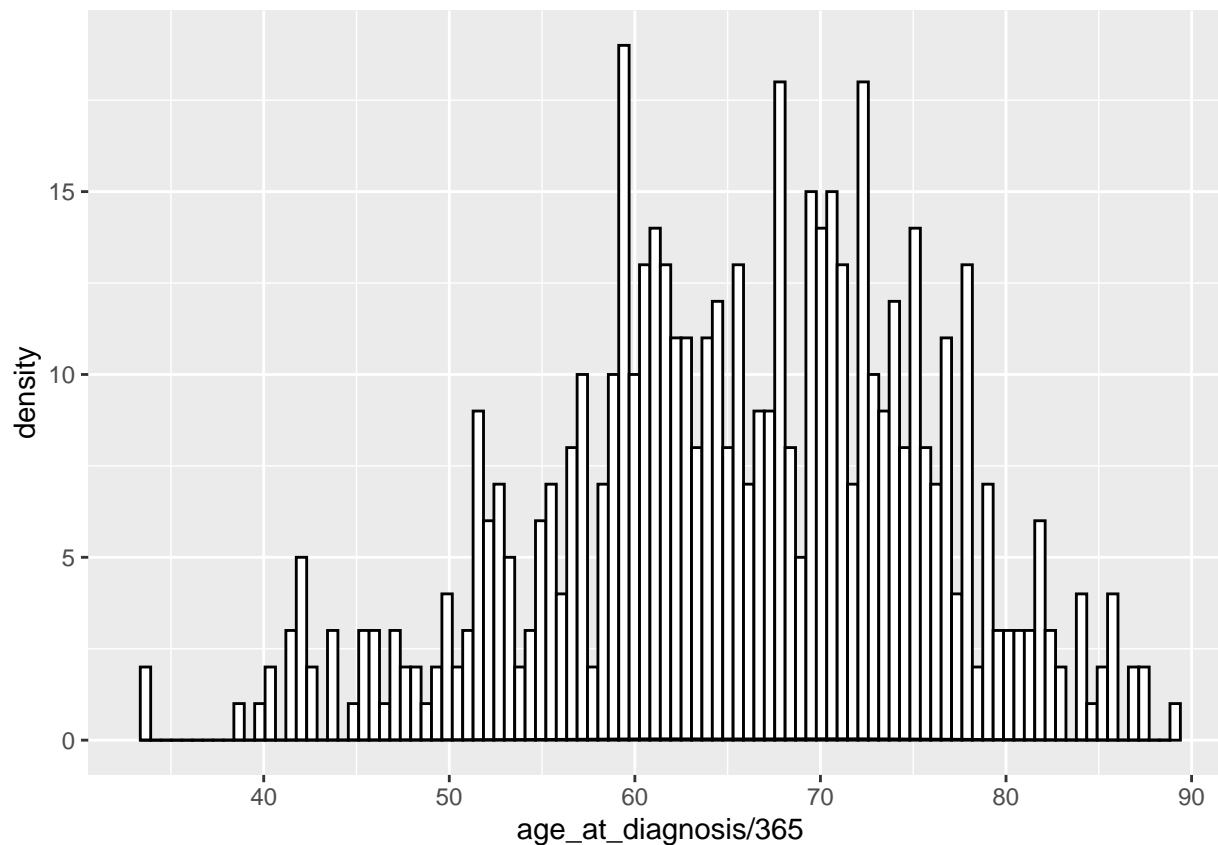


More plot types on distribution. First we can try the plot for both histogram and density.

```
# Plot both histogram and density plot  
ggplot(d_luad, aes(age_at_diagnosis/365)) +  
  geom_histogram(bins=100, colour="black", fill="white") +  
  geom_density(alpha=.2, fill="#FF6666")
```

```
## Warning: Removed 37 rows containing non-finite values (stat_bin).
```

```
## Warning: Removed 37 rows containing non-finite values (stat_density).
```



What did you miss from this?

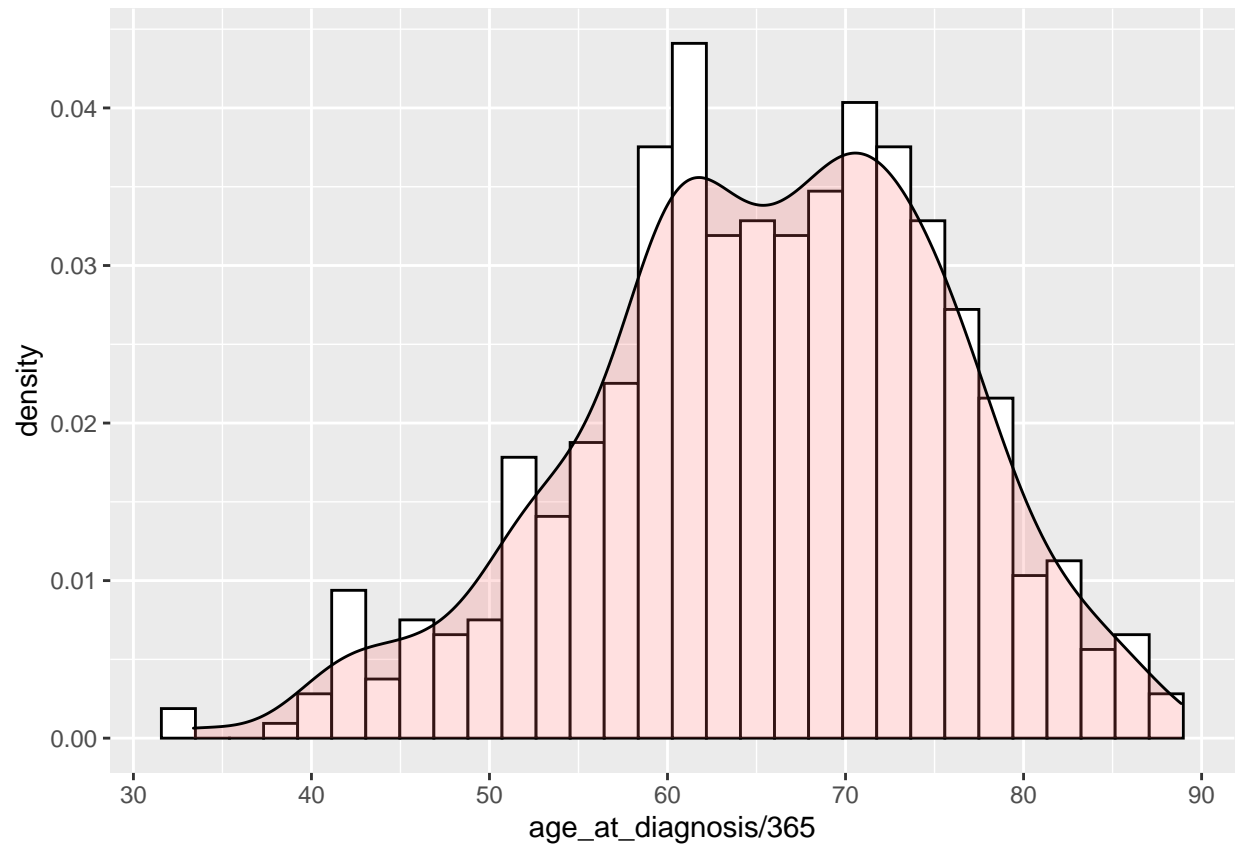
Let's plot a bit different version. We will replace the y-axis of the histogram with the y-axis of the density plot.

```
# Histogram with density plot
ggplot(d_luad, aes(age_at_diagnosis/365)) +
  geom_histogram(aes(y=..density..), colour="black", fill="white")+
  geom_density(alpha=.2, fill="#FF6666")

## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.

## Warning: Removed 37 rows containing non-finite values (stat_bin).

## Warning: Removed 37 rows containing non-finite values (stat_density).
```



Q: Do you think whether it is good visualization for the distribution?

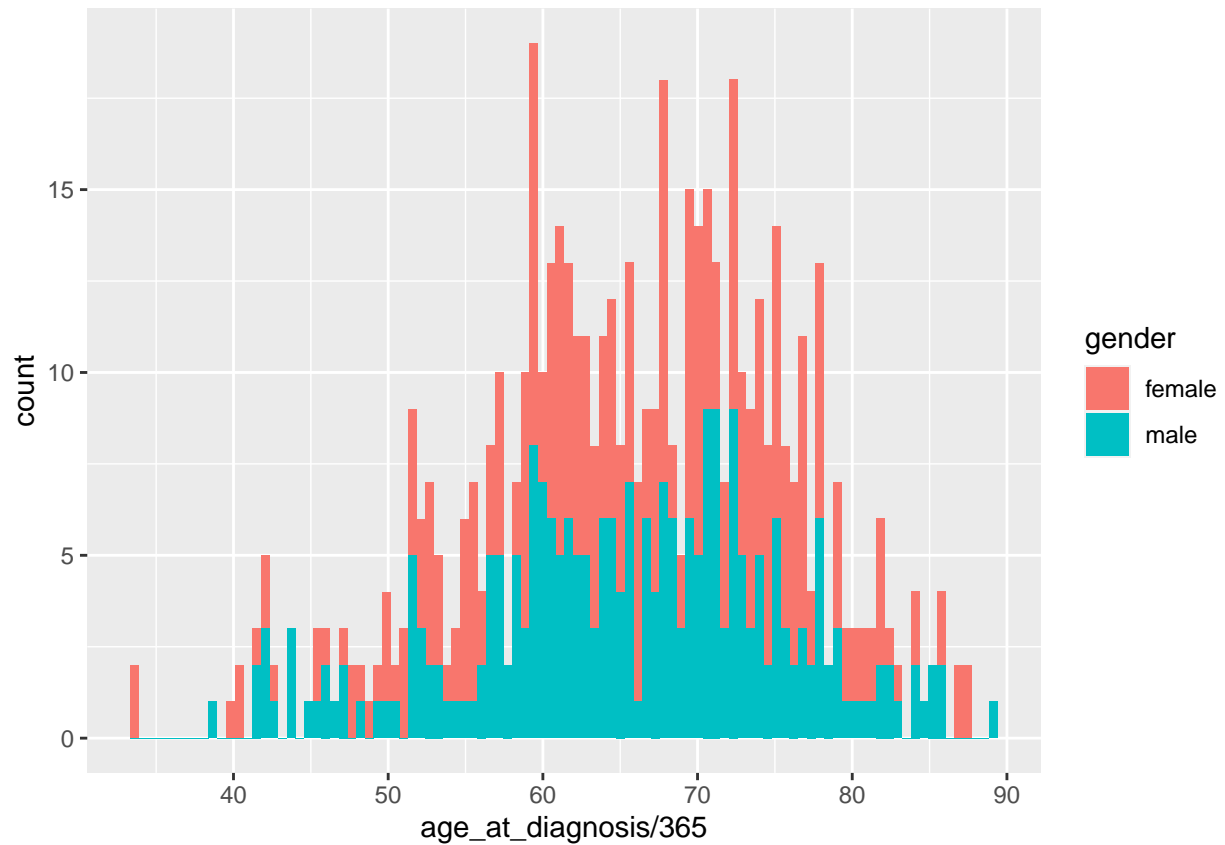
Yes??

