Presentations use info from:

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Chapter IV: Basic Local Alignment Sequence Tool (BLAST)

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What will you learn?

perform BLAST searches at the NCBI website;

understand how to vary optional BLAST search parameters;

explain the three phases of a BLAST search (compile, scan/extend, trace-back);

define the mathematical relationship between expect values and scores;

outline strategies for BLAST searching.

Outline

Introduction

BLAST search steps

Step 1: Specifying Sequence of interest

Step 2: Selecting BLAST Program

Step 3: Selecting a Database

Step 4: Selecting Search Parameters and Formatting Parameters

Stand-Alone BLAST

BLAST algorithm uses local alignment search strategy

BLAST algorithm parts: list, scan, extend

BLAST algorithm: local alignment search statistics and E value

Making sense of raw scores with bit scores

BLAST algorithm: Relation Between *E* and *p* values

BLAST search strategies

General concepts; principles of BLAST searching

How to evaluate the significance of results

How to handle too many or few results

BLAST searching with multidomain protein: HIV-1 Pol

Using BLAST for gene discovery: Find-a-Gene

Perspective

BLAST

BLAST (Basic Local Alignment Search Tool) allows rapid sequence comparison of a query sequence against a database.

The **BLAST** algorithm is <u>fast</u>, <u>accurate</u>, and <u>accessible</u> both via the web and the command line.

Why use BLAST?

BLAST searching is fundamental to understanding the relatedness of any favorite query sequence to other known proteins or DNA sequences.

Applications include

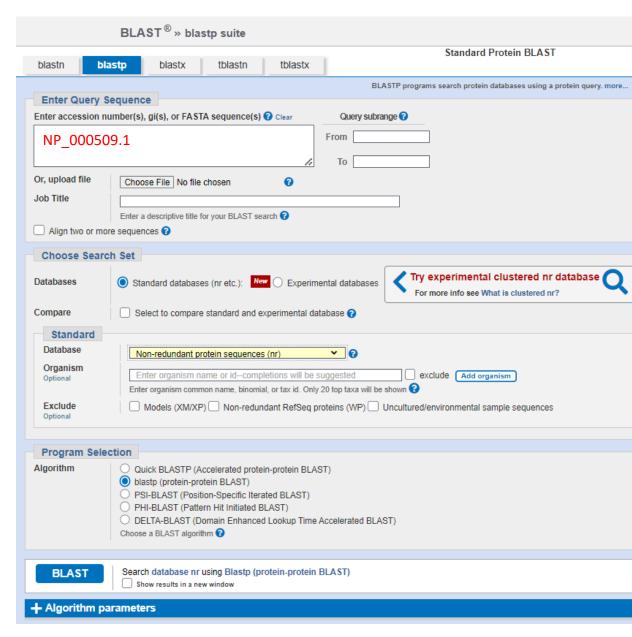
- identifying orthologs and paralogs
- what proteins or genes are present in organism
- discovering new genes or proteins
- discovering variants of genes or proteins
- investigating expressed sequence tags (ESTs)
- exploring protein structure and function

The programs produce high-scoring segment pairs (HSPs) that represent local alignments between your query and database sequences.

BLASTP search at NCBI: overview of web-based search

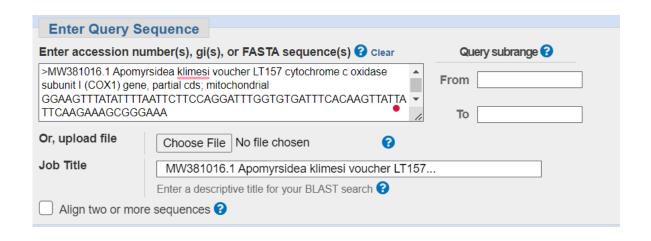
4 components to performing any web-based BLAST search:

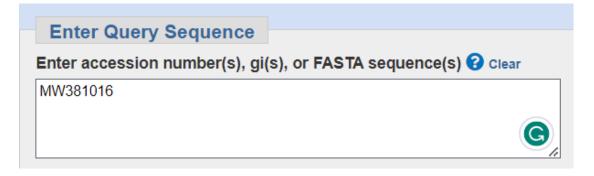
query: FASTA format or accession database algorithm parameters



Step 1: Choose your sequence

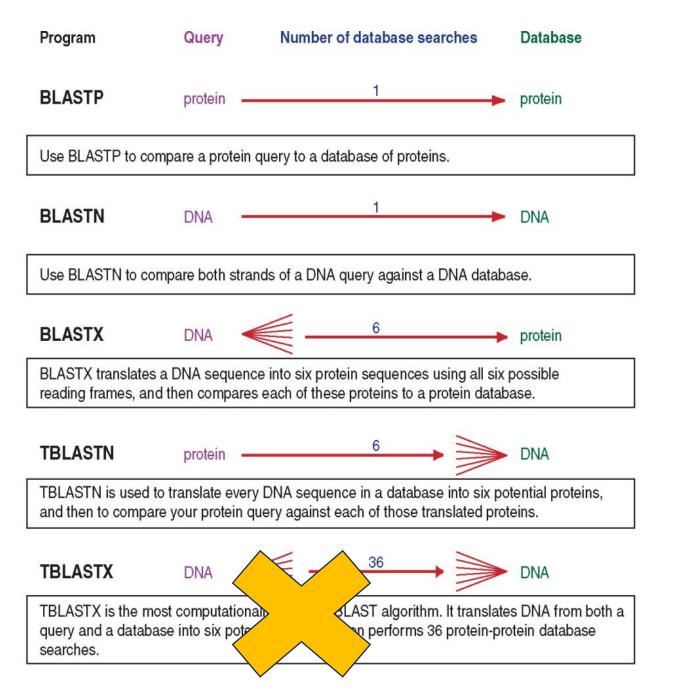
Sequence can be input in FASTA format or as accession number



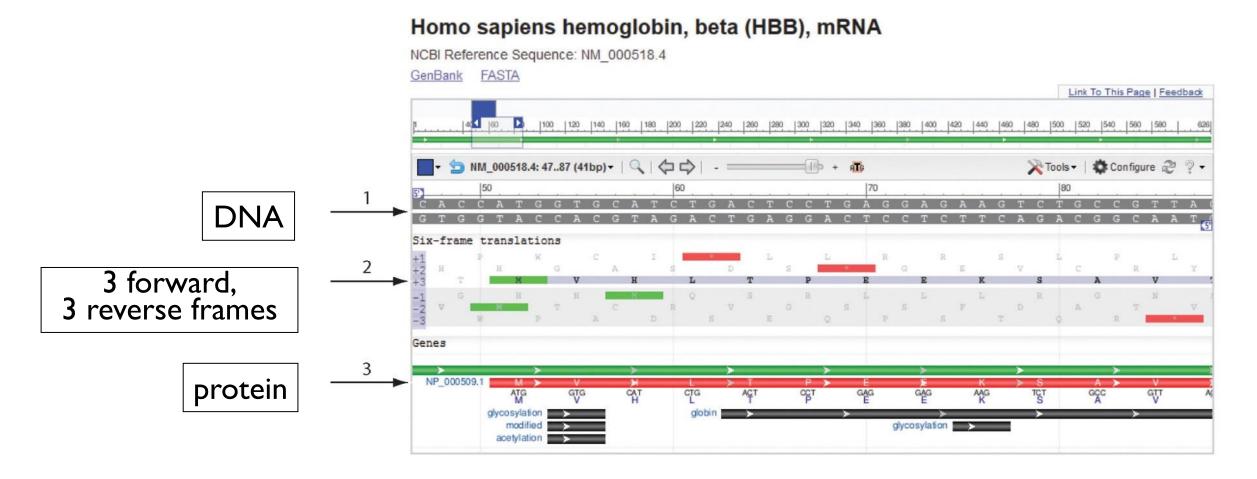


BLAST step 2: Choose Program

N refers to nucleotideX refers to a DNAP refers to proteinT refers to "translating"



