

Genome assembly

Genome assembly

Genome assembly is the process of converting short reads into a detailed set of sequences corresponding to the chromosome(s) of an organism.

To learn more about assembly visit
<http://www.ncbi.nlm.nih.gov/assembly/>
<http://www.ncbi.nlm.nih.gov/assembly/basics/>

Assembly

Genome assembly organization and additional information.

Using Assembly

[Assembly Help](#)
[Browse by Organism](#)
[NCBI Assembly Data Model](#)
[Assembly Basics](#)
[Genomes Download FAQ](#)
[Genomes FTP Site](#)

Submitting an Assembly

[Submission Information](#)
[Submission FAQ](#)
[AGP Specifications](#)
[AGP Validation](#)

Related Resources

[Genome](#)
[Genome Reference Consortium](#)
[Genome Remapping Service \(Remap\)](#)

NCBI Datasets



NCBI Datasets: Easy Access and Download Sequence Data and Metadata
 Effective June 2014, NCBI Datasets will replace legacy Genome and Assembly web resources.

As part of our ongoing efforts to streamline our resources and modernize our website, NCBI will gradually replace the legacy Genome and Assembly resources with the new, improved NCBI Datasets resource. NCBI Datasets is a centralized, easy-to-use platform designed to simplify access to NCBI's vast genomic data.

NCBI Datasets

A one-stop shop for finding, browsing, and downloading genomic data.

Find data by organism, assembly, or project.

Download data in a variety of formats.

View data in a variety of ways.

Filter data by a variety of criteria.

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How to use NCBI Datasets

The best way to start is to use the search bar below. But here's an overview of the steps to resources and data we offer:

Genome assembly: relevance

- Genome assembly is needed when a genome is first sequenced. We can relate reads to chromosomes.
- For the human genome, the assembly is "frozen" as a snapshot every few years. The current assembly is GRCh38. (GRC refers to Genome Reference Consortium at <http://www.ncbi.nlm.nih.gov/projects/genome/assembly/gvc/>)
- For most human genome work we do not need to do "de novo" (from anew) assembly. Instead we map reads to a reference genome—one that is already assembled.
- Genome assembly is a crucial behind-the-scenes part of calling human genome (or other) variants.

Whereas early genome assembly projects were often aided by clone maps or other mapping data, many current assembly projects forego these scaffolding data and only assemble genomes into smaller segments. Recently, new technologies have been invented that allow chromosome-scale assembly at a lower cost and faster speed than traditional methods.

Many new technologies can now be used to create chromosome-scale assemblies without costly and time-consuming methods such as BAC-end sequencing and physical mapping.

Rice and Green, 2019. New Approaches for Genome Assembly and Scaffolding

Consequently, the contiguity of new genome assemblies decreased as high-throughput sequencing was widely adopted (Figure 1b,c)

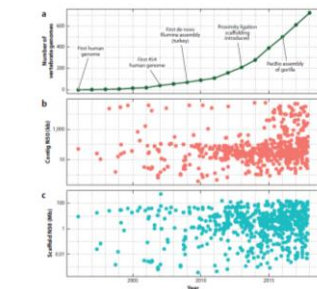


Figure 1
Trends and statistics of genome assemblies deposited to the National Center for Biotechnology Information's GenBank. Although second-generation sequencing has allowed more genomes to be published each year by making sequencing faster and cheaper, it has not increased the contiguity of published genomes. (a) Number of genome assemblies available on GenBank at the end of each year, showing accelerating growth over the past decade. (b) Contig and (c) scaffold N50s of all genome assemblies deposited to GenBank per year.

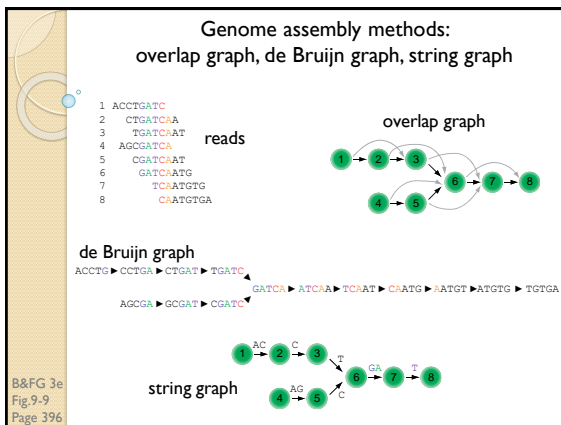
Genome Contig Assembly

No technology currently exists that can read DNA from one end to the other of even moderately sized chromosomes, which are typically tens or hundreds of millions of base pairs long. All current approaches for genome assembly read many segments that are considerably shorter than chromosomes.