From the plot above, we still found the bump around the center. What would contribute to this stratification? Let's look at some variables from other columns. First we can try information from the gender column.

```
ggplot(d_luad, aes(age_at_diagnosis/365, fill=gender)) + geom_histogram(bins=100)
```

```
## Warning: Removed 37 rows containing non-finite values (stat_bin).
```

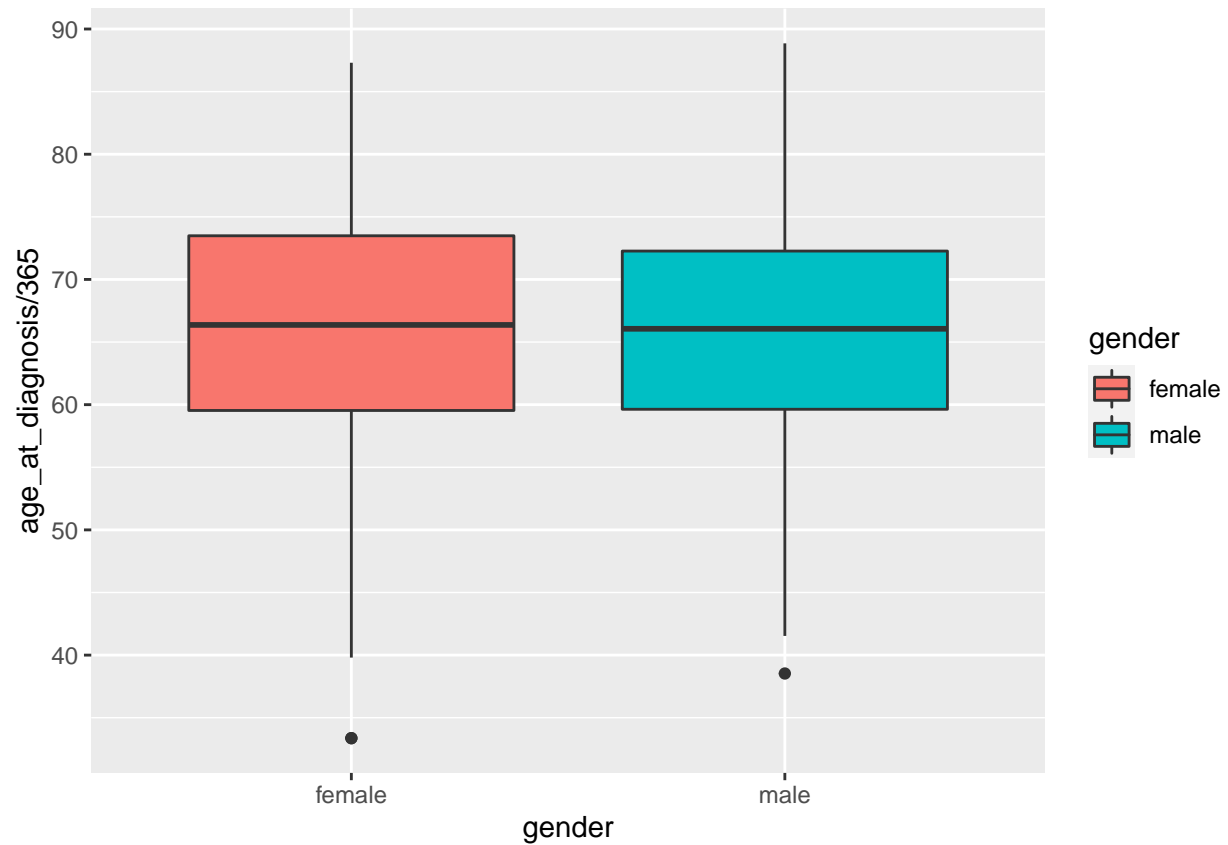




Histogram would be okay but not best visualization. What else we can try?

```
ggplot(d_luad, aes(gender, age_at_diagnosis/365, fill=gender)) +  
  geom_boxplot()
```

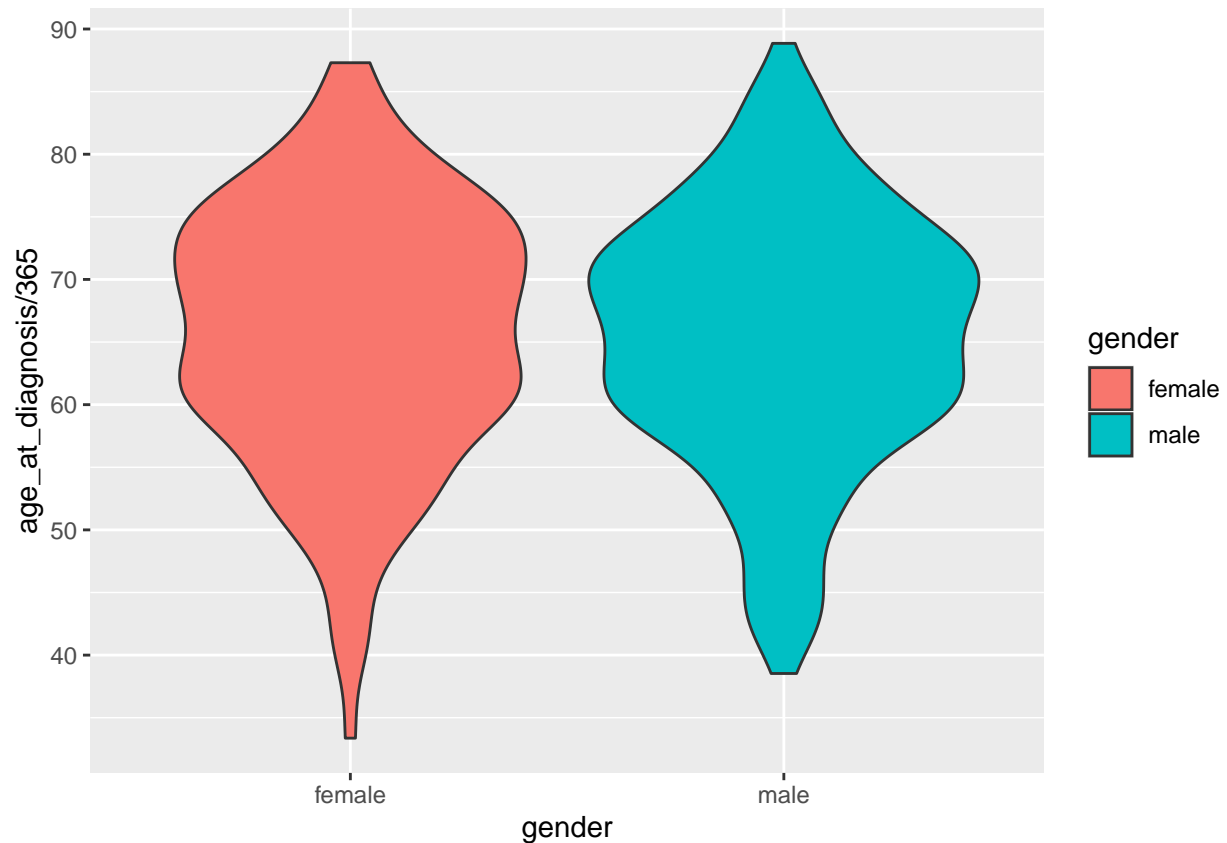
```
## Warning: Removed 37 rows containing non-finite values (stat_boxplot).
```



Can you tell the difference between boxplot and density plot? We can try a violin plot.

```
ggplot(d_luad, aes(gender, age_at_diagnosis/365, fill=gender)) +  
  geom_violin()
```

```
## Warning: Removed 37 rows containing non-finite values (stat_ydensity).
```



Can you tell the difference between boxplot and violin plot?

```
# ggplot(d_luad, aes(gender, age_at_diagnosis/365, fill=gender)) +
#   geom_boxplot() +
#   geom_violin()

# ggplot(d_luad, aes(gender, age_at_diagnosis/365, fill=gender)) +
#   geom_violin() +
#   geom_boxplot()

# ggplot(d_luad, aes(gender, age_at_diagnosis/365)) +
#   geom_violin(aes(fill=gender)) +
#   geom_boxplot(fill='white')

ggplot(d_luad, aes(gender, age_at_diagnosis/365)) +
  geom_violin(aes(fill=gender)) +
  geom_boxplot(fill='white', width=0.25)
```

```
## Warning: Removed 37 rows containing non-finite values (stat_ydensity).
```

```
## Warning: Removed 37 rows containing non-finite values (stat_boxplot).
```