Step 2 (choosing the BLAST program): DNA can be translated into six reading frames



This image is from the NCBI Nucleotide entry for HBB

Double-stranded DNA -> Amino Acids -> Codons (3 nucleotides)

- The ribosome starts at a start codon and continues reading until it reaches a stop codon.
- The ribosome can start reading the DNA sequence at any point on the strand.
- Forward and Reverse x3 = 6 reading frames

GenBank FASTA

DNA

3 forward, 3 reverse frames

protein



NCBI Nucleotide entry for HBB

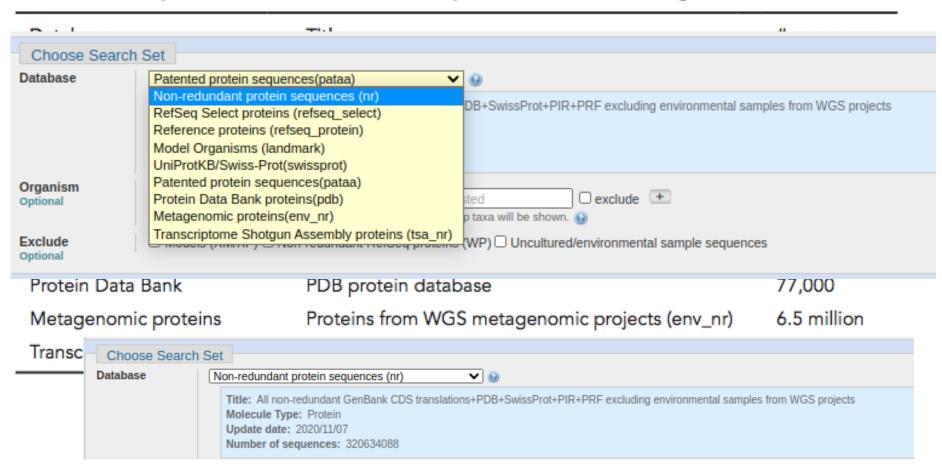
Step 3: choose a database to search (protein databases)

TABLE 4.1 Protein sequence databases that can be searched by BLAST searching at NCBI. PDB, Protein Data Bank. # indicates approximate number of sequences in database. Adapted from BLAST, NCBI, # http://blast.ncbi.nlm.nih.gov/.

Database	Title	# sequences
nr	All nonredundant GenBank CDS translations + PDB + SwissProt + PIR + PRF excluding environmental samples from WGS projects	65 million
Reference proteins	NCBI protein reference sequences	50 million
UniProtKB/SwissProt	Nonredundant UniProtKB/SwissProt sequences	450,000
Patented protein sequences	Protein sequences derived from the Patent division of GenBank	1.3 million
Protein Data Bank	PDB protein database	77,000
Metagenomic proteins	Proteins from WGS metagenomic projects (env_nr)	6.5 million
Transcriptome	Transcriptome Shotgun Assembly (TSA) sequences	770,000

Step 3: choose a database to search (protein databases)

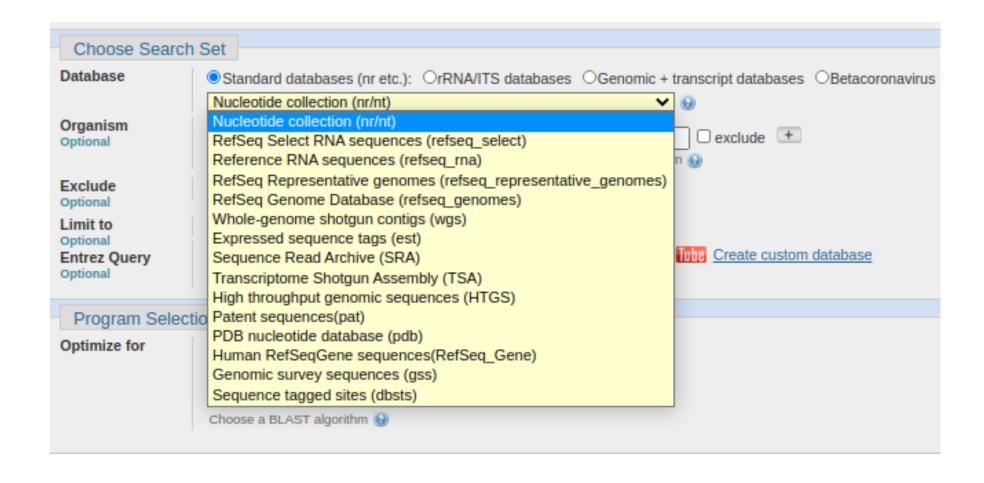
TABLE 4.1 Protein sequence databases that can be searched by BLAST searching at NCBI. PDB, Protein Data Bank. # indicates approximate number of sequences in database. Adapted from BLAST, NCBI, # http://blast.ncbi.nlm.nih.gov/.



Step 3: choose a database to search (nucleotide)

Database	Title	# sequences
Human Genomic + Transcript	Homo sapiens NCBI Annotation Release 104 RNAs; Homo sapiens all assemblies	
Mouse Genomic + Transcript	Mus musculus NCBI Annotation RNAs; Mus musculus all assemblies	N/A
nr/nt	All GenBank+EMBL+DDBJ+PDB+RefSec sequences, but excludes EST, STS, GSS WGS, TSA, patent segmences as well as se 0, 1, and 2	,
refseq_rna	anscrip ences	3.5 million
refseq_genomic	ces	2.7 million
NCBI Genomes	∠s	28,000
Expressed sequence tags (EST)	MBL+DDBJ visions	75 million
Genomic survey sequences (gss)	includes on-trapped ces	36 million d
High-throughput genomic sequences (HTGS)	ed hig Iomic	153,000 2
Patent sequences	vucleotide sequence erived from the Patent division of GenBank	21 million
Protein Data Bank	PDB nucleotide database	8000
alu	Human Alu repeat elements	325
Sequence tagged sites (STS)	Database of GenBank+EMBL+DDBJ sequences from STS Divisions	1.3 million
Whole-genome shotgun (wgs)	Whole-genome-shotgun contigs	116 million
Transcriptome Shotgun Assembly (TSA)	Transcriptome shotgun assembly (TSA) sequences	15 million
16S ribosomal RNA sequences (Bacteria and Archaea)	16S ribosomal RNA sequences (bacteria and archaea)	7500