Both long-read sequencing technologies implement single-molecule sequencing methods and generate reads with a distribution of lengths that, for assembly purposes, target a range of tens to hundreds of kilobases (kb)– typically 10–25 kb for PacBio HiFi reads (also circular consensus sequencing, CCS), 10–40 kb for PacBio continuous long reads (CLR) and 10 kb–2 Megabases (Mb) for ONT, where the upper limit is constrained principally by properties of the input material (Payne et al., 2018).

Assembly algorithms

- overlap-layout-consensus: input DNA sequence reads are compared, all versus all, in the overlap step. The overlap-layout-consensus algorithm is based on identifying overlapping regions between reads and using these overlaps to construct longer contiguous sequences (contigs).
- de Bruijn graph: short words (k-mers) that are observed in the reads are the nodes of the graph, and edges are added when these k-mers are adjacent in sequence reads. In this process, each read is used to populate the graph but not compared directly to all the other reads.
- hybrid assembly



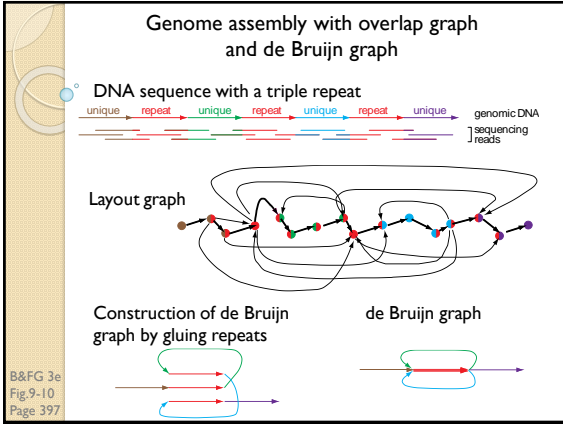


Table 1 Commonly used assembly software

Software	URL and reference	Description
Short-read assembly software		
Velvet	https://github.com/derebas/velvet (166)	Original de Bruijn graph assembler
SOAPdenovo	http://soap.genomics.org.cn/ (169)	De Bruijn graph assembler with error-correction step
Mercator	https://git.doc.gov/data-and-tools/mercator/ (170)	Hybrid k-mer/overlap-based
ALLPATHS-LG	http://software.broadinstitute.org/paths-lp/blog/ (171)	Uses overlap graph to collapse repeats
BGA	https://github.com/jsga (172)	Uses overlap graphs
AdSS	https://github.com/ncwgs/adss (173)	Represents de Bruijn graph with a Bloom filter
DISCOVAR de novo	https://software.broadinstitute.org/software/discovar/ (174)	Requires 250-bp PCR-free reads
SuperNova	https://github.com/10XGenomics/supernova (149)	Assembles 10x linked reads
Long-read assembly software		
TRAP	https://github.com/PacificBiosciences/Bioinformatics-Training/wiki/TRAP (124)	Error correction, overlap-layout-consensus assembly, and polishing workflow
Canu	https://github.com/marbl/canu (125)	K-mer-based overlap computation
EL CON	https://github.com/PacificBiosciences/TALCON (103)	Assembles phased diploid genomes
Flye	https://github.com/raiserghis/flye (126)	Uses a de Bruijn graph
Minimus	https://github.com/ikhram/minimus (128)	Fast, but no error correction
Polishing software		
Pilon	https://github.com/broadinstitute/pilon (133)	Uses short-read alignments to correct errors
Arrow	https://github.com/PacificBiosciences/GenomicConsensus	Hidden Markov model and long-read alignment
Nanopolish	https://github.com/raiserghis/nanopolish (115)	Nanopore only; uses original voltage data to correct errors

Spades??