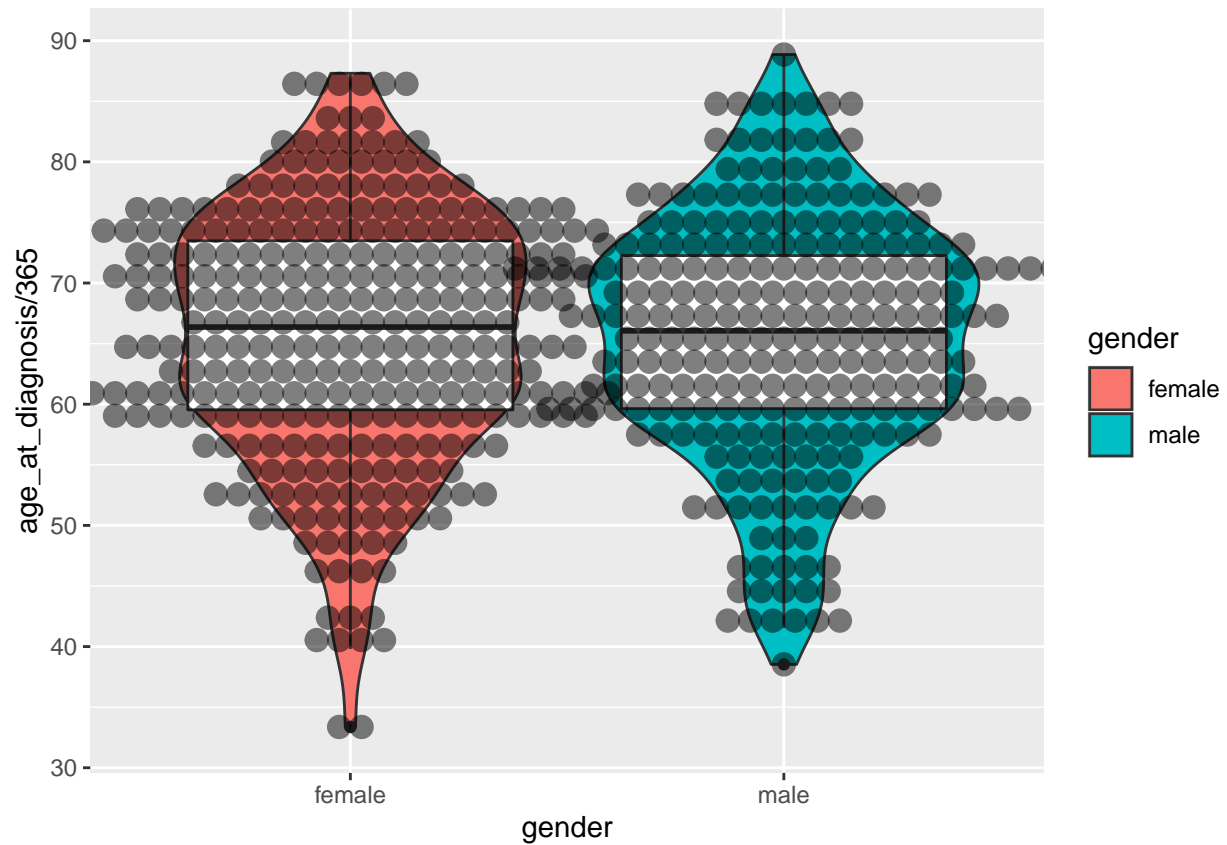
```
ggplot(d_luad, aes(gender, age_at_diagnosis/365)) +  
  geom_violin(aes(fill=gender)) +  
  geom_boxplot(fill='white') +  
  geom_dotplot(binaxis='y', stackdir='center', dotsize=1, alpha=0.5)
```

```
## Warning: Removed 37 rows containing non-finite values (stat_ydensity).
```

```
## Warning: Removed 37 rows containing non-finite values (stat_boxplot).
```

```
## Bin width defaults to 1/30 of the range of the data. Pick better value with `binwidth`.
```

```
## Warning: Removed 37 rows containing non-finite values (stat_bindot).
```

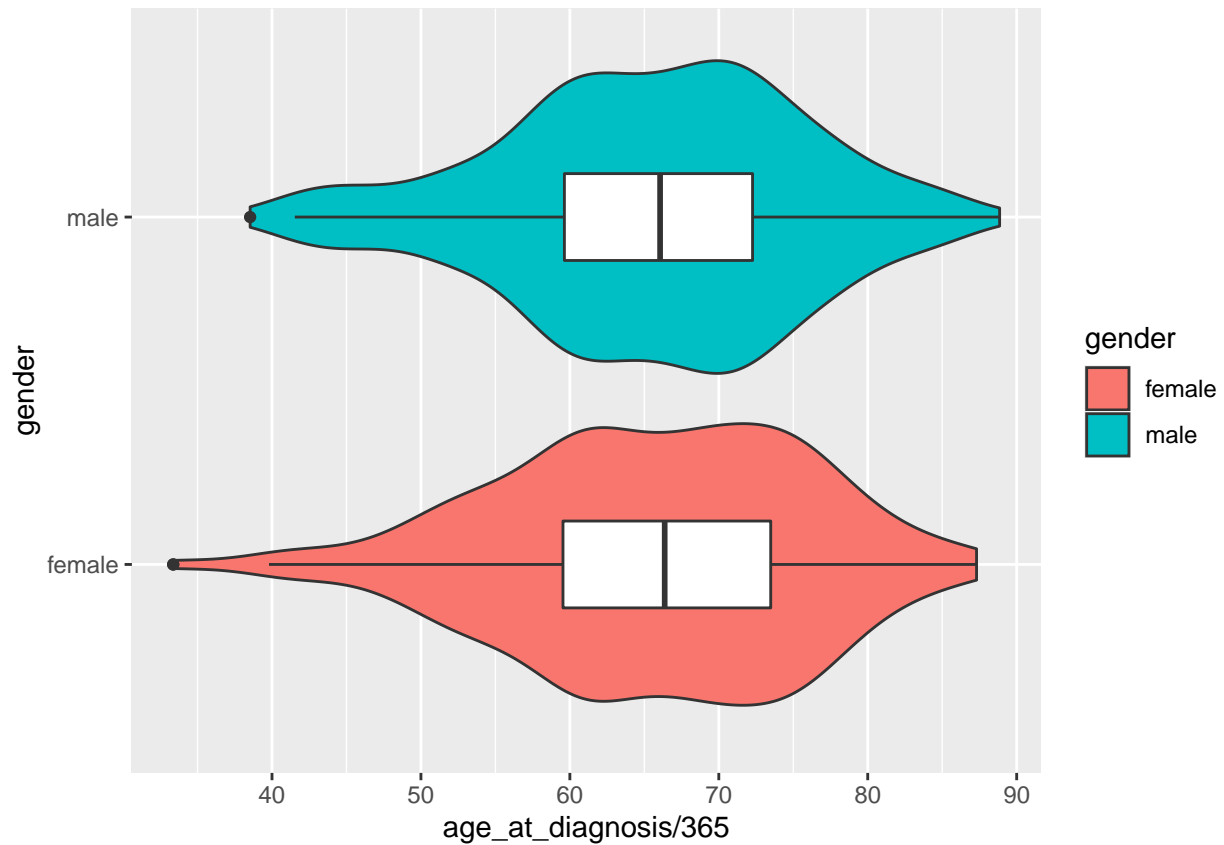


```
# Try different options from the manual
# http://www.sthda.com/english/wiki/ggplot2-violin-plot-quick-start-guide-r-software-and-data-visualization
```

```
#coord_flip
ggplot(d_luad, aes(gender, age_at_diagnosis/365)) +
  geom_violin(aes(fill=gender)) +
  geom_boxplot(fill='white', width=0.25) +
  coord_flip()
```

```
## Warning: Removed 37 rows containing non-finite values (stat_ydensity).
```