Step 3: choose a database to search (nucleotide)



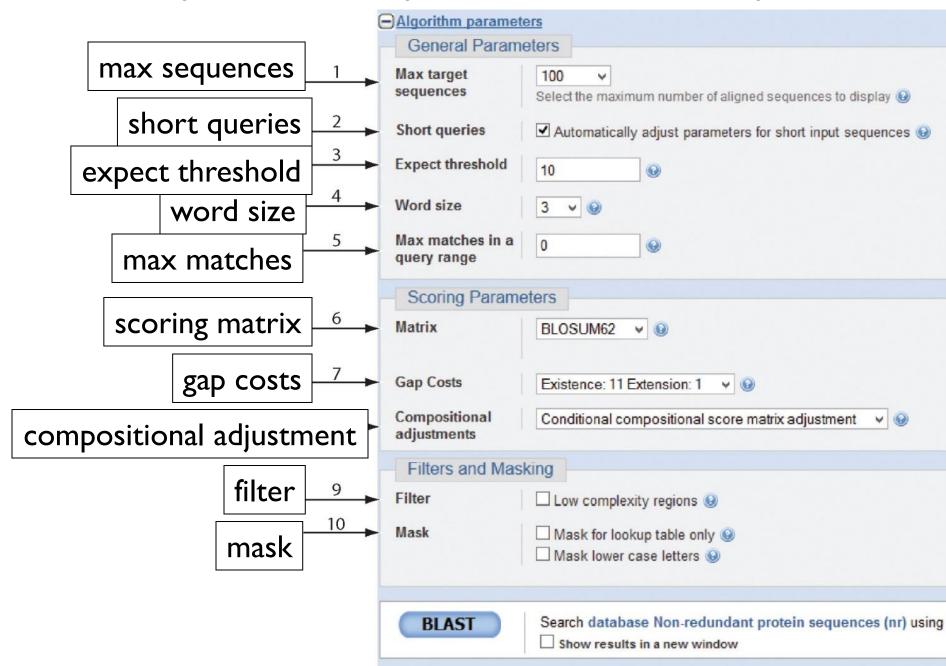
Step 4: optional parameters

You can...

- choose the organism to search
- turn filtering on/off
- change the substitution matrix
- change the expect (e) value
- change the word size

Example: BLASTP human insulin (NP_000198) against a *C. elegans* RefSeq database. Varying some parameters (filtering, compositional adjustments) can greatly affect the alignment itself.

Step 4a: choose optional BLASTP search parameters



Q: What is the Expect (E) value?

The Expect value (E) is a parameter that describes the number of hits one can "expect" to see by chance when searching a database of a particular size. It decreases exponentially as the Score (S) of the match increases.

E value describes the random background noise.

For example, an E value of 1 assigned to a hit can be interpreted as meaning that in a database of the current size one might expect to see 1 match with a similar score simply by chance.

Step 4a: compositional adjustment influences score, expect value search results

expect = 0.05

Default: conditional compositional score matrix adjustment

expect = 0.09

no adjustment

(a) Default: conditional compositional score matrix adjustment

Insulin-like peptide 3 [Drosophila melanogaster]

Sequence ID: ref|NP 648360.2| Length: 120 Number of Matches: 1

Range 1: 32 to 114 GenPept Graphics

Score		Expect	Method	Identities	Positives	Gaps	
31.6 bi	ts(70)	0.050	Compositional matrix adjus	t. 21/88(24%)	40/88(45%)	12/88(13	3%)
Query	29		LVEALYLVCGERGFFYTPKTRRE	AEDLQVGQVELGGG			87
Sbjct	32		PETLSKLCVYGFNAMTKRT				86
Query	88		RRGIVEQCCTSICSLYQLENYC	109			
Shict	87	LKTPRLE	DOWEDECCI.KSCTMDEWI.RVC	114			

(b) No adjustment (by default, filter low complexity regions)

Insulin-like peptide 3 [Drosophila melanogaster]

Sequence ID: ref|NP 648360.2| Length: 120 Number of Matches: 1

Range 1: 33 to 114 GenPept Graphics

Score		Expect	Identities	Positives	Gaps	
33.5 b	its(75	0.009	21/87(24%)	40/87(45%)	12/87(13%)	
Query	30	LCGSHLVEALYLVC		EDLQVGQVELGGGPGAG	SSLQPLALEGSLQ L+ L + S+O	87
Sbjct	33			DPVNFNQIDGFEDRS		87
Query	88	KRGIVEQCC		109		
Sbjct	88	KTRRLRDGVFDECC	40.000.NF/121814.dl/11184.00057774	114		

(c) Composition-based statistics

Insulin-like peptide 3 [Drosophila melanogaster]

Sequence ID: ref[NP 648360.2] Length: 120 Number of Matches: 1

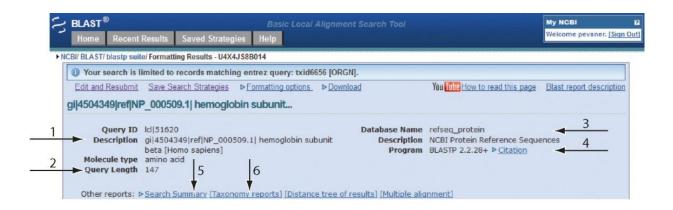
Range 1: 33 to 114 GenPept Graphics

Score		Expect	Method			Identit	ies	Positive	5	Gaps	
30.4 b	its(67)	1e-04	Composition	on-based	stats.	21/87(24%)	40/87(4	15%)	12/87(1	.3%)
Query	30		EALYLVCGE					2000		300 September 1	87
Sbjct	33		E L +C ETLSKLCV-			+ Q+ PVNFNQI				+ S+Q DSSVQML	87
Query	88		GIVEQCCTS:			.09					
Sbjct	88	20.0	GVFDECCLK:		T0000	.14					

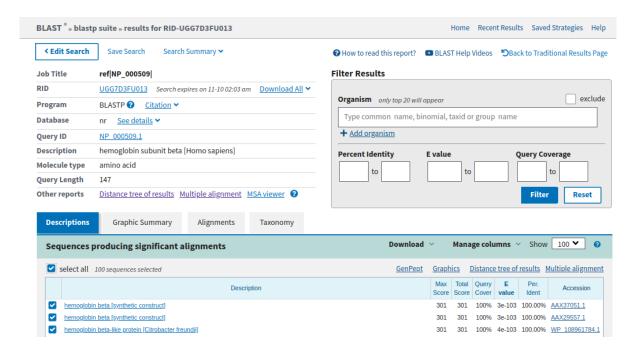
expect = Ie-04

composition-based statistics

Step 4b: formatting options



The top of the BLAST output summarizes the query, database, and BLAST algorithm.
Click to access a summary of the search parameters or a taxonomic report.