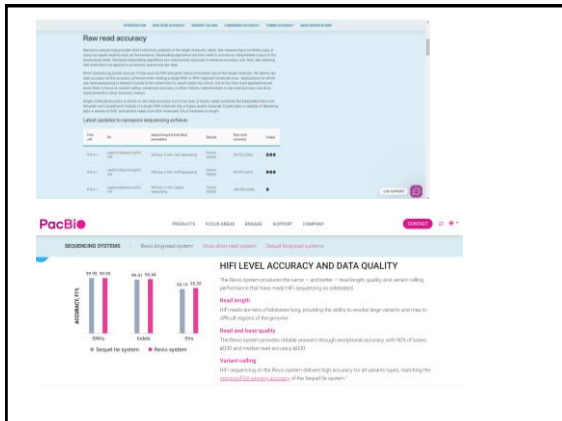
CREATING MORE CONTIGUOUS ASSEMBLIES WITH LONG READS

- Pacific Biosciences (SMRT, 2009)
 - The incorporation of fluorescently labeled nucleotides is detected and reveals the sequence of the analysed DNA strand.
 - PacBio offers Continuous Long Reads (CLR) and Circular Consensus Sequencing (CCS) reads also called High-Fidelity (HiFi).
- Oxford Nanopore Sequencing (2005, 1 channel flow cell, etc.)
 - It works by monitoring changes to an electrical current as nucleic acids are passed through a protein nanopore. The resulting signal is decoded to provide the specific DNA or RNA sequence.

Hybrid assembly

- The accuracy of the short reads is used to decrease the error rate of the long reads from up to 20% to as low as 0.1%. Then, the corrected long reads are assembled using an algorithm such as overlap-layout-consensus.

Is it still necessary with new chemistry used by ONT and PacBio?





<https://gigabytejournal.com/articles/122>

NEW APPROACHES FOR LONG-RANGE GENOME SCAFFOLDING

- method called Hi-C, Omni-C (Hi-C is a chromosome conformation capture (3C)-based technology to detect pair-wise chromatin interactions genome-wide)
- Linked-Read Sequencing (single-tube long fragment reads (stLFR) and haplo-tagging (Meier et al., 2020; Wang et al., 2019)
- Optical maps
- Synteny-Based Methods

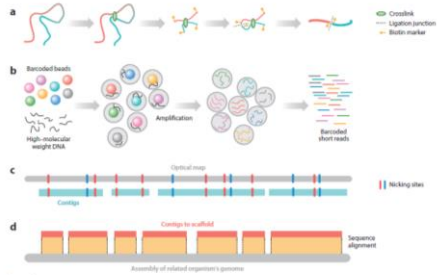
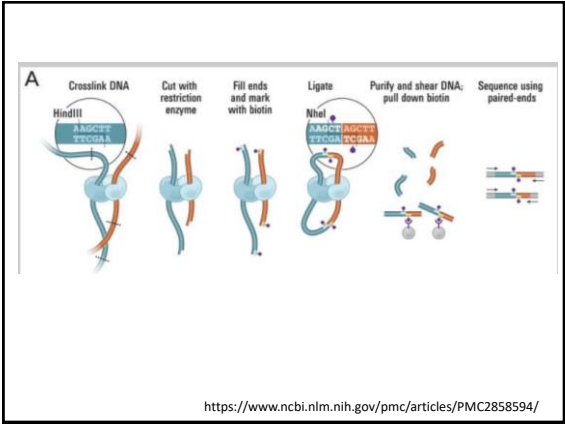
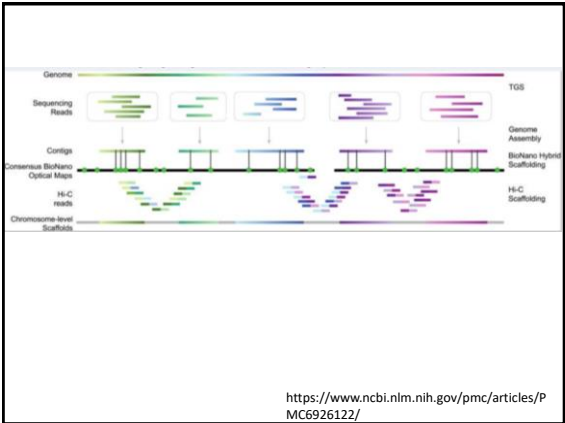
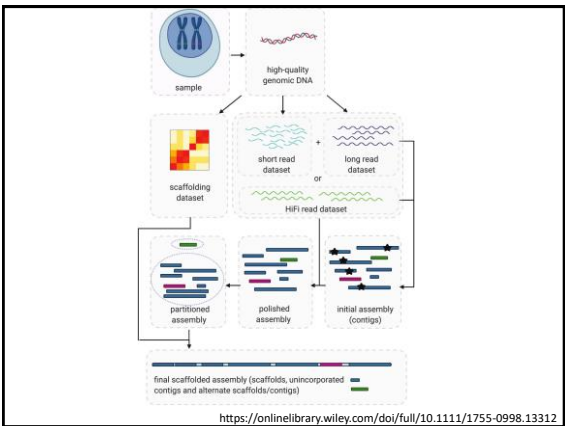