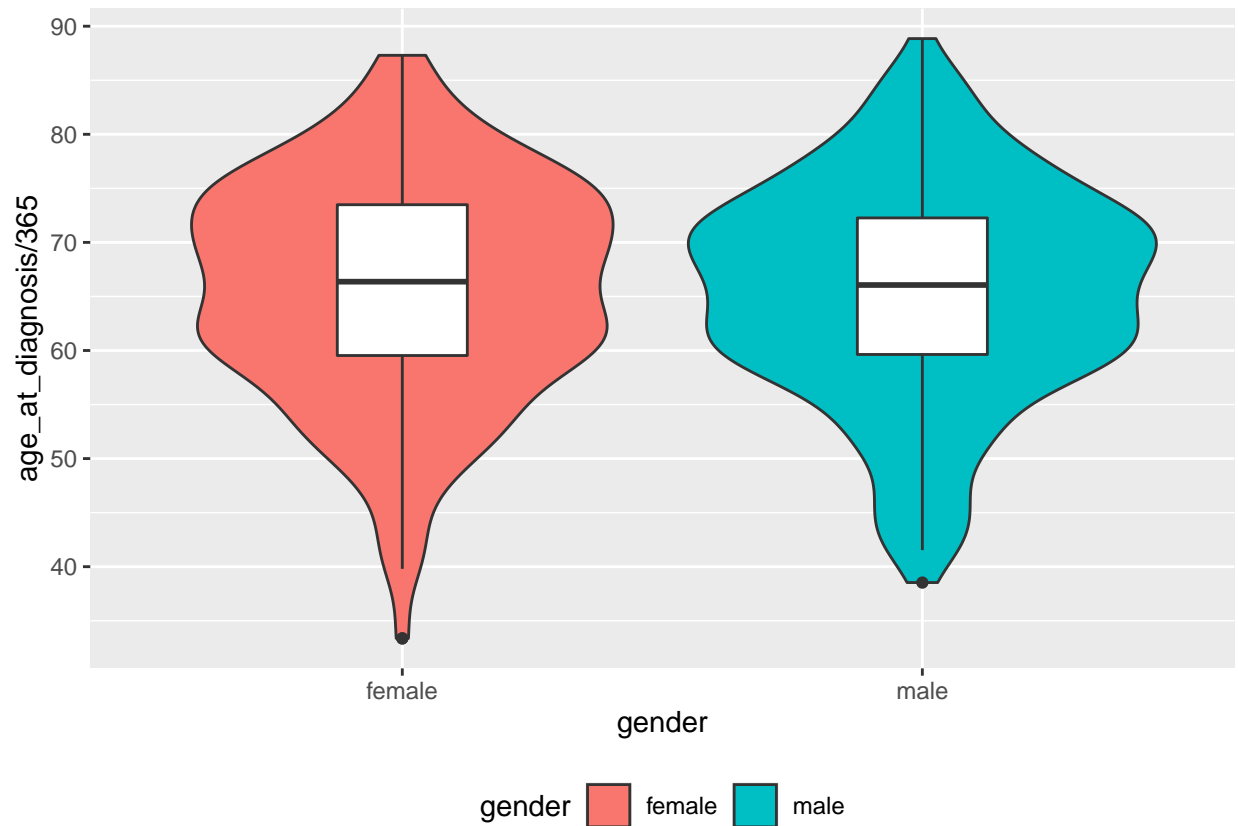
```
## Warning: Removed 37 rows containing non-finite values (stat_boxplot).
```



```
#change legend position
ggplot(d_luad, aes(gender, age_at_diagnosis/365)) +
  geom_violin(aes(fill=gender)) +
  geom_boxplot(fill='white', width=0.25) +
  theme(legend.position="bottom")
```

```
## Warning: Removed 37 rows containing non-finite values (stat_ydensity).
```

```
## Warning: Removed 37 rows containing non-finite values (stat_boxplot).
```

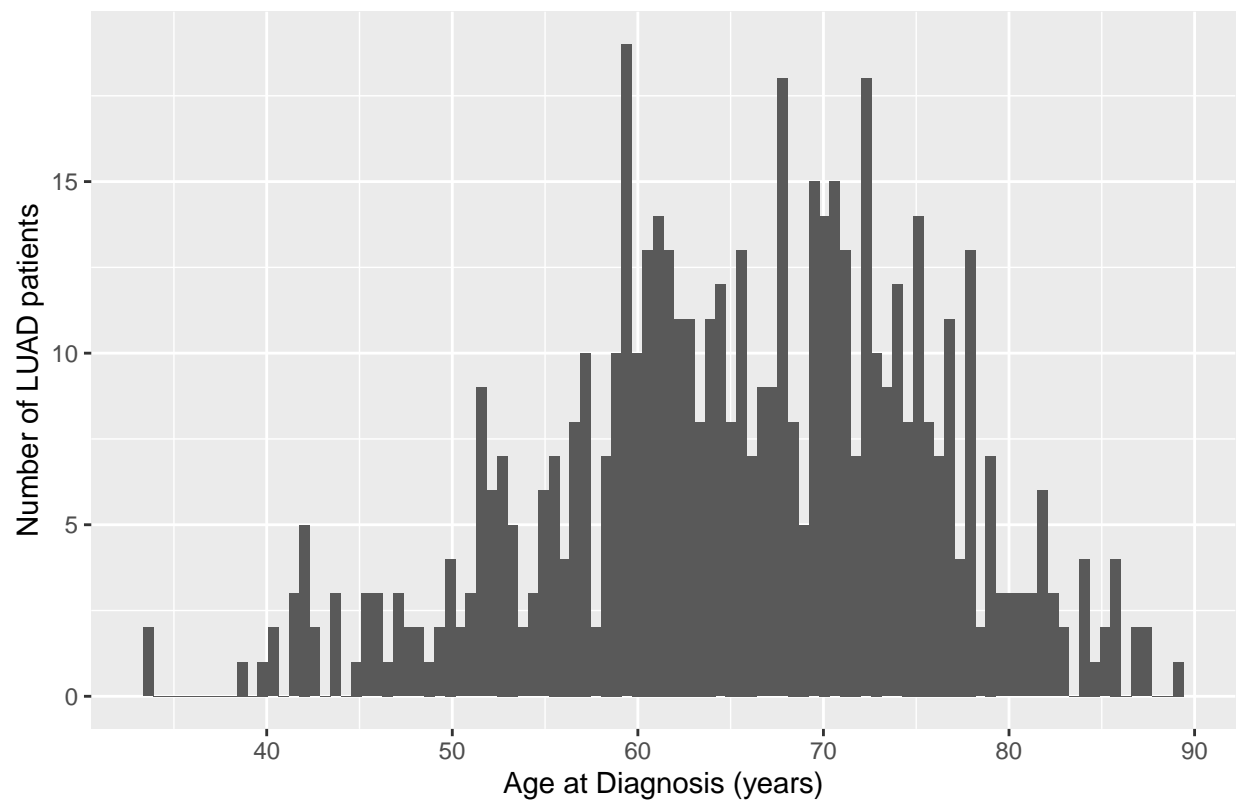


Do you think gender is the reason? If not, how about tumor stages?

```
ggplot(d_luad, aes(age_at_diagnosis/365)) +
  labs(x = 'Age at Diagnosis (years)', y = 'Number of LUAD patients',
       title = 'TCGA: Lung cancer adenocarcinoma') +
  geom_histogram(bins=100)
```

```
## Warning: Removed 37 rows containing non-finite values (stat_bin).
```

## TCGA: Lung cancer adenocarcinoma



More tasks for the class.

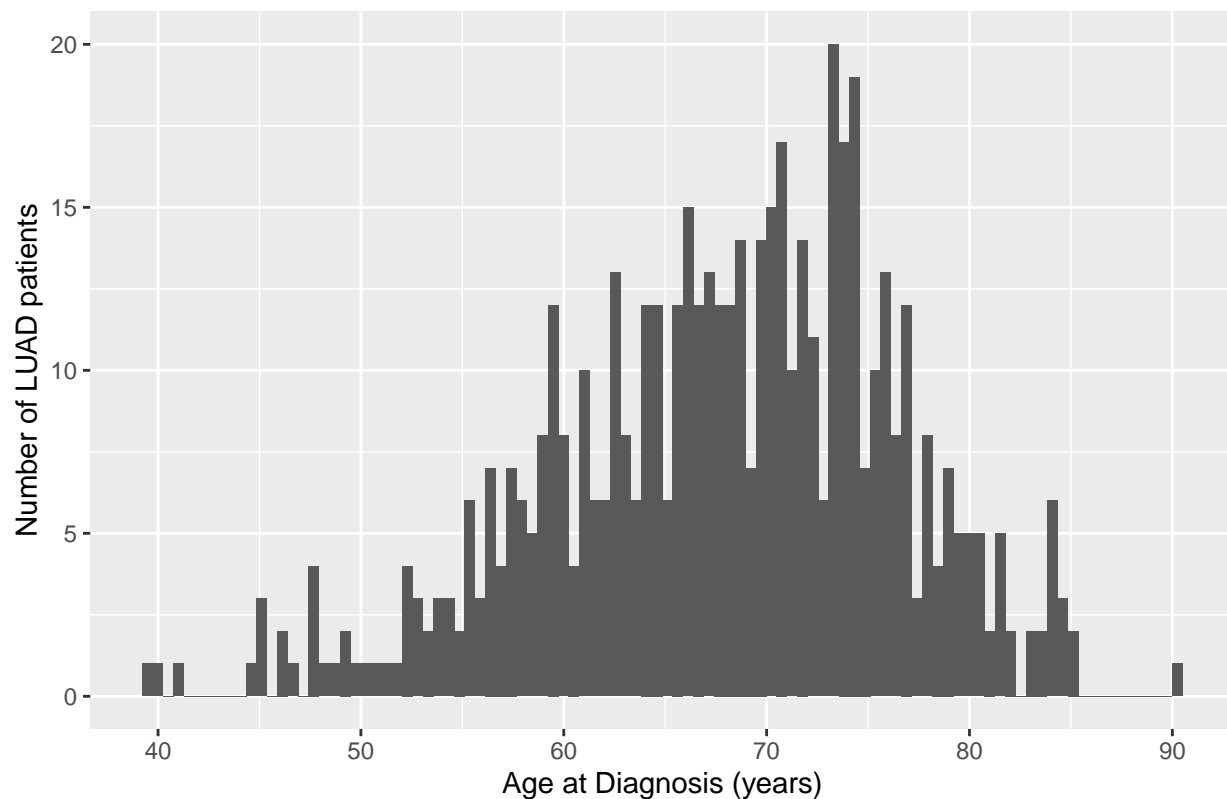
- Q1. Add labels for the plot.

```
ggplot(d_lusc, aes(age_at_diagnosis/365)) +  
  labs(x = 'Age at Diagnosis (years)', y = 'Number of LUAD patients',  
       title = 'TCGA: Lung squamous Cell Carcinoma') +  
  geom_histogram(bins=100)
```

```
## Warning: Removed 10 rows containing non-finite values (stat_bin).
```