Step 4b: formatting options (you can view search parameters)

Search Parameters						
Program	blastp					
Word size	3					
Expect value	10 -1					
Hitlist size	100					
Gapcosts	11,1					
Matrix	BLOSUM62 ← 2					
Filter string	F					
Genetic Code	1					
Window Size	40					
Threshold	11					
Composition-based stats	2					

Database							
Posted date	Jun 12, 2013 10:46 AM						
Number of letters	6,910,040,5394						
Number of sequences	19,996,853						
Entrez guery	tvid10090 [ORGN]						

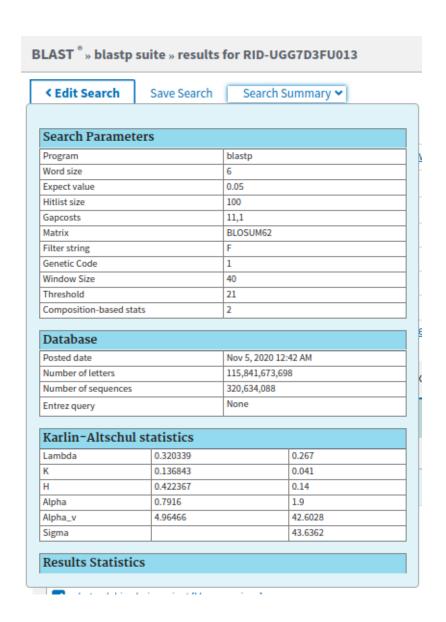
Karlin-Altschul statistics						
Lambda	0.320339	0.267				
K	0.136843	0.041				
Н	0.422367	0.14				
Alpha	0.7916	1.9				
Alpha_v	4.96466	42.6028				
Sigma		43.6362				

Expect value

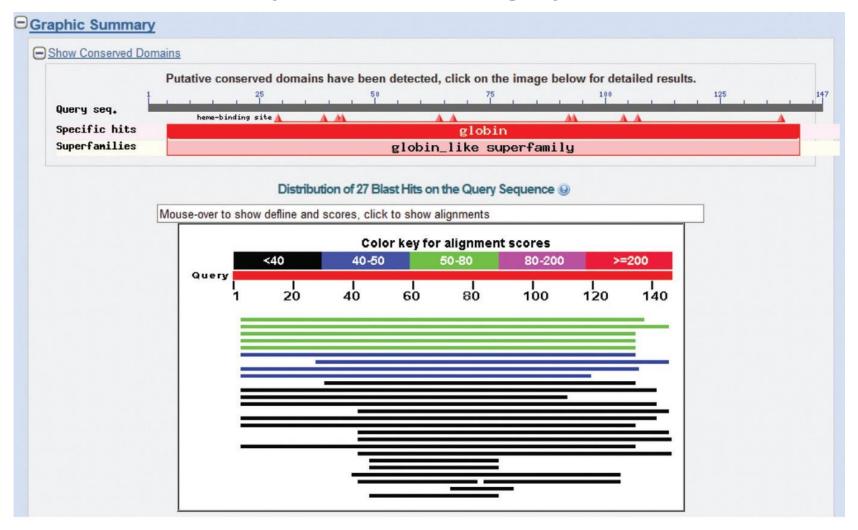
BLOSUM62 matrix

Threshold value T

Size of database



Step 4b: formatting options



Graphic summary of the results shows the alignment scores (coded by color) and the length of the alignment (given by the length of the horizontal bars)

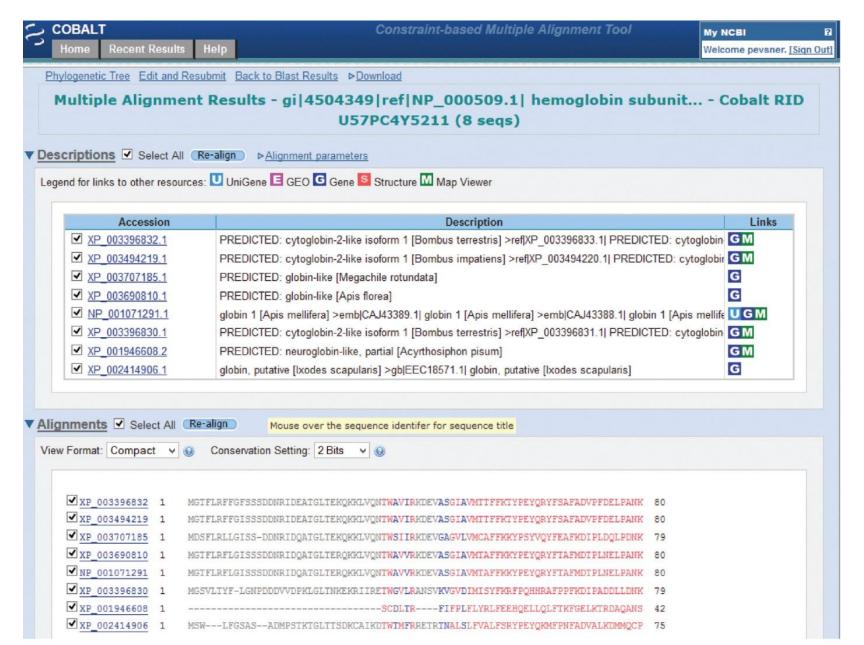
BLASTP output includes list of matches; links to the NCBI protein entry; bit score and E value; and download options

Sequences producing significant alignments:

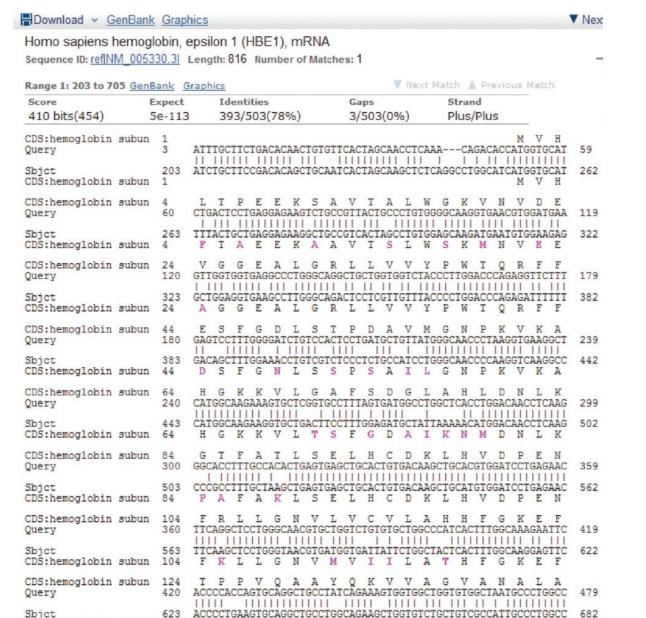
globin, putative [Ixodes scapularis]

