FISEVIER

Contents lists available at ScienceDirect

Ecological Engineering

journal homepage: www.elsevier.com/locate/ecoleng



Short communication

Effects of inorganic nitrogen form on growth, morphology, N uptake, and nutrient allocation in hybrid Napier grass (*Pennisetum purpureum* × *Pennisetum americanum* cv. Pakchong1)



Arunothai Jampeetong ^{a,*}, Hans Brix ^b, Suwasa Kantawanichkul ^c

- ^a Department of Biology, Faculty of Science, Chiang Mai University, Meuang, Chiang Mai 50202, Thailand
- ^b Department of Bioscience, Plant Biology, Aarhus University, Ole Worms Allé 1, 8000 Aarhus C, Denmark
- ^c Department of Environmental Engineering, Faculty of Engineering, Chiang Mai University, Chiang Mai 50200, Thailand

ARTICLE INFO

Article history: Received 24 March 2014 Received in revised form 21 August 2014 Accepted 14 September 2014 Available online xxx

Keywords: Leaf transpiration Napier grass Nitrogen nutrition

ABSTRACT

Plant cultivars with high biomass production may have a high potential for being used in integrated water treatment and plant production system. The highly productive hybrid Napier grass cultivar, *Pennisetum purpureum* × *Pennisetum americanum* cv. Pakchong1, may be a candidate species for being used in such systems. We studied the effects of inorganic nitrogen form (NH₄+, NH₄NO₃ or NO₃-) on growth, morphology, N uptake, water content and mineral allocation in this species under hydroponic conditions at equimolar concentrations (500 μ mol NL⁻¹). Generally, the N-form significantly affected growth, biomass allocation and tissue nutrient and mineral composition of the plants. The hybrid Napier grass grew better on NH₄+ compared to NO₃-, and the plants supplied with NH₄+ contained three times more chlorophylls than plants supplied with NO₃- alone or NO₃- combined with NH₄+. The morphology of the plants was, however, not affected by N source, except for the shoot to root ratio, which was lower in NH₄+fed plants. The relative water content of the leaves was lowest in the NH₄+fed plants, but the transpiration rate was not affected, indicating that NH₄+ nutrition and the associated low tissue concentration of K had negative effects on the water use efficiency of the plants.

The study suggests that this hybrid Napier grass cultivar may be a new candidate species for use in integrated water treatment and plant production systems.

© 2014 Published by Elsevier B.V.

1. Introduction

Plant species or plant cultivars with high biomass production are widely used as forage crops for animal feed, as bio-energy crops for fuel production and for bio-remediation of contaminated soil or water (Dzantor et al., 2000; Somerville et al., 2010). In recent years, there has been an increasing interest in perennial C_4 grasses, such as Napier grass in South-East Asia, switch grass in the US and Miscanthus in Europe, as promising energy crops because of their high biomass yield potential and high water-use efficiency. However, these C_4 species have rarely been used in constructed wetland (CW) systems for the treatment of wastewaters. Instead high productive wetland plants species such as Phragmites, Typha, Schoenoplectus and Cyperus that are adapted to the wetland growth

conditions, but with a biomass that have limited use-potential, are the most commonly used species (Leto et al., 2013; Vymazal, 2013).

Recently, there is an increased interest in integrating water treatment systems and plant production systems based on the common-sense approach of conversion of wastes into products. By integrating water treatment and plant production, it is possible to reduce wastes and associated environmental impacts, and at the same time generate an additional crop (Naegel, 1977; Quillere et al., 1995; Rakocy et al., 2006). In China, Coix lacryma-jobi L. has been used in CWs to remove nitrogen (N) and phosphorus (P) from polluted water and at the same time produce an edible crop (Xu and Li, 2007), and different species of leaf vegetables have been integrated with the treatment of fish aquaculture (Trang et al., 2010; Trang and Brix, 2014). The high nutrient supply from nonheavy metal contaminated wastewaters is an important source of fertilizer enhancing plant growth and increasing N content in the plant tissue, making the plants have a high quality as animal feed or green manure. Most species used for water treatments are aquatic or wetland plants. Ammonium (NH₄⁺), which is the major N form in water-saturated soils, is the preferred N-form taken up

^{*} Corresponding author. Tel.: +66 53 943346 51; fax: +66 53 892259. E-mail addresses: ajampeetong@yahoo.com, arunothai.2519@gmail.com (A. Jampeetong).

by wetland species like *Phragmites australis* (Cav.) Trin. ex Steudel (Tylova-Munzarova et al., 2005), whereas nitrate (NO₃⁻), which is the dominant N-form in well-drained soils (Ponnamperuma, 1972; Bloom et al., 2003), is preferred by most terrestrial plant species. The form of N taken up may, however, interact with the water relations of plants (Guo et al., 2007a). Studies comparing wetland plant species to crop and terrestrial plant species have found that the N use efficiency of the plants are closely related to their water use efficiency (Caviglia and Sadras, 2001; Fan et al., 2002; Ripullone et al., 2004). However, variations in the N nutrition may affect the water relations of plants in different ways for different N sources. (NO₃⁻, NH₄⁺ or both) It is important to understand the interaction between plant N nutrition and water relations in order to be able to evaluate the potential applicability of an untried species in CW systems.