## TCGA: Lung squamous Cell Carcinoma

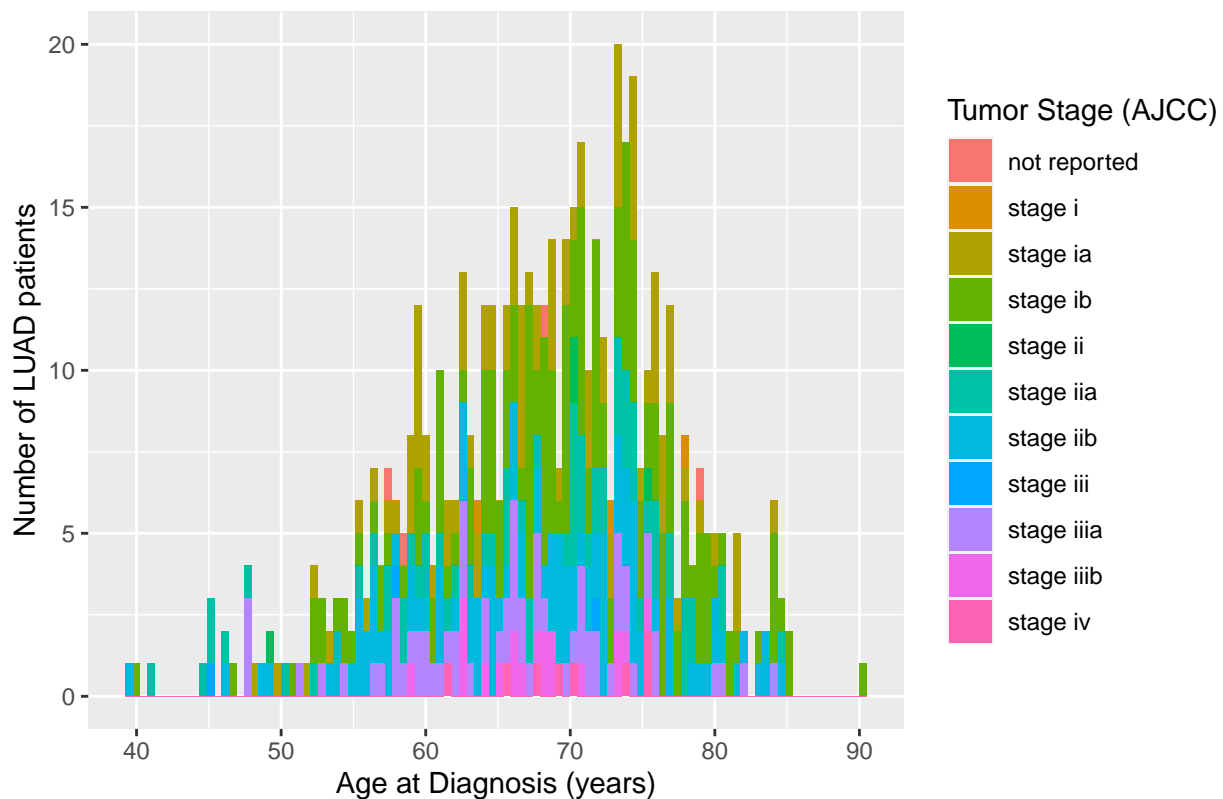


- Q2. Change the color for categories.

```
ggplot(d_lusc, aes(age_at_diagnosis/365, fill = tumor_stage)) +  
  labs(x = 'Age at Diagnosis (years)', y = 'Number of LUAD patients',  
       title = 'TCGA: Lung squamous Cell Carcinoma') +  
  geom_histogram(bins=100) +  
  labs(fill="Tumor Stage (AJCC)")
```

```
## Warning: Removed 10 rows containing non-finite values (stat_bin).
```

## TCGA: Lung squamous Cell Carcinoma



- Q3. Save to PDF file

```
# assign p for plot
p <- ggplot(d_lusc, aes(age_at_diagnosis/365, fill = tumor_stage)) +
  labs(x = 'Age at Diagnosis (years)', y = 'Number of LUAD patients',
       title = 'TCGA: Lung squamous Cell Carcinoma') +
  geom_histogram(bins=100) +
  labs(fill="Tumor Stage (AJCC)")

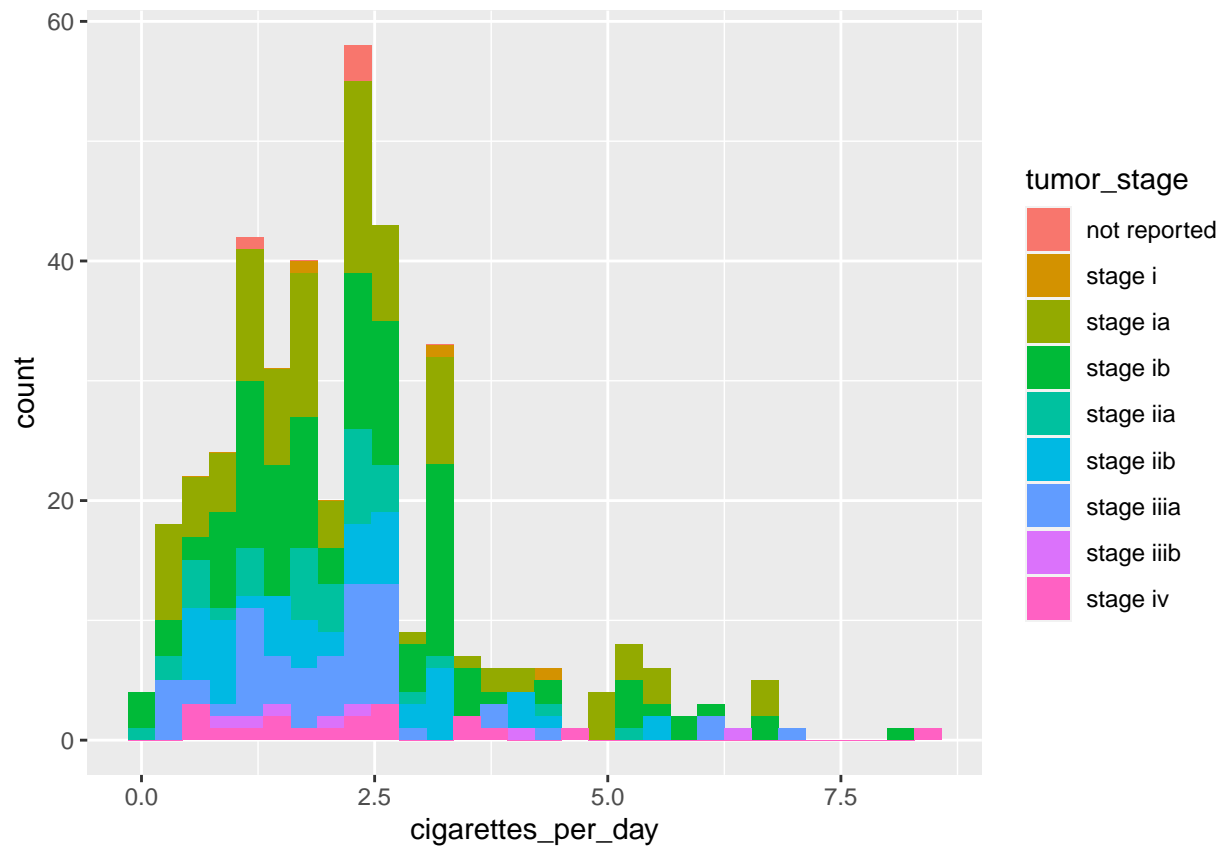
# save to file
ggsave('plot.TCGA_LUSC.histogram_AgeOfDX.20191002.pdf', p, width = 16, height = 9)
```

```
## Warning: Removed 10 rows containing non-finite values (stat_bin).
```

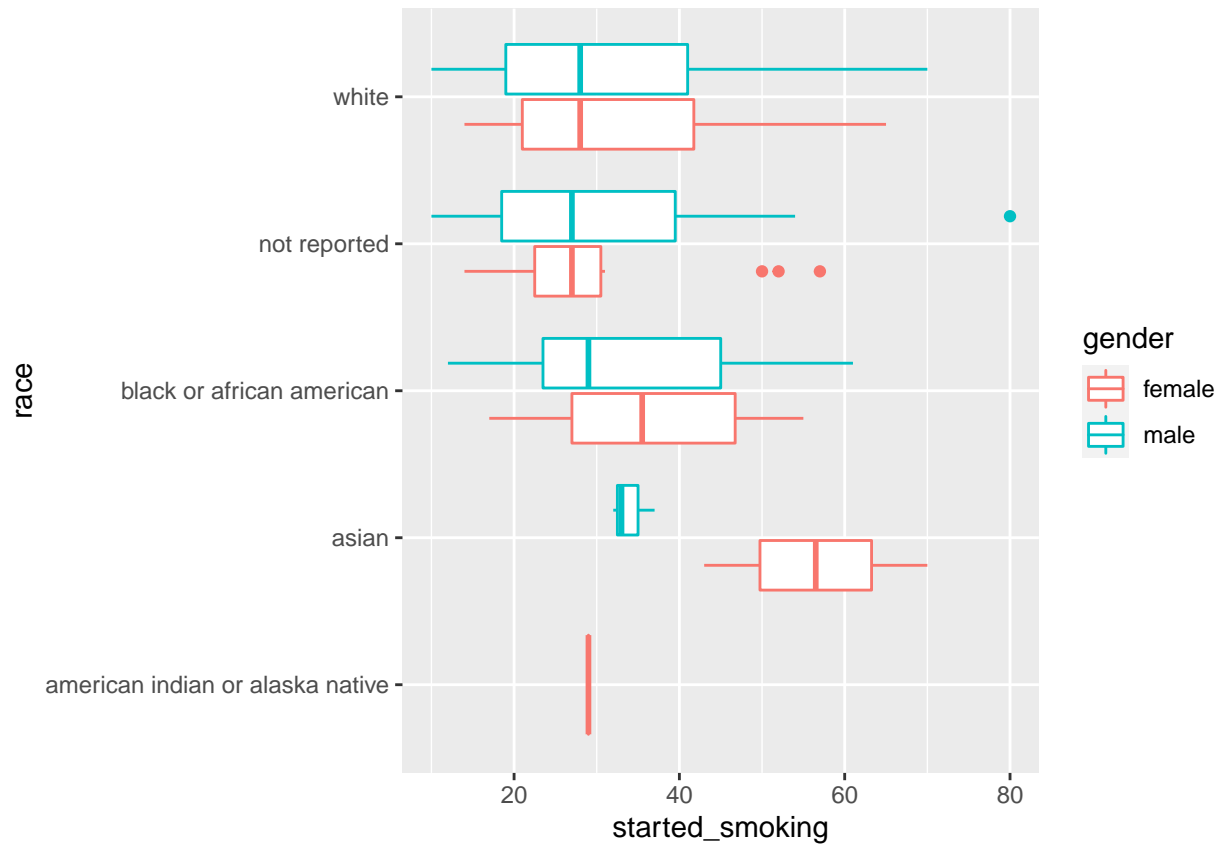
- Q4. Select the column with continuous information and plot the distribution by yourself.

```
#cigarettes_per_day & tumor_stage
d_luad %>% filter(!is.na(cigarettes_per_day)) %>%
  ggplot(aes(cigarettes_per_day, fill = tumor_stage)) +
  geom_histogram()
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
#started_smoking & race
bind_rows(
  d_luad %>% mutate(started_smoking = age_at_index - years_smoked) %>%
    filter(!is.na(started_smoking)),
  d_lusc %>% mutate(started_smoking = age_at_index - years_smoked) %>%
    filter(!is.na(started_smoking))) %>%
  ggplot(aes(race, started_smoking, col = gender)) +
  geom_boxplot() +
  coord_flip()
```



