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**Algorithm 1** Loading and Preprocessing

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1: procedure LOAD_FASTA_FILES(folder)
2:   list all “.fasta”/“.fas” files in folder
3:   initialize empty dict sequences_dict
4:   for each file in files do
5:     parse sequences into list
6:     sequences_dict[file]  $\leftarrow$  list of sequences
7:   end for
8:   return sequences_dict
9: end procedure
10: procedure ADJUST_THRESHOLD(sequences, init_threshold)
11:    $n \leftarrow$  number of sequences
12:   if  $n < 2$  then
13:     return init_threshold
14:   end if
15:   compute avg_sim = mean pairwise similarity
16:    $thr \leftarrow init\_threshold \times (1 + avg\_sim)$ 
17:    $thr \leftarrow \min(\max(thr, 0.2), 0.8)$ 
18:   return thr
19: end procedure
20: procedure PROCESS_GAPS(sequences, gap_threshold)
21:   convert sequences to char matrix
22:   compute gap fraction per column
23:   delete columns where fraction  $>$  gap_threshold
24:   fill remaining gaps with most common base
25:   return original_sequences, processed_sequences
26: end procedure
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**Algorithm 2** Conserved Region Detection

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1: procedure FIND_CONSERVED_REGIONS(proc_seqs, min_len, max_len, thr)
2:   pad all seqs to equal length and transpose
3:   for each column compute base frequency and label (freq > thr)
4:   scale features; train linear SVM; predict labels
5:   extract contiguous regions where label==1
6:   filter regions with length in [min_len, max_len]
7:   return regions, predictions, avg_accuracy
8: end procedure
9: procedure EXPORT_RESULTS(regions, out_folder, thr, preds, fname, acc,
   orig_seqs)
10:   create output file path
11:   for each region in regions do
12:     write header with index, fname, thr, original start, validation
13:     write region sequence
14:   end for
15:   write "Average accuracy: " acc
16:   return output_file_path
17: end procedure
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**Algorithm 3** Parsing and Difference Calculation

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1: procedure PARSE_FASTA(lines)
2:   initialize empty list seqs
3:   for each line in lines do
4:     if line starts with “i” then
5:       start new sequence entry
6:     else
7:       append to current sequence
8:     end if
9:   end for
10:  return list of (ID, sequence)
11: end procedure
12: procedure MODIFY_AND_WRITE_SEQUENCES(test_seqs, cons_seqs,
13:   out_folder, in_file)
14:  compute gap indices from cons_seqs
15:  for each test_seq in test_seqs do
16:    remove chars at gap indices
17:    write to new file in out_folder
18:  end for
19:  return list of output file paths
20: end procedure
21: procedure CALCULATE_DIFFERENCE(test_seq, species_seqs)
22:  filter species_seqs length  $\geq$  len(test_seq)
23:  compute mismatches and p_obs per sequence
24:   $avg\_diff \leftarrow \overline{p\_obs} \times 100$ 
25:  build pct_distribution and base_distribution
26:  return avg_diff, pct_distribution, base_distribution
27: end procedure
28: procedure PROCESS_DIFFERENCE(test_file, species_folder, out_xlsx)
29:  read test segments
30:  for each segment do
31:    call CALCULATE_DIFFERENCE
32:    collect summary, pct_details, base_details
33:  end for
34:  write DataFrames to Excel
35:  generate heatmaps for pct and base_details
36: end procedure
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**Algorithm 4** Identity Calculation and Utilities

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1: procedure CALCULATE_IDENTITY(test_seq, species_seqs)
2:   filter species_seqs length  $\geq$  len(test_seq)
3:   compute single_base identity rates
4:   build consensus sequence
5:   compute avg_percent_identity and percent_distribution
6:   return avg_percent_identity, percent_distribution, single_base_rates
7: end procedure
8: procedure PROCESS_IDENTITY(test_file, species_folder, out_xlsx)
9:   same as PROCESS_DIFFERENCE but use CALCULATE_IDENTITY
10: end procedure
11: procedure READ_ACCESSION_NUMBERS(txt_file)
12:   read each nonempty line as accession
13:   return list of accession numbers
14: end procedure
15: procedure FETCH_GENOME_FROM_ACCESSION(acc)
16:   call Entrez.efetch for FASTA
17:   return SeqRecord or None
18: end procedure
19: procedure FETCH_CDS_FROM_ACCESSION(acc)
20:   call Entrez.efetch for GenBank
21:   extract features with type "CDS"
22:   return list of CDS features
23: end procedure
24: procedure PROCESS_FASTA_FILES(folder, modify_titles, new_title, re-
    name_flag)
25:   list .fasta/.fas files
26:   for each file do
27:     clean degenerate bases; modify title per flags
28:     overwrite file
29:   end for
30: end procedure
31: procedure PROCESS_FASTA_FOR_PRIMERS(in_file, out_xlsx, primer_len)
32:   parse FASTA into name→seq dict
33:   for each seq do
34:     extract forward/reverse primers
35:     compute GC% and Tm
36:   end for
37:   write results to Excel
38: end procedure
39: procedure REPLACE_BASES_AND_FORMAT(in_file, out_docx)
40:   read text from txt or docx
41:   split into chunks of max_capacity
42:   for each chunk do
43:     generate QR code with rounded modules and gradient
44:     compress and save image
45:   end for
46:   create Word doc; replace bases with styled runs
47:   save doc; return image paths
48: end procedure
49: procedure GENERATE_HEATMAP(data, title, prefix, annotate)
50:   choose rasterization if large
51:   call seaborn.heatmap with vmin=0, vmax=100
52:   customize ticks for num and prop heatmaps
53:   save PNG and SVG
54: end procedure
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