Pennisetum species are annual or perennial grasses belonging to the family Poaceae. They are native to Africa, but have been introduced to most tropical and subtropical countries where they have become naturalized and used as forage crops (Farrell et al., 2002). The perennial Pennisetum purpureum Schumach (Napier or elephant grass) readily cross with the annual Pennisetum americanum (L.) Leeke (pearl millet) and the resultant interspecific hybrids are more vigorous than the parent species. Hence, several hybrid Napier grass cultivars (*P. purpureum* \times *P. americanum*) have been developed (Premaratne and Premalal, 2006). Because of the high biomass production and very high forage quality for livestock, growth of these hybrids is widely promoted in subtropical and tropical countries including Thailand (Tudsri et al., 2002; Premaratne and Premalal, 2006). Recently, a very productive cultivar (Pakchong1) of this hybrid grass has been developed and marketed by Thailand's Department of Livestock Development. The Pakchong1 cultivar is claimed to have several advantages over other Napier grass cultivar, as it grows taller, has a higher content of crude protein, has an annual yield exceeding 500 t year⁻¹, and can be harvested 5-6 times per year.

Both of the parent species of this hybrid Napier, P. purpureum and P. americanum, have been used in CWs for wastewater treatments (Chikafumbwa, 1996; Kivaisi, 2001; Yang et al., 2001, 2007; Prasad, 2010), But information about growth and physiological responses under different conditions is still lacking, and in particular the N nutrition and its interaction with water relations are not well understood. The hybrid Napier grass is presently being tested for use in CWs in Thailand (unpublished data), but the basic ecophysiology of this cultivar has barely been studied. Therefore, this study aimed to assess growth, morphological characteristics and water relations of hybrid Napier grass (P. purpureum \times P. americanum cv. Pakchong1), as affected by N source (either NO₃or $\mathrm{NH_4}^+$ and combined $\mathrm{NH_4NO_3})$ in order to gain further knowledge on its nutritional ecology. This is, to our knowledge, the first study to investigate the N nutrition of this Napier grass cultivar.

2. Materials and methods

2.1. Plant material and experimental design

Plant stocks of hybrid Napier grass (P.purpureum Schumach $\times P.americanum$ (L.) Leeke cv. Pakchong1) were obtained from the Energy Research and Development Institute–Nakornping, Chiang Mai University, Thailand. In order to produce similar-sized plants for the experiment, the stems were cut into 200–300 mm sections and laid in shallow water until new shoots and adventitious roots were initiated from the stem nodes. Then the stems were cut at both sides of the nodes, and 18 similar-sized plants (approximately 70–90 mm tall; 14–18 g fresh mass) were placed in 5-L plastic buckets (diameter 0.25 m; height 0.22 m) containing a full strength

standard N and P free growth medium prepared according to Smart and Barko (1985) to which 100 μ M KHPO₄ $^{2-}$ and a commercial plant micronutrient solution (Tropica, Egaa, Denmark) were added. pH was adjusted to 6.5 ± 0.2 using HCl and NaOH. Nitrogen was added as $(NH_4^+)_2SO_4$ and KNO₃ at equimolar (500 μ mol N L $^{-1}$) concentrations to create the following three treatments: (i) 500 μ M NH₄ $^+$ (ii) 250 μ M NH₄NO₃ and (iii) 500 μ M NO₃ $^-$ (n = 6). The plants were placed in the greenhouse at the Department of Biology, Faculty of Science, Chiang Mai University, Thailand. During the experimental period, the temperature and light regimes in the greenhouse were $26\pm10\,^{\circ}$ C:14 $\pm5\,^{\circ}$ C day:night temperature and approximately 10 h light/14h dark. The growth medium was renewed every 3 days in order to minimize the depletion of nutrients from the growth medium.

2.2. Growth and morphological study

After 48 days, all plants were harvested and cleaned. The total biomass, plant height, number of new shoots, number of leaves and average area of four mature leaves, number of roots and the maximum root length were recorded. Then all plants were fragmented into shoots and roots and freeze-dried. The relative growth rate (RGR; g g⁻¹ days⁻¹) in each treatment was calculated by the formula: RGR = $(\ln W_2 - \ln W_1)/(t_2 - t_1)$, where W_1 and W_2 are the initial and final dry mass (DM), and t_1 and t_2 are initial and final time (days). The shoot elongation rate (SER; mm days⁻¹) was calculated as the total increase in shoot length throughout the experiment divided by the number of days.