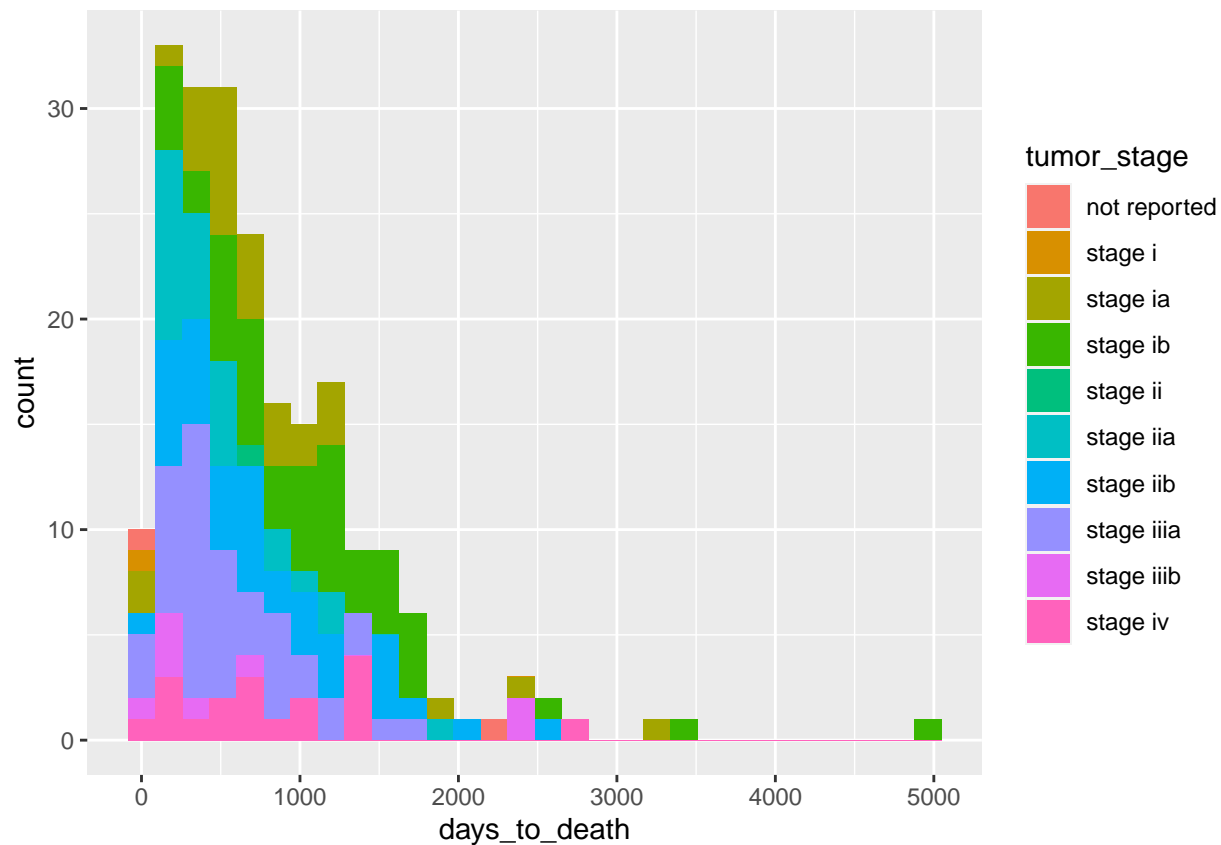
```
#days_to_death & tumor stage
```

```
d_luad %>%
```

```
  ggplot(aes(days_to_death, fill = tumor_stage)) + geom_histogram()
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

```
## Warning: Removed 380 rows containing non-finite values (stat_bin).
```



```
#tumor_stage & days_to_death(boxplot)
```

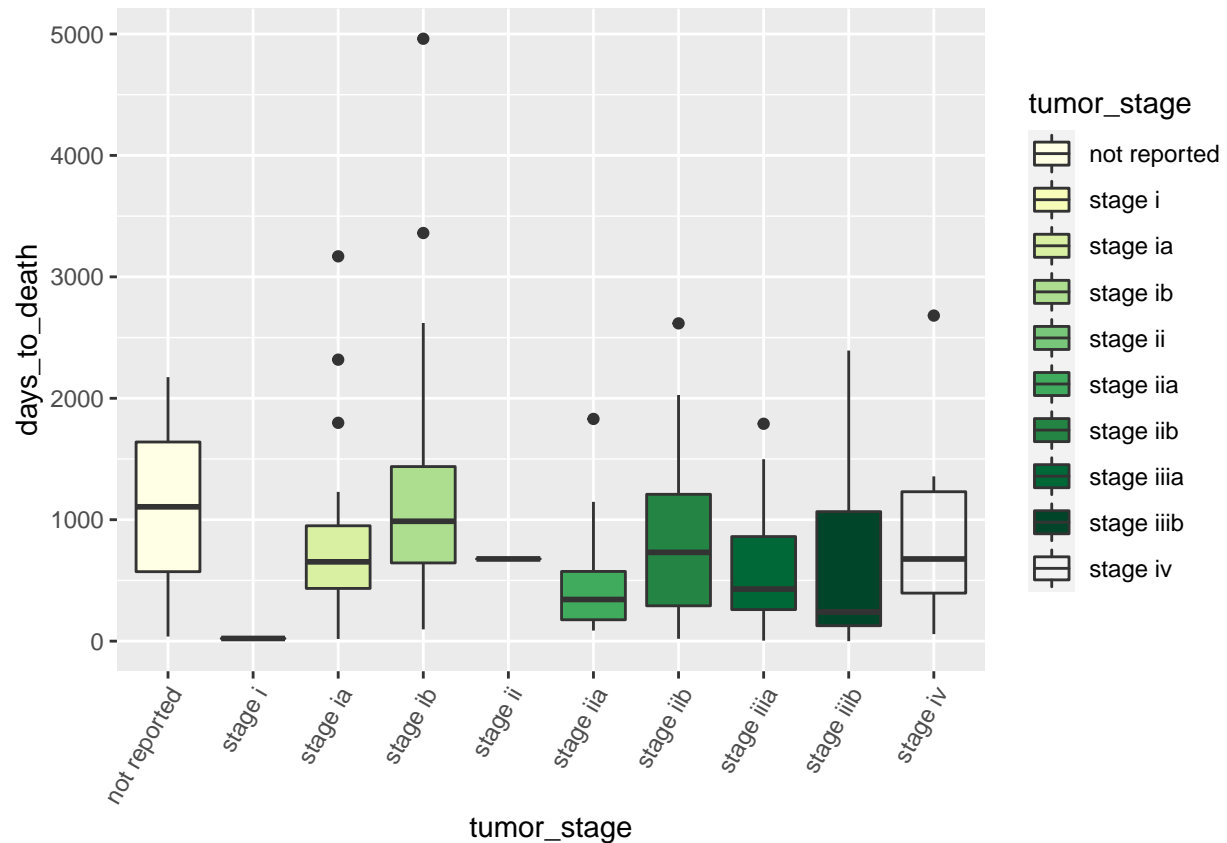
```
d_luad %>%
```

```
ggplot(aes(tumor_stage, days_to_death, fill = tumor_stage)) +  
  geom_boxplot() +  
  theme(axis.text.x = element_text(angle = 60, hjust = 1)) +  
  scale_fill_brewer(palette = "YlGn")
```

```
## Warning: Removed 380 rows containing non-finite values (stat_boxplot).
```

```
## Warning in RColorBrewer::brewer.pal(n, pal): n too large, allowed maximum for palette YlGn is 9
```

