

Uterine Leiomyoma Beadchip Gene Expressions

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This is to re-examine the UL and non-UL samples from the Gene Expression Omnibus online data repository (GEO) for genotypes in the ULs compared to those samples without tumor tissue in them. The accession IDs for the Series is [GSE95101](#) and for the platform is [GPL13376](#)

Lets look at some of these copy number variants of one gene with seven [copy number variants](#) or CNVs and see where the changes in the nucleotide sequences occur. Copy number variations in nucleotides can have short repeats, jumps in sequence, or deletions of a gene. I have been calling these CNVs [genotypes](#), which are the traits and alleles responsible for the physical traits or phenotypes of an organism. Some CNVs are responsible for diseases, and in tumors there are many different CNVs that are found to be responsible. A uterine leiomyoma or fibroid is a benign tumor. These samples were taken from uterus tissue with these uterine tumors and the same neighboring uterine tissue without uterine tumors.

```
library(dplyr)
library(tidyr)
library(e1071)
library(caret)
library(randomForest)
library(MASS)
library(gbm)

UL1a <- read.csv('UL1a.csv', sep=',',
                header=T, na.strings=c('', ' '))
UL1b <- read.csv('UL1b.csv', sep=',',
                header=T, na.strings=c('', ' '))
UL1c <- read.csv('UL1c.csv', sep=',',
                header=T, na.strings=c('', ' '))
UL1d <- read.csv('UL1d.csv', sep=',',
                header=T, na.strings=c('', ' '))

UL1 <- rbind(UL1a,UL1b,UL1c,UL1d)
rm(UL1a,UL1b,UL1c,UL1d)
str(UL1)

## 'data.frame':    48701 obs. of  51 variables:
## $ X              : int  1 2 3 4 5 6 7 8 9 10 ...
## $ ID             : Factor w/ 48701 levels "ILMN_1343289",...: 1 2 3
4 5 6 7 8 9 10 ...
## $ Species        : Factor w/ 1 level "Homo sapiens": 1 1 1 1 1 1 1
```

```

1 1 1 ...
## $ Source          : Factor w/ 3 levels "ILMN_Controls",...: 1 1 2 1 1
2 2 2 2 2 ...
## $ Search_Key       : Factor w/ 46721 levels
"ILMN_10001","ILMN_10014",...: 11664 11664 11664 11664 11664 11664 11664 9848
11306 1417 ...
## $ Transcript       : Factor w/ 46724 levels
"ILMN_10001","ILMN_10014",...: 2290 2291 1377 2292 2293 4903 1218 9858 11314
1421 ...
## $ ILMN_Gene        : Factor w/ 43186 levels "1-Dec","1-Mar",...: 8904
10174 2074 10143 10156 88 2713 4872 9250 2029 ...
## $ Source_Reference_ID : Factor w/ 46721 levels "NM_000015.1",...: 776
4636 1225 1123 1827 1130 1438 9379 7158 10894 ...
## $ RefSeq_ID         : Factor w/ 28570 levels "NM_000015.1",...: 776
4636 1225 1123 1827 1130 1438 9379 7158 10894 ...
## $ Unigene_ID        : Factor w/ 18153 levels "NA","Hs.100554",...: 1 1
1 1 1 1 1 1 1 1 ...
## $ Entrez_Gene_ID     : Factor w/ 16063 levels "10","10000","10001",...:
10744 10744 1487 10744 10744 5871 2331 6516 10718 5545 ...
## $ GI                 : int  14141192 20149305 25453469 4507728 4507744
5016088 7669491 88954077 33469136 89070645 ...
## $ Accession          : Factor w/ 46721 levels "NM_000015.1",...: 776
4636 1225 1123 1827 1130 1438 9379 7158 10894 ...
## $ Symbol             : Factor w/ 25036 levels "1-Dec","1-Mar",...: 8904
10174 2074 10143 10156 88 2713 4872 9250 2029 ...
## $ Protein_Product    : Factor w/ 28173 levels "NA","NP_000006.1",...: 1
1 1224 1 1 1129 1437 9300 7155 10815 ...
## $ Probe_Id          : Factor w/ 48701 levels "ILMN_1343289",...: 1 2 3
4 5 6 7 8 9 10 ...
## $ Array_Address_Id   : int  2140735 6550370 2690379 4590356 4260048
5860528 1770601 50270 3310274 7040079 ...
## $ Probe_Type         : Factor w/ 3 levels "A","I","S": 3 3 3 3 3 3 3 3
3 2 ...
## $ Probe_Start        : int  416 1856 1293 1408 72 1725 930 1 1103 2975
...
## $ SEQUENCE           : Factor w/ 48701 levels
"AAAAAACAGGAATAGCTCTAGGAGTCCTTACACAGGTCCGAGGGACCAGC",...: 9453 9026 11512 4877
6702 9532 5532 1967 10077 11476 ...
## $ Chromosome         : Factor w/ 56 levels "1","10","11",...: 27 27 22
27 27 23 4 14 1 27 ...
## $ Probe_Chr_Orientation: Factor w/ 3 levels "-","+","NA": 3 3 1 3 3 1 2 2
1 3 ...
## $ Probe_Coordinates  : Factor w/ 41351 levels "100000925-100000974",...:
10161 10161 8915 10161 10161 7449 8192 4231 2830 10161 ...
## $ Cytoband           : Factor w/ 3676 levels "10p11.1d","10p11.21a",...:
2501 2501 2039 2501 2501 2165 288 1490 1036 2033 ...
## $ Definition          : Factor w/ 46614 levels "Homo sapiens 1-
acylglycerol-3-phosphate O-acyltransferase 1 (lysophosphatidic acid
acyltransferase, alpha) (AGP"| __truncated__,...: 5692 7144 2221 7047 6692 98
2786 8195 6195 7895 ...

```

```
## $ Ontology_Component : Factor w/ 7849 levels "A 20S multiprotein
assembly of total mass about 1.2 MDa that activates dynein-based activity in
vivo. A large s"| __truncated__,...: 2614 1893 1304 1321 1266 2112 1468 1893
1763 269 ...
## $ Ontology_Process : Factor w/ 8950 levels "[goid 6069] [pmid
1755855] [evidence IDA]; A change in state or activity of a cell or an
organism (in terms of "| __truncated__,...: 2067 2257 3716 1085 547 417 1784
1091 1091 962 ...
## $ Ontology_Function : Factor w/ 9453 levels "[goid 16505] [pmid
10426319] [evidence NAS]",...: 3847 2486 1975 1911 2750 2514 95 3637 3637 828
...
## $ Synonyms : Factor w/ 16472 levels "0610037N12Rik; RPP20;
RPP2",...: 4591 4591 5325 4591 4591 5300 2402 4591 3649 4591 ...
## $ Obsolete_Probe_Id : Factor w/ 16878 levels "0610037N12Rik; RPP20;
RPP2",...: 4784 4784 645 6450 1251 5508 2490 4784 3785 4784 ...
## $ GB_ACC : Factor w/ 46717 levels "NA","NM_000015.1",...:
777 4635 1225 1123 1827 1130 1438 9377 7156 10892 ...
## $ GSM2496185 : num 13942 23759 27434 3092 6857 ...
## $ GSM2496186 : num 12934 15091 26473 4269 7799 ...
## $ GSM2496187 : num 11909 22609 23964 3455 7954 ...
## $ GSM2496188 : num 12147 18225 27823 4258 7380 ...
## $ GSM2496189 : num 14142 20728 24486 3333 6445 ...
## $ GSM2496190 : num 11650 19582 26225 4545 9215 ...
## $ GSM2496191 : num 12786 19105 28200 3413 10031 ...
## $ GSM2496192 : num 9383 10008 27997 3191 7428 ...
## $ GSM2496193 : num 11481 10575 23172 3597 7712 ...
## $ GSM2496203 : num 4136 1028 16324 4994 6466 ...
## $ GSM2496204 : num 11458 17921 26664 3095 7471 ...
## $ GSM2496205 : num 15445 18186 25687 3138 7047 ...
## $ GSM2496206 : num 11098 8905 22094 2473 6307 ...
## $ GSM2496207 : num 11510 9721 21161 4353 4826 ...
## $ GSM2496208 : num 11446 11451 26427 3863 7069 ...
## $ GSM2496209 : num 9945 16387 27837 3027 6462 ...
## $ GSM2496217 : num 12707 18456 28792 3251 8407 ...
## $ GSM2496218 : num 12261 19342 25018 2322 6925 ...
## $ GSM2496219 : num 11087 9198 27179 4554 9100 ...
## $ GSM2496220 : num 11746 21023 29030 4131 7771 ...
```

```
nonUL1a <- read.csv('nonUL1a.csv', sep=',',
                    header=T, na.strings=c('', ' '))
nonUL1b <- read.csv('nonUL1b.csv', sep=',',
                    header=T, na.strings=c('', ' '))
nonUL1c <- read.csv('nonUL1c.csv', sep=',',
                    header=T, na.strings=c('', ' '))
nonUL1d <- read.csv('nonUL1d.csv', sep=',',
                    header=T, na.strings=c('', ' '))

nonUL1 <- rbind(nonUL1a,nonUL1b,nonUL1c,nonUL1d)
```

```
rm(nonUL1a,nonUL1b,nonUL1c,nonUL1d)
str(nonUL1)
```

```
## 'data.frame':    48701 obs. of  49 variables:
## $ X                : int  1 2 3 4 5 6 7 8 9 10 ...
## $ ID               : Factor w/ 48701 levels "ILMN_1343289",...: 1 2 3
4 5 6 7 8 9 10 ...
## $ Species          : Factor w/ 1 level "Homo sapiens": 1 1 1 1 1 1 1
1 1 1 ...
## $ Source           : Factor w/ 3 levels "ILMN_Controls",...: 1 1 2 1 1
2 2 2 2 2 ...
## $ Search_Key       : Factor w/ 46721 levels
"ILMN_10001","ILMN_10014",...: 11664 11664 11664 11664 11664 11664 11664 9848
11306 1417 ...
## $ Transcript       : Factor w/ 46724 levels
"ILMN_10001","ILMN_10014",...: 2290 2291 1377 2292 2293 4903 1218 9858 11314
1421 ...
## $ ILMN_Gene        : Factor w/ 43186 levels "1-Dec","1-Mar",...: 8904
10174 2074 10143 10156 88 2713 4872 9250 2029 ...
## $ Source_Reference_ID : Factor w/ 46721 levels "NM_000015.1",...: 776
4636 1225 1123 1827 1130 1438 9379 7158 10894 ...
## $ RefSeq_ID        : Factor w/ 28570 levels "NM_000015.1",...: 776
4636 1225 1123 1827 1130 1438 9379 7158 10894 ...
## $ Unigene_ID       : Factor w/ 18153 levels "NA","Hs.100554",...: 1 1
1 1 1 1 1 1 1 1 ...
## $ Entrez_Gene_ID   : Factor w/ 16063 levels "10","10000","10001",...:
10744 10744 1487 10744 10744 5871 2331 6516 10718 5545 ...
## $ GI               : int  14141192 20149305 25453469 4507728 4507744
5016088 7669491 88954077 33469136 89070645 ...
## $ Accession        : Factor w/ 46721 levels "NM_000015.1",...: 776
4636 1225 1123 1827 1130 1438 9379 7158 10894 ...
## $ Symbol           : Factor w/ 25036 levels "1-Dec","1-Mar",...: 8904
10174 2074 10143 10156 88 2713 4872 9250 2029 ...
## $ Protein_Product  : Factor w/ 28173 levels "NA","NP_000006.1",...: 1
1 1224 1 1 1129 1437 9300 7155 10815 ...
## $ Probe_Id        : Factor w/ 48701 levels "ILMN_1343289",...: 1 2 3
4 5 6 7 8 9 10 ...
## $ Array_Address_Id : int  2140735 6550370 2690379 4590356 4260048
5860528 1770601 50270 3310274 7040079 ...
## $ Probe_Type       : Factor w/ 3 levels "A","I","S": 3 3 3 3 3 3 3 3
3 2 ...
## $ Probe_Start      : int  416 1856 1293 1408 72 1725 930 1 1103 2975
...
## $ SEQUENCE         : Factor w/ 48701 levels
"AAAAAACAGGAATAGCTCTAGGAGTCCTTACACAGGTCCGAGGGACCAGC",...: 9453 9026 11512 4877
6702 9532 5532 1967 10077 11476 ...
## $ Chromosome       : Factor w/ 56 levels "1","10","11",...: 27 27 22
27 27 23 4 14 1 27 ...
## $ Probe_Chromosome : Factor w/ 3 levels "-", "+", "NA": 3 3 1 3 3 1 2 2
1 3 ...
```

```

## $ Probe_Coordinates : Factor w/ 41351 levels "100000925-100000974",...:
10161 10161 8915 10161 10161 7449 8192 4231 2830 10161 ...
## $ Cytoband : Factor w/ 3676 levels "10p11.1d","10p11.21a",...:
2501 2501 2039 2501 2501 2165 288 1490 1036 2033 ...
## $ Definition : Factor w/ 46614 levels "Homo sapiens 1-
acylglycerol-3-phosphate O-acyltransferase 1 (lysophosphatidic acid
acyltransferase, alpha) (AGP"| __truncated__,...: 5692 7144 2221 7047 6692 98
2786 8195 6195 7895 ...
## $ Ontology_Component : Factor w/ 7849 levels "A 20S multiprotein
assembly of total mass about 1.2 MDa that activates dynein-based activity in
vivo. A large s"| __truncated__,...: 2614 1893 1304 1321 1266 2112 1468 1893
1763 269 ...
## $ Ontology_Process : Factor w/ 8950 levels "[goid 6069] [pmid
1755855] [evidence IDA]; A change in state or activity of a cell or an
organism (in terms of "| __truncated__,...: 2067 2257 3716 1085 547 417 1784
1091 1091 962 ...
## $ Ontology_Function : Factor w/ 9453 levels "[goid 16505] [pmid
10426319] [evidence NAS]",...: 3847 2486 1975 1911 2750 2514 95 3637 3637 828
...
## $ Synonyms : Factor w/ 16472 levels "0610037N12Rik; RPP20;
RPP2",...: 4591 4591 5325 4591 4591 5300 2402 4591 3649 4591 ...
## $ Obsolete_Probe_Id : Factor w/ 16878 levels "0610037N12Rik; RPP20;
RPP2",...: 4784 4784 645 6450 1251 5508 2490 4784 3785 4784 ...
## $ GB_ACC : Factor w/ 46717 levels "NA","NM_000015.1",...:
777 4635 1225 1123 1827 1130 1438 9377 7156 10892 ...
## $ GSM2496194 : num 9823 18157 27796 3428 7706 ...
## $ GSM2496195 : num 11265 20893 24042 4279 9407 ...
## $ GSM2496196 : num 13016 20943 24368 3049 9110 ...
## $ GSM2496197 : num 11698 18242 24179 3574 7935 ...
## $ GSM2496198 : num 11448 20998 25276 2178 7307 ...
## $ GSM2496199 : num 11454 21756 26935 3768 8928 ...
## $ GSM2496200 : num 11514 21849 26969 3170 9457 ...
## $ GSM2496201 : num 10621 10200 24231 2292 7765 ...
## $ GSM2496202 : num 11066 9349 22945 4513 8454 ...
## $ GSM2496210 : num 10189 21816 29280 4816 8773 ...
## $ GSM2496211 : num 9998 18435 26231 4683 8579 ...
## $ GSM2496212 : num 11407 23942 27389 3589 7977 ...
## $ GSM2496213 : num 9476 10440 23432 5444 8126 ...
## $ GSM2496214 : num 11708 9478 22640 4533 8418 ...
## $ GSM2496215 : num 11457 11803 24008 5839 8549 ...
## $ GSM2496216 : num 12900 20375 28086 4044 7252 ...
## $ GSM2496221 : num 10020 16842 25324 2469 7225 ...
## $ GSM2496222 : num 13409 9030 22273 3315 7844 ...

UL <- UL1[, -c(1:13,15:19,21,29:31)]
nonUL <- nonUL1[, -c(1:13,15:19,21,29:31)]

write.csv(UL, 'UL.csv', row.names=FALSE)
write.csv(nonUL, 'nonUL.csv', row.names=FALSE)

```

```

fibroid <- read.csv('UL.csv', sep=',', header=T, na.strings=c('', ' '))
nonFibroid <- read.csv('nonUL.csv', sep=',', header=T, na.strings=c('', ' '))

fibroid_gene_n <- fibroid %>% group_by(Symbol) %>% count(n())
narm <- grep('^NA$', fibroid_gene_n$Symbol)

fibroid1 <- fibroid_gene_n[-narm, -2]
colnames(fibroid1)[2] <- 'gene_count'

NONfibroid_gene_n <- nonFibroid %>% group_by(Symbol) %>% count(n())
narm1 <- grep('^NA$', NONfibroid_gene_n$Symbol)

nonFibroid1 <- NONfibroid_gene_n[-narm1, -2]
colnames(nonFibroid1)[2] <- 'gene_count'

GeneCopyNumberVariants <-
fibroid1[order(fibroid1$gene_count, decreasing=TRUE)[1:10], ]
GeneCopyNumberVariants

## # A tibble: 10 x 2
## # Groups:   Symbol [10]
##   Symbol      gene_count
##   <fct>         <int>
## 1 DDX12             10
## 2 KIAA0692           9
## 3 LOC23117           8
## 4 PLEC1             8
## 5 BDNF              7
## 6 CTNNB1            7
## 7 DMD               7
## 8 LOC202134          7
## 9 LOC339047          7
## 10 LOC653086         7