2.3. Nitrogen uptake

Three days before harvest, the N uptake of plants from each treatment was determined. Selected plants (n=4) were preincubated in a N-free growth medium for 18 h in the same growth conditions as the growth study. After pre-incubation, the plants were placed in 500-mL beakers containing the basic growth medium with either $500 \,\mu\text{M} \, \text{NH}_4^+$ or NO_3^- . The solutions in the beakers were mixed by continuous air bubbling. The NO₃⁻ and NH₄⁺ uptake rates were estimated based on the N depletion in the solution during 6h (Konnerup and Brix, 2010). The NH₄⁺ concentration in the samples was analyzed using a modified salicylate method (Quikchem Method no. 10-107-06-3-B; Lachat Instruments, Milwaukee, WI, USA). The NO₃⁻ concentration was analyzed from the absorbance at 202 nm and 250 nm (Cedergreen and Madsen, 2002). After the uptake experiment, all plants were separated into shoots and roots, freeze-dried and weighed. The N uptake rates were calculated from the depletion curves with linear regression analyses and related to root DM.

2.4. Leaf desiccation

The transpiration rate and relative water content (RWC) of leaves were determined according to Nejad and van Meeteren (2005). The 3rd or 4th leaf of each plant counted from the apical shoot was sampled (n=4). All leaves were soaked with distilled water under dark conditions for 2h to bring them to maximum fresh mass. The leaves were then gently dried using blotting paper, weighed and placed on a sieve in light conditions at room temperature. The fresh mass of the leaves were then recorded every 5 min for 3h. The transpiration rates were calculated from the weight loss between two consecutive measurements and expressed per leaf area. The RWC was calculated according to González and González-Vilar (2001): RWC (%) = [(fresh mass – dry mass)]/(saturated mass – dry mass)] × 100. The RWC express the water content in percent as related to the water content at full turgor.

2.5. Leaf epidermal studies

Mature leaves (3rd or 4th counted from the apical shoot) were taken for epidermal study. The leaves were cut into small pieces $(25 \times 25 \, \text{mm})$ and placed into vials containing bleaching solution (6% sodium hypochlorite: distilled water; 1:1) for 24h. The samples were then rinsed with tap water and preserved in 70% ethanol. The density of epidermal cells and stomata as well as the length of guard cells on both the upper and lower sides of the leaves were determined using 40× light transmission-microscopy according to Hansen et al. (2007). The stomatal index was calculated as the number of stomata per unit leaf area divided by the number of epidermal cells plus guard cells per unit leaf area, then multiplied by 100 (Willmer and Fricker, 1996).

2.6. Pigments

5 mg samples of freeze-dried leaves were weighed, cut into small pieces and extracted in 96% ethanol in the dark at room temperature. After 24 h, the extracts were thoroughly mixed by vortexing and their absorbance measured at 648.6 nm and 664.2 nm using a UV-vis spectrophotometer (Lambda 25 version 2.85.04, USA). Chlorophyll a (Chl a), chlorophyll b (Chl b) and total chlorophyll (total Chl a+b) were calculated according to Lichtenthaler (1987).

2.7. Tissue mineral and nutrient concentrations

The concentrations of Ca, Fe, K, Mg, Mn, Na and P in leaves and roots were analyzed in subsamples (approximately 110–120 mg) of finely ground freeze-dried plant materials. The subsamples were digested with 4 mL concentrated HNO3 and 2 mL H2O2 in a microwave sample preparation system (Multiwave 3000, Anton Paar GmbH, Austria), and the element concentrations analyzed using inductively coupled plasmaspectrometry (Optima 2000 DV, PerkinElmer Instruments Inc., CT, USA). The total carbon (C) and nitrogen (N) contents of the leaves and roots were analyzed in the ground plant samples (1.8–3.0 mg) using a CHN analyzer (Na2000, Carlo Erba, Italy).

Table 1 Growth, morphological characteristics and chlorophyll contents (mean ± SD) of hybrid Napier grass (*Pennisetum purpureum* × *P. americanum* cv. Pakchong1) grown with either NO₃⁻, NH₄⁺ or NH₄NO₃ at equimolar N concentration (500 μM N) and results of one-way ANOVA (F-ratios).

	N source			F- ratio
	NO ₃ ⁻	NH ₄ NO ₃	NH ₄ ⁺	ratio
Relative growth rate $(g g^{-1} days^{-1})$	0.023 ± 0.003^a	0.030 ± 0.001^b	0.030 ± 0.001^b	15.4
Shoot elongate rate (mm days ⁻¹)	2.8 ± 0.1	3.3 ± 0.7	3.1 ± 0.2	1.6
Shoot:root ratio	5.3 ± 1.3^{c}	4.4 ± 0.7^{ab}	$3.8 \pm 0.4^{\text{a}}$	4.4°
Average leaf area (cm²)	37 ± 1	42 ± 14	35 ± 7	0.5
Leaf number	$\textbf{7.2} \pm \textbf{1.4}$	$\textbf{7.2} \pm \textbf{0.7}$	$\textbf{7.8} \pm \textbf{0.7}$	0.8
Root number	23 ± 4	25 ± 5	22 ± 4	1.2
Root length (cm)	17 ± 4	18 ± 3	28 ± 15	2.7
New shoot	$\boldsymbol{0.16 \pm 0.04}$	$\boldsymbol{0.5 \pm 0.8}$	0.0 ± 0.0	1.4
Chl $a \text{ (mg g}^{-1} \text{ DM)}$	$2.1\pm1.3^{\text{a}}$	2.5 ± 1.2^{a}	$10.7\pm1.4^{\rm b}$	77.8
Chl $b \text{ (mg g}^{-1} \text{ DM)}$	0.7 ± 0.4^a	0.9 ± 0.4^a	$3.6\pm0.5^{\rm b}$	86.8*
Total Chl $a + b$ (mg g ⁻¹ DM)	2.8 ± 1.8^{a}	3.4 ± 1.5^a	14.3 ± 1.9^{b}	80.4