```
## Returning the palette you asked for with that many colors
```

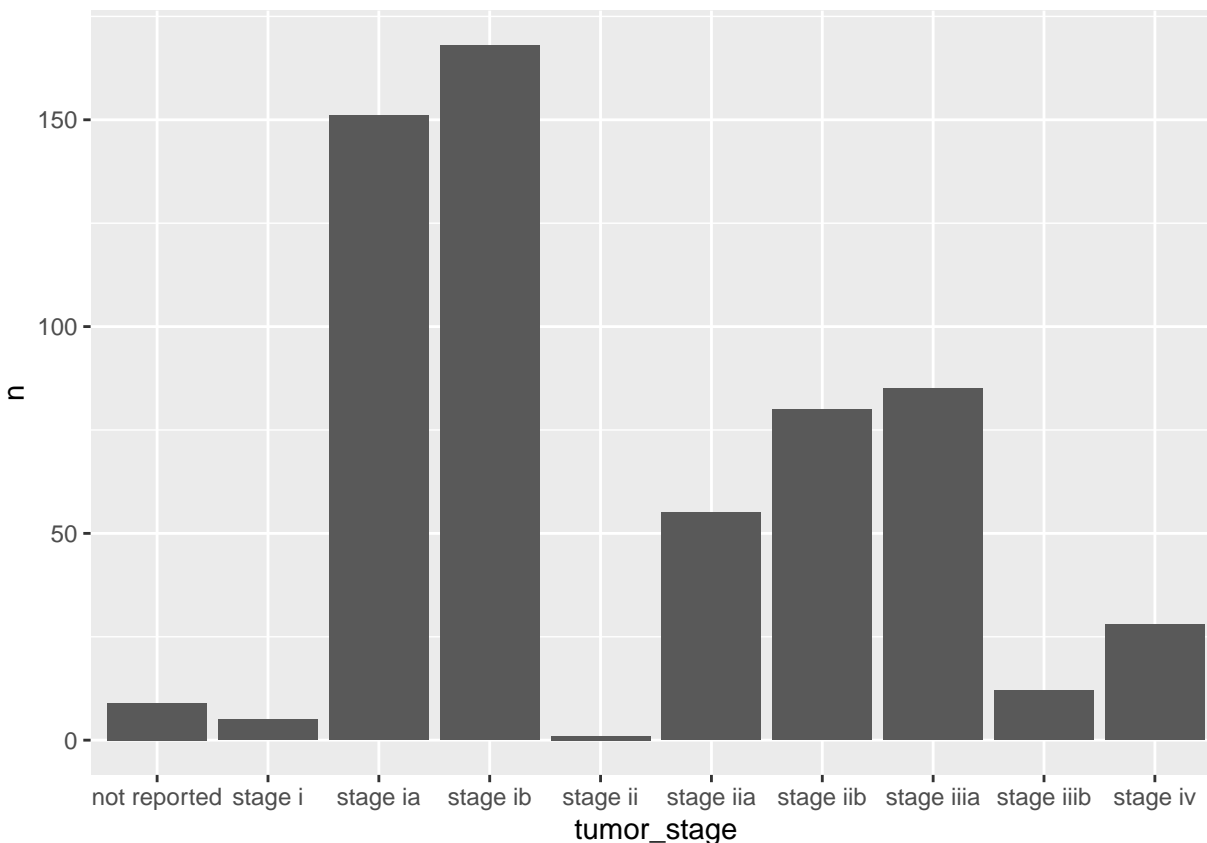


### 3.3. Tumor stages of lung cancers

First information you might want to explore is staging cancers. Different types of staging systems are used for different types of cancer. You can read further information in general staging rules, or specifics in lung cancers (e.g. stage IA).

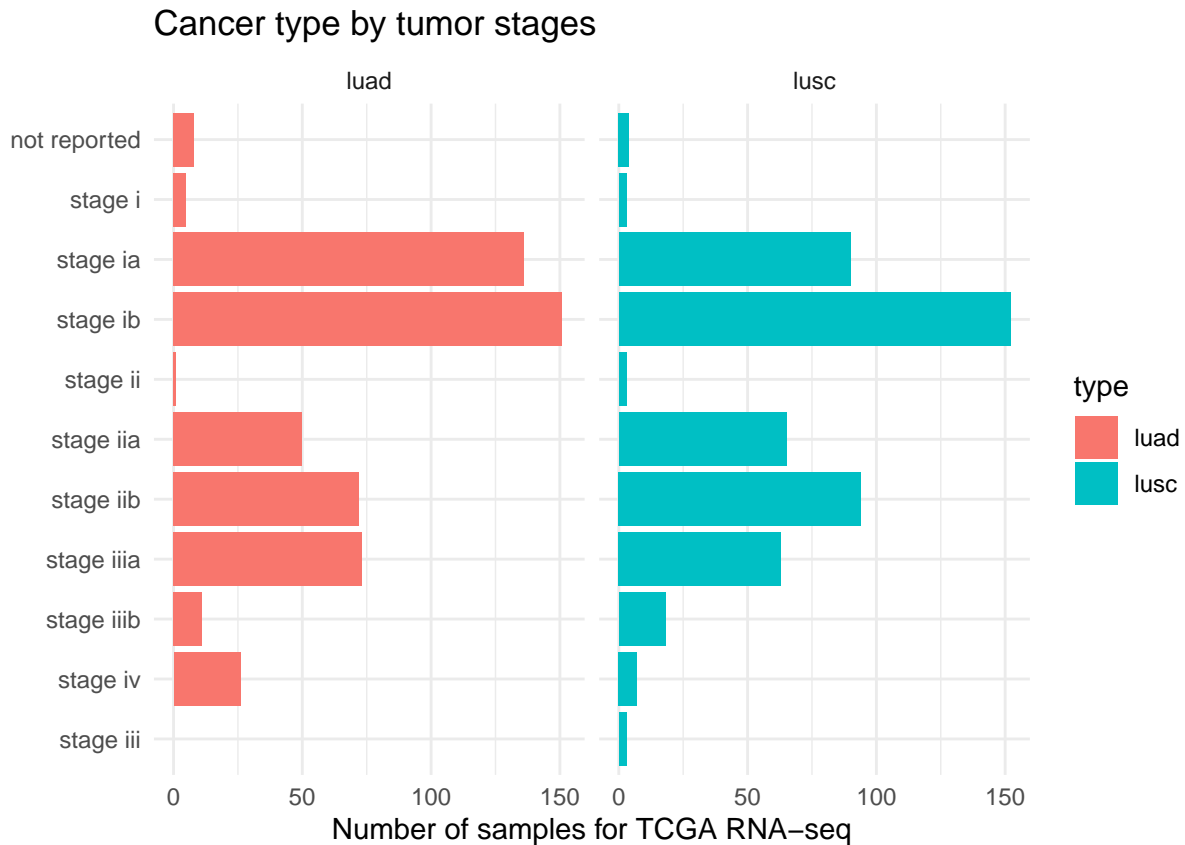
```
# Count the number of tumors in the LUAD dataset
counts_tumor <- d_luad %>% dplyr::count(tumor_stage)

# Try bar plot
ggplot(counts_tumor, aes(tumor_stage, n)) + geom_bar(stat="identity")
```



Now you can combine both for visualization.