```

Combine the gene counts with the tables of samples for each type of UL or nonUL.

```

Fibroid_count <- merge(fibroid1, fibroid, by.x='Symbol', by.y='Symbol')
nonFibroid_count <- merge(nonFibroid1, nonFibroid, by.x='Symbol',
by.y='Symbol')
Fibroid_count[order(Fibroid_count$gene_count, decreasing=TRUE)[1:20], 1:3]

##           Symbol gene_count
SEQUENCE
## 5646      DDX12          10
CCAGTCCCTGACTACAGAGGATTTCCCAAAGTCCCTGGCTGTGAGGTTC
## 5647      DDX12          10
TTACTGGGGATGGTATTTAGGAGCCAGGAAAGCCGGTGCATTCTAGTGA
## 5648      DDX12          10
TCTCCTGCCCCCTCCGGAAGCTTGGATGCCCCCTCCACACCCTCTTGATCT
## 5649      DDX12          10
CAGACTTCTCGCTTCCTTTCTGCTGGGCCTCTGAGGGGTCATGGGGCCAT

```

```

## 5650      DDX12      10
ACATGTGCTGTCCTGGAACCTTGCTCTTTTCACTCAGCAGCCAGAGGGTC
## 5651      DDX12      10
AAACGTTACAGTGTTCCGATGAGACACAGTAGGCAGTACTTGGGAGGGTC
## 5652      DDX12      10
CAGGGCAGGAACCACGTCTTTACAGTTTGATGTTCCCAGAGCTGACCCAG
## 5653      DDX12      10
GCAGGGGAGATTGGGTTTAGGGGCTTTCCTGGTCTGCATTCTGCTACAGC
## 5654      DDX12      10
CCGCCGGGCTGCTTTTTCTTGATGCCCATCAGGACGCCTCAGTTCTCT
## 5655      DDX12      10
CGTTGCTACAAGCTGTTTTTTGAATGTCTCTACACAGTCCAGGCAGGAAG
## 10983 KIAA0692      9
AAGTGGTGCCTGGCTGTCCCTATACTGTGCTGCTGGGTGTTCCAGCCTGT
## 10984 KIAA0692      9
TAAGTGCAGTGAGCTCTGGCGGAAACCACCCTCTGCCCCGTCTGTTGGAT
## 10985 KIAA0692      9
CATTGTAATGATAAGGAAATGTTGCGATCAAATAAGATTTAGACACACTT
## 10986 KIAA0692      9
GATCACAGGCACAGGGAAGCCACAAGGAGCTCTGTATGAGTTGTGTTTGC
## 10987 KIAA0692      9
CAGGCGACTGGGTAGCAGATGTGGAAGCTGATGGTTAGGCCCAGGGCATG
## 10988 KIAA0692      9
GTTGTTCTGGACGATCTTCGGGATCCTCTGGGGCACTGTGACACTCGGAG
## 10989 KIAA0692      9
GAGTGCTGGGAAGGTTAATGTTAAATGGGTTGTGTGTCGGGGAGGGTACA
## 10990 KIAA0692      9
AGCTCCACCTTGACCCAGCCTCACAACAAAAAGTTTGTGTATGACCAGGC
## 10991 KIAA0692      9
GCAAATGTAAGTCAAGGGGTTTGGGGCCAGAGGAAGAGGGAGAAGGTGGCC
## 12174 LOC23117      8
CTGGCCTTCCCTCATCAGCCGTAAATGATGATTTACTGCTGTTACCATCA

```

Add a mean, median, min, and max column to these tables.

```

Fibroid_count$Fibroid_Mean <- rowMeans(Fibroid_count[,11:30])
nonFibroid_count$nonFibroid_Mean <- rowMeans(nonFibroid_count[,11:28])

```

Use the tidyr package to group by sample ID by gathering those columns into one.

```

UL_3 <- gather(Fibroid_count, 'UL_Sample_ID', 'Value', 11:30)
nonUL_3 <- gather(nonFibroid_count, 'nonUL_Sample_ID', 'Value', 11:28)

```

Create the stat tables then combine for the UL and nonUL sample sets using the dplyr package.

```

UL_median <- UL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value),
median)
colnames(UL_median)[2] <- 'Fibroid_Median'

nonUL_median <- nonUL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value),

```

```

median)
colnames(nonUL_median)[2] <- 'nonFibroid_Median'

UL_max <- UL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value), max)
colnames(UL_max)[2] <- 'Fibroid_max'

nonUL_max <- nonUL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value),
max)
colnames(nonUL_max)[2] <- 'nonFibroid_max'

UL_min <- UL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value), min)
colnames(UL_min)[2] <- 'Fibroid_min'

nonUL_min <- nonUL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value),
min)
colnames(nonUL_min)[2] <- 'nonFibroid_min'

UL_sd <- UL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value), sd)
colnames(UL_sd)[2] <- 'Fibroid_stdError'

nonUL_sd <- nonUL_3 %>% group_by(SEQUENCE) %>% summarise_at(vars(Value), sd)
colnames(nonUL_sd)[2] <- 'nonFibroid_stdError'

```

Combine these four tables together.

```

Fibroid_stats <- merge(UL_median, UL_max, by.x='SEQUENCE', by.y='SEQUENCE')
Fibroid_stats1 <- merge(Fibroid_stats, UL_min, by.x='SEQUENCE',
by.y='SEQUENCE')
Fibroid_stats2 <- merge(Fibroid_count, Fibroid_stats1, by.x='SEQUENCE',
by.y='SEQUENCE')
Fibroid_stats3 <- merge(Fibroid_stats2, UL_sd, by.x='SEQUENCE',
by.y='SEQUENCE')
colnames(Fibroid_stats3)[11:30] <- paste('UL_',
colnames(Fibroid_stats3)[11:30], sep='')

nonFibroid_stats <- merge(nonUL_median, nonUL_max, by.x='SEQUENCE',
by.y='SEQUENCE')
nonFibroid_stats1 <- merge(nonFibroid_stats, nonUL_min, by.x='SEQUENCE',
by.y='SEQUENCE')
nonFibroid_stats2 <- merge(nonFibroid_count, nonFibroid_stats1,
by.x='SEQUENCE', by.y='SEQUENCE')
nonFibroid_stats3 <- merge(nonFibroid_stats2, nonUL_sd, by.x='SEQUENCE',
by.y='SEQUENCE')
colnames(nonFibroid_stats3)[11:28] <- paste('nonUL_',
colnames(nonFibroid_stats3)[11:28], sep='')

nonfibroid <- nonFibroid_stats3[,c(1,11:33)]

```



```

all <- merge(Fibroid_stats3, nonfibroid, by.x='SEQUENCE', by.y='SEQUENCE')
str(all)

## 'data.frame':    30549 obs. of  58 variables:
## $ SEQUENCE          : Factor w/ 48701 levels
"AAAAAACAAACCGCGCAGCGGAGAACCGGTGCCTGAGTCTCCAGGGAC",...: 1 4 5 7 8 10 12 13
15 18 ...
## $ Symbol            : Factor w/ 25036 levels "1-Dec","1-Mar",...: 12031
11383 13002 14397 15611 12721 12474 10953 24822 18000 ...
## $ gene_count        : int  1 1 1 1 1 1 1 1 1 1 ...
## $ Probe_Chrl_Orientation: Factor w/ 3 levels "-","+", "NA": 2 1 2 3 3 2 2 3
1 2 ...
## $ Probe_Coordinates  : Factor w/ 41351 levels "100000925-100000974",...:
33600 3552 19430 41351 41351 36542 28872 41351 31503 41119 ...
## $ Cytoband          : Factor w/ 3676 levels "10p11.1d","10p11.21a",...:
3472 3472 3472 3472 3472 293 3472 3472 1261 2180 ...
## $ Definition         : Factor w/ 46614 levels "{3 region, probe S2}
[human, 76N, mammary epithelial cells, mRNA Partial, 339 nt]",...: 33743 33388
39229 37744 38664 34103 33968 35730 29449 21454 ...
## $ Ontology_Component : Factor w/ 7849 levels "A 20S multiprotein
assembly of total mass about 1.2 MDa that activates dynein-based activity in
vivo. A large s"| __truncated__,...: 4150 4150 4150 4150 4150 4150 4150 4150
5434 6267 ...
## $ Ontology_Process   : Factor w/ 8950 levels "[goid 19642] [evidence
IEA]; The chemical reactions and pathways involving carbohydrates, any of a
group of org"| __truncated__,...: 2492 2492 2492 2492 2492 2492 2492 2492 8502
3316 ...
## $ Ontology_Function  : Factor w/ 9453 levels "[goid 15280] [evidence
IEA]; Interacting selectively with sodium ions (Na+) [goid 31402] [evidence
IEA]",...: 8111 8111 8111 8111 8111 8111 8111 8111 7071 2705 ...
## $ UL_GSM2496185      : num  49.5 51.2 59.9 312.3 52.3 ...
## $ UL_GSM2496186      : num  51.5 52.3 64.5 333.3 53.7 ...
## $ UL_GSM2496187      : num  46.9 49 50.3 331.8 56.4 ...
## $ UL_GSM2496188      : num  50.1 48.4 52 360.9 51.8 ...
## $ UL_GSM2496189      : num  54 49.8 52.8 339 52.6 ...
## $ UL_GSM2496190      : num  50.1 48.6 52 411.7 51.9 ...
## $ UL_GSM2496191      : num  49.5 46 54.7 424.3 54.8 ...
## $ UL_GSM2496192      : num  46.5 50.6 50.2 517.3 53.6 ...
## $ UL_GSM2496193      : num  48 46.7 52.9 631.7 54.1 ...
## $ UL_GSM2496203      : num  48.1 51.5 55.1 576.7 55 ...
## $ UL_GSM2496204      : num  51.7 50.8 51.6 300.4 52.3 ...
## $ UL_GSM2496205      : num  46.9 47.5 52.3 348.9 53.2 ...
## $ UL_GSM2496206      : num  50.5 43.9 54.9 722.6 56.1 ...
## $ UL_GSM2496207      : num  44 46.5 56.5 1218 55.9 ...
## $ UL_GSM2496208      : num  51.1 46 51.9 535.5 55.5 ...
## $ UL_GSM2496209      : num  55.2 46.9 50.6 669 53.3 ...
## $ UL_GSM2496217      : num  48.9 45.8 49.6 361.9 49.8 ...
## $ UL_GSM2496218      : num  50 49.9 53.9 377.2 53.6 ...
## $ UL_GSM2496219      : num  48.3 49.5 50.4 544.1 58.1 ...
## $ UL_GSM2496220      : num  47.7 46.7 51.7 406.1 56.3 ...

```

```
## $ Fibroid_Mean      : num  49.4 48.4 53.4 486.1 54 ...
## $ Fibroid_Median    : num  49.5 48.5 52.1 408.9 53.7 ...
## $ Fibroid_max        : num  55.2 52.3 64.5 1218 58.1 ...
## $ Fibroid_min        : num  44 43.9 49.6 300.4 49.8 ...
## $ Fibroid_stdError   : num  2.59 2.3 3.61 214.26 2 ...
## $ nonUL_GSM2496194   : num  42.7 42.7 56.6 442 57 ...
## $ nonUL_GSM2496195   : num  49.1 45.3 56 505.4 55.3 ...
## $ nonUL_GSM2496196   : num  51.1 45.3 55.1 416.5 57.4 ...
## $ nonUL_GSM2496197   : num  46 52.6 56.2 475.3 54.6 ...
## $ nonUL_GSM2496198   : num  49.1 54.1 53.7 408.1 50.8 ...
## $ nonUL_GSM2496199   : num  52.6 47.9 51.1 510.7 52.5 ...
## $ nonUL_GSM2496200   : num  46 47.1 52.5 681.9 50.3 ...
## $ nonUL_GSM2496201   : num  45.9 46.8 48.3 689.3 56.7 ...
## $ nonUL_GSM2496202   : num  50.6 45.3 56.5 735.2 48.7 ...
## $ nonUL_GSM2496210   : num  48.3 49.3 56.3 426.9 52.5 ...
## $ nonUL_GSM2496211   : num  46.8 46 54.6 567.3 47.4 ...
## $ nonUL_GSM2496212   : num  50.6 48.7 54.3 493.8 51.5 ...
## $ nonUL_GSM2496213   : num  49.1 49.5 56.7 792.8 48.8 ...
## $ nonUL_GSM2496214   : num  48.8 49.9 54 710.5 50.6 ...
## $ nonUL_GSM2496215   : num  48 47.1 52.9 795.4 53.1 ...
## $ nonUL_GSM2496216   : num  47.9 49.2 48 574.5 51.1 ...
## $ nonUL_GSM2496221   : num  48.8 48.2 50.6 507.4 54.7 ...
## $ nonUL_GSM2496222   : num  44.6 49.5 50.4 590 56 ...
## $ nonFibroid_Mean    : num  48.1 48 53.5 573.5 52.7 ...
## $ nonFibroid_Median  : num  48.5 48.1 54.1 539 52.5 ...
## $ nonFibroid_max      : num  52.6 54.1 56.7 795.4 57.4 ...
## $ nonFibroid_min      : num  42.7 42.7 48 408.1 47.4 ...
## $ nonFibroid_stdError : num  2.46 2.75 2.85 130.03 3.07 ...
```

Lets change the 'fibroid' in the column names to 'UL' for uterine leiomyoma.

```
colnames(all) <- gsub('Fibroid', 'UL', colnames(all))
```

Reorder the table so that the stats are at the end of the columns.

```
All <- all[,c(1:10,11:30, 36:53,31:35,54:58)]
str(All)

## 'data.frame': 30549 obs. of 58 variables:
## $ SEQUENCE : Factor w/ 48701 levels
"AAAAAACAAAACCGCGCAGCGGAGAACCGGTGCCTGAGTCTCCAGGGAC",...: 1 4 5 7 8 10 12 13
15 18 ...
## $ Symbol : Factor w/ 25036 levels "1-Dec","1-Mar",...: 12031
11383 13002 14397 15611 12721 12474 10953 24822 18000 ...
## $ gene_count : int 1 1 1 1 1 1 1 1 1 1 ...
## $ Probe_Chr_Orientation: Factor w/ 3 levels "-", "+", "NA": 2 1 2 3 3 2 2 3
1 2 ...
## $ Probe_Coordinates : Factor w/ 41351 levels "100000925-100000974",...:
33600 3552 19430 41351 41351 36542 28872 41351 31503 41119 ...
## $ Cytoband : Factor w/ 3676 levels "10p11.1d","10p11.21a",...:
3472 3472 3472 3472 3472 293 3472 3472 1261 2180 ...
```

```

## $ Definition      : Factor w/ 46614 levels "{3 region, probe S2}
[human, 76N, mammary epithelial cells, mRNA Partial, 339 nt]",...: 33743 33388
39229 37744 38664 34103 33968 35730 29449 21454 ...
## $ Ontology_Component : Factor w/ 7849 levels "A 20S multiprotein
assembly of total mass about 1.2 MDa that activates dynein-based activity in
vivo. A large s"| __truncated__,...: 4150 4150 4150 4150 4150 4150 4150 4150
5434 6267 ...
## $ Ontology_Process   : Factor w/ 8950 levels "[goid 19642] [evidence
IEA]; The chemical reactions and pathways involving carbohydrates, any of a
group of org"| __truncated__,...: 2492 2492 2492 2492 2492 2492 2492 2492 8502
3316 ...
## $ Ontology_Function   : Factor w/ 9453 levels "[goid 15280] [evidence
IEA]; Interacting selectively with sodium ions (Na+) [goid 31402] [evidence
IEA]",...: 8111 8111 8111 8111 8111 8111 8111 8111 7071 2705 ...
## $ UL_GSM2496185      : num  49.5 51.2 59.9 312.3 52.3 ...
## $ UL_GSM2496186      : num  51.5 52.3 64.5 333.3 53.7 ...
## $ UL_GSM2496187      : num  46.9 49 50.3 331.8 56.4 ...
## $ UL_GSM2496188      : num  50.1 48.4 52 360.9 51.8 ...
## $ UL_GSM2496189      : num  54 49.8 52.8 339 52.6 ...
## $ UL_GSM2496190      : num  50.1 48.6 52 411.7 51.9 ...
## $ UL_GSM2496191      : num  49.5 46 54.7 424.3 54.8 ...
## $ UL_GSM2496192      : num  46.5 50.6 50.2 517.3 53.6 ...
## $ UL_GSM2496193      : num  48 46.7 52.9 631.7 54.1 ...
## $ UL_GSM2496203      : num  48.1 51.5 55.1 576.7 55 ...
## $ UL_GSM2496204      : num  51.7 50.8 51.6 300.4 52.3 ...
## $ UL_GSM2496205      : num  46.9 47.5 52.3 348.9 53.2 ...
## $ UL_GSM2496206      : num  50.5 43.9 54.9 722.6 56.1 ...
## $ UL_GSM2496207      : num  44 46.5 56.5 1218 55.9 ...
## $ UL_GSM2496208      : num  51.1 46 51.9 535.5 55.5 ...
## $ UL_GSM2496209      : num  55.2 46.9 50.6 669 53.3 ...
## $ UL_GSM2496217      : num  48.9 45.8 49.6 361.9 49.8 ...
## $ UL_GSM2496218      : num  50 49.9 53.9 377.2 53.6 ...
## $ UL_GSM2496219      : num  48.3 49.5 50.4 544.1 58.1 ...
## $ UL_GSM2496220      : num  47.7 46.7 51.7 406.1 56.3 ...
## $ nonUL_GSM2496194    : num  42.7 42.7 56.6 442 57 ...
## $ nonUL_GSM2496195    : num  49.1 45.3 56 505.4 55.3 ...
## $ nonUL_GSM2496196    : num  51.1 45.3 55.1 416.5 57.4 ...
## $ nonUL_GSM2496197    : num  46 52.6 56.2 475.3 54.6 ...
## $ nonUL_GSM2496198    : num  49.1 54.1 53.7 408.1 50.8 ...
## $ nonUL_GSM2496199    : num  52.6 47.9 51.1 510.7 52.5 ...
## $ nonUL_GSM2496200    : num  46 47.1 52.5 681.9 50.3 ...
## $ nonUL_GSM2496201    : num  45.9 46.8 48.3 689.3 56.7 ...
## $ nonUL_GSM2496202    : num  50.6 45.3 56.5 735.2 48.7 ...
## $ nonUL_GSM2496210    : num  48.3 49.3 56.3 426.9 52.5 ...
## $ nonUL_GSM2496211    : num  46.8 46 54.6 567.3 47.4 ...
## $ nonUL_GSM2496212    : num  50.6 48.7 54.3 493.8 51.5 ...
## $ nonUL_GSM2496213    : num  49.1 49.5 56.7 792.8 48.8 ...
## $ nonUL_GSM2496214    : num  48.8 49.9 54 710.5 50.6 ...
## $ nonUL_GSM2496215    : num  48 47.1 52.9 795.4 53.1 ...
## $ nonUL_GSM2496216    : num  47.9 49.2 48 574.5 51.1 ...