Different letters superscripts between columns indicate significant differences between treatments.

2.8. Data analysis

Statistical analysis was performed using Statgraphics Plus version 4.1 (Manugistics, Inc., MD, USA). One-way analysis of variance (ANOVA) was used to determine the effects of treatments. Differences between treatments were identified using the Tukev's HSD post hoc procedure at the 5% significance level.

3. Results

3.1. Growth and tissue composition

The N form supplied to the plants affected the growth of hybrid Napier grass significantly (Table 1). The plants supplied with NH₄⁺ or NH₄NO₃ had higher RGRs than the NO₃⁻ fed plants. The number of roots, the root length and the average leaf area were not affected by N form. But the shoot; root ratio was affected, as plants supplied with NH₄⁺ had relatively more biomass allocated to roots than plants supplied with NO₃⁻. The concentrations of chlorophylls in the leaves, however, were approximately four times higher in NH₄⁺-fed plants than in NO₃⁻ and NH₄NO₃-fed plants.

Table 2 Total C, total N, C/N ratio and concentrations of Ca, K, Mg, Fe, Mn, P and Na (mean \pm SD) in leaves and roots of hybrid Napier grass (*Pennisetum purpureum* \times *P.* americanum cv. Pakchong 1) grown with either NO₃-, NH₄+ or NH₄NO₃ at equimolar N concentration (500 µM N) and results of one-way ANOVA (F-ratios).

	N source	F-ratio						
	NO ₃ ⁻	NH ₄ NO ₃	NH ₄ ⁺					
Total C (%)								
Leaves	34.0 ± 1.2^{a}	36.5 ± 0.8^{b}	38.9 ± 0.6^{c}	42.5				
Roots	40.6 ± 0.2^a	40.7 ± 1.5^a	41.6 ± 0.6^{b}	6.7				
Total N (%)								
Leaves	3.1 ± 0.3^a	4.0 ± 0.6^{b}	4.0 ± 0.4^{b}	7.8				
Roots	2.4 ± 0.0^a	$2.7 \pm 0.2^{\rm b}$	3.8 ± 0.2^{b}	8.9				
C/N ratio								
Leaves	11.1 ± 1.2^{b}	9.2 ± 1.2^a	10.0 ± 0.9^{ab}	4.6				
Roots	16.9 ± 0.3^b	15.1 ± 0.8^a	14.9 ± 0.5^a	9.7				
$Ca (mg g^{-1} DM)$								
Leaves	8.5 ± 3.2^{b}	8.3 ± 3.0^{b}	6.6 ± 2.0^a	0.9				
Roots	3.2 ± 0.3^{b}	$3.0\pm0.5^{\rm b}$	1.9 ± 0.2^a	10.7				
K $(mgg^{-1}D$								
Leaves	86.4 ± 7.2^{c}	63.4 ± 10.8^{b}	10.0 ± 0.9^{ab}	4.6				
Roots	16.9 ± 0.3^b	$15.1\pm0.8^{\text{a}}$	14.9 ± 0.5^a	9.7				
${ m Mg}~({ m mg}{ m g}^{-1}$								
Leaves	$3.6\pm0.8^{\rm b}$	3.4 ± 1.0^{ab}	2.6 ± 0.6^a	2.8				
Roots	1.6 ± 0.1^a	1.6 ± 0.1^a	$2.0\pm0.1^{\rm b}$	14.8**				
Fe $(mg g^{-1} DM)$								
Leaves	0.2 ± 0.1	$\textbf{0.2} \pm \textbf{0.1}$	0.2 ± 0.0	0.2				
Roots	2.1 ± 0.7	4.1 ± 1.5	3.0 ± 1.0	2.2				
$Mn (mg g^{-1} DM)$								
Leaves	0.09 ± 0.03	0.11 ± 0.05	$\boldsymbol{0.08 \pm 0.02}$	0.8				
Roots	4.5 ± 1.2^{b}	6.5 ± 2.5^{b}	0.7 ± 0.2^a	10.3				
$P (mg g^{-1} DM)$								
Leaves	23.9 ± 4.7	26.2 ± 4.3	24.7 ± 1.7	0.6				
Roots	15.9 ± 1.1^{ab}	$17.8\pm1.8^{\mathrm{b}}$	15.0 ± 0.5^{a}	3.9				
Na $(mgg^{-1}DM)$								
Leaves	1.1 ± 0.3^{ab}	$1.4\pm0.8^{\mathrm{b}}$	0.7 ± 0.4^{a}	2.4				
Roots	1.0 ± 0.1^a	1.1 ± 0.1^{a}	2.1 ± 0.4^{b}	19.9				

Different letters superscripts between columns indicate significant differences between treatments.

