PARSING A FASTQ AND COMPUTING QUALITY STATISTICS

check_fastq.ipynb download fastq from aula.esci.upf (inside reads/unknown_illumina_2024.fastq)

- 1. Use SeqIO.parse to print the first record in the fastq
- 2. What's the read length?
- 3. How many reads are stored in this file?
- 4. Can you print the quality score? (check https://biopython.org/wiki/SeqRecord)
- 5. Plot the mean quality at every position in the reads: start with mean quality at position 1 across all reads, then at position 2, and so on until N (that is the length of the reads).
- 6. Show lines with the mean quality score and the 95% conficence interval (2 s.d.)
- 7. Convert the qualities to error probabilities using the Phred Quality Score equations. Plot them, at which positions is higher? what's the expected error rate of them?
- 8. Practical Assessment: Identify the reads origin. (How would you find out from which genome come these reads? To which species they belong? Please describe the method used and the reliability of the results)

Consulting materials: https://en.wikipedia.org/wiki/FASTQ_format

https://biopython.org/wiki/SegIO

https://biopython.org/wiki/SeqRecord

```
from Bio import SeqIO
```

1. Use SegIO.parse to print the first record in the fastq

```
record = list(SeqIO.parse("example.fastq", "fastq"))
```

1. What's the read length?

```
print(f'The length of the read is: {len(record[0].seq)}')
The length of the read is: 151
```

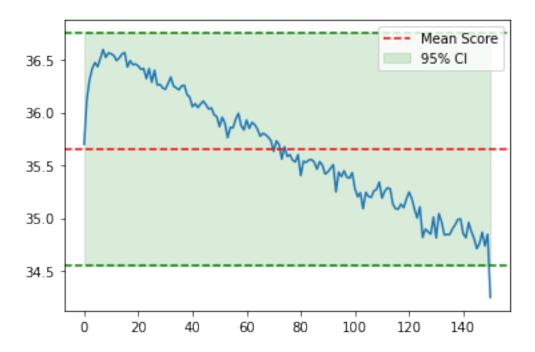
1. How many reads are stored in this file?

```
print(f'There are {len(record)} reads.')
There are 10000 reads.
```

1. Can you print the quality score? (check https://biopython.org/wiki/SeqRecord)

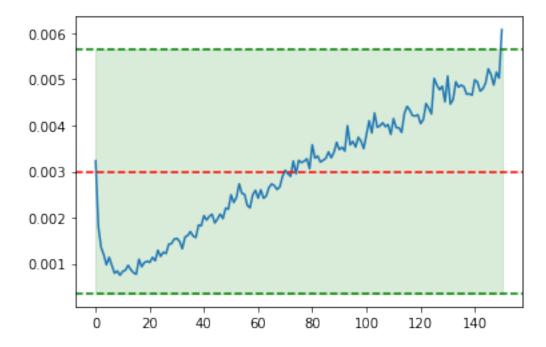
1. Plot the mean quality at every position in the reads: start with mean quality at position 1 across all reads, then at position 2, and so on until N (that is the length of the reads).

```
import matplotlib.pyplot as plt
import numpy as np
max length = max([len(record[num].seq) for num in range(len(record))])
#get maximum length
pos score = [0 \text{ for } x \text{ in } range(max length)]
for x in range(len(record)):
    list = [num for num in
record[x].letter annotations['phred quality']]
    for pos in range(len(record[x])):
        pos score[pos] += list[pos]/10000
mean = np.mean(pos score)
sd = np.std(pos score)
ci = 1.96 * sd
plt.plot(pos score)
plt.axhline(y=mean, color='red', linestyle='--', label='Mean Score')
plt.axhline(y=mean-ci, color='green', linestyle='--')
plt.axhline(y=mean+ci, color='green', linestyle='--')
plt.fill_between(range(max_length), mean-ci, ci+mean, alpha = 0.15,
color = 'g', label = '95% CI')
plt.legend()
<matplotlib.legend.Legend at 0x7fc499382670>
```



1. Convert the qualities to error probabilities using the Phred Quality Score equations. Plot them, at which positions is higher? what's the expected error rate of them?

```
import matplotlib.pyplot as plt
import numpy as np
\max length = \max([len(record[num].seq) for num in range(len(record))])
#get maximum length
error_score = [0 for x in range(max_length)]
for x in range(len(record)):
    list = [num for num in
record[x].letter annotations['phred quality']]
    for pos in range(len(record[x])):
        error score[pos] += (10**(list[pos]/-10))/10000
mean = np.mean(error score)
sd = np.std(error score)
ci = 1.96 * sd
print('mean: ',mean, 'sd: ',sd, 'ci:',ci)
plt.plot(error_score)
plt.axhline(y=mean, color='red', linestyle='--', label='Mean Score')
plt.axhline(y=mean-ci, color='green', linestyle='--')
plt.axhline(y=mean+ci, color='green', linestyle='--')
plt.fill between(range(max length), mean-ci, ci+mean, alpha = 0.15,
color = 'g', label = '95% \overline{CI}')
       0.003000667251991566 sd: 0.0013501553541598577 ci:
mean:
0.002646304494153321
```