```
# Cancer type by tumor stages
bind_rows(d_luad %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='luad') %>%
  select(type, tumor_stage),
d_lusc %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='lusc') %>%
  select(type, tumor_stage)) %>%
dplyr::count(type, tumor_stage) %>%
mutate(tumor_stage = factor(tumor_stage, levels=rev(unique(tumor_stage)))) %>%
ggplot(aes( tumor_stage, n, fill=type)) +
labs(title = 'Cancer type by tumor stages',
      x = '', y='Number of samples for TCGA RNA-seq') +
theme_minimal() + geom_bar(stat="identity") +
facet_wrap(~type) + coord_flip()
```

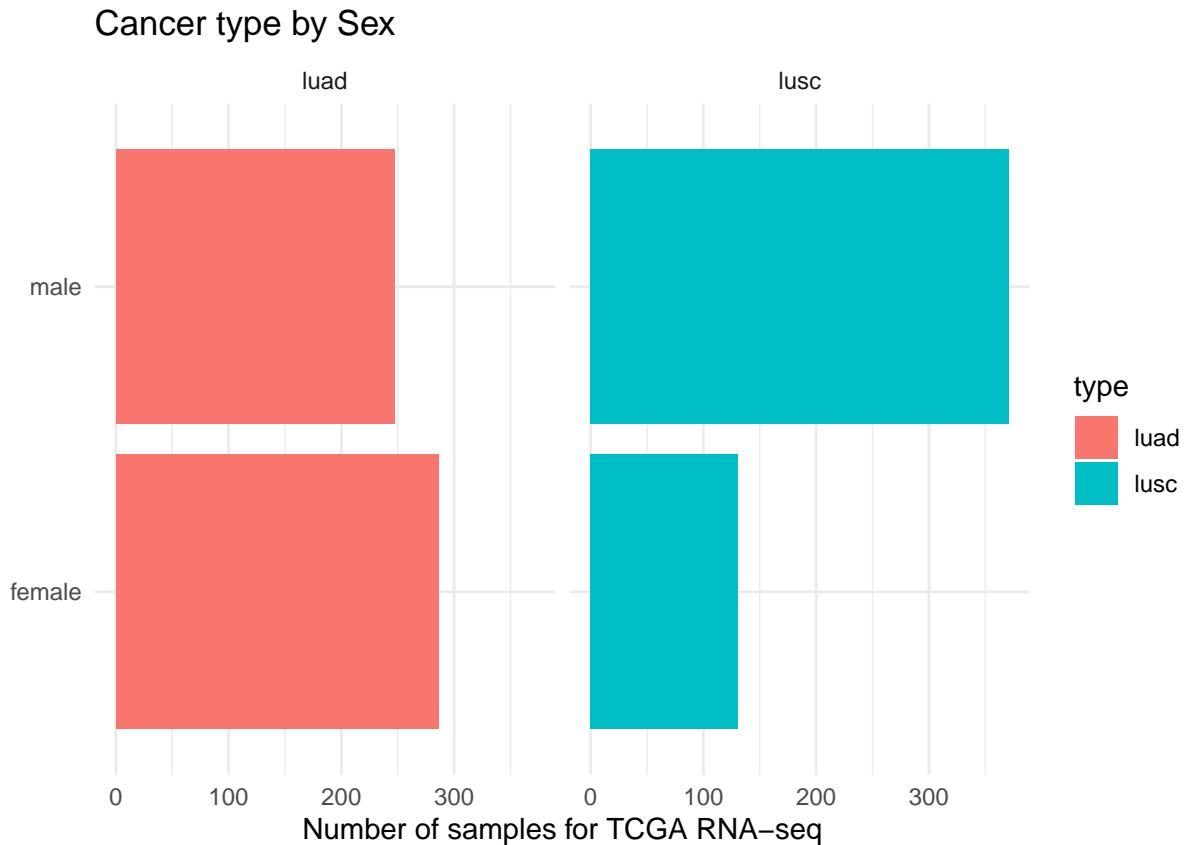


### 3.4. Cancer type by gender

Let's try another information - gender. We will describe this by a bar plot like above. After plotting, which information you can read from this? NB: I am not pretty sure why TCGA put gender, not sex in the dataset, in addition to mixed use of female/male with gender.

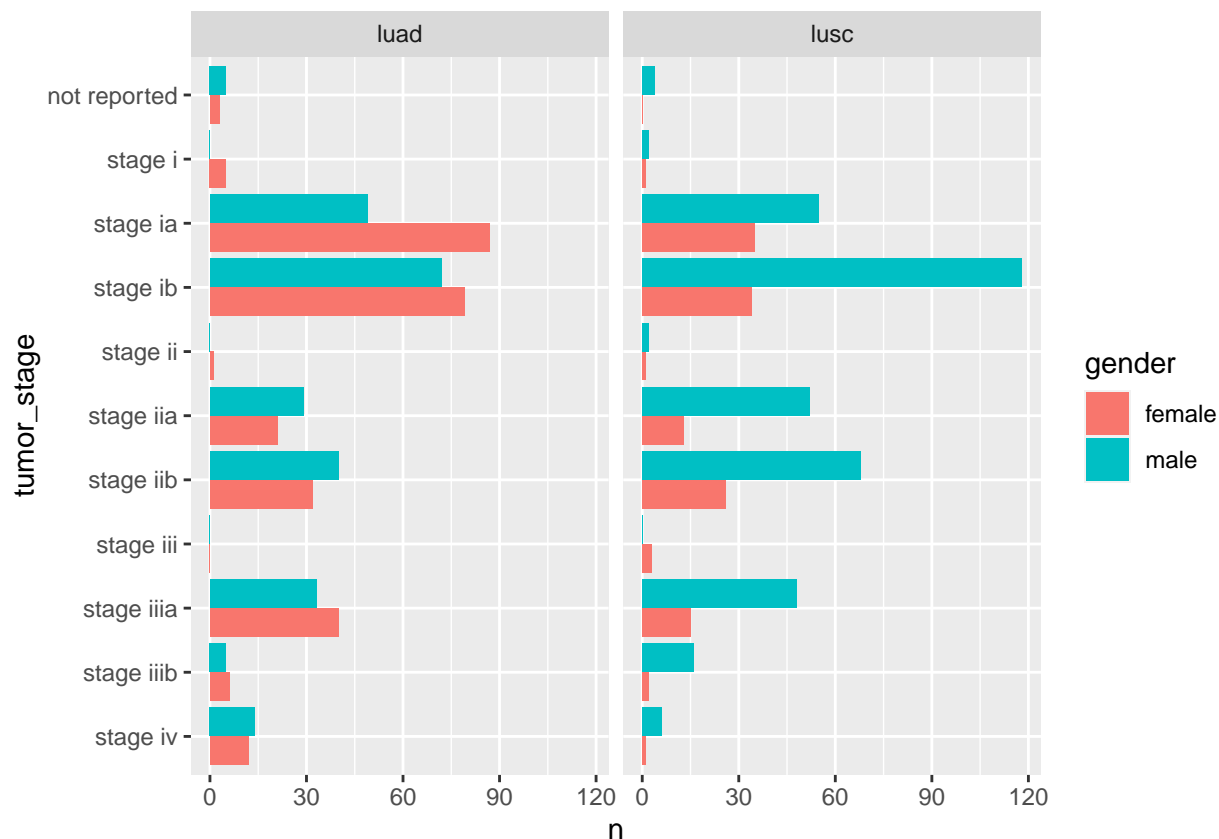
```
# Cancer type by Sex
bind_rows(d_luad %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='luad') %>%
  select(type, gender),
  d_lusc %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='lusc') %>%
  select(type, gender)) %>%
dplyr::count(type, gender) %>%
ggplot(aes( gender, n, fill=type)) +
labs(title = 'Cancer type by Sex',
  x = '', y='Number of samples for TCGA RNA-seq') +
theme_minimal() + geom_bar(stat="identity") +
facet_wrap(~type) + coord_flip()
```





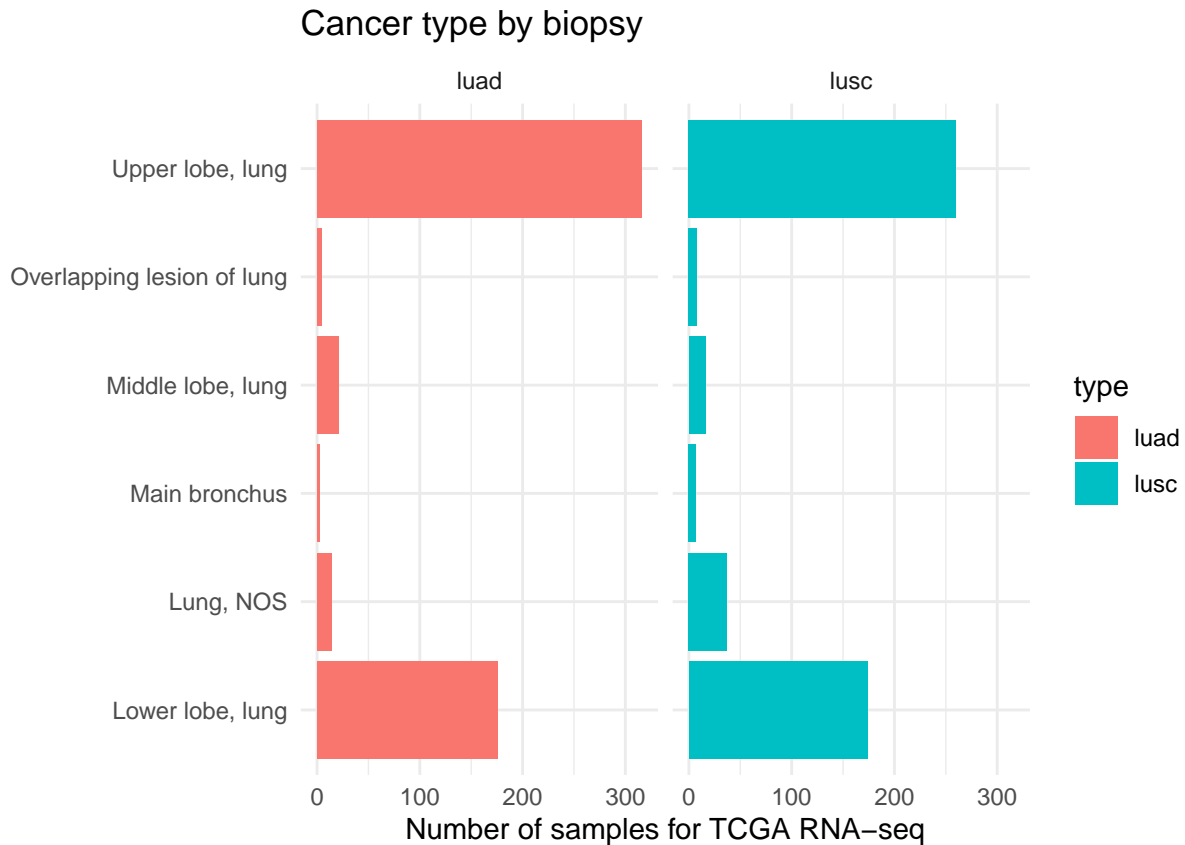
Is there difference in tumor stages by gender? Let's check with LUSC.

```
# Add gender with stages
bind_rows(d_luad %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='luad') %>%
  select(type, tumor_stage, gender),
d_lusc %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='lusc') %>%
  select(type, tumor_stage, gender)) %>%
dplyr::count(type, tumor_stage, gender) %>%
complete(gender, type, tumor_stage, fill = list(n = 0)) %>%
mutate(tumor_stage = factor(tumor_stage, levels=rev(unique(tumor_stage))),
  gender = factor(gender)) %>%
ggplot(., aes(tumor_stage, n, fill=gender)) +
geom_bar(stat="identity", position=position_dodge()) +
facet_wrap(~type) + coord_flip()
```



### 3.5. Site of biopsy

```
# Cancer type by biopsy
bind_rows(d_luad %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='luad') %>%
  select(type, site_of_resection_or_biopsy),
d_lusc %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='lusc') %>%
  select(type, site_of_resection_or_biopsy)) %>%
dplyr::count(type, site_of_resection_or_biopsy) %>%
ggplot(aes( site_of_resection_or_biopsy, n, fill=type)) +
labs(title = 'Cancer type by biopsy',
  x = '', y='Number of samples for TCGA RNA-seq') +
theme_minimal() + geom_bar(stat="identity") +
facet_wrap(~type) + coord_flip()
```



Plot the years at diagnosis by tissue or organ of origin. We would also include difference by gender.

```
bind_rows(d_luad %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='luad') %>%
  select(type, gender, age_at_diagnosis, tissue_or_organ_of_origin),
d_lusc %>%
  filter(shortLetterCode == 'TP') %>%
  mutate(type='lusc') %>%
  select(type, gender, age_at_diagnosis, tissue_or_organ_of_origin)) %>%
ggplot(., aes(gender, age_at_diagnosis/365, fill=gender)) +
geom_boxplot() + labs(y='year at diagnosis') +
facet_wrap(~tissue_or_organ_of_origin)
```

```
## Warning: Removed 40 rows containing non-finite values (stat_boxplot).
```