P < 0.05.

P < 0.01

^{***} P < 0.001

^{*} P < 0.05.

P < 0.01

^{***} P < 0.001.

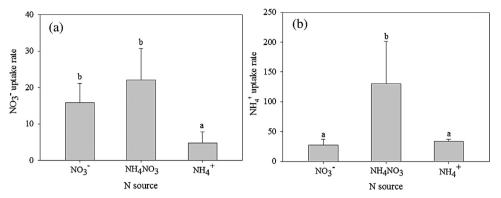


Fig. 1. (a) Average $(\pm SD) NO_3^-$ uptake rate $(\mu mol NO_3^- g^{-1} root DM h^{-1})$ and $(b) NH_4^+$ uptake rate $(\mu mol NH_4^+ g^{-1} root DM h^{-1})$ of hybrid Napier grass (Pennisetum purpureum Purp× P. americanum cv. Pakchong 1) grown with either NO₃⁻, NH₄⁺ or NH₄NO₃ at equimolar N concentration (500 µM N). Different letters above columns indicate significant differences between treatments.

The tissue concentrations C and N were significantly affected by N source as NH₄⁺-fed plants had higher concentrations of both C and N than NO₃⁻ fed plants (Table 2). The concentrations of Ca, K, Mn and Mg generally were lower in the plants supplied with NH₄⁺ compared with plants supplied with NO₃⁻. In particular, the K concentration was affected, as the K concentration in leaves of NH₄⁺-fed plants was eight times lower than the concentrations in NO₃⁻ fed plants. Na in roots of NH₄⁺-fed plants was higher than in the two other treatments, and tissue P and Fe concentrations were unaffected by the N supply.

3.2. Nitrogen uptake

The uptake rates of NH₄⁺ were generally much higher than the uptake rates of NO₃⁻, particularly in plants supplied with NH₄⁺ in combination with NO₃⁻ (Fig. 1). The highest NH₄⁺ uptake rate $(130\pm71~\mu\text{mol NH}_4{}^+\text{g}^{-1}~\text{root DM}~\text{h}^{-1})$ found in NH₄NO₃-fed plants, was approximately four times higher than the uptake rates in NH4+ and NO₃⁻ fed plants. The plants supplied with NO₃⁻ had higher NO₃⁻ uptake rates than plants supplied with NH₄⁺ alone.

3.3. Leaf desiccation

The leaf water loss rates decreased over time (Fig. 2) and were consistently higher in NH₄NO₃-fed plant than in NH₄⁺ and NO₃⁻ fed

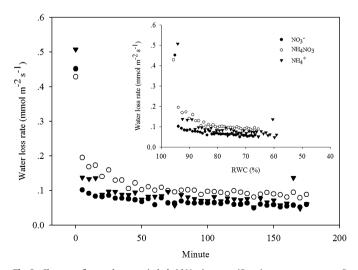


Fig. 2. Changes of water loss rate in hybrid Napier grass (*Pennisetum purpureum* \times *P.* americanum cv. Pakchong1) grown with either NO₃⁻, NH₄⁺ or NH₄NO₃ at equimolar N concentration (500 $\mu\text{M}\,\text{N}).$ The relationship between water loss rate and relative water content (RWC) is shown as an insert.

Table 3 Epidermal density, stomatal density, stomatal index and stomatal length of the upper and lower leaf surface (mean ± SD) of hybrid Napier grass (Pennisetum purpureum × P. americanum cv. Pakchong1) grown with either NO₃-, NH₄+ or NH_4NO_3 at equimolar N concentration (500 μ M N) and results of one-way ANOVA (F-ratios).

		N source			F-ratio		
		NO ₃ -	NH ₄ NO ₃	NH ₄ ⁺			
1.	Epidermal density (1	idermal density (mm ⁻²)					
	Upper leaf surface	338 ± 26^{b}	265 ± 74^a	309 ± 23^{ab}	3.00		
	Lower leaf surface	333 ± 67	281 ± 30	316 ± 52	1.34		
2.	Stomatal density (m	m^{-2})					
	Upper leaf surface	74 ± 22	67 ± 15	65 ± 13	0.39		
	Lower leaf surface	73 ± 12	73 ± 11	70 ± 13	0.07		
3.	Stomatal index (%)						
	Upper leaf surface	18 ± 4	21 ± 6	18 ± 4	0.71		
	Lower leaf surface	18 ± 4	21 ± 3	19 ± 5	0.52		
4.	Guard cell length (µ						
	Upper leaf surface	30 ± 3^a	44 ± 3^c	$37\pm5^{\rm b}$	83.6***		
	Lower leaf surface	29 ± 4^a	42 ± 2^c	39 ± 2^b	114.9***		

Different letters superscripts between columns indicate significant differences between treatments.

plants (Fig. 2). At the termination of the desiccation experiment after 3 h, the RWC of the NH₄⁺-fed plants (59.0%) was significantly lower than the RWC of the NO₃⁻ fed plants (66.6%) and NH₄NO₃-fed plants (65.5%).