```

```
## $ nonUL_GSM2496221 : num 48.8 48.2 50.6 507.4 54.7 ...
## $ nonUL_GSM2496222 : num 44.6 49.5 50.4 590 56 ...
## $ UL_Mean : num 49.4 48.4 53.4 486.1 54 ...
## $ UL_Median : num 49.5 48.5 52.1 408.9 53.7 ...
## $ UL_max : num 55.2 52.3 64.5 1218 58.1 ...
## $ UL_min : num 44 43.9 49.6 300.4 49.8 ...
## $ UL_stdError : num 2.59 2.3 3.61 214.26 2 ...
## $ nonUL_Mean : num 48.1 48 53.5 573.5 52.7 ...
## $ nonUL_Median : num 48.5 48.1 54.1 539 52.5 ...
## $ nonUL_max : num 52.6 54.1 56.7 795.4 57.4 ...
## $ nonUL_min : num 42.7 42.7 48 408.1 47.4 ...
## $ nonUL_stdError : num 2.46 2.75 2.85 130.03 3.07 ...
```

```
All_stats_only <- All[,c(1,2,3,49:58)]
stats_all <- All_stats_only[!duplicated(All_stats_only),]
```

```
stats_all$foldChangeMean_UL_to_nonUL <-
stats_all$UL_Mean/stats_all$nonUL_Mean
```

```
FoldChangeGenes <- stats_all[order(stats_all$foldChangeMean_UL_to_nonUL,
decreasing=TRUE)[c(1:5,30545:30549)],]
```

FoldChangeGenes

##		SEQUENCE	Symbol
##	gene_count		
##	16709	GCAACGCTCCTCTGAAATGCTTGTCTTTTTCTGTTGCCGAAATAGCTGG	KIAA1199
1			
##	17948	GCCCCAGCAAGCCTCCCTCCATCCTCCAGTGGGAAACTGTTGATGGTGT	PENK
1			
##	22790	GGTATTGCTGATCGTATGCAGAAGGAAATCACTGCTCTGGCTCCTAGCAC	ACTC
1			
##	28162	TGCGAGACCTGGGTGTCCAACCTGCGCTACAACCACATGCTGCGGAAGAA	DLK1
2			
##	3569	AGGCCCTGGAGGCTGCAACATACCTCAATCCTGTCCCAGGCCGGATCCTC	MMP11
1			
##	17565	GCCAACCTCCTCTCACAGCCTCTGTATCTCTGCAGGCCATACTGGTTCCA	ABCA8
1			
##	6582	CAGATGTTTTCCCTTGTGGCAGTCTTCAGCCTCCTCTACCCTACATGATC	ADH1A
1			
##	8958	CCCAGTGACACTTCAGAGAGCTGGTAGTTAGTAGCATGTTGAGCCAGGCC	FOS
1			
##	27542	TGACTGTCCCTGCCAATGCTCCAGCTGTCGTCTGACTCTGGGTTCGTTGG	FOSB
1			
##	6881	CAGCTGGGCGATGTGCGAGCTGATAGTGAGCGGCAGAATCAGGAGTACCA	KRT19
1			
##		UL_Mean UL_Median UL_max UL_min UL_stdError nonUL_Mean	
##	16709	3029.4710 2566.7096 6548.8160 156.8375 1948.88193 167.75283	
##	17948	1798.4359 237.7790 26134.7105 48.2000 5783.33114 120.28719	
##	22790	4693.5182 4880.4535 13992.2647 110.2923 3328.78611 392.30767	

```

## 28162  946.4365  249.4440  5799.7170  54.7250  1542.06289  87.41913
## 3569  4767.6862 3652.1430 15286.3475 235.9674 4029.71487 442.19991
## 17565  143.2650  99.0957  628.2257  56.0500  126.17477  724.33959
## 6582   608.7258 545.4995 1947.6415  54.3500  476.16852 3102.90066
## 8958 1287.1220 805.1551 7335.1691 220.6810 1573.12379 6916.73077
## 27542 205.9075  70.8273 2290.1261  53.1000  493.97933 1142.67872
## 6881  130.4870  85.6123  311.2020  45.6000   90.60451  815.61118
##      nonUL_Median  nonUL_max nonUL_min nonUL_stdError
## 16709    116.83410    514.9222   63.1000    127.90943
## 17948     71.88335    431.7050   54.2000     98.67055
## 22790    332.26735    830.2335  169.4130    197.47324
## 28162     52.70835    669.4027   47.3000    145.29680
## 3569     308.24915   1683.5592   75.9750    430.96362
## 17565    643.88430   1493.2380  125.9636    317.38002
## 6582    2961.70150   7727.8238  275.2883   1833.54639
## 8958    6722.92010 17362.0317 2406.0706   3676.04993
## 27542    595.09105   7604.0221   92.4250   1864.52510
## 6881     738.20585  2049.6529   57.4000    498.12489
##      foldChangeMean_UL_to_nonUL
## 16709                18.0591346
## 17948                14.9511840
## 22790                11.9638706
## 28162                10.8264232
## 3569                 10.7817440
## 17565                 0.1977871
## 6582                 0.1961796
## 8958                 0.1860882
## 27542                 0.1801972
## 6881                 0.1599868

```

```
str(stats_all)
```

```

## 'data.frame':  30549 obs. of  14 variables:
## $ SEQUENCE : Factor w/ 48701 levels
"AAAAAACAAACCGCGCAGCGGAGAACCGGTGCCTGAGTCTCCAGGGAC",...: 1 4 5 7 8 10 12 13
15 18 ...
## $ Symbol : Factor w/ 25036 levels "1-Dec","1-Mar",...:
12031 11383 13002 14397 15611 12721 12474 10953 24822 18000 ...
## $ gene_count : int 1 1 1 1 1 1 1 1 1 1 ...
## $ UL_Mean : num 49.4 48.4 53.4 486.1 54 ...
## $ UL_Median : num 49.5 48.5 52.1 408.9 53.7 ...
## $ UL_max : num 55.2 52.3 64.5 1218 58.1 ...
## $ UL_min : num 44 43.9 49.6 300.4 49.8 ...
## $ UL_stdError : num 2.59 2.3 3.61 214.26 2 ...
## $ nonUL_Mean : num 48.1 48 53.5 573.5 52.7 ...
## $ nonUL_Median : num 48.5 48.1 54.1 539 52.5 ...
## $ nonUL_max : num 52.6 54.1 56.7 795.4 57.4 ...
## $ nonUL_min : num 42.7 42.7 48 408.1 47.4 ...
## $ nonUL_stdError : num 2.46 2.75 2.85 130.03 3.07 ...
## $ foldChangeMean_UL_to_nonUL: num 1.027 1.007 0.997 0.848 1.024 ...

```

```
write.csv(stats_all, 'stats_only_UL_nonUL.csv', row.names=FALSE)
```

Combine the table of top and bottom five genes in fold change values of the ratio of UL to non-UL sample means, FoldChangeGenes, with the table of the ten genes having the highest number of copy number variations or genotypes, GeneCopyNumberVariants.

```
ontology <- nonFibroid[,c(1,6:9)]
gnc <- as.data.frame(GeneCopyNumberVariants)[1]

keyGenes1 <- merge(gnc, stats_all, by.x='Symbol', by.y='Symbol')

keyGenes1a <- merge(keyGenes1, ontology, by.x='Symbol', by.y='Symbol')

keyGenes2 <- merge(FoldChangeGenes, ontology, by.x='Symbol', by.y='Symbol')
keyGenes2a <- keyGenes2[,c(1:3,15:18,4:14)]
keyGenes1b <- keyGenes1a[,c(1:3,15:18,4:14)]
```

```
KeyGenes <- rbind(keyGenes2a, keyGenes1b)
KG <- KeyGenes[!duplicated(KeyGenes$SEQUENCE),]
KG1 <- KG[order(KG$foldChangeMean_UL_to_nonUL, decreasing=TRUE),]
KG1[,c(1:3,18)]
```

```
##          Symbol                               SEQUENCE
gene_count
## 8      KIAA1199  GCAACGCTCCTCTGAAATGCTTGTCTTTTTCTGTTGCCGAAATAGCTGG
1
## 11      PENK    GCCCCAGCAAGCCTCCCTCCATCCTCCAGTGGGAACTGTTGATGGTGT
1
## 2       ACTC    GGTATTGCTGATCGTATGCAGAAGGAAATCACTGCTCTGGCTCCTAGCAC
1
## 4       DLK1    TGCGAGACCTGGGTGTCCAACCTGCGCTACAACCACATGCTGCGGAAGAA
2
## 10      MMP11   AGGCCCTGGAGGCTGCAACATACCTCAATCCTGTCCCAGGCCGGATCCTC
1
## 68      CTNNB1  AGCTGCAGGGGTCCTCTGTGAACTTGCTCAGGACAAGGAAGCTGCAGAAG
7
## 89      CTNNB1  CTGCAGGGGTCCTCTGTGAACTTGCTCAGGACAAGGAAGCTGCAGAAGCT
7
## 82      CTNNB1  AGTCTCTCGTAGTGTTAAGTTATAGTGAATACTGCTACAGCAATTTCTAA
7
## 231     DMD     CAGTGTTGGGATCACTCACTTTCCCCCTACAGGACTCAGATCTGGGAGGC
7
## 210     DMD     CTCCTCTCAGCTGAACACCCTCCTTTCACTCCCAAATGCAAACAGTCTCT
7
## 96      CTNNB1  GCCTCTTGCACTCTGAATTGGGAATGTTTGCACCACAGTGGGGGGCTTGC
7
## 140     DDX12   CCGCCGGGCTGCTTTTTCTTTGGATGCCCATCAGGACGCCTCAGTTCTCT
10
## 397     LOC23117 TCAACCACATCCTTCAAAAGGACTATGCCTGTTTATAAGCCCAGCTGTTT
8
```

361 LOC202134 GCCAAAGGAATGGGCTCCAGACACCCCCTCTTCCAGAGCAAGGATGAAGG
7
286 KIAA0692 TAAGTGCAGTGAGCTCTGGCGGAAACCACCCCTCTGCCCCGTCTGTTGGAT
9
61 CTNNB1 CAAACTTTACAGAGGAGAATGCCCTGTTTGTTAACCATGTTTCTTTTGGC
7
516 LOC653086 TGATGTGTCACGCCACTGTACTCCAGCCTGACGGCAGAGCGAGACTCCAT
7
33 BDNF CCCTCCACCTCCTGCTCGGGGGGCTTTAATGAGACACCCACCGCTGCTGT
7
551 PLEC1 CCCGACGAGCAGGACTTCATCCAGGCCTACGAGGAGGTGCGCGAGAAGTA
8
445 LOC23117 TGTCGTTTCCTCCATTCTTCACCAAAACATCAGCGTACATAGGCACATGG
8
474 LOC339047 ACTGCCTGTGTGGCTCCTTGAGTGCGCGGAGGCCAAAGCTGAGATGACTT
7
331 KIAA0692 CATTGTAATGATAAGGAAATGTTGCGATCAAATAAGATTTAGACACACTT
9
559 PLEC1 CCCTCGGGCAGCCTGTTTCCCTCCCTGGTGGTTGTGGGTACGTTGTCAC
8
530 LOC653086 GGTGTGCTCTGGTATGTAATGACAATATGTGAACAAACCTGTGGAATTAA
7
259 KIAA0692 GAGTGCTGGGAAGGTTAATGTTAAATGGGTTGTGTGTCGGGGAGGGTACA
9
429 LOC23117 GGCTCCTCTTTGGGCTCCTACTGGAATTTATCAGCCATCAGTGCATCTCT
8
252 DMD TCTATCAACAGAGCTGAATGAGTGCCAGGAAGCTGCGAAATCTGTCTTAC
7
268 KIAA0692 GCAAATGTAACCTCAGGGGTTTGGGGCCAGAGGAAGAGGGAGAAGGTGGCC
9
19 BDNF AATAATAGAGTGTGGGAGTTTTGGGGCCGAAGTCTTTCCCGGAGCAGCTG
7
340 LOC202134 CAAACCCCTTGAAGACATTTAGGGCCATGCTCACTTGGGAGGGTTTGAGG
7
405 LOC23117 AAAGCAGTGTTTTTCTGCTGCCAGAGGCCTGAGAGAGTTTGGGCATACTC
8
245 DMD CCATTGAGAAGAATGATAAATGCCACAAGCATTTGGAAACAGGCTTCCCT
7
322 KIAA0692 AGCTCCACCTTGACCCAGCCTCACAACAAAAAGTTTGTGTATGACCAGGC
9
375 LOC202134 AGTGGGCAGAATGATGAGGGAAGTGGGCACGTGCCCATGTTCTTCTTGGC
7
382 LOC202134 TTCATCCAGGCCTGCGCCGGTGTTACAGTGGTCTCATCTAAGCCAGCC
7
591 PLEC1 CCGGGCCTTCTCGTGGTACCCTGCCTGCTGCCTTTGCCCCGCACTGACT
8
47 BDNF GCTCGCTGAAGTTGGCTTCCTAGCGGTGTAGGCTGGAATAGACTCTTGGC
7
575 PLEC1 GGCGCAGACATGGACCCCTCGCGAGCCATCCAGAACGAGATCAGCTCCCT
8

313 KIAA0692 GATCACAGGCACAGGGAAGCCACAAGGAGCTCTGTATGAGTTGTGTTTGC
9
495 LOC339047 TTTCAGGCCCATGGCAGAGGGTGGGCTCAGGAGGGCCATCGTGGGTGTCC
7
170 DDX12 GCAGGGGAGATTGGGTTTAGGGGCTTTCTGGTCTGCATTCTGCTACAGC
10
460 LOC339047 AGTGCCACATCACACAGCATCTAGCACGTAAGTGCACCCCGGGAGTCGT
7
437 LOC23117 GGCTCTGTTGGAATCCGCATAGTGTGGAAATGAGTTTGCCCTGGAAAGGG
8
502 LOC653086 AGTGTTGGGACTACAGGTGTGTGTTACTGCTCCCAGCTGGGAGGCAGGCT
7
509 LOC653086 GTGAGCCTGTTTCATCATCTGTAACTTTGAATAATGATACCTACCCCGC
7
467 LOC339047 CGCCCTGAAAGGACCAGGACATGCGGGTGCGGTGGCTGCTCTTTTGGCTC
7
150 DDX12 CAGGGCAGGAACCACGTCTTTACAGTTTGATGTTCCCAGAGCTGACCCAG
10
75 CTNNB1 CAGGAATCTAGTCTGGATGACTGCTTCTGGAGCCTGGATGCAGTACCATT
7
295 KIAA0692 GTTGTTCTGGACGATCTTCGGGATCCTCTGGGGCACTGTGACACTCGGAG
9
180 DDX12 TCTCCTGCCCCCTCCGGAAGCTTGGATGCCCCCTCCACACCCTCTTGATCT
10
389 LOC23117 CTGGCCTTCCCTCATCAGCCGTAAATGATGATTTACTGCTGTTACCATCA
8
103 CTNNB1 GCAATTTGCCAAGTTTCTTTAGCATTTGGCCCTGGATTACGCTGGACCCC
7
413 LOC23117 CCCTTCCTACATTCTTGTTTTCATTTTTTCGGAGGAAGAGGAGTTGCTAG
8
523 LOC653086 AGCAGCACATCGTCATTTTACAATTGAGAAACATGGAGACTCCAAATGGA
7
421 LOC23117 GGGAAGTACATGGGGCAGATGGAAGAACCTGAGATAATCGCAAGGATGGC
8
40 BDNF TCAGACCCCTCAGGCCACTGCTGTTCTGTCTCACACATTCCTGCAAAGGAC
7
607 PLEC1 CTCCGTCTGCCCCGTGGGCTCCTGCCACCGTCCCCGATGAAGATCGTGCC
8
453 LOC339047 TTTCTGAAATGGAGCTTTGCTCTTGTTGCCAGGCCGTAGTGCAATGGC
7
599 PLEC1 GCCTTTGCCTCGCCGAGGGAGGTCTTGCTGGAGCGGCCGTGCTGGCTGGA
8
12 BDNF ATGTACGTGGGGGATTCTTGACTCGGGTAGTCTCTGGGGATGCAGAGCC
7
583 PLEC1 CAGCCCTGGGGACACACTGCCCTGGAACCTTGGGAAAACGCAGCGGAGCC
8
190 DDX12 CGTTGCTACAAGCTGTTTTTTGAATGTCTCTACACAGTCCAGGCAGGAAG
10
481 LOC339047 TCTGTATGGACCCTGCCAAGCTCTGCCCCCTGCCCCCTGCATTGGGGCGC
7


```

## 54      BDNF  TGGGGAGACGAGATTTTAAGACACTTGAGTCTCCAGGACAGCAAAGGCAC
7
## 160     DDX12 CAGACTTCTCGCTTCCTTTCTGCTGGGCCTCTGAGGGGTCATGGGGCCAT
10
## 304     KIAA0692 AAGTGGTGCCTGGCTGTCCCTATACTGTGCTGCTGGGTGTTCCAGCCTGT
9
## 224     DMD   GCAGCCAACTTATTGGCATGATGGAGTGACAGGAAAAACAGCTGGCATGG
7
## 200     DDX12 TTACTGGGGATGGTATTTAGGAGCCAGGAAAGCCGGTGCAATTCCTAGTGA
10
## 544     LOC653086 TACCTGGCCTATCTTTCATAGGTTATATAAATTCCTTGGTTCCCAGTTTT
7
## 217     DMD   GGGTTTTCTCAGGATTGCTATGCAACAGGATCAGTGCTGTAGTGCCCGGT
7
## 120     DDX12 ACATGTGCTGTCACTGGAACCTTGCTCTTTTCACTCAGCAGCCAGAGGGTC
10
## 110     DDX12 CCAGTCCCTGACTACAGAGGATTTCCCCAAAGTCCCTGGCTGTGAGGTTC
10
## 130     DDX12 AAACGTTACAGTGTTCCGATGAGACACAGTAGGCAGTACTTGGGAGGGTC
10
## 368     LOC202134 GACCAAAGCAGGACAATTGCTTGATCCCAGGAGTTTAAGACCAGCCGGGG
7
## 537     LOC653086 AAGGACTCAGATGCAGGGTCTTCTCTGCTCCCCGTCACACAGAGGGTGCC
7
## 277     KIAA0692 CAGGCGACTGGGTAGCAGATGTGGAAGCTGATGGTTAGGCCCAGGGCATG
9
## 488     LOC339047 GACCTGTAGCTAAACCTTCCACCAGCGCTTGAGAACTTAATTTGAACCGG
7
## 238     DMD   GCACTCCGACTACATCAGGAGAAGATGTTTCGAGACTTTGCCAAGGTACTA
7
## 354     LOC202134 CCACGCCGGCAAAGAAATTGGAAGACTCCACCATTACAGGCAGCCACCAG
7
## 26      BDNF  CTTGCTGTGGTCTCTTTGTGGCAGAAGTGTTTCATGCATGGCAGCAGGCC
7
## 567     PLEC1  AGCCTCTGTTCCCCTAGTAAGTGCCTTCCATGTCGGCCTCTAACCCCAGG
8
## 347     LOC202134 CCTGTTTGGATCACATGGTCTTGTCCTGATAACTTGGAAGAGGTTGCTTC
7
## 1       ABCA8  GCCAACCTCCTCTCACAGCCTCTGTATCTCTGCAGGCCATACTGGTTCCA
1
## 3       ADH1A  CAGATGTTTTCCCTTGTGGCAGTCTTCAGCCTCCTCTACCCTACATGATC
1
## 6       FOS   CCCAGTGACACTTCAGAGAGCTGGTAGTTAGTAGCATGTTGAGCCAGGCC
1
## 7       FOSB  TGA CTGTCCCTGCCAATGCTCCAGCTGTCGTCTGACTCTGGGTTTCGTTGG
1
## 9       KRT19  CAGCTGGGCGATGTGCGAGCTGATAGTGAGCGGCAGAATCAGGAGTACCA
1
##      foldChangeMean_UL_to_nonUL
## 8      18.0591346