Sele	ect: All None Selected:2						
AT	Alignments Download - GenPept Graphics Distance tree of results Multiple align	ment					0
	Description	Max score	Total score	Query	E value	Max ident	Accession
•	PREDICTED: cytoglobin-2-like isoform 1 [Bombus terrestris] >ref[XP 003396833.1] PREDIC	59.7	59.7	91%	1e-10	29%	XP 003396832.1
•	PREDICTED: cytoglobin-2-like isoform 1 [Bombus impatiens] >ref[XP 003494220.1] PREDI	58.5	58.5	97%	3e-10	28%	XP 003494219.1
	PREDICTED: globin-like [Megachile rotundata]	57.8	57.8	89%	6e-10	29%	XP 003707185.1
	PREDICTED: globin-like [Apis florea]	53.9	53.9	89%	1e-08	30%	XP 003690810.1
	globin 1 [Apis mellifera]	52.8	52.8	89%	4e-08	30%	NP 001071291.1
	PREDICTED: cytoglobin-2-like isoform 1 [Bombus terrestris] >ref[XP 003396831.1] PREDIC	45.1	45.1	89%	2e-05	26%	XP 003396830.1
П	PREDICTED: neuroglobin-like, partial [Acvrthosiphon pisum]	42.4	42.4	80%	2e-04	23%	XP 001946608.2

BLAST output can be formatted to display multiple alignment



For BLASTN, CDS output displays amino acids above DNA sequence of query and subject



T P E V Q A A W Q K L V S A V A I A L A

CDS:hemoglobin subun 124

CDS:hemoglobin subun 144

CDS:hemoglobin subun 144

Query

Sbjct

HKYH

HKYH

CACAAGTATCACTAAGCTCGCTT 502

CATAAGTACCACTGAGTTCTCTT 705

11 11111 1111 11 11 111

Command-line BLAST+

Visit the BLAST site at NCBI ("help" tab) to find the URL for the BLAST+ download.

Three steps:

- (1) Obtain a protein database (we'll use a perl script included in the BLAST+ installation);
- (2) Obtain a query protein (we'll use EDirect);
- (3) Perform the search

Command-line BLAST+ (Step 1: obtain a database)

Visit the BLAST site at NCBI ("help" tab) to find the URL for the BLAST+ download.

```
$ mkdir database # this creates a new directory
$ cd database/ # we navigate into that directory
# Enter the following, without arguments, to see a help document.
$ update_blastdb.pl
# Next get a list of all available databases
$ update_blastdb.pl --showall
$ update_blastdb.pl --showall | less
```

```
$ update_blastdb.pl refseq_protein
$ tar -zxvf refseq_protein.00.tar.gz
```

You will also need to install the EDirect command-line utility if you wish to look up TAXIDs via the command-line script

(You will explore command-line workflow during exercises)

Command-line BLAST+ (Step 2: obtain a query protein)

Use EDirect to obtain a globin protein.

```
$ esearch -db protein -query "NP_000509" | efetch -format fasta > hbb.
txt
$ cat hbb.txt # cat is the concatenate utility that we use to print the
# file
>gi|4504349|ref|NP_000509.1| hemoglobin subunit beta [Homo sapiens]
MVHLTPEEKSAVTALWGKVNVDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPKVKAHGKKVLG
AFSDGLAHLDNLKGTFATLSELHCDKLHVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVAN
ALAHKYH
```

Command-line BLAST+ (Step 3: perform a search!)

Do the search:

```
$ blastp --h # Get help
$ blastp -query hbb.txt -db ./database/refseq_protein -out mysearch1
# Note that we use ./ to specify the directory location of the
# executable which is within the executable directory
```

View the results:

```
$ less mysearch1
```

Try repeating the search, e.g. changing the database size:

```
$ blastp -query hbb.txt -db ./database/refseq_protein -dbsize 9750000 -out
mysearch2
```

(You will explore command-line workflow during exercises)

How a BLAST search works

"The central idea of the BLAST algorithm is to confine attention to segment pairs that contain a word pair of length w with a score of at least T."

Altschul et al. (1990)

How the original BLAST algorithm works: three phases

Phase I: compile a list of word pairs (w=3) above threshold T

Example: for a human RBP queryFSGTWYA... (query word is in green)

A list of $\underline{\text{words}}$ (w=3) is:

FSG SGT GTW TWY WYA YSG TGT ATW SWY WFA FTG SVT GSW TWF WYS

• • •

Phase 1: compile a list of words (w=3)

	GTW	6,5,11	22
neighborhood	GSW	6,1,11	18
word hits	ATW	0,5,11	16
> threshold	NTW	0,5,11	16
(T=11)	GTY	6,5,2	13
(I-II)	GNW		10
neighborhood	GAW		9
word hits			
< below thresho	old		

A threshold value **T** is established for the score of aligned words.

Phase 1: Setup: compile a list of words (w=3) above threshold T

Query sequence: human beta globin NP_000509.1 (includes ...VTALWGKVNVD...).
 This sequence is read; low complexity or other filtering is applied; a "lookup" table is built.

Words derived from query sequence (HBB): VTA TAL ALW LWG WGK GKV KVN VNV NVD
 Generate a list of words matching query (both above and below T). Consider LWG in the query and the scores (derived from a

 Generate similar lists of words spanning the query (e.g. words for wgw, gwg, wgk...).

threshold

BLOSUM62 matrix) for various words.

MWG 2+11+6=19 1+11+6=18 VWG examples of FWG 0+11+6=17 words >= 0+11+6=17 AWG threshold 12 4+11+0=15 LWS 4+11+0=15 LWN LWA 4+11+0=15 4+ 2+6=12 LYG 4+ 1+6=11 LFG examples of FWS 0+11+0=11

EFG 4+ 1+6=11
examples of FWS 0+11+0=11
words below AWS -1+11+0=10
threshold CWS -1+11+0=10
IWC 2+11-3=10

Phase 2: scan the database for matches and extend

Phase 2: Scanning and extensions

- Select all the words above threshold T (LWG, IWG, MWG, VWG, FWG, AWG, LWS, LWN, LWA, LYG)
- Scan the database for entries ("hits") that match the compiled list
- Create a hash table index with the locations of all the hits for each word
- Perform gap free extensions
- Perform gapped extensions

```
LTPEEKSAVTALWGKV--NVDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPKV HBB

L+P +K+ V A WGKV + E G EAL R+ + +P T+ +F F D G+ +V

LSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHF------DLSHGSAQV HBA

extension extension

first phases of search

"hits" alpha globin,

triggers extension
```

Phase 3: Traceback to generate gapped alignment

Phase 3: Traceback

- Calculate locations of insertions, deletions, and matches (for alignments saved in Phase 2)
- Apply composition-based statistics (for BLASTP, TBLASTN)
- · Generate gapped alignment

How a BLAST search works: threshold

You can locally install BLAST and modify the threshold parameter.

The default value for BLASTP is 11.

To change it, enter "-f 16" or "-f 5" in the advanced options of BLAST+.

For **BLASTN**, the word size is typically **7**, **11**, **or 15** (EXACT match). Changing word size is like changing threshold of proteins. w=15 gives fewer matches and is faster than w=11 or w=7.

For megaBLAST the word size is 28 and can be adjusted to 64. What will this do? MegaBLAST is VERY fast for finding closely related DNA sequences!