3.4. Leaf epidermal studies

The density of epidermal cells on the upper side of the leaves was affected by N form, but there was no difference in epidermal density on the lower side of the leaves (Table 3). Stomatal density and stomatal index of both sides of the leaves were also not affected by the treatments. However, the length of stomata (guard cell length) differed significantly between the treatments both on the upper and lower side of the leaves. The longest guard cells were found in NH₄NO₃-fed plants followed by NH₄⁺-fed plants and NO₃⁻ fed plants, respectively.

4. Discussion

The form of N supplied to the hybrid Napier grass cultivar (P. purpureum × P. americanum cv. Pakchong1) significantly affected growth, biomass allocation and tissue nutrient and mineral

P < 0.05.

P < 0.001.

composition of the plants. The parent species, P. americanum, is very drought tolerant and susceptible to water logging (Premaratne and Premalal, 2006; Ayub et al., 2009), whereas the other parent species, P. purpureum can grow in soils that are water saturated for most of the year (Cheng et al., 2009). The hybrid Napier grass have inherited traits from both parent species and thus may be able to acclimate to the different soil conditions. including the prevailing form of N present in different soils. The hybrid Napier grass grew better on $\mathrm{NH_4}^+$ compared to $\mathrm{NO_3}^-$ as the RGR of plants supplied with NH₄⁺ alone or NH₄⁺ combined with NO₃⁻ was higher than the RGR of plants supplied with NO₃⁻ alone. Also the plants supplied with NH₄⁺ contained much more chlorophylls than plants supplied with NO₃⁻ alone or NO₃⁻ combined with NH₄⁺. The morphology of the plants was, however, not affected by N source, except for the shoot to root ratio, as NH₄⁺fed plants had relatively more biomass allocated to roots than plants supplied with NO₃⁻.

Other studies have reported contrasting results concerning N nutrition preference of plants. For instance, the free-floating water fern Salvinia natans (L.) All. (Jampeetong and Brix, 2009) and the smooth flatsedge Cyperus laevigatus L. (Piwpuan et al., 2013) are both reported to prefer NH₄⁺ over NO₃⁻ as the main N source. However, the growth rate of the common reed, *P. australis* (Cav.) Trin. Ex Steudel, were unaffected by the N-form available, but the plant had a higher uptake capacity for NH₄⁺ compared to NO₃⁻, even in fully induced NO₃⁻ fed plants (Tylova-Munzarova et al., 2005). Similar results are reported for Canna indica L. (Konnerup and Brix, 2010). The present study showed that the hybrid Napier grass cultivar (*P. purpureum* × *P. americanum* cv. Pakchong1) grew well with NH₄⁺ or NH₄NO₃, but the uptake rate of NH₄⁺ was higher than that of NO₃⁻ and resulted in higher concentrations of N in the tissues of NH₄⁺-fed plants compared to plants supplied with NO₃⁻ alone. Hence, this hybrid Napier grass cultivar has a preference for NH₄⁺ over NO₃⁻, which is consistent with the fact, that most C₄ grasses are reported to prefer NH₄⁺ (Guo et al., 2007a; Taylor et al., 2009).

The relative water content (RWC) of NH₄⁺-fed plants was lower than that of the NO₃⁻ and NH₄NO₃-fed plants. The NH₄NO₃-fed plants had the largest stomata size, and the plants maintained a higher RWC compared to NH₄⁺-fed plants with smaller stomata. Thus, the stomatal size seems not related to the RWC. Raab and Terry (1994) reported that the RWC in leaves of the beet Beta vulgaris L. supplied with NH₄⁺ was lower than in plants supplied with NO₃⁻. Several studies have reported negative effects of NH₄⁺ nutrition on plant water uptake rate (Guo et al., 2007a,b,c,c,c). This may be associated with the reduced cation uptake, especially K, of NH₄⁺-fed as the osmotic regulation of the leaves as well as stomata movement is partly controlled by K. The concentration of K in leaves of NH₄⁺-fed hybrid Napier grass was 6-8 times lower than the concentration in the leaves of plants supplied with NO₃⁻, and the concentrations of other cations (Mg, Na) were also lower. Hence, the N nutrition likely interacted with water uptake as Guo et al. (2007b) showed that NH₄⁺ nutrition significantly affected aquaporin expression in the common bean Phaseolus vulgaris L.