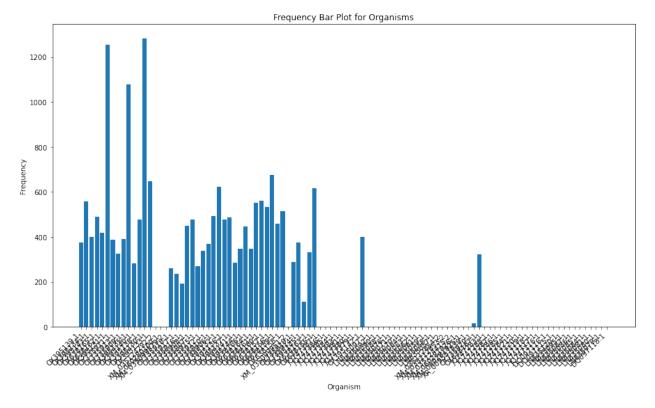


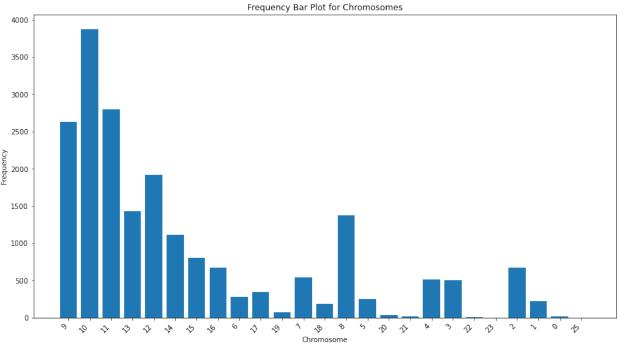
1. Practical Assessment: Identify the reads origin. (How would you find out from which genome come these reads? To which species they belong? Please describe the method used and the reliability of the results)

```
#One sample scenario:
#Check maximum quality score.
def get sample(index, scores, max score):
    if scores[index] == max_score:
        return index
    return get sample(index+1, scores, max score)
sample_position = get_sample(0,pos_score, max(pos_score)) #Get
position in fasta record
seq sample = record[sample position].seq #Get sequence in that
position
temp seq file = "temp sequence.fasta"
with open(temp_seq_file, "w") as f:
    f.write(f">sequence\n{seq sample}")
#makeblastdb -in nt.fasta -dbtype nucl -out nt
blastn command = f"blastn -query temp sequence.fasta -db nt -out
blastn output.xml"
!{blastn_command}
```

```
#USE BLAST+ TO NOT DOWNLOAD THE NT DATABASE.
BLAST Database error: No alias or index file found for nucleotide
database [nt] in search path
[/Users/eloivilellaescolano/ESCI/ASAB/session2::]
top 10 scores = sorted(pos score, reverse=True)[:10]
with open("temp sequence.fasta", "w") as f:
    for top score in top 10 scores:
        sample position = get sample(0, pos score, top score)
        seq sample = record[sample position].seq
        f.write(f">sequence{top score}\n{seq sample}\n")
    # Run blastn
    #blastn command = f"blastn -query {temp seq file} -db nt -remote -
out blastn output {top score}.xml"
    #!{blastn command}
    #USE BLAST+ TO NOT DOWNLOAD THE NT DATABASE.
#We import Blast results to csv and we plot to find an answer
import csv
import matplotlib.pyplot as plt
organism frequency = {}
chromosome frequency = {}
with open('UV52MXC1013-Alignment-HitTable.csv', 'r') as csvfile:
    csv_reader = csv.reader(csvfile, delimiter=',')
    next(csv reader)
    for row in csv reader:
        if row:
            _, organism, _, _, chromosome, _, _, _, _, _, _ = row
            if organism not in organism frequency:
                organism frequency[organism] = 0
            organism frequency[organism] += 1
            if chromosome not in chromosome frequency:
                chromosome frequency[chromosome] = 0
            chromosome frequency[chromosome] += 1
organisms, organism counts = zip(*organism frequency.items())
chromosomes, chromosome counts = zip(*chromosome frequency.items())
plt.figure(figsize=(15, 8))
plt.bar(organisms, organism counts)
```

```
plt.xlabel('Organism')
plt.ylabel('Frequency')
plt.title('Frequency Bar Plot for Organisms')
plt.xticks(rotation=45, ha='right')
plt.show()
plt.figure(figsize=(15, 8))
plt.bar(chromosomes, chromosome counts)
plt.xlabel('Chromosome')
plt.ylabel('Frequency')
plt.title('Frequency Bar Plot for Chromosomes')
plt.xticks(rotation=45, ha='right')
plt.show()
\max \text{ organism} = \max(\text{organism frequency}, \text{ key=organism frequency.get})
max organism count = organism frequency[max organism]
max chromosome = max(chromosome frequency,
key=chromosome frequency.get)
max chromosome count = chromosome frequency[max chromosome]
print(f"Most probable organism: {max organism}, Count:
{max organism count}")
print(f"Most probable chromosome: {max chromosome}, Count:
{max chromosome count}")
print(f'The method used uses the blastn output of the 10 top quality
reads and creates plots to show the most probable genome of those
reads.\n\n To maximize accuracy we use: \n\tMultiple samples\n\
tMaximizing quality ')
```





Most probable organism: 0X395130.1, Count: 1282 Most probable chromosome: 10, Count: 3880

The method used uses the blastn output of the 10 top quality reads and creates plots to show the most probable genome of those reads.

To maximize accuracy we use:
 Multiple samples
 Maximizing quality