How to interpret a BLAST search: expect value

The **expect value** *E* is the number of alignments with scores greater than or equal to **score** *S* that are expected to occur by chance in a database search.

An **E** value is related to a **probability value p**.

The key equation describing an *E* value is:

 $E = Kmn e^{-lS}$

$$E = Kmn e^{-\lambda S}$$

This equation is derived from a description of the extreme value distribution

S = the score

E = the expect value = the number of high-scoring segment pairs (HSPs) expected to occur with a score of at least S

m, *n* = the length of two sequences

I, K = Karlin Altschul statistics

The E value depends on the score and on λ , which is a parameter that scales the scoring system. E also depends on the length of the query sequence and the length of the database. The parameter K is a scaling factor for the search space. The parameters K and λ are described by Karlin and Altschul (1990), and are often referred to as Karlin–Altschul statistics.

From raw scores to bit scores

- There are two kinds of scores: raw scores (calculated from a substitution matrix) and bit scores (normalized scores)
- Bit scores are comparable between different searches because they are normalized to account for the use of different scoring matrices and different database sizes

$$S' = bit score = (IS - InK) / In2$$

The *E* value corresponding to a given bit score is: $E = mn \ 2^{-S'}$

Bit scores allow you to compare results between different database searches, even using different scoring matrices.

How to interpret BLAST: *E* values and *p* values

The **expect value** *E* is the number of alignments with scores greater than or equal to **score** *S* that are expected to occur by chance in a database search. A *p* value is a different way of representing the significance of an alignment.

$$p = I - e^{-E}$$

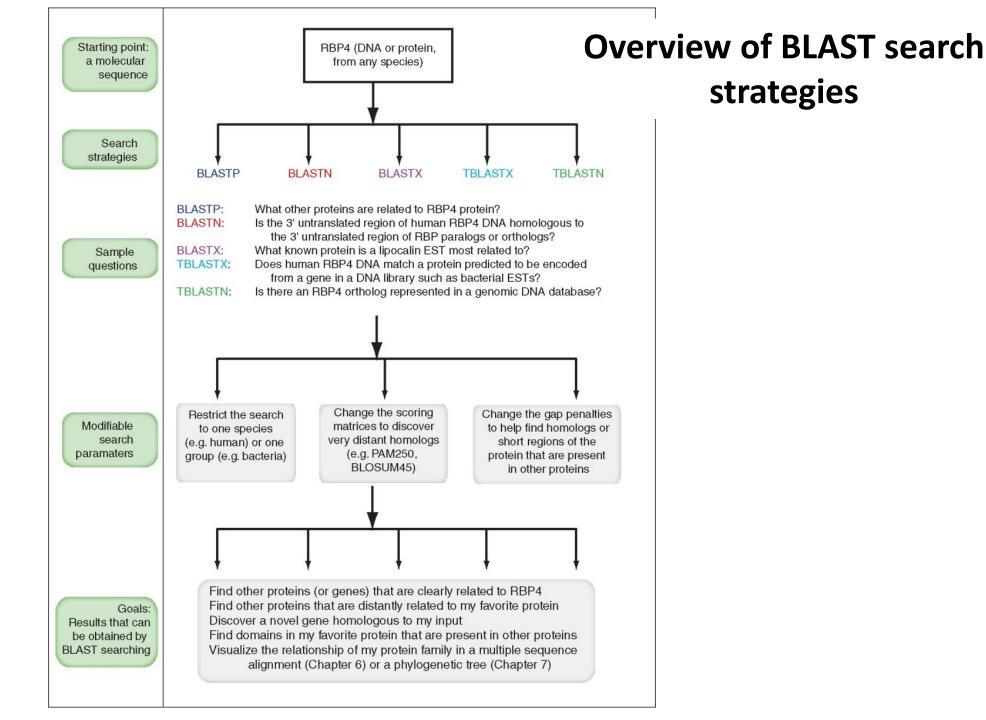
How to interpret BLAST: *E* values and *p* values

E values of about I to 10 are far easier to interpret than corresponding p values.

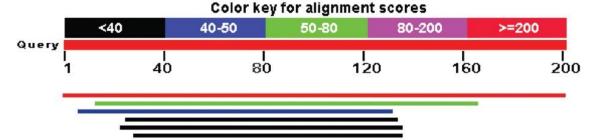
Very small E values are very similar to p values.

<u> </u>	<u>Þ</u>
10	0.99995460
5	0.99326205
2	0.86466472
1	0.63212056
0.1	0.09516258 (about 0.1)
0.05	0.04877058 (about 0.05)
0.001	0.00099950 (about 0.001
0.0001	0.00010000

E values are comparable to p values and are designed to be more convenient to interpret.



(a) Graphical overview



(b) List of alignments

Sequences producing significant alignments:

Select: All None Selected:6

BLASTP search: human RBP4 query, human RefSeq database

	Description	Max score	Total score	Query	E value	Max ident	Accession	
⊽	retinol-binding protein 4 precursor [Homo sapiens]	420	420	100%	1e-150	100%	NP 006735.2	
V	apolipoprotein D precursor [Homo sapiens]	55.5	55.5	76%	1e-09	28%	NP 001638.1	
V	glycodelin precursor [Homo sapiens] >ref[NP 002562.2] glycodelin precursor [Homo s	40.0	40.0	62%	5e-04	26%	NP 001018059	
V	protein AMBP preproprotein [Homo sapiens]	35.0	35.0	54%	0.034	23%	NP 001624.1	
V	complement component C8 gamma chain precursor [Homo sapiens]	32.3	32.3	56%	0.18	25%	NP 000597.2	
V	lipocalin-15 precursor [Homo sapiens]	28.5	28.5	53%	3.4	23%	NP 976222.1	

Results include matches (such as CG8) with high E values and limited identity to the query

(c) Pairwise alignment of RBP4 and C8G

complement component C8 gamma chair precursor (nomo sapiens)
Sequence ID: ref[NP 000597.2] Length: 202 Number of Matches: 1

Range 1: 33 to 139 GenPept Graphics

Score Expect Method Identities Positives Gaps
32.3 bits(72) 0.18 Compositional matrix adjust. 28/114(25%) 49/114(42%) 8/114(7%)

Query 24 VSSFRVKENFDKARFSGTWYAMAKKDPEGLFLQDNIVAEFSVDETG-QMSATAKGRVRLL 82
+5+ + K NFD +F+GTW +A + AE + Q +A A R L

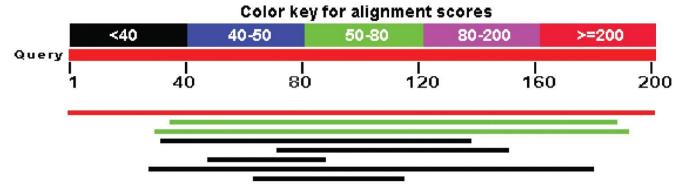
Sbjct 33 ISTIQPKANFDAQQFAGTWLLVAVGSACRFLQEQGHRAEATTLHVAPQGTAMAVSTFRKL 92