As the hybrid Napier grass (P. purpureum \times P. americanu cv. Pakchong1) grew very well on NH_4^+ , this hybrid Napier grass cultivar may have great potential for bioremediation in CW systems used to treat various types of wastewaters. Its extensive root system and high growth rate may result in a significant uptake of nutrients from the polluted water, and at the same time the large amount of biomass produced can be harvested and used for varies purposes. The species seems to have potential for being used in an integrated water treatment and plant production system. However, further studies with this hybrid Napier cultivar under realistic CW conditions must be carried out to assess the real potential of using this plant species in CWs.

Acknowledgments

This research was funded by the Institute for the Promotion of Teaching Science and Technology (IPST), Thailand. Finally, thanks to the Danish Council for Independent Research - Natural Sciences, via a grant to Hans Brix.

References

- Ayub, M., Nadeem, M.A., Tahir, M., Ibrahim, M., Aslam, M.N., 2009. Effect of nitrogen application and harvesting intervals on forage yield and quality of pearl millet (*Pennisetum americanum* L.). Pak. J. Life Soc. Sci. 7, 185–189.
- Bloom, A.J., Meyerhoff, P.A., Taylor, A.R., Rost, T.J., 2003. Root development and absorption of ammonium and nitrate from the rhizosphere. J. Plant Growth Regul. 21, 416–431.
- Caviglia, O.P., Sadras, V.O., 2001. Effect of nitrogen supply on crop conductance: water- and radiation-use efficiency of wheat. Field Crops Res. 69, 259–266.
- Cedergreen, N., Madsen, T.V., 2002. Nitrogen uptake by the floating macrophyte Lemna minor. New Phytol. 155, 285–292.
- Cheng, X.-Y., Chen, W.-Y., Gu, B.-H., Liu, X.-C., Chen, F., Chen, Z.-H., Zhou, X.-Y., Li, Y.-X., Huang, H., Chen, Y.-J., 2009. Morphology, ecology, and contaminant removal efficiency of eight wetland plants with different root systems. Hydrobiologia 623, 77–85
- Chikafumbwa, F.J.K., 1996. The use of Napier grass (*Pennisetum purpureum*) and maize (*Zea mays*) bran as low-cost tilapia aquaculture inputs. Aquaculture 146, 101–107.
- Dzantor, E.K., Chekol, T., Vough, L.R., 2000. Feasibility of using forage grasses and legumes for phytoremediation of organic pollutants. J. Environ. Sci. Health 35, 1645–1661.
- Fan, X., Zhang, J., Wu, P., 2002. Water and nitrogen use efficiency of lowland rice in ground covering rice production system in south China. J. Plant Nutr. 25, 1855–1862.
- Farrell, G., Simons, S.A., Hillocks, R.J., 2002. Pests, diseases, and weeds of Napier grass, *Pennisetum purpureum*: a review. Int. J. Pest Manage. 48, 39–48.
- González, L., González-Vilar, M., 2001. Determination of relative water content. In: Reigosa Roger, M.J. (Ed.), Handbook of Plant Ecophysiology Technique. Kluwer Academic Publishers, Netherlands, pp. 207–212.
- Guo, S., Zhou, Y., Shen, Q., Zhang, F., 2007a. Effect of ammonium and nitrate nutrition on some physiological processes in higher plants-growth photosynthesis, photorespiration, and water relations. Plant Biol. 9, 21–29.
- Guo, S., Kaldenhoff, R., sattelmacher, B., Brueck, H., 2007b. Relationship between water and nitrogen uptake in nitrate- and ammonium-supplied *Phaseolus vulgaris* L. plants. J. Plant Nutr. Soil Sci. 170, 73–80.
- Guo, S., Shen, Q., Brueck, H., 2007c. Effects of local nitrogen supply on water uptake of bean plants in a split root system. J. Integr. Plant Biol. 49, 472–480.
- Hansen, D.L., Lambertini, C., Jampeetong, A., Brix, H., 2007. Clone-specific differences in *Phragmites australis*: effects of ploidy level and geographic origin. Aquat. Bot. 86, 269–279.
- Jampeetong, A., Brix, H., 2009. Nitrogen nutrition of Salvinia natans: effect of inorganic nitrogen form on growth, morphology, nitrate reductase activity and uptake kinetics of ammonium and nitrate. Aquat. Bot. 90, 67–73.
- Kivaisi, A.K., 2001. The potential for constructed wetlands for wastewater treatment and reuse in developing countries: a review. Ecol. Eng. 16, 545–560.
- Konnerup, D., Brix, H., 2010. Nitrogen nutrition of *Canna indica*: effects of ammonium versus nitrate on growth, biomass allocation, photosynthesis, nitrate reductase activity and N uptake rates. Aquat. Bot. 92, 142–148.
- Leto, C., Tuttolomondo, T., La Bella, S., Leone, R., 2013. Effects of plant species in a horizontal subsurface flow constructed wetland–phytoremediation of treated urban wastewater with Cyperus alternifolius L. and Typha latifolia L. in the West of Sicily (Italy). Ecol. Eng. 61, 282–291.
- Lichtenthaler, H.K., 1987. Chlorophylls and carotenoids: pigments of photosynthetic biomembranes. Methods Enzymol. 148. 350–382.
- Naegel, L.C.A., 1977. Combined production of fish and plants in recirculating water. Aquaculture 10, 17–24.
- Nejad, A.R., van Meeteren, U., 2005. Stomatal response characteristics of Tradescantia virginiana grown at high relative air humidity. Physiol. Plant. 125, 324–332.
- Piwpuan, N., Zhai, X., Brix, H., 2013. Nitrogen nutrition of *Cyperus laevigatus* and *Phormium tenax*: effects of ammonium versus nitrate on growth, nitrate reductase activity and N uptake kinetics. Aquat. Bot. 106, 42–51.
- Ponnamperuma, F.N., 1972. The chemistry of submerged soils. Adv. Agron. 24, 29–96.
- Prasad, M.N.V., 2010. Exploring the potential of wetland plants for clean up of hazardous waste. J. Basic Appl. Biol. 4, 18–28.
- Premaratne, S., Premalal, G.G.C., 2006. Hybrid Napier (*P. purpureum* × *P. americanum*) var. CO-3: a resourceful fodder grass for dairy development in Sri Lanka. J. Agric. Sci. 2, 22–33.
- Quillere, I., Roux, L., Marie, D., Roux, Y., Gosse, F., Morotgaudry, J.F., 1995. An artificial productive ecosystem based on a fish/bacteria/plant association. 2. Performance. Agric. Ecosyst. Environ. 53, 19–30.
- Raab, T.K., Terry, N., 1994. Nitrogen source regulation of growth and photosynthesis in *Beta vulgaris* L. Plant Physiol. 105, 1159–1166.
- Rakocy, J.E., Masser, M.P., Losordo, T.M., 2006. Recirculating Aquaculture Tank Production Systems: Aquaponics Integrating Fish and Plant Culture. SRAC