```

## 11	14.9511840
## 2	11.9638706
## 4	10.8264232
## 10	10.7817440
## 68	1.5689829
## 89	1.5248323
## 82	1.2503004
## 231	1.1977828
## 210	1.1332249
## 96	1.1098758
## 140	1.1076674
## 397	1.0971923
## 361	1.0775532
## 286	1.0762308
## 61	1.0740152
## 516	1.0680820
## 33	1.0451334
## 551	1.0433765
## 445	1.0317830
## 474	1.0291369
## 331	1.0250419
## 559	1.0243757
## 530	1.0237370
## 259	1.0212595
## 429	1.0204791
## 252	1.0202573
## 268	1.0150804
## 19	1.0137555
## 340	1.0132932
## 405	1.0108892
## 245	1.0100160
## 322	1.0098454
## 375	1.0093062
## 382	1.0091169
## 591	1.0071980
## 47	1.0067778
## 575	1.0042547
## 313	1.0038554
## 495	1.0036545
## 170	1.0019589
## 460	1.0005056
## 437	0.9998782
## 502	0.9991520
## 509	0.9982094
## 467	0.9981707
## 150	0.9980314
## 75	0.9977081
## 295	0.9976800
## 180	0.9964084
## 389	0.9937542

```
## 103          0.9930219
## 413          0.9927896
## 523          0.9924044
## 421          0.9921585
## 40           0.9911345
## 607          0.9868791
## 453          0.9867059
## 599          0.9860355
## 12           0.9821732
## 583          0.9816619
## 190          0.9793547
## 481          0.9791535
## 54           0.9767836
## 160          0.9762911
## 304          0.9737579
## 224          0.9735481
## 200          0.9725752
## 544          0.9708972
## 217          0.9699886
## 120          0.9694418
## 110          0.9672238
## 130          0.9664926
## 368          0.9545894
## 537          0.9529309
## 277          0.9406746
## 488          0.9392117
## 238          0.9047898
## 354          0.8995475
## 26           0.8520654
## 567          0.7581050
## 347          0.4746867
## 1            0.1977871
## 3            0.1961796
## 6            0.1860882
## 7            0.1801972
## 9            0.1599868
```

```
write.csv(KG1, 'keyGenes_UL_FCs_CNVs.csv', row.names=FALSE)
```

Order by gene count, then by fold change.

```
KG2 <- KG1[with(KG1, order(gene_count, foldChangeMean_UL_to_nonUL,
decreasing=TRUE)),]
```

Lets add in a fold change of the median value ratios of UL to non-UL samples to compare.

```
colnames(KG2)[18] <- 'foldChange_Mean'
KG2$foldChange_Median <- KG2$UL_Median/KG2$nonUL_Median
```

Lets look at some of these copy number variants of one gene with seven [copy number variants](#) or CNVs and see where the changes in the nucleotide sequences occur. Copy

number variations in nucleotides can have short repeats, jumps in sequence, insertions, or deletions of a gene. I have been calling these CNVs [genotypes](#), which are the traits and alleles responsible for the physical traits or phenotypes of an organism. Some CNVs are responsible for diseases, and in tumors there are many different CNVs that are found to be responsible. A uterine leiomyoma or fibroid is a benign tumor. These samples were taken from uterus tissue with these uterine tumors and the same neighboring uterine tissue without uterine tumors.

```
CTNNB1 <- subset(KG2, KG2$Symbol=='CTNNB1')
CTNNB1_seq <- CTNNB1[,1:2]
```

Add in a column to describe the length of the nucleotides in each copy number variant nucleotide strand.

```
CTNNB1_seq$SEQUENCE <- as.character(CTNNB1$SEQUENCE)
CTNNB1_seq$nChar <- nchar(CTNNB1_seq$SEQUENCE)
```

Lets look at the CNVs of the CTNNB1 gene.

```
CTNNB1_seq
```

##	Symbol	SEQUENCE	nChar
## 68	CTNNB1	AGCTGCAGGGGTCCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAG	50
## 89	CTNNB1	CTGCAGGGGTCCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAGCT	50
## 82	CTNNB1	AGTCTCTCGTAGTGTTAAGTTATAGTGAATACTGCTACAGCAATTTCTAA	50
## 96	CTNNB1	GCCTCTTGCACTCTGAATTGGGAATGTTTGCACCACAGTGGGGGGCTTGC	50
## 61	CTNNB1	CAAACCTTACAGAGGAGAATGCCCTGTTTGTAAACCATGTTTCTTTTGGC	50
## 75	CTNNB1	CAGGAATCTAGTCTGGATGACTGCTTCTGGAGCCTGGATGCAGTACCATT	50
## 103	CTNNB1	GCAATTTGCCAAGTTTCTTTAGCATTTGGCCCTGGATTACGCTGGACCCC	50

From the above, some of the CNVs make you wonder if they are even the same gene. The first two have the same pattern of 'CTGCAGGG' then some variations. Then its not obvious what the other sequence alignments are. We could go back to the cytoband location and where the gene starts to see if there is more information.

Lets get the SEQUENCE, protein product, and cytoband columns from the original UL1 table.

```
cytoband <- UL1[,c(15,20,24)]
```

Now combine with the CTNNB1_seq and the KG2 table.

```
CTNNB1_cyto <- merge(cytoband, CTNNB1_seq, by.x='SEQUENCE', by.y='SEQUENCE')
KG2_cyto <- merge(cytoband, KG2, by.x='SEQUENCE', by.y='SEQUENCE')
```

Now lets look at the KG2_cyto table to see where these CNVs are located within the cytoband of each gene location.

```
KG3 <- KG2_cyto[with(KG2_cyto, order(gene_count, foldChange_Mean, decreasing =
TRUE)),]
KG3[,1:5]
```

	SEQUENCE	Protein_Product
## 37	CCGCCGGGCTGCTTTTTCTTGATGCCCATCAGGACGCCTCAGTTCTCT	XP_936926.1
## 56	GCAGGGGAGATTGGGTTTAGGGCTTTCCTGGTCTGCATTCTGCTACAGC	XP_937020.1
## 26	CAGGGCAGGAACCACGTCTTTACAGTTTGTATGTTCCAGAGCTGACCCAG	XP_936919.1
## 77	TCTCCTGCCCCCTCCGGAAGCTTGGATGCCCCCTCCACACCCTCTTGATCT	XP_936947.1
## 41	CGTTGCTACAAGCTGTTTTTTGAATGTCTCTACACAGTCCAGGCAGGAAG	XP_937000.1
## 20	CAGACTTCTCGCTTCCTTTCTGCTGGGCCTCTGAGGGGTCATGGGGCCAT	XP_936988.1
## 84	TTACTGGGGATGGTATTTAGGAGCCAGGAAAGCCGGTGCATTCTAGTGA	XP_936932.1
## 6	ACATGTGCTGTCACTGGAACCTGCTCTTTTCACTCAGCAGCCAGAGGGTC	XP_936976.1
## 30	CCAGTCCCTGACTACAGAGGATTTCCCCAAAGTCCCTGGCTGTGAGGTTC	XP_936952.1
## 1	AAACGTTACAGTGTTCCGATGAGACACAGTAGGCAGTACTTGGGAGGGTC	XP_936980.1
## 72	TAAGTGCAGTGAGCTCTGGCGGAAACCACCCTCTGCCCCGTCTGTTGGAT	XP_935983.1
## 28	CATTGTAATGATAAGGAAATGTTGCGATCAAATAAGATTTAGACACACTT	XP_935991.1
## 49	GAGTGCTGGGAAGGTTAATGTTAAATGGGTTGTGTGTCGGGGAGGGTACA	XP_935974.1
## 51	GCAAATGTAACCTCAGGGGTTTGGGGCCAGAGGAAGAGGGAGAAGGTGGCC	XP_935936.1
## 10	AGCTCCACCTTGACCCAGCCTCACAACAAAAAGTTTGTGTATGACCAGGC	XP_935967.1
## 50	GATCACAGGCACAGGGAAGCCACAAGGAGCTCTGTATGAGTTGTGTTTGC	XP_935893.1
## 71	GTTGTTCTGGACGATCTTCGGGATCCTCTGGGGCACTGTGACACTCGGAG	XP_936004.1
## 4	AAGTGGTGCCTGGCTGTCCCTATACTGTGCTGCTGGGTGTTCCAGCCTGT	XP_935903.1
## 25	CAGGCGACTGGGTAGCAGATGTGGAAGCTGATGGTTAGGCCAGGGCATG	XP_935881.1
## 74	TCAACCACATCCTTCAAAGGACTATGCCTGTTTATAAGCCCAGCTGTTT	XP_938957.1
## 33	CCCGACGAGCAGGACTTCATCCAGGCCTACGAGGAGGTGCGCGAGAAGTA	NP_958780.1
## 83	TGTCGTTTTCTCCATTCTTCACAAAAACATCAGCGTACATAGGCACATGG	XP_938806.1
## 35	CCCTCGGGCAGCCTGTTTTCCCTCCCTGGTGGTTGTGGGGTCACGTTGTCAC	NP_958784.1
## 64	GGCTCCTCTTTGGGCTCCTACTGGAATTTATCAGCCATCAGTGCATCTCT	XP_938917.1
## 2	AAAGCAGTGGTTTTTCAGCTGCCAGAGGCCTGAGAGAGTTTGGGCATACTC	XP_938927.1
## 38	CCGGGCCTTCTCGTGGTACCCTGCCTGCTGCCTTTGCCCGCACTGACT	NP_958782.1
## 63	GGCGCAGACATGGACCCCTCGCGAGCCATCCAGAACGAGATCAGCTCCCT	NP_958781.1
## 65	GGCTCTGTTGGAATCCGCATAGTGTGGAATGAGTTTGCCCTGGAAAGGG	XP_938916.1
## 45	CTGGCCTTCCCTCATCAGCCGTAAATGATGATTTACTGCTGTTACCATCA	XP_939002.1
## 36	CCCTTCTACATTCTTGTTTTTCATTTTTTTCGGAGGAAGAGGAGTTGCTAG	XP_938960.1
## 66	GGGAAGTACATGGGGCAGATGGAAGAACCTGAGATAATCGCAAGGATGGC	XP_938807.1
## 42	CTCCGTCTGCCCCGTGGGCTCCTGCCACCGTCCCCGATGAAGATCGTGCC	NP_958783.1
## 61	GCCTTTGCCTCGCCGAGGGAGGTCTTGCTGGAGCGGCCGTGCTGGCTGGA	NP_958785.1
## 22	CAGCCCTGGGGACACACTGCCCTGGAACCTTGGGAAAACGCAGCGGAGCC	NP_000436.2
## 9	AGCCTCTGTTCCCTAGTAAGTGCCTTCCATGTGCGGCTCTAACCCAGG	NP_958786.1
## 11	AGCTGCAGGGGTCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAG	XP_950743.1
## 44	CTGCAGGGGTCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAGCT	NP_001895.1
## 13	AGTCTCTCGTAGTGTTAAGTTATAGTGAATACTGCTACAGCAATTTCTAA	NP_001895.1
## 27	CAGTGTTGGGATCACTCACTTTCCCCCTACAGGACTCAGATCTGGGAGGC	NP_003997.1
## 43	CTCCTCTCAGCTGAACACCCTCCTTTCACTCCCAAATGCAAACAGTCTCT	NP_004010.1
## 60	GCCTCTTGCACTCTGAATTGGGAATGTTTGACCACAGTGGGGGGCTTGC	XP_950747.1
## 57	GCCAAAGGAATGGGCTCCAGACACCCCTCTTCCAGAGCAAGGATGAAGG	XP_937236.1
## 19	CAAACCTTTACAGAGGAGAATGCCCTGTTTGTTAACCATGTTTCTTTTGGC	XP_950746.1
## 80	TGATGTGTACGCCACTGTACTCCAGCCTGACGGCAGAGCGAGACTCCAT	XP_936088.1
## 34	CCCTCCACCTCCTGCTCGGGGGGCTTTAATGAGACACCCACCGCTGCTGT	NP_001700.2