Query 83 NNWDVCADMVGTFTDTEDPAKFKMKYWGVASFLQKGNDDHWIVDTDYDTYAVQY 136
+ +C + + DT +F ++ +G + +TDY ++AV Y

Sbjct 93 DG--ICWQVRQLYGDTGVLGRFLLQARDA-----RGAVHVVVAETDYQSFAVLY 139

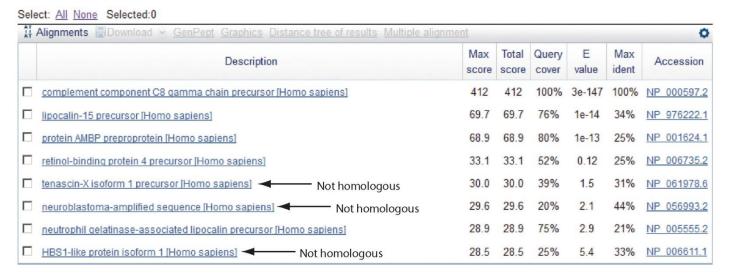
"Reciprocal" BLASTP search with CG8 as query includes RBP4 and other lipocalins

(a) Graphical overview



(b) List of alignments

Sequences producing significant alignments:



This confirms that the finding of CG8 using RBP4 as a query was a true positive

Sequence analysis

Choosing BLAST options for better detection of orthologs as reciprocal best hits

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ABSTRAC

Motivation: The analyses of the increasing number of genome sequences requires shortcuts for the detection of orthologs, such as Reciprocal Best Hits (RBH), where orthologs are assumed if two genes each in a different genome find each other as the best hit in the other genome. Two BLAST options seem to affect alignment scores the most, and thus the choice of a best hit: the filtering of low information sequence segments and the algorithm used to produce the final alignment. Thus, we decided to test whether such options would help better detect orthologs.

Results: Using Escherichia coli K12 as an example, we compared the number and quality of orthologs detected as RBH. We tested four different conditions derived from two options: filtering of low-information segments, hard (default) versus soft; and alignment algorithm, default (based on matching words) versus Smith-Waterman. All options resulted in significant differences in the number of orthologs detected, with the highest numbers obtained with the combination of soft filtering with Smith-Waterman alignments. We compared these results with those of Reciprocal Shortest Distances (RSD), supposed to be superior to RBH because it uses an evolutionary measure of distance, rather than BLAST statistics. to rank homologs and thus detect orthologs. RSD barely increased the number of orthologs detected over those found with RBH. Error estimates, based on analyses of conservation of gene order, found small differences in the quality of orthologs detected using RBH. However, RSD showed the highest error rates. Thus, RSD have no

Availability: Orthologs detected as Reciprocal Best Hits using soft masking and Smith-Waterman alignments can be downloaded from http://popolvuh.wlu.ca/Orthologs.

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computational biologists often need their own orthologous sets for variable reasons such as: (1) a newly sequenced genome that needs annotation; (2) a need for updated mappings not available in published ortholog databases; (3) the lack of agreement about the genome annotations to use, for instance, those provided by the authors of a genome, corrections such as those within the RefSeq database (Maglott et al., 2000; Pruitt et al., 2005), the Genome Reviews (http://www.ebi.ac.uk/GenomeReviews/), the HAMAP project (Boeckmann et al., 2003; Gattiker et al., 2003) or even those re-annotations produced by particular research groups (Besemer et al., 2001).

Orthologs are defined as genes that have diverged after a speciation event (Fitch, 2000). Another way to define them might be as the 'same genes' in different organisms. This evolutionary relationship implies that products of orthologous genes should tend to keep their original functions. Paralogs, on the other hand, are defined as genes that have diverged after a duplication event (Fitch, 2000). These have been proposed as a source of functional innovation (Francino, 2005; Ohno, 1970) and are less expected to have similar functions. It is therefore very important to be able to differentiate between orthologs and extra-paralogs, paralogous genes residing in different organisms (Janga and Moreno-Hagelsieb, 2004).

The definitions above are based on the event separating the histories of the homologous genes in question. In practice, one has to rely on sequence similarity and suitable statistics to detect homologs. Once putative homologs have been detected, evolutionary models such as phylogenetic trees, would be too computationally intensive to run for orthology detection, especially given the growth rate of genome sequence databases. Thus, most research in comparative genomics relies on some sort of shortcut, or working definition, to detect orthology.

Essentially, if gene a in genome A finds gene b as its best, highest scoring, match in genome B; and gene b finds gene a as its best match in genome A, they are RBH and thus inferred to be orthologs.

Sequence analysis

Progress in quickly finding orthologs as reciprocal best hits

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Abstract

Motivation: While the authors of software for the quick comparison of protein sequences evaluate the speed of their software and compare their results against the most usual software for the task, it is not common for them to evaluate their software for more particular uses, such as finding orthologs as reciprocal best hits (RBH). Here we focused on comparing RBH results using software that runs faster than blastp. Namely, lastal, diamond, and MMseqs2.

Results: Of the programs tested, lastal required the least time to produce results. However, it produced fewer results than any other program when comparing evolutionarily distant genomes. The program producing the most similar number of RBH as blastp was MMseqs2. This program was also resulted in the lowest error estimates compared to any of the programs used. The results with diamond were very close to those obtained with MMseqs2, with diamond running much faster. Our results suggest that the best of the programs tested was diamond, ran with the "sensitive" option, which took 7% of the time as blastp to run, and produced results with lower error rates than blastp.

Availability: A program to obtain reciprocal best hits using each sequence comparison program is maintained at https://github.com/Computational-conSequences/SequenceTools

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Supplementary information: There is no Supplementary data associated with this manuscript.

...two somewhat recently developed programs for sequence comparison include **diamond (Buchfink et al., 2015)** and **MMseqs2 (Steinegger and Söding, 2017)** (from now on mmseqs). Here we use lastal as a reference to the previous report (Ward and Moreno-Hagelsieb, 2014), where **lastal** was the program producing the most-similar-to-blastp results, and test these two new programs in terms of the proportion of RBH found and their quality compared to blastp.

Standard workflow:

DIAMOND or MMseqs2 (recommended, although BLAST+ can be used instead)

The MCL graph clustering algorithm

FastME (The appropriate version for your system, e.g. 'fastme-2.1.5-linux64', should be renamed `fastme', see instructions below.)

