# N-Mixture/CMR Exploratory - Pennsylvania

# **Load Packages**

```
if (!require(librarian)){
  install.packages("librarian")
  library(librarian)
}
librarian::shelf(tidyverse, RPostgres, DBI, unmarked, here, lubridate, kableExtra)
```

# Connect to survey\_data schema in ribbitr database

```
tryCatch({
    drv <- dbDriver("Postgres")</pre>
    print("Connecting to Database...")
    connection <- dbConnect(drv,</pre>
                 dbname = Sys.getenv("aws_dbname"),
                 host = Sys.getenv("aws_host"),
                 port = Sys.getenv("aws_port"),
                 user = Sys.getenv("aws_user"),
                 password = Sys.getenv("aws_password"),
                 timezone=NULL)
    print("Database Connected!")
    },
    error=function(cond) {
            print("Unable to connect to Database.")
    })
#search path
dbExecute(connection, "set search_path to survey_data")
```

# Query 2022 CMR Penn data

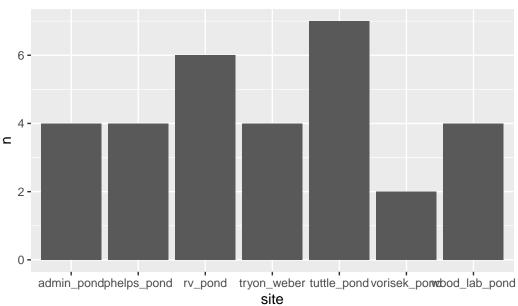
```
# Data
cmr_q <- "select r.region, s.site, v.date, v.survey_time, s2.duration_minutes,</pre>
          c.species_capture, c.capture_type
          from region r
          join site s on r.region_id = s.region_id
          full join visit v on s.site_id = v.site_id
          join survey s2 on v.visit_id = s2.visit_id
          join capture c on s2.survey_id = c.survey_id
          where r.region = 'pennsylvania'
          and v.date > '2022-01-01';"
raw_data <- dbGetQuery(connection, cmr_q) %>%
  select(!c(region, survey_time, duration_minutes)) %>%
  arrange(date)
# find all visits
visit_cmr_q <- "select r.region, s.site, v.date, v.survey_time</pre>
                from region r
                join site s on r.region_id = s.region_id
                join visit v on s.site_id = v.site_id
                where r.region = 'pennsylvania'
                and v.date > '2022-01-01';"
raw_visits <-dbGetQuery(connection, visit_cmr_q) %>%
  arrange(date) %>%
  select(site, date)
```

#### Visualize all Visits

```
viz <- raw_visits %>%
  group_by(site) %>%
  summarise(n = n())

ggplot(data = viz) +
  geom_col(aes(x=site, y = n)) +
  ggtitle("Raw Visits")
```





## Clean up species list with counts

Filter for CMR focal species and summarize species counts. Then populate zeros for 3 focal species into the data set for visits when captures did not occur.

`summarise()` has grouped output by 'date', 'site'. You can override using the `.groups` argument.

```
Joining with `by = join_by(site, date)`

Warning in left_join(., mid_clean_up): Detected an unexpected many-to-many relationship betweet
i Row 1 of `x` matches multiple rows in `y`.
i Row 34 of `y` matches multiple rows in `x`.
i If a many-to-many relationship is expected, set `relationship =
   "many-to-many"` to silence this warning.
```

# rana\_catesbeiana

#### Table formatting

Filter data for rana\_catesbeiana, tally the numbner of visits, pivot data frame into correct matrix form, and finally populate zeros into the NAs if sites were visited.

site	visit_1	visit_2	visit_3	visit_4	visit_5	visit_6	visit_7
wood_lab_pond	2	4	3	0	NA	NA	NA
phelps_pond	4	3	6	0	NA	NA	NA
admin_pond	9	15	20	0	NA	NA	NA
rv_pond	1	7	5	13	29	11	NA
tuttle_pond	15	22	0	39	31	29	20
tryon_weber	0	0	0	NA	NA	NA	NA
vorisek_pond	14	13	NA	NA	NA	NA	NA