## 7	ACTGCCTGTGTGGCTCCTTGAGTGCGCGGAGGCCAAAGCTGAGATGACTT	XP_937640.1
## 69	GGTGTGCTCTGGTATGTAATGACAATATGTGAACAAACCTGTGGAATTAA	XP_936056.1
## 76	TCTATCAACAGAGCTGAATGAGTGCCAGGAAGCTGCGAAATCTGTCTTAC	NP_004003.1
## 5	AATAATAGAGTGTGGGAGTTTTGGGGCCGAAGCTTTCCCGGAGCAGCTG	NP_733929.1
## 18	CAAACCTTTGAAGACATTTTCAGGGCCATGCTCACTTGGGAGGGTTTTGAGG	XP_937228.1
## 31	CCATTGAGAAGAATGATAAATGCCACAAGCATTTGGAAACAGGCTTCCCT	NP_004001.1
## 15	AGTGGGCAGAATGATGAGGGAAGTGGGCACGTGCCCATGTTCTTCTTGGC	XP_937222.1
## 85	TTCATCCAGGCCTGCGCCGGTGTTACAGTGGTCCTCATCTAAGCCAGCC	XP_937214.1
## 62	GCTCGCTGAAGTTGGCTTCCTAGCGGTGTAGGCTGGAATAGACTCTTGGC	NP_733928.1
## 86	TTTCAGGCCCATGGCAGAGGGTGGGCTCAGGAGGGCCATCGTGGGTGTCC	XP_937694.1
## 14	AGTGCCACATCACACAGCATCTAGCACGTAACCTGCACCCCGGGAGTCGT	XP_937456.1
## 16	AGTGTTGGGACTACAGGTGTGTGTTACTGCTCCCAGCTGGGAGGCAGGCT	XP_936104.1
## 70	GTGAGCCTGTTTCATCATCTGTAACTTTGAATAATGATACCTACCCCGC	XP_936038.1
## 40	CGCCCTGAAAGGACCAGGACATGCGGGTGCGGTGGCTGCTCTTTTGGCTC	XP_937724.1
## 24	CAGGAATCTAGTCTGGATGACTGCTTCTGGAGCCTGGATGCAGTACCATT	XP_950748.1
## 53	GCAATTTGCCAAGTTTTCTTTAGCATTTGGCCCTGGATTACGCTGGACCCC	XP_947138.1
## 8	AGCAGCACATCGTCATTTTACAATTGAGAAACATGGAGACTCCAAATGGA	XP_936080.1
## 75	TCAGACCCCTCAGGCCACTGCTGTTCTGTACACATTCTGCAAAGGAC	NP_733931.1
## 87	TTTCCTGAAATGGAGCTTTGCTCTTGTTGCCAGGCCGTAGTGCAATGGC	XP_937537.1
## 17	ATGTACGTGGGGGATTCTTGACTCGGGTTAGTCTCTGGGGATGCAGAGCC	NP_733930.1
## 78	TCTGTATGGACCCTGCCAAGCTCTGCCCTCTGCCCTGCATTGGGGCGC	XP_937505.1
## 82	TGGGGAGACGAGATTTTAAGACACTTGAGTCTCCAGGACAGCAAAGGCAC	NP_733927.1
## 55	GCAGCCAACCTATTGGCATGATGGAGTGACAGGAAAAACAGCTGGCATGG	NP_000100.2
## 73	TACCTGGCCTATCTTTCATAGGTTATATAAATTCCTTGGTTCCCAGTTTT	XP_936049.1
## 67	GGGTTTTTCTCAGGATTGCTATGCAACAGGATCAGTGCTGTAGTGCCCGGT	NP_004005.1
## 47	GACCAAAGCAGGACAATTGCTTGATCCCAGGAGTTTAAGACCAGCCGGGG	XP_932593.1
## 3	AAGGACTCAGATGCAGGGTCTTCTCTGCTCCCCGTACACAGAGGGTGGC	XP_936046.1
## 48	GACCTGTAGCTAAACCTTCCACCAGCGCTTGAGAAGTTAATTTGAACCGG	XP_937490.1
## 54	GCACTCCGACTACATCAGGAGAAGATGTTTCGAGACTTTGCCAAGGTACTA	NP_004010.1
## 29	CCACGCCGGCAAAGAAATTGGAAGACTCCACCATTACAGGCAGCCACCAG	XP_937214.1
## 46	CTTGCTGTGGTCTCTTTGTGGCAGAAGTGTTTCATGCATGGCAGCAGGCC	NP_001700.2
## 39	CCTGTTTGGATCACATGGTCTTGTCCTGATAACTTGAAGAGGTTGCTTC	XP_371783.3
## 81	TGCGAGACCTGGGTGTCCAACCTGCGCTACAACCACATGCTGCGGAAGAA	NP_003827.3
## 52	GCAACGCTCCTCTGAAATGCTTGCTTTTTTCTGTTGCCGAAATAGCTGG	NP_061159.1
## 59	GCCCCAGCAAGCCTCCCTCCATCCTCCAGTGGGAAACTGTTGATGGTGTT	NP_006202.1
## 68	GGTATTGCTGATCGTATGCAGAAGGAAATCACTGCTCTGGCTCCTAGCAC	NP_005150.1
## 12	AGGCCCTGGAGGCTGCAACATACCTCAATCCTGTCCCAGGCCGGATCCTC	NP_005931.2
## 58	GCCAACCTCCTCTCACAGCCTCTGTATCTCTGCAGGCCATACTGGTTCCA	NP_009099.1
## 21	CAGATGTTTTCCCTTGTTGGCAGTCTTCAGCCTCCTCTACCCTACATGATC	NP_000658.1
## 32	CCCAGTGACACTTCAGAGAGCTGGTAGTTAGTAGCATGTTGAGCCAGGCC	NP_005243.1
## 79	TGACTGTCCCTGCCAATGCTCCAGCTGTCTGCTGACTCTGGGTTCTGTTGG	NP_006723.1
## 23	CAGCTGGGCGATGTGCGAGCTGATAGTGAGCGGCAGAATCAGGAGTACCA	NP_002267.2

##	Cytoband	Symbol	gene_count
## 37	12p13.31a	DDX12	10
## 56	12p13.31a	DDX12	10
## 26	12p13.31a	DDX12	10
## 77	12p13.31a	DDX12	10
## 41	12p13.31a	DDX12	10
## 20	12p13.31a	DDX12	10
## 84	12p13.31a	DDX12	10

## 6	12p13.31a	DDX12	10
## 30	12p13.31a	DDX12	10
## 1	12p13.31a	DDX12	10
## 72	12q24.33d	KIAA0692	9
## 28	12q24.33d	KIAA0692	9
## 49	12q24.33d	KIAA0692	9
## 51	12q24.33d	KIAA0692	9
## 10	12q24.33d	KIAA0692	9
## 50	12q24.33d	KIAA0692	9
## 71	12q24.33d	KIAA0692	9
## 4	12q24.33d	KIAA0692	9
## 25	12q24.33d	KIAA0692	9
## 74	16p12.2a	LOC23117	8
## 33	8q24.3g	PLEC1	8
## 83	16p12.2a	LOC23117	8
## 35	8q24.3g	PLEC1	8
## 64	16p12.2a	LOC23117	8
## 2	16p12.2a	LOC23117	8
## 38	8q24.3g	PLEC1	8
## 63	8q24.3g	PLEC1	8
## 65	16p12.2a	LOC23117	8
## 45	16p12.2a	LOC23117	8
## 36	16p12.2a	LOC23117	8
## 66	16p12.2a	LOC23117	8
## 42	8q24.3g	PLEC1	8
## 61	8q24.3g	PLEC1	8
## 22	8q24.3g	PLEC1	8
## 9	8q24.3g	PLEC1	8
## 11	3p22.1b	CTNNB1	7
## 44	3p22.1b	CTNNB1	7
## 13	3p22.1b	CTNNB1	7
## 27	Xp21.2a-p21.1d	DMD	7
## 43	Xp21.2a-p21.1d	DMD	7
## 60	3p22.1b	CTNNB1	7
## 57	5q35.2d	LOC202134	7
## 19	3p22.1b	CTNNB1	7
## 80	NA	LOC653086	7
## 34	11p14.1d	BDNF	7
## 7	16p13.11b	LOC339047	7
## 69	NA	LOC653086	7
## 76	Xp21.2a-p21.1d	DMD	7
## 5	11p14.1d	BDNF	7
## 18	5q35.2d	LOC202134	7
## 31	Xp21.2a-p21.1d	DMD	7
## 15	5q35.2d	LOC202134	7
## 85	5q35.2d	LOC202134	7
## 62	11p14.1d	BDNF	7
## 86	16p13.11b	LOC339047	7
## 14	16p13.11b	LOC339047	7
## 16	NA	LOC653086	7

## 70	NA	LOC653086	7
## 40	16p13.11b	LOC339047	7
## 24	3p22.1b	CTNNB1	7
## 53	3p22.1b	CTNNB1	7
## 8	NA	LOC653086	7
## 75	11p14.1d	BDNF	7
## 87	16p13.11b	LOC339047	7
## 17	11p14.1d	BDNF	7
## 78	16p13.11b	LOC339047	7
## 82	11p14.1d	BDNF	7
## 55	Xp21.2a-p21.1d	DMD	7
## 73	NA	LOC653086	7
## 67	Xp21.2a-p21.1d	DMD	7
## 47	5q35.2d	LOC202134	7
## 3	NA	LOC653086	7
## 48	16p13.11b	LOC339047	7
## 54	Xp21.2a-p21.1d	DMD	7
## 29	5q35.2d	LOC202134	7
## 46	11p14.1d	BDNF	7
## 39	5q35.2d	LOC202134	7
## 81	14q32.2b	DLK1	2
## 52	15q25.1b	KIAA1199	1
## 59	8q12.1b	PENK	1
## 68	15q14a	ACTC	1
## 12	22q11.23a	MMP11	1
## 58	17q24.2c	ABCA8	1
## 21	4q23b	ADH1A	1
## 32	14q24.3b	FOS	1
## 79	19q13.32a	FOSB	1
## 23	17q21.2b	KRT19	1

```
CTNNB1_b <- subset(KG3, KG3$Symbol=='CTNNB1')
CTNNB1_b[,c(1:5,20:21)]
```

##	SEQUENCE	Protein_Product
##	Cytoband	
## 11	AGCTGCAGGGTCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAG	XP_950743.1
	3p22.1b	
## 44	CTGCAGGGTCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAGCT	NP_001895.1
	3p22.1b	
## 13	AGTCTCTCGTAGTGTTAAGTTATAGTGAATACTGCTACAGCAATTTCTAA	NP_001895.1
	3p22.1b	
## 60	GCCTCTTGCACTCTGAATTGGGAATGTTTGCACCACAGTGGGGGGCTTGC	XP_950747.1
	3p22.1b	
## 19	CAAACCTTACAGAGGAGAATGCCCTGTTTGTTAACCATGTTTCTTTTGGC	XP_950746.1
	3p22.1b	
## 24	CAGGAATCTAGTCTGGATGACTGCTTCTGGAGCCTGGATGCAGTACCATT	XP_950748.1
	3p22.1b	
## 53	GCAATTTGCCAAGTTTCTTTAGCATTTGGCCCTGGATTACGCTGGACCCC	XP_947138.1
	3p22.1b	

##	Symbol	gene_count	foldChange_Mean	foldChange_Median
## 11	CTNNB1	7	1.5689829	1.5744314
## 44	CTNNB1	7	1.5248323	1.4655024
## 13	CTNNB1	7	1.2503004	1.1602376
## 60	CTNNB1	7	1.1098758	1.0850642
## 19	CTNNB1	7	1.0740152	1.0471422
## 24	CTNNB1	7	0.9977081	0.9924948
## 53	CTNNB1	7	0.9930219	0.9818097

The cytoband location of each of these CNVs for CTNNB1 is the same location on chromosome 3 on the p strand/direction along 22.1b. Also, the fold change for the mean and median values for the first listed CNVs changed by 16-57 percent more in UL compared to non-UL samples. This could mean that these four CNVs of the gene CTNNB1 offer some clues as to what mutations or changes impact risk in developing uterine leiomyomas for some females.

Lets order the key genes by fold change median then by CNVs.

```
KG4 <- KG3[with(KG3, order(foldChange_Median, gene_count, decreasing =
TRUE)),]
KG4[,c(1:5,21)]
```

##	SEQUENCE	Protein_Product
## 52	GCAACGCTCCTCTGAAATGCTTGCTTTTTTCTGTTGCCGAAATAGCTGG	NP_061159.1
## 68	GGTATTGCTGATCGTATGCAGAAGGAAATCACTGCTCTGGCTCCTAGCAC	NP_005150.1
## 12	AGGCCCTGGAGGCTGCAACATACCTCAATCCTGTCCCAGGCCGGATCCTC	NP_005931.2
## 81	TGCGAGACCTGGGTGTCCAACCTGCGCTACAACCACATGCTGCGGAAGAA	NP_003827.3
## 59	GCCCCAGCAAGCCTCCCTCCATCCTCCAGTGGGAAACTGTTGATGGTGTT	NP_006202.1
## 11	AGCTGCAGGGGTCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAG	XP_950743.1
## 44	CTGCAGGGGTCTCTGTGAACCTTGCTCAGGACAAGGAAGCTGCAGAAGCT	NP_001895.1
## 43	CTCCTCTCAGCTGAACACCCTCCTTTCACTCCCAAATGCAAACAGTCTCT	NP_004010.1
## 13	AGTCTCTCGTAGTGTTAAGTTATAGTGAATACTGCTACAGCAATTTCTAA	NP_001895.1
## 27	CAGTGTGGGATCACTCACTTTCCCCCTACAGGACTCAGATCTGGGAGGC	NP_003997.1
## 60	GCCTCTTGCACTCTGAATTGGAATGTTTGACCACAGTGGGGGGCTTGC	XP_950747.1
## 74	TCAACCACATCCTTCAAAAGGACTATGCCTGTTTATAAGCCCAGCTGTTT	XP_938957.1
## 10	AGCTCCACCTTGACCCAGCCTCACAACAAAAAGTTTGTGTATGACCAGGC	XP_935967.1
## 19	CAAACCTTACAGAGGAGAATGCCCTGTTTGTTAACCATGTTTCTTTTGGC	XP_950746.1
## 33	CCCGACGAGCAGGACTTCATCCAGGCCTACGAGGAGGTGCGCGAGAAGTA	NP_958780.1
## 80	TGATGTGTCACGCCACTGTACTCCAGCCTGACGGCAGAGCGAGACTCCAT	XP_936088.1
## 31	CCATTGAGAAGAATGATAAATGCCACAAGCATTTGGAAACAGGCTTCCCT	NP_004001.1
## 64	GGCTCCTCTTTGGGCTCCTACTGGAATTTATCAGCCATCAGTGCATCTCT	XP_938917.1
## 69	GGTGTGCTCTGGTATGTAATGACAATATGTGAACAAACCTGTGGAATTAA	XP_936056.1
## 28	CATTGTAATGATAAGGAAATGTTGCGATCAAATAAGATTTAGACACACTT	XP_935991.1
## 76	TCTATCAACAGAGCTGAATGAGTGCCAGGAAGCTGCGAAATCTGTCTTAC	NP_004003.1
## 16	AGTGTGGGACTACAGGTGTGTGTTACTGCTCCCAGCTGGGAGGCAGGCT	XP_936104.1
## 37	CCGCCGGGCTGCTTTTTCTTGATGCCCATCAGGACGCCTCAGTTCTCT	XP_936926.1
## 86	TTTCAGGCCCATGGCAGAGGGTGGGCTCAGGAGGGCCATCGTGGGTGTCC	XP_937694.1
## 49	GAGTGCTGGGAAGGTTAATGTTAAATGGGTTGTGTGTCGGGGAGGGTACA	XP_935974.1
## 83	TGTCGTTTCTCCATTCTTCACCAAAACATCAGCGTACATAGGCACATGG	XP_938806.1
## 35	CCCTCGGGCAGCCTGTTTCCCTCCCTGGTGGTTGTGGGTCACGTTGTCAC	NP_958784.1

## 57	GCCAAAGGAATGGGCTCCAGACACCCCTCTTCCAGAGCAAGGATGAAGG	XP_937236.1
## 14	AGTGCCACATCACACAGCATCTAGCACGTAAGTGCACCCGGGAGTCGT	XP_937456.1
## 50	GATCACAGGCACAGGGAAGCCACAAGGAGCTCTGTATGAGTTGTGTTTGC	XP_935893.1
## 70	GTGAGCCTGTTTCATCATCTGTAACTTTGAATAATGATACCTACCCCGC	XP_936038.1
## 38	CCGGCCTTCTCGTGGTACCCTGCCTGCTGCCTTTGCCCCCGCACTGACT	NP_958782.1
## 65	GGCTCTGTTGGAATCCGCATAGTGTGAAATGAGTTTGCCCTGGAAAGGG	XP_938916.1
## 78	TCTGTATGGACCCTGCCAAGCTCTGCCCCTCTGCCCCTGCATTGGGGCGC	XP_937505.1
## 85	TTCATCCAGGCCTGCGCCGGTGTTACAGTGGTCCTCATCTAAGCCAGCC	XP_937214.1
## 7	ACTGCCTGTGTGGCTCCTTGAGTGC GCGGAGGCCAAAGCTGAGATGACTT	XP_937640.1
## 5	AATAATAGAGTGTGGGAGTTTTGGGGCCGAAGTCTTTCCCGGAGCAGCTG	NP_733929.1
## 18	CAAACCTTGAAGACATTTTCAGGGCCATGCTCACTTGGGAGGGTTTGAGG	XP_937228.1
## 62	GCTCGCTGAAGTTGGCTTCTAGCGGTGTAGGCTGGAATAGACTCTTGGC	NP_733928.1
## 66	GGGAAGTACATGGGGCAGATGGAAGAACCTGAGATAATCGCAAGGATGGC	XP_938807.1
## 41	CGTTGCTACAAGCTGTTTTTTGAATGTCTCTACACAGTCCAGGCAGGAAG	XP_937000.1
## 45	CTGGCCTTCCCTCATCAGCCGTAAATGATGATTTACTGCTGTTACCATCA	XP_939002.1
## 51	GCAAATGTAAGTCAAGGGGTTTGGGGCCAGAGGAAGAGGGAGAAGGTGGCC	XP_935936.1
## 40	CGCCCTGAAAGGACCAGGACATGCGGGTGCGGTGGCTGCTCTTTTGGCTC	XP_937724.1
## 84	TTACTGGGGATGGTATTTTAGGAGCCAGGAAAGCCGGTGCAATTCCTAGTGA	XP_936932.1
## 17	ATGTACGTGGGGGATTCTTGACTCGGGTTAGTCTCTGGGGATGCAGAGCC	NP_733930.1
## 20	CAGACTTCTCGCTTCTCTTCTGCTGGGCCTCTGAGGGGTGATGGGGCCAT	XP_936988.1
## 24	CAGGAATCTAGTCTGGATGACTGCTTCTGGAGCCTGGATGCAGTACCATT	XP_950748.1
## 36	CCCTTCTACATTCTTGTTTTTCAATTTTTTCGGAGGAAGAGGAGTTGCTAG	XP_938960.1
## 15	AGTGGGCAGAATGATGAGGGAAGTGGGCACGTGCCCATGTTCTTCTTGGC	XP_937222.1
## 26	CAGGGCAGGAACCACGTCTTTACAGTTTGATGTTCCAGAGCTGACCCAG	XP_936919.1
## 2	AAAGCAGTGGTTTTTCAGCTGCCAGAGGCCTGAGAGAGTTTGGGCATACTC	XP_938927.1
## 75	TCAGACCCCTCAGGCCACTGCTGTTCTGTGCACACATTCTGCAAAGGAC	NP_733931.1
## 56	GCAGGGGAGATTGGGTTTAGGGGCTTTCCTGGTCTGCATTCTGCTACAGC	XP_937020.1
## 55	GCAGCCAACCTATTGGCATGATGGAGTGACAGGAAAAACAGCTGGCATGG	NP_000100.2
## 8	AGCAGCACATCGTCATTTTACAATTGAGAAACATGGAGACTCCAAATGGA	XP_936080.1
## 42	CTCCGTCTGCCCCGTGGGCTCCTGCCACCGTCCCCGATGAAGATCGTGCC	NP_958783.1
## 53	GCAATTTGCCAAGTTTTCTTTAGCATTTGGCCCTGGATTACGCTGGACCCC	XP_947138.1
## 61	GCCTTTGCCTCGCCGAGGGAGGTCTTGCTGGAGCGGCCGTGCTGGCTGGA	NP_958785.1
## 72	TAAGTGCAGTGAGCTCTGGCGGAAACCACCCTCTGCCCCGTCTGTTGGAT	XP_935983.1
## 71	GTTGTTCTGGACGATCTTCGGGATCCTCTGGGGCACTGTGACACTCGGAG	XP_936004.1
## 22	CAGCCCTGGGGACACACTGCCCTGGAACCTTGGGAAAACGCAGCGGAGCC	NP_000436.2
## 73	TACCTGGCCTATCTTTCATAGGTTATATAAATTCCTTGGTTCCCAGTTTT	XP_936049.1
## 77	TCTCCTGCCCCCTCCGGAAGCTTGATGCCCCCTCCACACCCTCTTGATCT	XP_936947.1
## 63	GGCGCAGACATGGACCCCTCGCGAGCCATCCAGAACGAGATCAGCTCCCT	NP_958781.1
## 67	GGGTTTTCTCAGGATTGCTATGCAACAGGATCAGTGCTGTAGTGCCCGGT	NP_004005.1
## 82	TGGGGAGACGAGATTTTAAGACACTTGAGTCTCCAGGACAGCAAAGGCAC	NP_733927.1
## 4	AAGTGGTGCCTGGCTGTCCCTATACTGTGCTGCTGGGTGTTCCAGCCTGT	XP_935903.1
## 48	GACCTGTAGCTAAACCTTCCACCAGCGCTTGAGAACTTAATTTGAACCGG	XP_937490.1
## 87	TTTCTGAAATGGAGCTTTGCTCTTGTTGCCAGGCCGTAGTGCAATGGC	XP_937537.1
## 6	ACATGTGCTGTCACTGGAACCTGCTCTTTTCACTCAGCAGCCAGAGGGTC	XP_936976.1
## 34	CCCTCCACCTCCTGCTCGGGGGGCTTTAATGAGACACCCACCGCTGCTGT	NP_001700.2
## 30	CCAGTCCCTGACTACAGAGGATTTCCCCAAAGTCCCTGGCTGTGAGGTTT	XP_936952.1
## 1	AAACGTTACAGTGTTCCGATGAGACACAGTAGGCAGTACTTGGGAGGGTC	XP_936980.1
## 3	AAGGACTCAGATGCAGGGTCTTCTCTGCTCCCCGTACACAGAGGGTGGC	XP_936046.1
## 47	GACCAAAGCAGGACAATTGCTTGATCCCAGGAGTTTAAGACCAGCCGGGG	XP_932593.1
## 46	CTTGCTGTGGTCTCTTTGTGGCAGAAGTGTTTCATGCATGGCAGCAGGCC	NP_001700.2