OrthoFinder: phylogenetic orthology inference for comparative genomics

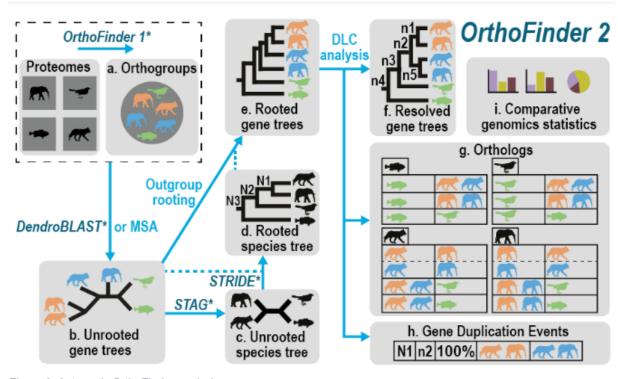


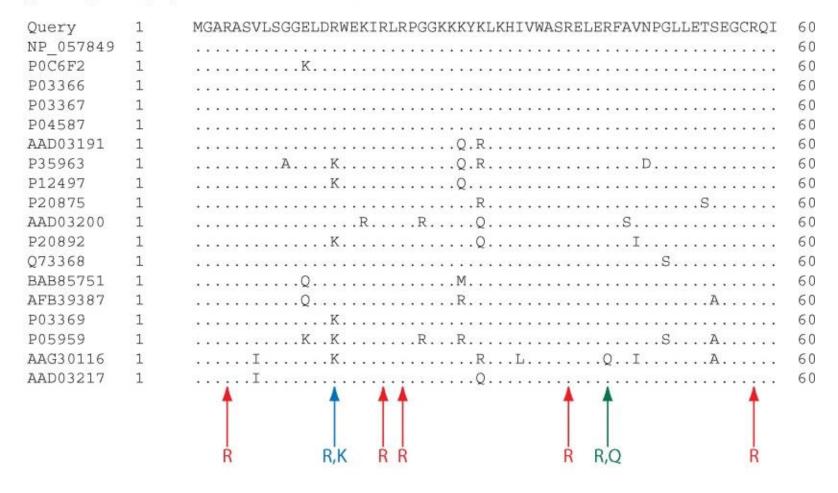
Figure 1: Automatic OrthoFinder analysis

What does OrthoFinder do?

OrthoFinder is a fast, accurate and comprehensive platform for comparative genomics. It finds **orthogroups** and **orthologs**, infers **rooted gene trees** for all orthogroups and identifies all of the **gene duplication events** in those gene trees. It also infers a **rooted species tree** for the species being analysed and maps the gene duplication events from the gene trees to branches in the species tree. OrthoFinder also provides **comprehensive statistics** for comparative genomic analyses. OrthoFinder is simple to use and all you need to run it is a set of protein sequence files (one per species) in FASTA format.

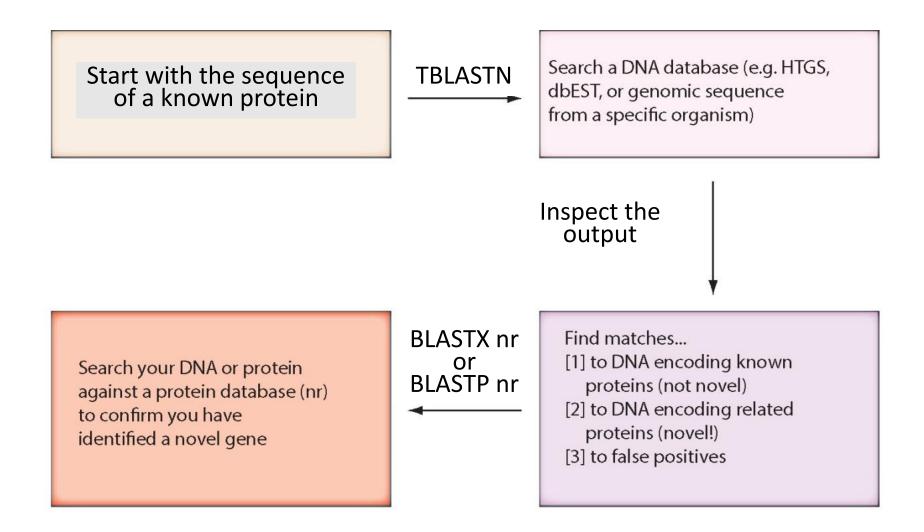
BLAST searching a multidomain protein: HIV-1 pol

(b) List of alignments (query-anchored with dots for identities)



This output shows identical residues as a dot ".". Note that the column positions that contain an arginine (R) can sometimes also contain a lysine (K) or glutamine (Q) in a position-specific pattern. This is a preview of the concept of position-specific scoring matrices.

"Find-a-gene project" to practice BLAST



"Find-a-gene project" example: novel globin

(a) Result of TBLASTN against nematode ESTs using human beta globin as a guery

Ac_EH1r_01A07_M13 Adult Anguillicola crassus Anguillicola crassus cDNA clone Ac_EH1r_01A07
Sequence ID: gblJK511422.1| Length: 559 Number of Matches: 1

Range 1: 40 to 483 GenBank Graphics V Next Match A Previous Match								
Score	Exp	pect	Method	Identities	Positives	Gaps	Frame	
149 bits	(375) 6e-	-44	Compositional matrix adjus	t. 69/148(47%)	97/148(65%)	1/148(0%)	+1	
Query 1			KSAVTALWGKVNVDEVGGEALGRLL +A+ +LW K+NV+E+G +A+ RLL			60		
Sbjct 4			HTAILSLWKKINVEEIGPQAMRRLL			219		
Query 6	1 VKAHO		TLGAFSDGLAHLDNLKGTFATLSELH			120		
Sbjct 2	Spells Statement	-	MGGLDRAIQNMDDIKNAYRELSVMH			399		
Query 1			QAAYQKVVAGVANALAHKYH 147 Q A+QK + V +AL +YH					
Sbjct 4			QEAWQKFLMAVTSALGRQYH 483					

Query: NP_000509 Program: TBLASTN Database: EST

(nematodes)

Match: novel globin

(b) BLASTX result with a nematode EST showing its closest known protein match is in a vertebrate

RecName: Full=Hemoglobin anodic subunit beta; AltName: Full=Hemoglobin anodic beta chain Sequence ID: splP80946.1|HBBA_ANGAN_Length: 147_Number of Matches: 1

Score		Expect	Method	Identities	Positives	Gaps	Frame
290 bi	ts(742	2) 2e-97	Compositional matrix	adjust. 136/147(93%)	141/147(95%)	0/147(0%)	+1
Query	43			RRLLIVCPWTQRHFANFGNLS RRLLIVCPWTORHFANFGNLS		22	
Sbjct	1			RRLLIVCPWTQRHFANFGNLS		0	
Query	223			SVMHSEKLHVDPDNFRLLSEH SVMHSEKLHVDPDNFRLL+EH		02	
Sbjct	61			SVMHSEKLHVDPDNFRLLAEF		20	
Query	403		EAWQKFLMAVTSALGRQYH EAWQKFLMAVTSAL RQYH	483			
Sbict	121	TEFTADVÖ		147			

Confirmation
Query: nematode EST
Program: BLASTX
Best match: a globin, but
not a previously
annotated globin

"Find-a-gene project"

- The find-a-gene project is meant to be a very focused, specific project to help you understand how to use various BLAST tools (e.g. TBLASTN, BLASTX, BLASTP) and various databases.
- You can start with (almost) any protein, from the organism of your choice, and discover a "novel" gene in another organism that is homologous but has never been annotated before as related to your query. Therefore, you are discovering a new gene.
- You can take your new gene/protein, name it, then search it against databases to confirm it has not been described before.
- You can further perform multiple sequence alignment, phylogeny, and predict its protein structure and its function.