### static n-mixture unmakred models no covariates

```
bull_frog_unmarked_nmixture <- bull_frog_visits %>%
  ungroup() %>%
  select(!c(site))

umf <- unmarkedFramePCount(y = bull_frog_unmarked_nmixture)
summary(umf)</pre>
```

# unmarkedFrame Object

## 7 sites

Maximum number of observations per site: 7 Mean number of observations per site: 4.29

Sites with at least one detection: 6

Tabulation of y observations:

```
0
     1
          2
               3
                             6
                                 7
                                      9 11
                                             13
                                                  14
                                                        15
                                                             20
                                                                 22
                                                                      29
7
                   2
                             1
                                      1
                                          1
          1
31
    39 <NA>
1
     1
         19
```

```
fm1 <- pcount(~1 ~1, data = umf)</pre>
```

Warning in pcount(~1 ~ 1, data = umf): K was not specified and was set to 139.

```
# Get estimates on link scale
backTransform(fm1, "state")
```

```
Backtransformed linear combination(s) of Abundance estimate(s)

Estimate SE LinComb (Intercept)
40.7 7.97 3.71 1

Transformation: exp

# Get estimates on natural scale
backTransform(fm1, "det")

Backtransformed linear combination(s) of Detection estimate(s)

Estimate SE LinComb (Intercept)
0.232 0.0421 -1.2 1

Transformation: logistic
```

Assuming the sites and enviro variables are exactly the same we can say there is 40.7 +-7.9 frogs at these sites and we are only detecting 23% of the frogs.

## pseudacris\_crucifer

### **Table formatting**

Filter data for pseudacris\_crucifer, tally the numbner of visits, pivot data frame into correct matrix form, and finally populate zeros into the NAs if sites were visited.

```
!is.na(lag(.)), 0, .)))
kable(peep_frog_visits)
```

site	visit_1	visit_2	visit_3	visit_4	visit_5	visit_6	visit_7
wood_lab_pond	2	0	0	0	NA	NA	NA
phelps_pond	4	0	0	0	NA	NA	NA
admin_pond	4	2	3	1	NA	NA	NA
rv_pond	16	0	0	0	0	0	NA
tuttle_pond	0	0	0	0	0	0	0
tryon_weber	0	0	0	NA	NA	NA	NA
vorisek_pond	0	0	NA	NA	NA	NA	NA

# static n-mixture unmakred models no covariates

```
peep_unmarked_nmixture <- peep_frog_visits %>%
  ungroup() %>%
  #filter(capture_type == "new") %>%
  select(!c(site))

umf <- unmarkedFramePCount(y = peep_unmarked_nmixture)
summary(umf)</pre>
```

unmarkedFrame Object

```
7 sites
```

Maximum number of observations per site: 7 Mean number of observations per site: 4.29 Sites with at least one detection: 4

Tabulation of y observations:

```
0 1 2 3 4 16 <NA> 23 1 2 1 19
```

```
fm1 <- pcount(~1 ~1, data = umf)</pre>
```

```
Warning in pcount(~1 ~ 1, data = umf): K was not specified and was set to 116.
```

```
backTransform(fm1, "state")

Backtransformed linear combination(s) of Abundance estimate(s)

Estimate SE LinComb (Intercept)
85.8 24.4 4.45 1

Transformation: exp

backTransform(fm1, "det")

Backtransformed linear combination(s) of Detection estimate(s)

Estimate SE LinComb (Intercept)
0.0124 0.00411 -4.38 1

Transformation: logistic
```

Assuming the sites and enviro variables are exactly the same we can estimate there is 85.7 +-24.4 frogs at these sites and we are only detecting 1.2% of the frogs.