## 29	CCACGCCGGCAAAGAAATTGGAAGACTCCACCATTACAGGCAGCCACCAG	XP_937214.1
## 25	CAGGCGACTGGGTAGCAGATGTGGAAGCTGATGGTTAGGCCCAGGGCATG	XP_935881.1
## 54	GCACTCCGACTACATCAGGAGAAGATGTTTCGAGACTTTGCCAAGGTACTA	NP_004010.1
## 9	AGCCTCTGTTCCCTAGTAAGTGCCTTCCATGTCGGCCTCTAACCCCAGG	NP_958786.1
## 39	CCTGTTTGGATCACATGGTCTTGTCTCTGATAACTTGAAGAGGTTGCTTC	XP_371783.3
## 21	CAGATGTTTTCCCTTGTGGCAGTCTTCAGCCTCCTCTACCCTACATGATC	NP_000658.1
## 58	GCCAACCTCCTCTCACAGCCTCTGTATCTCTGCAGGCCATACTGGTTCCA	NP_009099.1
## 32	CCCAGTGACACTTCAGAGAGCTGGTAGTTAGTAGCATGTTGAGCCAGGCC	NP_005243.1
## 79	TGACTGTCCCTGCCAATGCTCCAGCTGTCGTCTGACTCTGGGTTCGTTGG	NP_006723.1
## 23	CAGCTGGGCGATGTGCGAGCTGATAGTGAGCGGCAGAATCAGGAGTACCA	NP_002267.2
##	Cytoband Symbol gene_count foldChange_Median	
## 52	15q25.1b KIAA1199 1 21.9688396	
## 68	15q14a ACTC 1 14.6883331	
## 12	22q11.23a MMP11 1 11.8480229	
## 81	14q32.2b DLK1 2 4.7325319	
## 59	8q12.1b PENK 1 3.3078453	
## 11	3p22.1b CTNNB1 7 1.5744314	
## 44	3p22.1b CTNNB1 7 1.4655024	
## 43	Xp21.2a-p21.1d DMD 7 1.2286298	
## 13	3p22.1b CTNNB1 7 1.1602376	
## 27	Xp21.2a-p21.1d DMD 7 1.1029701	
## 60	3p22.1b CTNNB1 7 1.0850642	
## 74	16p12.2a LOC23117 8 1.0667330	
## 10	12q24.33d KIAA0692 9 1.0473309	
## 19	3p22.1b CTNNB1 7 1.0471422	
## 33	8q24.3g PLEC1 8 1.0462839	
## 80	NA LOC653086 7 1.0414456	
## 31	Xp21.2a-p21.1d DMD 7 1.0377049	
## 64	16p12.2a LOC23117 8 1.0286458	
## 69	NA LOC653086 7 1.0279570	
## 28	12q24.33d KIAA0692 9 1.0247769	
## 76	Xp21.2a-p21.1d DMD 7 1.0233573	
## 16	NA LOC653086 7 1.0191080	
## 37	12p13.31a DDX12 10 1.0179040	
## 86	16p13.11b LOC339047 7 1.0175470	
## 49	12q24.33d KIAA0692 9 1.0151057	
## 83	16p12.2a LOC23117 8 1.0146392	
## 35	8q24.3g PLEC1 8 1.0138161	
## 57	5q35.2d LOC202134 7 1.0137122	
## 14	16p13.11b LOC339047 7 1.0120598	
## 50	12q24.33d KIAA0692 9 1.0118272	
## 70	NA LOC653086 7 1.0087172	
## 38	8q24.3g PLEC1 8 1.0081202	
## 65	16p12.2a LOC23117 8 1.0071909	
## 78	16p13.11b LOC339047 7 1.0071241	
## 85	5q35.2d LOC202134 7 1.0066428	
## 7	16p13.11b LOC339047 7 1.0047835	
## 5	11p14.1d BDNF 7 1.0046246	
## 18	5q35.2d LOC202134 7 1.0037413	
## 62	11p14.1d BDNF 7 1.0016629	

## 66	16p12.2a	LOC23117	8	1.0009319
## 41	12p13.31a	DDX12	10	0.9972581
## 45	16p12.2a	LOC23117	8	0.9970746
## 51	12q24.33d	KIAA0692	9	0.9962676
## 40	16p13.11b	LOC339047	7	0.9950549
## 84	12p13.31a	DDX12	10	0.9945794
## 17	11p14.1d	BDNF	7	0.9937282
## 20	12p13.31a	DDX12	10	0.9930535
## 24	3p22.1b	CTNNB1	7	0.9924948
## 36	16p12.2a	LOC23117	8	0.9922770
## 15	5q35.2d	LOC202134	7	0.9917228
## 26	12p13.31a	DDX12	10	0.9915254
## 2	16p12.2a	LOC23117	8	0.9908288
## 75	11p14.1d	BDNF	7	0.9900315
## 56	12p13.31a	DDX12	10	0.9899665
## 55	Xp21.2a-p21.1d	DMD	7	0.9894561
## 8	NA	LOC653086	7	0.9847075
## 42	8q24.3g	PLEC1	8	0.9844314
## 53	3p22.1b	CTNNB1	7	0.9818097
## 61	8q24.3g	PLEC1	8	0.9800936
## 72	12q24.33d	KIAA0692	9	0.9784906
## 71	12q24.33d	KIAA0692	9	0.9774402
## 22	8q24.3g	PLEC1	8	0.9773820
## 73	NA	LOC653086	7	0.9770270
## 77	12p13.31a	DDX12	10	0.9757264
## 63	8q24.3g	PLEC1	8	0.9754829
## 67	Xp21.2a-p21.1d	DMD	7	0.9733874
## 82	11p14.1d	BDNF	7	0.9728164
## 4	12q24.33d	KIAA0692	9	0.9714923
## 48	16p13.11b	LOC339047	7	0.9696381
## 87	16p13.11b	LOC339047	7	0.9692033
## 6	12p13.31a	DDX12	10	0.9685507
## 34	11p14.1d	BDNF	7	0.9681280
## 30	12p13.31a	DDX12	10	0.9657570
## 1	12p13.31a	DDX12	10	0.9617268
## 3	NA	LOC653086	7	0.9575042
## 47	5q35.2d	LOC202134	7	0.9568036
## 46	11p14.1d	BDNF	7	0.9289947
## 29	5q35.2d	LOC202134	7	0.9285395
## 25	12q24.33d	KIAA0692	9	0.8823950
## 54	Xp21.2a-p21.1d	DMD	7	0.7956211
## 9	8q24.3g	PLEC1	8	0.7882088
## 39	5q35.2d	LOC202134	7	0.3998123
## 21	4q23b	ADH1A	1	0.1841845
## 58	17q24.2c	ABCA8	1	0.1539030
## 32	14q24.3b	FOS	1	0.1197627
## 79	19q13.32a	FOSB	1	0.1190193
## 23	17q21.2b	KRT19	1	0.1159735

The above table gives the protein products, the ontology function, the fold change of the median values of UL/nonUL, gene symbol, sequence of CNV, and cytoband location. The protein products can be found at genecards.org by entering the ID for the protein product into the search bar. A quick scan of a few of the protein products in genecards.org gave the following descriptions. The first listed protein NP_061159.1 says it is a colon cancer secreted protein. Many of the above CNVs are listed as proteins involved in the extracellular matrix like DMD and CTNNB1. There are also various neurological and synapses diseases associated with those proteins.

The site genecards.org has very useful properties in analyzing gene expression data from this research. If you are a member, you can download the network genes involved in diseases you query and compare to how the genes in certain tissues compare to those genes. Three out of the seven CNVs for CTNNB1 are in the top fold change median values in the ratio of UL/nonUL samples.

```
write.csv(KG4, 'keyGenes_topMedFCs.csv', row.names=FALSE)
```

Lets make a machine learning data set to test various algorithms on predicting if the sample is a UL or not. We will use the samples in this set, plus add in some microarray samples that have been studied by me elsewhere using this set of genes and sequences if available in any of the microarray studies.

Lets isolate those genes that are in our key genes of top picks for UL targets and combine the UL and nonUL sample information to those genes and sequences without the stats.

```
keyTargets <- KG4[,c(1,4)]

ULs <- UL[,c(2,10:29)]
colnames(ULs)[2:21] <- paste('UL', colnames(ULs)[2:21], sep='_')

nonULs <- nonUL[,c(2,10:27)]
colnames(nonULs)[2:19] <- paste('nonUL', colnames(nonULs)[2:19], sep='_')

keyULs <- merge(keyTargets, ULs, by.x='SEQUENCE', by.y='SEQUENCE')
keys <- merge(keyULs, nonULs, by.x='SEQUENCE', by.y='SEQUENCE')

write.csv(keys, 'keyGeneTargetsCNVs.csv', row.names=FALSE)
```

Lets create the matrix for machine learning.

```
keys_m1 <- as.data.frame(t(keys))
colnames(keys_m1) <- paste(keys$Symbol, keys$SEQUENCE, sep='_')
keys_m11 <- keys_m1[-(1:2),]
keys_m11$Type <-
as.factor(c(rep('UL', length(grep('^UL_', row.names(keys_m11)))),
            rep('nonUL', length(grep('^nonUL_', row.names(keys_m11))))))
keys_m12 <- keys_m11[,c(88,1:87)]
```

```
write.csv(keys_m12, 'm1_ready_UL_classes.csv', row.names=TRUE)
```

Now, let's pull in the other data sets that are from the microarray samples and see if we can get the genes and sequences that correspond to our key genes above in identifying a sample as UL or not with predictive analytics.

There is one study of the other studies that has Sequence, Gene symbol and a few UL and nonUL microarray samples to compare to this above beadchip UL and nonUL set. The GEO series ID is GSE68295 with the GEO platform of GPL6480. The files are 27 MB each in file size.

```
setwd('./microArray UL')

non <- read.csv('nonUL_GSE68295_GPL6480_table.csv', sep=',',
               header=T, na.strings=c('', ' '))
uls <- read.csv('UL_GSE68295_GPL6480_table.csv', sep=',',
               header=T, na.strings=c('', ' '))
setwd('./')
```

Keep only the needed columns.

```
uls_array <- uls[,c(8,18:21)]
colnames(uls_array)[3:5] <- paste('UL', colnames(uls_array)[3:5], sep='_')

non_array <- non[,c(8,18:21)]
colnames(non_array)[3:5] <- paste('nonUL', colnames(non_array)[3:5], sep='_')
```

The sequences don't align or match any in the microarrays with the beadchip UL samples.

```
uls_array0 <- merge(keyTargets, uls_array, by.x='SEQUENCE', by.y='SEQUENCE')
ulsArray0 <- uls_array0[, -2]
```

Match by gene symbol between the microarray and beadchip UL samples.

```
uls_array1 <- merge(keyTargets, uls_array, by.x='Symbol', by.y='GENE_SYMBOL')
ulsArray <- uls_array1[, -2]
```

The sequences don't align between the arrays and beadchip samples for nonULs.

```
non_array0 <- merge(keyTargets, non_array, by.x='SEQUENCE', by.y='SEQUENCE')
nonArray0 <- non_array0[, -(1:2)]
```

Match by Gene symbol between the microarray and beadchip samples of nonULs.

```
non_array1 <- merge(keyTargets, non_array, by.x='Symbol', by.y='GENE_SYMBOL')
nonArray <- non_array1[, -(1:2)]
```

Combine the UL and nonUL samples of the microarrays into one dataset.

```
microarrays <- merge(ulsArray, nonArray, by.x='SEQUENCE.y',
by.y='SEQUENCE.y')
Marrays <- microarrays[!duplicated(microarrays$SEQUENCE),]
```

Since these two expression types can't be compared by sequence, they should be compared by gene. Lets combine them into a study by gene expression values.

```
keys1 <- keys[, -1]
keys2 <- keys1 %>% group_by(Symbol) %>%
  summarise_at(vars(as.vector(colnames(keys1)[2:39])), mean)

Marrays1 <- Marrays[, -1]
Marrays2 <- Marrays1 %>% group_by(Symbol) %>%
  summarise_at(vars(as.vector(colnames(Marrays1)[2:7])), mean)

beadArrays <- merge(keys2, Marrays2, by.x='Symbol', by.y='Symbol')
```

There are only 12 genes in common among these combined samples of microarray and beadchip UL and nonUL samples.

```
beadArrays_ML <- as.data.frame(t(beadArrays))
colnames(beadArrays_ML) <- beadArrays$Symbol
beadArrays_ML2 <- beadArrays_ML[-1,]
beadArrays_ML2$Type <- as.factor(c(rep('UL_bead',20), rep('nonUL_bead',18),
  rep('UL_array',3), rep('nonUL_array',3)))
beadArrays_ML3 <- beadArrays_ML2[,c(13,1:12)]

UL_seq_ML <- keys_m12
UL_gene_ML <- beadArrays_ML3
```

There are two datasets to use for machine learning. The first is our beadchip samples of 88 sequences and 38 samples of 20 UL and 18 nonUL in the **UL_seq_ML** data set. The second dataset for machine learning is the mixed microarray and beadchip samples of UL and nonUL by gene in the **UL_gene_ML** data set, because there were no common sequence or copy number variants of the gene sequences between the beadchip and microarray sets of UL and nonUL samples.

The libraries were installed earlier.

```
set.seed(02242020)
```

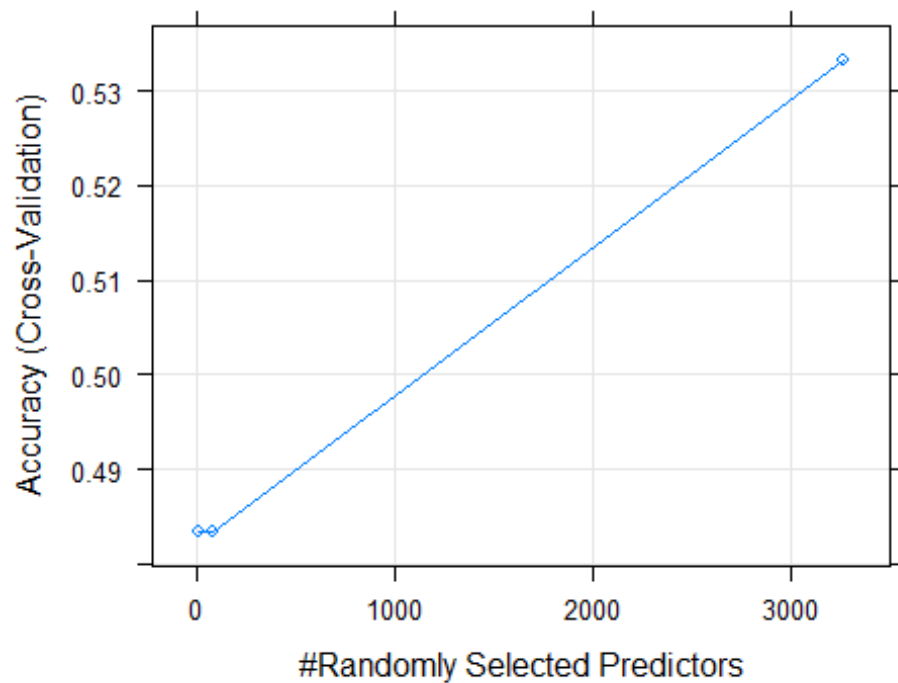
Create a partition of the data with a 70/30 split into training/testing sets of the first data set with two classes of UL or nonUL and 88 features of genes with their CNVs.

```
inTrain <- createDataPartition(y=UL_seq_ML$Type, p=0.7, list=FALSE)

trainingSet <- UL_seq_ML[inTrain,]
testingSet <- UL_seq_ML[-inTrain,]
```

RandomForest, cross-validation (cv) = 5

```
rfMod <- train(Type~., method='rf', data=(trainingSet),  
               trControl=trainControl(method='cv'), number=5)  
plot(rfMod)
```



Run predictions on the testing set

```
predRF <- predict(rfMod, testingSet)  
  
predDF <- data.frame(predRF, type=testingSet$Type)  
predDF  
  
##      predRF  type  
## 1      UL    UL  
## 2      UL    UL  
## 3      UL    UL  
## 4      UL    UL  
## 5      UL    UL  
## 6      UL    UL  
## 7      UL nonUL  
## 8      UL nonUL  
## 9      UL nonUL  
## 10     UL nonUL  
## 11     UL nonUL  
  
sum <- sum(predRF==testingSet$Type)  
length <- length(testingSet$Type)
```



```

accuracy_rfMod <- (sum/length)
accuracy_rfMod

## [1] 0.5454545

results <- c(round(accuracy_rfMod,2), round(100,2))
results <- as.factor(results)
results <- t(data.frame(results))

colnames(results) <- colnames(predDF)
Results <- rbind(predDF, results)
Results

##          predRF  type
## 1             UL    UL
## 2             UL    UL
## 3             UL    UL
## 4             UL    UL
## 5             UL    UL
## 6             UL    UL
## 7             UL nonUL
## 8             UL nonUL
## 9             UL nonUL
## 10            UL nonUL
## 11            UL nonUL
## results    0.55   100

```

The above shows that using the genes and the CNV of each gene totalling 88 features, makes a poor data set of results with only 27 observations to train and 11 to test on 2 classes of UL or non-UL.