Figure 6
Overview of methods for long-range scaffolding. (a) In proximity ligation, chromatin is crosslinked and then restriction digested, ligated, and fragmented to create reads containing sequence from two different parts of the same chromosome. (b) In 10x linked-read sequencing, high-molecular weight DNA is combined with barcoded beads in oil droplets and then undergoes barcoding and amplification inside the droplets, resulting in reads with the same barcode that came from the same initial fragment of DNA. (c) BioNano optical maps are created by nicking high-molecular weight DNA with multiple nicking enzymes and attaching fluorescent markers at the nick sites. Contigs can then be aligned to the optical map by lining up nicking sequences in the contigs with the locations of fluorescent markers in the map. (d) In synteny-based approaches, contigs are mapped to the assembled genomes of one or more related species. These alignments imply the order and orientation of the aligned contigs.

As of April 2021, four biochemical companies (Arima Genomics, Dovetail Genomics, Phase Genomics, and Qiagen) manufacture Hi-C kits, which are formulated with different components and protocols. In general, conventional Hi-C kits employ a restriction enzyme or a cocktail of multiple restriction enzymes, whereas Omni-C employs a sequence-independent endonuclease (Table 1). In Omni-C, to capture more proximal contacts, disuccinimidyl glutarate (DSG) and formaldehyde are used for sample fixation (Nowak et al., 2005), which is now provided as a kit by Dovetail Genomics.







Key approaches for genome assembly that are generally recommended in all species include the following:

- (a) Genome assemblies should include long-read sequencing except in rare cases where it is effectively impossible to acquire adequately preserved samples needed for HMW DNA standards.
- (b) At least one scaffolding approach should be included with genome assembly such as Hi-C mapping or optical mapping (linked-read data is also appropriate but may not be available for most future projects).
- (c) Short-read data should be included for genome polishing, error correction, k-mer analyses, and estimating the percent of reads that map back to assembly.

<https://onlinelibrary.wiley.com/doi/full/10.1111/1755-0998.13312>

Validation of genome assembly

- BUSCO (Benchmarking Universal SingleCopy Orthologs) OrthoDB (BUSCO uses set of genes which are present in 90 % of species in one copy only)
- QUAST

Genome annotation

- Genome annotation is the process of identifying and labeling functional elements within the genome, such as genes, regulatory regions, and repetitive elements.

Table 3. Commonly used genome annotation tools and programs.