### rana\_clamitans

#### Table formatting

Filter data for rana\_clamitans, tally the number of visits, pivot data frame into correct matrix form, and finally populate zeros into the NAs if sites were visited.

```
ungroup() %>%
    group_by(site, n_visit, ) %>%
    summarise(n = sum(n)) \%>\%
    ungroup() %>%
    pivot_wider(names_from = c("n_visit"), values_from = c("n")) %>%
    #add_row(site = "phelps_pond", capture_type = "recapture") %>%
    group by(site) %>%
    mutate(across(contains("visit"),
                  ~ifelse(is.na(.) &
                            !is.na(lag(.)), 0, .)))
`summarise()` has grouped output by 'site'. You can override using the
`.groups` argument.
static n-mixture with no co-variates
  green_frog_unmarked_nmixture <- green_frog_visits %>%
    ungroup() %>%
    select(!c(site))
  umf <- unmarkedFramePCount(y = green_frog_unmarked_nmixture)</pre>
  summary(umf)
unmarkedFrame Object
7 sites
Maximum number of observations per site: 7
Mean number of observations per site: 4.29
Sites with at least one detection: 7
Tabulation of y observations:
                       5 7
             2
                                 8 10 16 19 22 <NA>
                              1 1 1 1 1 19
  fm1 <- pcount(~1 ~1, data = umf)</pre>
```

Warning in pcount(~1 ~ 1, data = umf): K was not specified and was set to 122.

Assuming the sites and enviro variables are exactly the same we estimate there is 94.8 + 13.1 frogs at these sites and we are only detecting 4.3% of the frogs.

### **Table formating capture-recapture**

```
# mid clean up <- raw data %>%
   filter(species_capture %in% c("pseudacris_crucifer", "rana_catesbeiana",
                                  "rana clamitans")) %>%
   mutate(capture_type = if_else(is.na(capture_type), "new", capture_type)) %>%
   group_by(date, site, capture_type, species_capture) %>%
   summarise(n = n()) \%>\%
#
   ungroup()
# # populate zeros
# clean_up <- raw_visits %>%
   left_join(mid_clean_up) %>%
   complete(nesting(date, site),
#
             species_capture = unique(mid_clean_up$species_capture),
             fill = list(n = 0)) \%
#
#
   mutate(capture_type = if_else(is.na(capture_type), "new", capture_type))
```

## **Bullfrog CMR data**

```
# bull_frog_cmr <- clean_up %>%
   select(site, date, capture_type, species_capture, n) %>%
   filter(species_capture == "rana_catesbeiana") %>%
   select(!species_capture) %>%
#
   group_by(site) %>%
#
   mutate(n_visit = match(date, unique(date)),
           n_visit = paste0("visit_", n_visit, sep = "")) %>%
   select(!date) %>%
   ungroup() %>%
   pivot_wider(names_from = c("n_visit"), values_from = c("n")) %>%
   add_row(site = "phelps_pond", capture_type = "recapture") %>%
   add_row(site = "tryon_weber", capture_type = "recapture", visit_1 = 0, visit_2 = 0, vi
#
   group_by(site) %>%
   mutate(across(contains("visit"),
                  ~ifelse(is.na(.) &
#
                            !is.na(lag(.)), 0, .)))
# t <- clean_up %>%
   select(site, date, species_capture, n) %>%
   filter(species_capture == "rana_catesbeiana") %>%
   select(!species_capture) %>%
   group_by(site) %>%
   mutate(n_visit = match(date, unique(date)),
#
           n_visit = paste0("visit_", n_visit, sep = "")) %>%
#
   select(!date) %>%
   ungroup() %>%
   pivot_wider(names_from = c("n_visit"), values_from = c("n")) %>%
   group_by(site) %>%
   mutate(across(contains("visit"),
                  ~ifelse(is.na(.) &
                            !is.na(lag(.)), 0, .)))
#
```