What if we used random forest to only predict by gene in the first beadchip type data set? We can use the transpose of the keys2 data set made earlier when combining to make the 2nd ML dataset.

```

keys_t <- as.data.frame(t(keys2))
colnames(keys_t) <- keys2$Symbol
keys_t1 <- keys_t[-1,]
keys_t1$Type <- keys_m12$Type
keys_ML <- keys_t1[,c(21,1:20)]

```

Now we will use our new data set based on the beadchip genes and not the CNVs of those genes, in the keys_ML data set to predict with RandomForest.

```

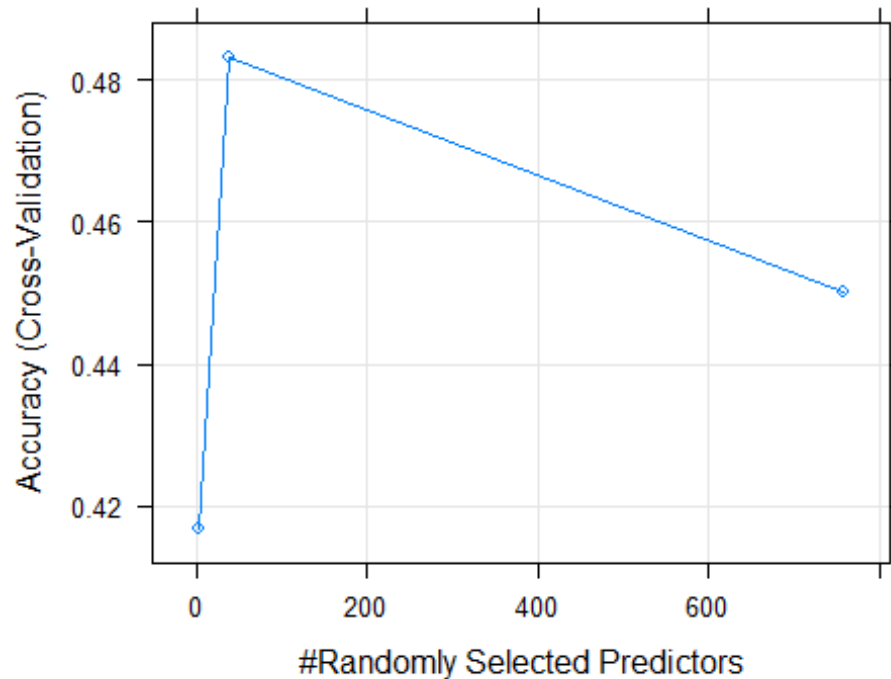
inTrain <- createDataPartition(y=keys_ML$Type, p=0.7, list=FALSE)

trainingSet <- keys_ML[inTrain,]
testingSet <- keys_ML[-inTrain,]

```

RandomForest, cross-validation (cv) = 5

```
rfMod <- train(Type~., method='rf', data=(trainingSet),
               trControl=trainControl(method='cv'), number=5)
plot(rfMod)
```



Run predictions on the testing set

```
predRF <- predict(rfMod, testingSet)

predDF <- data.frame(predRF, type=testingSet$Type)
predDF
```

##	predRF	type
## 1	UL	UL
## 2	UL	UL
## 3	UL	UL
## 4	UL	UL
## 5	UL	UL
## 6	UL	UL
## 7	UL	nonUL
## 8	UL	nonUL
## 9	UL	nonUL
## 10	UL	nonUL
## 11	UL	nonUL

```
sum <- sum(predRF==testingSet$Type)
length <- length(testingSet$Type)
```

```

accuracy_rfMod <- (sum/length)
accuracy_rfMod

## [1] 0.5454545

results <- c(round(accuracy_rfMod,2), round(100,2))
results <- as.factor(results)
results <- t(data.frame(results))

```

```

colnames(results) <- colnames(predDF)
Results <- rbind(predDF, results)
Results

```

```

##          predRF  type
## 1             UL    UL
## 2             UL    UL
## 3             UL    UL
## 4             UL    UL
## 5             UL    UL
## 6             UL    UL
## 7             UL nonUL
## 8             UL nonUL
## 9             UL nonUL
## 10            UL nonUL
## 11            UL nonUL
## results    0.55   100

```

How about with the KNN algorithm.

```

knnMod <- train(Type ~ .,
                 method='knn', preProcess=c('center','scale'),
                 tuneLength=10, trControl=trainControl(method='cv'),
                 data=trainingSet)

## Warning in preProcess.default(thresh = 0.95, k = 5, freqCut = 19,
## uniqueCut =
## 10, : These variables have zero variances: ABCA8 88.61720, ABCA8 77.21540,
## ABCA8
## 96.40000, ABCA8 110.85790, ABCA8 111.68890, ABCA8 125.96360, ABCA8
## 158.93000,
## ABCA8 206.22750, ABCA8 576.07270, ABCA8 586.46630, ABCA8 703.32080, ABCA8
## 761.93730, ABCA81083.18600, ABCA8ABCA8, ACTC 204.95590, ACTC 2077.75710,
## ACTC
## 216.30000, ACTC 277.35920, ACTC 320.94930, ACTC 350.72790, ACTC 830.23350,
## ACTC1205.77090, ACTC4004.56310, ACTC5434.41450, ACTC6205.93360,
## ACTC6278.26070,
## ACTC7325.32550, ACTC8327.85490, ACTCACTC, ADH1A 153.07270, ADH1A
## 260.63550,
## ADH1A 275.28830, ADH1A 517.43540, ADH1A 659.09590, ADH1A 660.45380, ADH1A
## 873.77740, ADH1A1157.79510, ADH1A1936.02330, ADH1A1997.93640,
## ADH1A1998.15530,

```

ADH1A3647.80130, ADH1A3728.12040, ADH1AADH1A, BDNF 73.08220, BDNF 58.65697,
BDNF 59.53214, BDNF 60.29837, BDNF 60.99053, BDNF 61.44807, BDNF 62.29957,
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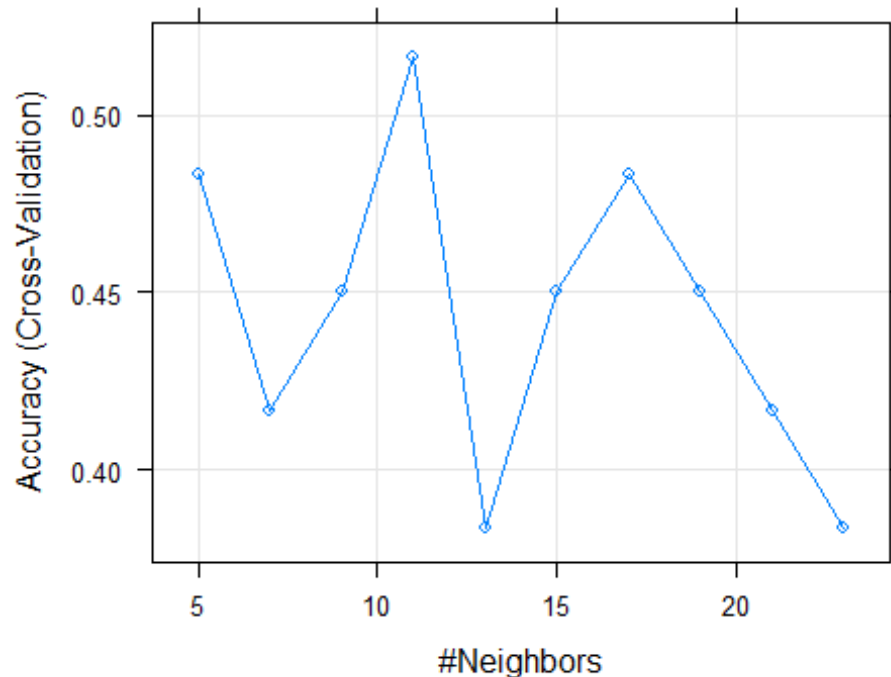
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168.43680, KRT19 229.34340, KRT19 232.75320, KRT19 656.00690, KRT19
773.91250,
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LOC202134
80.73967, LOC202134 80.93756, LOC202134 86.33883, LOC202134 91.90153,
LOC202134
99.14780, LOC202134 99.72284, LOC202134 100.08753, LOC202134 100.62161,
LOC202134 125.11937, LOC202134 153.49023, LOC202134LOC202134, LOC23117
58.45052, LOC23117 56.78304, LOC23117 57.00729, LOC23117 57.58250,
LOC23117

```
## 57.63400, LOC23117 57.87063, LOC23117 57.87225, LOC23117 58.25216,  
LOC23117  
## 59.70365, LOC23117 61.10820, LOC23117 64.34166, LOC23117LOC23117,  
LOC339047  
## 52.66400, LOC339047 53.39899, LOC339047 54.63439, LOC339047 54.96286,  
LOC339047  
## 55.83333, LOC339047 56.06596, LOC339047 56.25000, LOC339047 56.88054,  
LOC339047  
## 57.71951, LOC339047 58.38786, LOC339047LOC339047, LOC653086 909.83774,  
LOC653086  
## 390.88886, LOC653086 648.07329, LOC653086 660.33487, LOC653086 772.56214,  
## LOC653086 825.04971, LOC653086 852.31269, LOC653086 870.00770, LOC653086  
## 897.15661, LOC6530861065.10449, LOC6530861122.18651, LOC653086LOC653086,  
## MMP11 75.97500, MMP11 84.25000, MMP11 318.05450, MMP11 375.42970, MMP11  
## 443.38950, MMP111338.71330, MMP1115286.34750, MMP113590.97470,  
MMP113713.31130,  
## MMP115414.01170, MMP118858.71940, MMP11MMP11, PENK 54.20000, PENK  
59.48000,  
## PENK 60.30000, PENK 61.60000, PENK 99.13530, PENK 166.37800, PENK  
183.02000,  
## PENK 214.79190, PENK 628.35690, PENK 699.56800, PENK26134.71050, PENKPENK,  
## PLEC1 63.21976, PLEC1 68.78959, PLEC1 69.57295, PLEC1 71.50187, PLEC1  
76.19250,  
## PLEC1 78.75296, PLEC1 79.68368, PLEC1 85.84841, PLEC1 94.55256, PLEC1  
116.72813,  
## PLEC1PLEC1  
  
plot(knnMod)
```



The accuracy seems to be better between 8 and 9 neighbors for classification from what the above plot is displaying.

```
rpartMod <- train(Type ~ ., method='rpart', tuneLength=7, data=trainingSet)
glmMod <- train(Type ~ .,
                 method='glm', data=trainingSet)

predKNN <- predict(knnMod, testingSet)
predRPART <- predict(rpartMod, testingSet)
predGLM <- predict(glmMod, testingSet)

length=length(testingSet$Type)

sumKNN <- sum(predKNN==testingSet$Type)
sumRPart <- sum(predRPART==testingSet$Type)
sumGLM <- sum(predGLM==testingSet$Type)

accuracy_KNN <- sumKNN/length
accuracy_RPART <- sumRPart/length
accuracy_GLM <- sumGLM/length

predDF2 <- data.frame(predRF, predKNN, predRPART, predGLM,
                      TYPE=testingSet$Type)
colnames(predDF2) <- c('RandomForest', 'KNN', 'Rpart', 'GLM', 'TrueValue')

results <- c(round(accuracy_rfMod, 2),
```

```

round(accuracy_KNN,2),
round(accuracy_RPART,2),
round(accuracy_GLM,2),
round(100,2))

results <- as.factor(results)
results <- t(data.frame(results))
colnames(results) <- c('RandomForest', 'KNN', 'Rpart', 'GLM', 'TrueValue')
Results <- rbind(predDF2, results)
Results

##           RandomForest  KNN Rpart  GLM TrueValue
## 1                UL    UL    UL    UL         UL
## 2                UL    UL    UL    UL         UL
## 3                UL    UL    UL    UL         UL
## 4                UL    UL    UL    UL         UL
## 5                UL    UL    UL    UL         UL
## 6                UL    UL    UL    UL         UL
## 7                UL    UL    UL    UL        nonUL
## 8                UL    UL    UL    UL        nonUL
## 9                UL    UL    UL    UL        nonUL
## 10               UL    UL    UL    UL        nonUL
## 11               UL    UL    UL    UL        nonUL
## results          0.55 0.55  0.55 0.55         100

```

As far as the algorithms used above go, the prediction accuracy is not good at all. This could be because there aren't enough samples. This could also be because there are a lot of features and not enough samples. We could try just predicting the sample type using less features, like the genes with the highest fold change values. Those genes were KIAA1199, ACTC, MMP11, FOSB, KRT19, and FOSB with the three most over/under expressed in fold change mean values.

```

keys_ML_b <- keys_ML[,c(1,3,10,11,13,19)]

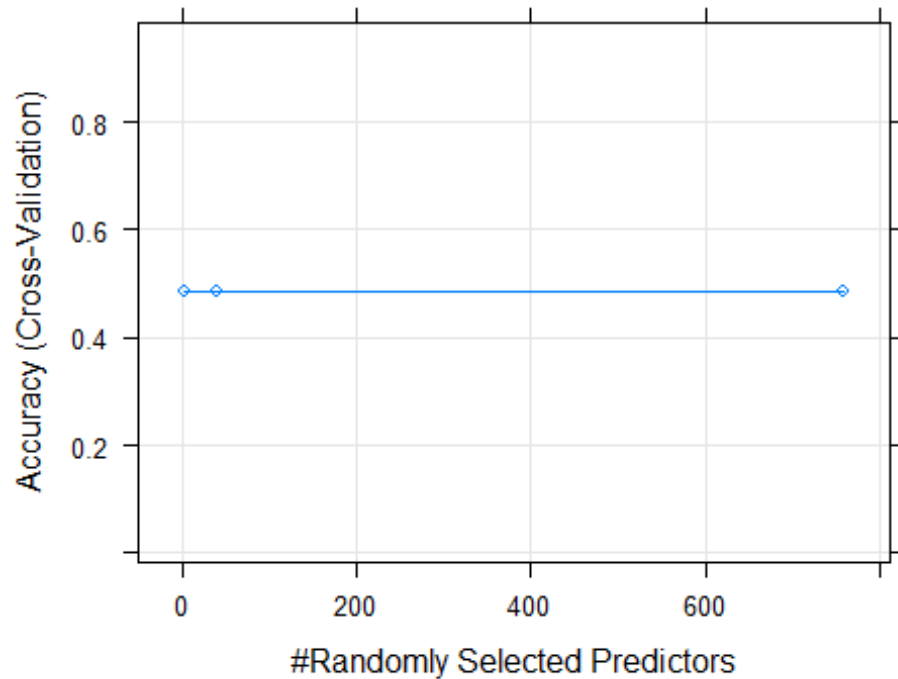
inTrain <- createDataPartition(y=keys_ML_b$Type, p=0.7, list=FALSE)

trainingSet <- keys_ML[inTrain,]
testingSet <- keys_ML[-inTrain,]

RandomForest, cross-validation (cv) = 5

rfMod <- train(Type~., method='rf', data=(trainingSet),
               trControl=trainControl(method='cv'), number=5)
plot(rfMod)

```



Run predictions on the testing set

```
predRF <- predict(rfMod, testingSet)

predDF <- data.frame(predRF, type=testingSet$Type)
predDF

##      predRF  type
## 1      UL    UL
## 2      UL    UL
## 3      UL    UL
## 4      UL    UL
## 5      UL    UL
## 6      UL    UL
## 7      UL nonUL
## 8      UL nonUL
## 9      UL nonUL
## 10     UL nonUL
## 11     UL nonUL

sum <- sum(predRF==testingSet$Type)
length <- length(testingSet$Type)
accuracy_rfMod <- (sum/length)
accuracy_rfMod

## [1] 0.5454545
```

```

results <- c(round(accuracy_rfMod,2), round(100,2))
results <- as.factor(results)
results <- t(data.frame(results))

```

```

colnames(results) <- colnames(predDF)
Results <- rbind(predDF, results)
Results

```

```

##      predRF  type
## 1      UL    UL
## 2      UL    UL
## 3      UL    UL
## 4      UL    UL
## 5      UL    UL
## 6      UL    UL
## 7      UL nonUL
## 8      UL nonUL
## 9      UL nonUL
## 10     UL nonUL
## 11     UL nonUL
## results 0.55  100