- Publication No. 454. Southern Regional Aquaculture Center. Texas A & M University, Texas, USA.
- Ripullone, F., Lauteri, M., Grassi, G., Amato, M., Borghetti, M., 2004. Variation in nitrogen supply change water-use efficiency of *Pseudotsuga menziesii* and *Populus* × *euroamericana*; a comparison of three approaches to determine water-use efficiency. Tree Physiol. 24, 671–679.
- Smart, R.M., Barko, J.W., 1985. Laboratory culture of submerged freshwater macrophytes on natural sediments. Aquat. Bot. 21, 251–263.
- Somerville, C., Youngs, H., Taylor, C., Davis, S.C., Long, S.P., 2010. Feed stocks for lignocellulosic biofuels. Science 329, 790–792.
- Taylor, S.H., Stephen, P.H., Rees, M., Ripley, B.S., Woodward, F.I., Osborne, C.P., 2009. Ecophysiology traits in C_3 and C_4 grass: a phylogenetically controlled screening experiment. New Phytol. 185, 780–791.
- Trang, N.T.D., Schierup, H.-H., Brix, H., 2010. Leaf vegetables for use in intergrated hydroponics and aquaculture systems: effects of flooding on growth mineral composition and nutrient uptake. Afr. J. Biotechnol. 9, 4186–4196.
- Trang, N.T.D., Brix, H., 2014. Use of planted biofilters in integrated recirculating aquaculture–hydroponics systems in the Mekong Delta, Vietnam. Aquacult. Res. 45, 460–469.

- Tudsri, S., Jorgensen, S.T., Riddach, P., Pookpakdi, A., 2002. Effect of cutting height and dry season closing date on yield and quality of five Napier grass cultivars in Thailand. Trop. Grasslands 36, 248–252.
- Tylova-Munzarova, E., Lorenzen, B., Brix, H., Votrubova, O., 2005. The effects of NH₄⁺ and NO₃⁻ on growth, resource allocation and nitrogen uptake kinetics of *Phragmites australis* and *Glyceria maxima*. Aquat. Bot. 81, 326–342.
- Vymazal, J., 2013. Emergent plants used in free water surface constructed wetlands: a review. Ecol. Eng. 61P, 582–592.
- Willmer, C., Fricker, M., 1996. Stomata. Topics in Plant Functional Biology: 2. London, Chapman & Hall.
- Xu, D.F., Li, Y.X., 2007. Screen plants and substrates of the constructed wetland for treatment of wastewater. Wetland Sci. 5, 32–38.
- Yang, L., Chang, H.-T., Huang, M.-N.L., 2001. Nutrient removal in gravel- and soil-based wetland microcosms with and without vegetation. Ecol. Eng. 18, 91–105.
- Yang, Q., Chen, Z.-H., Zhao, J.-G., Gu, B.H., 2007. Contaminant removal of domestic wastewater by constructed wetlands: effects of plant species. J. Integr. Plant Biol. 49, 437–446.