Name	GitHub link	Main feature
Prokka pipeline		
Prokka	https://www.sdu.se/cib/cib-genome/genome/annotation_prokka.html	Fast-track genome annotation. An automatic pipeline with flexibility and speed. Good for beginners and easy to use.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Prokaryotic genome annotation. An automatic pipeline with flexibility and speed. Good for beginners.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Genome annotation. An automatic pipeline for importing external data or using predictive algorithms. Good for beginners and easy to use.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Annotation and prediction.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Integration with Bioinformatics and Bioinformatics. An automatic pipeline and pipeline for genome structural and functional annotation. A user-friendly interactive portal that includes visualization and editing. Good for beginners and easy to use.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Genome and transcriptome. A rapid automatic platform for functional annotation and enrichment. A user-friendly pipeline that can export results in different output formats. Good for beginners and easy to use.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Functional annotation. An automatic platform as a standalone application that has high throughput and is interactive. A user-friendly pipeline with easy setup and low maintenance. Good for beginners.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	GO and GO enrichment analysis. A user-friendly web-based platform. Requires some configuration of public databases with fast, flexible and easy for the standalone application. A good web resource for beginners, but local installation requires bioinformatics support.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Detection of orthologous groups and functional annotation. An automatic pipeline and pipeline for any genome for the sake of good and quality (11 and 1.5 times faster than BLAST and BLASTX, respectively). Requires some configuration of public databases with various computer languages for a standalone application. A good web resource for beginners, but local installation requires bioinformatics support.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Orthologous assignment and pathway mapping. An automatic pipeline that has a limited number of query sequences. A good web resource for beginners, but local installation requires bioinformatics support.
GeneMark	https://www.sdu.se/cib/cib-genome/genome/annotation_geneMark.html	Gene structure annotation and annotation using de novo and transcript-based prediction. An automatic pipeline and pipeline for any genome for the sake of good and quality (11 and 1.5 times faster than BLAST and BLASTX, respectively). Requires some configuration of public databases with various computer languages and dependencies for a standalone application. A good web resource for beginners, but local installation requires bioinformatics support.
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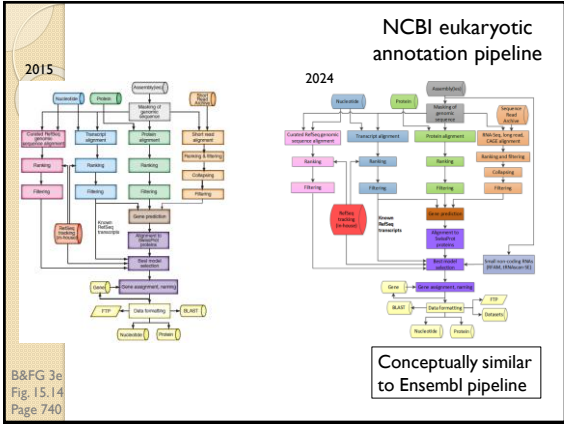
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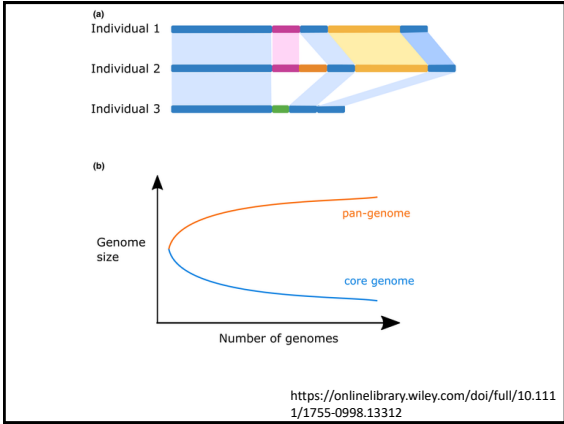
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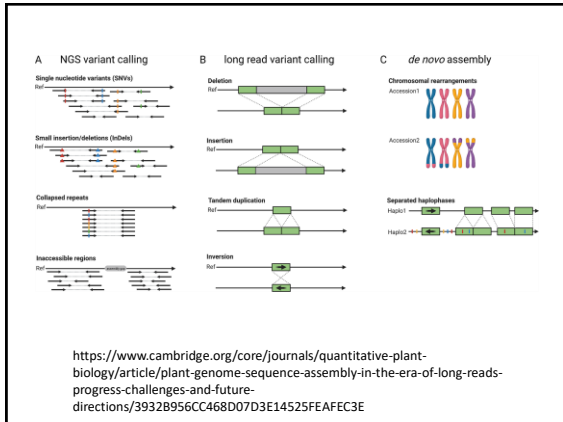


Biological challenges

- Repetitive regions (expanded gene families, complex repeats, highly repetitive regions such as centromeres and telomeres, and sex chromosomes, or at least portions of them.)
- Ploidy
- Pan and core genomes (The pan-genome represents all sequences among all of the DNA sequences that occur in a species whereas the core-genome is the DNA that is shared among all sequenced individuals.)
- For example, the comparison of eight chromosome-level assemblies of *Arabidopsis thaliana* accessions revealed a core-genome, shared by all accessions, of ~105 Mb and ~24,000 genes, whereas the pan-genome was ~135 Mb in length and included ~30,000 genes (Liao & Schneeberger, 2020), highlighting the vast amount of sequence data, including genes, that are missed by a single reference genome assembly.

<https://onlinelibrary.wiley.com/doi/full/10.1111/1755-0998.13312>





Why Is Chromosome-Scale Assembly Important?

- Cis-regulatory elements and the complexity of regulatory architecture
- Recombination
- Genetic association studies
- Chromosome evolution