```

How about with the KNN algorithm.

```

knnMod <- train(Type ~ .,
                 method='knn', preProcess=c('center','scale'),
                 tuneLength=10, trControl=trainControl(method='cv'),
                 data=trainingSet)

## Warning in preProcess.default(thresh = 0.95, k = 5, freqCut = 19,
## uniqueCut
## = 10, : These variables have zero variances: ABCA8 155.80000, ABCA8
## 56.05000,
## ABCA8 681.04640, ABCA8 92.13060, ABCA8 96.40000, ABCA8 125.96360, ABCA8
## 132.38820, ABCA8 292.93330, ABCA8 455.64560, ABCA8 562.68870, ABCA8
## 576.07270,
## ABCA8 830.87020, ABCA81493.23800, ABCA8ABCA8, ACTC 478.79120, ACTC
## 1452.70000,
## ACTC 169.41300, ACTC 222.50000, ACTC 320.94930, ACTC 350.72790, ACTC
## 622.15940,
## ACTC 710.61260, ACTC13992.26470, ACTC2718.71890, ACTC4004.56310,
## ACTC5419.39300,
## ACTC6894.49490, ACTC7028.38050, ACTC8327.85490, ACTCACTC, ADH1A 171.55770,
## ADH1A 54.35000, ADH1A 631.70000, ADH1A 1797.60150, ADH1A 260.63550, ADH1A
## 275.28830, ADH1A 450.46120, ADH1A 640.42100, ADH1A 873.77740,
## ADH1A1104.32570,
## ADH1A1503.13120, ADH1A1944.23980, ADH1A1998.15530, ADH1A2893.83880,
## ADH1A7727.82380, ADH1AADH1A, BDNF 66.79751, BDNF 68.67747, BDNF 59.53214,
## BDNF 61.34184, BDNF 62.38321, BDNF 62.61744, BDNF 62.63791, BDNF 63.00849,
## BDNF 63.68421, BDNF 64.23757, BDNF 64.79746, BDNF 64.93227, BDNF 66.47571,
## BDNF 82.74799, BDNFBDNF, CTNNB1 430.51867, CTNNB1 80.07464, CTNNB1

```

1367.28571,
CTNNB1 427.31916, CTNNB1 441.03484, CTNNB1 506.44616, CTNNB1 599.45751,
CTNNB1 626.21914, CTNNB1 629.60446, CTNNB1 656.16527, CTNNB1 661.17213,
CTNNB1
701.44551, CTNNB1 736.31490, CTNNB1 762.08883, CTNNB1CTNNB1, DDX12
55.51556,
DDX12 56.68058, DDX12 53.20944, DDX12 53.62324, DDX12 54.58100, DDX12
55.09119,
DDX12 55.16945, DDX12 55.82984, DDX12 57.06467, DDX12 57.38962, DDX12
58.59496,
DDX12 59.08089, DDX12 59.42375, DDX12 66.08112, DDX12DDX12, DLK1
121.15160,
DLK1 226.30000, DLK1 47.30000, DLK1 49.90000, DLK1 51.00000, DLK1
51.72500,
DLK1 55.00000, DLK1 58.50000, DLK1 60.50000, DLK1 74.35000, DLK1
150.24510,
DLK1 384.38110, DLK1 475.95320, DLK14126.77500, DLK1DLK1, DMD 113.91429,
DMD 132.51420, DMD 173.72703, DMD 60.21667, DMD 87.96567, DMD 100.27049,
DMD
103.15579, DMD 119.81640, DMD 131.68924, DMD 136.18699, DMD 152.12489, DMD
157.94423, DMD 181.68309, DMD 213.71983, DMD 223.08834, DMDDMD, FOS
621.78120,
FOS 874.10000, FOS 220.68100, FOS 386.81900, FOS 417.75610, FOS 577.79010,
FOS 612.89650, FOS1298.03740, FOS17362.03170, FOS2406.07060,
FOS3514.50890,
FOS3868.98980, FOS6603.47780, FOS8590.64340, FOS9819.01050, FOSFOS, FOSB
95.39480, FOSB 121.20000, FOSB 57.90000, FOSB 60.92000, FOSB 61.65000,
FOSB
64.62220, FOSB 110.60000, FOSB 125.27830, FOSB 207.68810, FOSB 243.48700,
FOSB
4247.93770, FOSB 445.09370, FOSB 601.59350, FOSB 680.53840,
FOSB7604.02210,
FOSBFOSB, KIAA0692 62.88800, KIAA0692 67.28364, KIAA0692 58.06852,
KIAA0692
60.28206, KIAA0692 62.29838, KIAA0692 63.64361, KIAA0692 64.27382,
KIAA0692
64.75153, KIAA0692 64.89243, KIAA0692 65.02993, KIAA0692 66.84581,
KIAA0692
67.47397, KIAA0692 69.12237, KIAA0692 88.99077, KIAA0692KIAA0692, KIAA1199
65.80000, KIAA1199 861.20000, KIAA1199 93.08750, KIAA1199 200.75370,
KIAA1199
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KIAA11991798.94770, KIAA11994272.82880, KIAA11994624.54100,
KIAA11995074.68360,
KIAA11996160.16650, KIAA11996548.81600, KIAA1199KIAA1199, KRT19 225.70000,
KRT19
45.60000, KRT19 53.70000, KRT19 57.40000, KRT19 577.13950, KRT19 58.18000,
KRT19
173.69500, KRT19 180.82220, KRT19 232.75320, KRT19 293.32220, KRT19
773.91250,

```

## KRT19 992.90020, KRT191098.19990, KRT192049.65290, KRT19KRT19, LOC202134
## 90.27716, LOC202134 116.28997, LOC202134 80.93756, LOC202134 81.95797,
LOC202134
## 95.92870, LOC202134 96.08817, LOC202134 97.86973, LOC202134 98.63219,
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LOC23117
## 56.92304, LOC23117 57.00729, LOC23117 57.14000, LOC23117 57.58250,
LOC23117
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LOC23117
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LOC339047
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## LOC6530861066.72823, LOC6530861203.33167, LOC6530861519.48689,
## LOC653086LOC653086, MMP11 75.97500, MMP11 136.19640, MMP11 199.35870,
MMP11
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MMP11113478.50000,
## MMP1111683.55920, MMP1111875.99000, MMP112082.50560, MMP113135.01660,
## MMP114141.86420, MMP118858.71940, MMP11MMP11, PENK 158.16150, PENK
248.20700,
## PENK 442.00000, PENK 48.20000, PENK 54.20000, PENK 55.70000, PENK
60.10000,
## PENK 60.38330, PENK 61.60000, PENK 89.07890, PENK 133.11190, PENK
255.79600,
## PENK 313.38190, PENK 699.56800, PENK2788.93790, PENKPENK, PLEC1 98.52500,
PLEC1
## 117.83730, PLEC1 187.59291, PLEC1 53.04029, PLEC1 66.99375, PLEC1
68.78959,
## PLEC1 70.30182, PLEC1 71.20156, PLEC1 74.33759, PLEC1 76.05800, PLEC1
79.68368,
## PLEC1 94.55256, PLEC1 94.90675, PLEC1 104.29716, PLEC1 137.53784,
PLEC1PLEC1

## Warning in preProcess.default(thresh = 0.95, k = 5, freqCut = 19,
uniqueCut
## = 10, : These variables have zero variances: ABCA8 155.80000, ABCA8

```


56.05000,
ABCA8 681.04640, ABCA8 92.13060, ABCA8 96.40000, ABCA8 125.96360, ABCA8
132.38820, ABCA8 292.93330, ABCA8 455.64560, ABCA8 562.68870, ABCA8
576.07270,
ABCA8 830.87020, ABCA81493.23800, ABCA8ABCA8, ACTC 478.79120, ACTC
1452.70000,
ACTC 169.41300, ACTC 222.50000, ACTC 320.94930, ACTC 350.72790, ACTC
622.15940,
ACTC 710.61260, ACTC13992.26470, ACTC2718.71890, ACTC4004.56310,
ACTC5419.39300,
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ADH1A 54.35000, ADH1A 631.70000, ADH1A 1797.60150, ADH1A 260.63550, ADH1A
275.28830, ADH1A 450.46120, ADH1A 640.42100, ADH1A 873.77740,
ADH1A1104.32570,
ADH1A1503.13120, ADH1A1944.23980, ADH1A1998.15530, ADH1A2893.83880,
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CTNNB1 626.21914, CTNNB1 629.60446, CTNNB1 656.16527, CTNNB1 661.17213,
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DMD 132.51420, DMD 173.72703, DMD 60.21667, DMD 87.96567, DMD 100.27049,
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FOSB
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54.22306, LOC339047 54.32881, LOC339047 54.90714, LOC339047 54.96286,
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LOC653086 764.01074, LOC653086 798.87956, LOC653086 870.00770, LOC653086
897.15661, LOC653086 922.11347, LOC653086 925.40631, LOC6530861014.19773,
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LOC653086LOC653086, MMP11 75.97500, MMP11 136.19640, MMP11 199.35870,
MMP11
443.38950, MMP11 5787.43480, MMP11 627.16670, MMP11 669.80360,
MMP1113478.50000,

```

## MMP111683.55920, MMP111875.99000, MMP112082.50560, MMP113135.01660,
## MMP114141.86420, MMP118858.71940, MMP11MMP11, PENK 158.16150, PENK
248.20700,
## PENK 442.00000, PENK 48.20000, PENK 54.20000, PENK 55.70000, PENK
60.10000,
## PENK 60.38330, PENK 61.60000, PENK 89.07890, PENK 133.11190, PENK
255.79600,
## PENK 313.38190, PENK 699.56800, PENK2788.93790, PENKPENK, PLEC1 98.52500,
PLEC1
## 117.83730, PLEC1 187.59291, PLEC1 53.04029, PLEC1 66.99375, PLEC1
68.78959,
## PLEC1 70.30182, PLEC1 71.20156, PLEC1 74.33759, PLEC1 76.05800, PLEC1
79.68368,
## PLEC1 94.55256, PLEC1 94.90675, PLEC1 104.29716, PLEC1 137.53784,
PLEC1PLEC1

## Warning in preProcess.default(thresh = 0.95, k = 5, freqCut = 19,
uniqueCut
## = 10, : These variables have zero variances: ABCA8 155.80000, ABCA8
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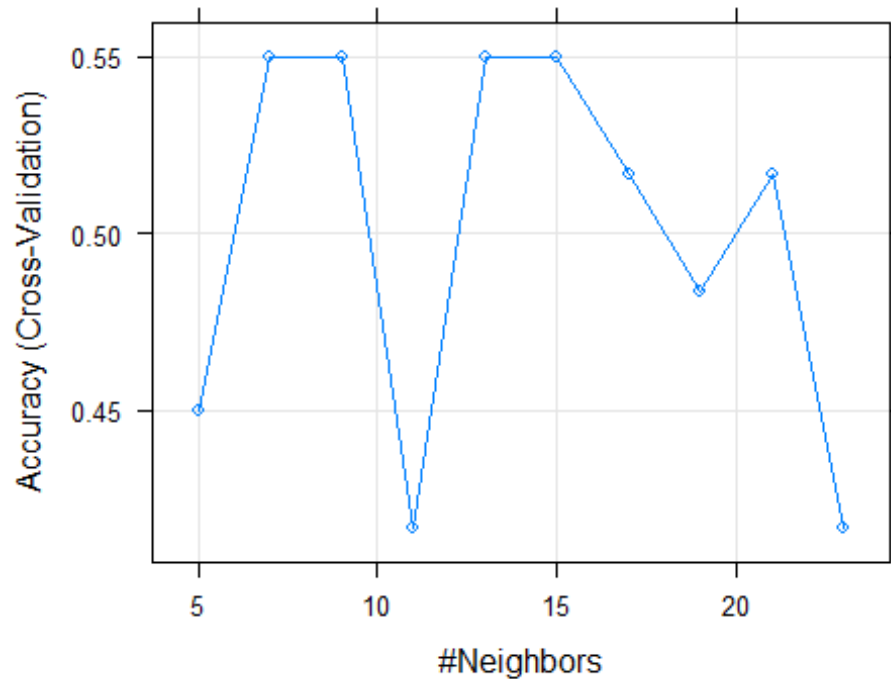
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207.68810, FOSB 4247.93770, FOSB 601.59350, FOSB 680.53840,
FOSB7604.02210,
FOSBFOSB, KIAA0692 62.88800, KIAA0692 67.28364, KIAA0692 58.06852,
KIAA0692 60.28206, KIAA0692 64.27382, KIAA0692 64.75153, KIAA0692
64.89243,
KIAA0692 65.02993, KIAA0692 66.84581, KIAA0692 67.47397, KIAA0692
69.12237,
KIAA0692KIAA0692, KIAA1199 65.80000, KIAA1199 93.08750, KIAA1199
200.75370,
KIAA1199 212.18080, KIAA1199 5039.24990, KIAA11991798.94770,
KIAA11994272.82880,
KIAA11994624.54100, KIAA11996160.16650, KIAA11996548.81600,
KIAA1199KIAA1199,
KRT19 45.60000, KRT19 57.40000, KRT19 577.13950, KRT19 58.18000, KRT19
173.69500, KRT19 180.82220, KRT19 232.75320, KRT19 293.32220, KRT19
773.91250,
KRT19 992.90020, KRT19KRT19, LOC202134 90.27716, LOC202134 116.28997,
LOC202134
80.93756, LOC202134 81.95797, LOC202134 95.92870, LOC202134 97.86973,
LOC202134
99.14780, LOC202134 103.06420, LOC202134 103.91630, LOC202134 109.43154,

```
## LOC202134 153.49023, LOC202134LOC202134, LOC23117 58.95793, LOC23117
63.41986,
## LOC23117 56.92304, LOC23117 57.00729, LOC23117 57.14000, LOC23117
57.58250,
## LOC23117 58.72856, LOC23117 59.18544, LOC23117 59.26586, LOC23117
59.70365,
## LOC23117 60.51843, LOC23117LOC23117, LOC339047 54.32226, LOC339047
55.18711,
## LOC339047 53.41786, LOC339047 54.22306, LOC339047 54.90714, LOC339047
54.96286,
## LOC339047 54.99321, LOC339047 55.83333, LOC339047 56.07547, LOC339047
57.69556,
## LOC339047 57.71951, LOC339047LOC339047, LOC653086 754.31924, LOC653086
## 841.71641, LOC653086 390.88886, LOC653086 735.69129, LOC653086 764.01074,
## LOC653086 870.00770, LOC653086 897.15661, LOC653086 922.11347, LOC653086
## 925.40631, LOC6530861066.72823, LOC6530861203.33167, LOC653086LOC653086,
## MMP11 75.97500, MMP11 136.19640, MMP11 199.35870, MMP11 443.38950, MMP11
## 5787.43480, MMP11 669.80360, MMP111875.99000, MMP112082.50560,
MMP114141.86420,
## MMP118858.71940, MMP11MMP11, PENK 158.16150, PENK 248.20700, PENK
48.20000,
## PENK 54.20000, PENK 55.70000, PENK 60.38330, PENK 61.60000, PENK 89.07890,
PENK
## 255.79600, PENK 699.56800, PENK2788.93790, PENKPENK, PLEC1 117.83730,
PLEC1
## 187.59291, PLEC1 66.99375, PLEC1 68.78959, PLEC1 70.30182, PLEC1 76.05800,
PLEC1
## 79.68368, PLEC1 94.55256, PLEC1 94.90675, PLEC1 104.29716, PLEC1
137.53784,
## PLEC1PLEC1
```

```
plot(knnMod)
```



The accuracy seems to be better between 8 and 9 neighbors for classification from what the above plot is displaying.

```
rpartMod <- train(Type ~ ., method='rpart', tuneLength=7, data=trainingSet)
glmMod <- train(Type ~ .,
                 method='glm', data=trainingSet)

predKNN <- predict(knnMod, testingSet)
predRPART <- predict(rpartMod, testingSet)
predGLM <- predict(glmMod, testingSet)

length=length(testingSet$Type)

sumKNN <- sum(predKNN==testingSet$Type)
sumRPart <- sum(predRPART==testingSet$Type)
sumGLM <- sum(predGLM==testingSet$Type)

accuracy_KNN <- sumKNN/length
accuracy_RPART <- sumRPart/length
accuracy_GLM <- sumGLM/length

predDF2 <- data.frame(predRF, predKNN, predRPART, predGLM,
                      TYPE=testingSet$Type)
colnames(predDF2) <- c('RandomForest', 'KNN', 'Rpart', 'GLM', 'TrueValue')

results <- c(round(accuracy_rfMod, 2),
```

```

round(accuracy_KNN,2),
round(accuracy_RPART,2),
round(accuracy_GLM,2),
round(100,2))

results <- as.factor(results)
results <- t(data.frame(results))
colnames(results) <- c('RandomForest', 'KNN', 'Rpart', 'GLM', 'TrueValue')
Results <- rbind(predDF2, results)
Results

##           RandomForest  KNN Rpart   GLM TrueValue
## 1                UL    UL    UL nonUL         UL
## 2                UL    UL    UL nonUL         UL
## 3                UL    UL    UL nonUL         UL
## 4                UL    UL    UL nonUL         UL
## 5                UL    UL    UL nonUL         UL
## 6                UL    UL    UL   UL         UL
## 7                UL    UL    UL nonUL       nonUL
## 8                UL    UL    UL nonUL       nonUL
## 9                UL    UL    UL nonUL       nonUL
## 10               UL    UL    UL nonUL       nonUL
## 11               UL    UL    UL nonUL       nonUL
## results          0.55 0.55  0.55  0.55         100

```

I find it difficult to see that there are no changes in values, there are only two classes to predict on 38 samples with 6-20 features, yet every data set and algorithm is producing the same result of 55%. I will see if my other scripts with these algorithms are producing the same results and see if there is a bug in the package libraries or it could be that the mean values in this set were taken from sequence expression values. But I have never seen this type of result using R ever.