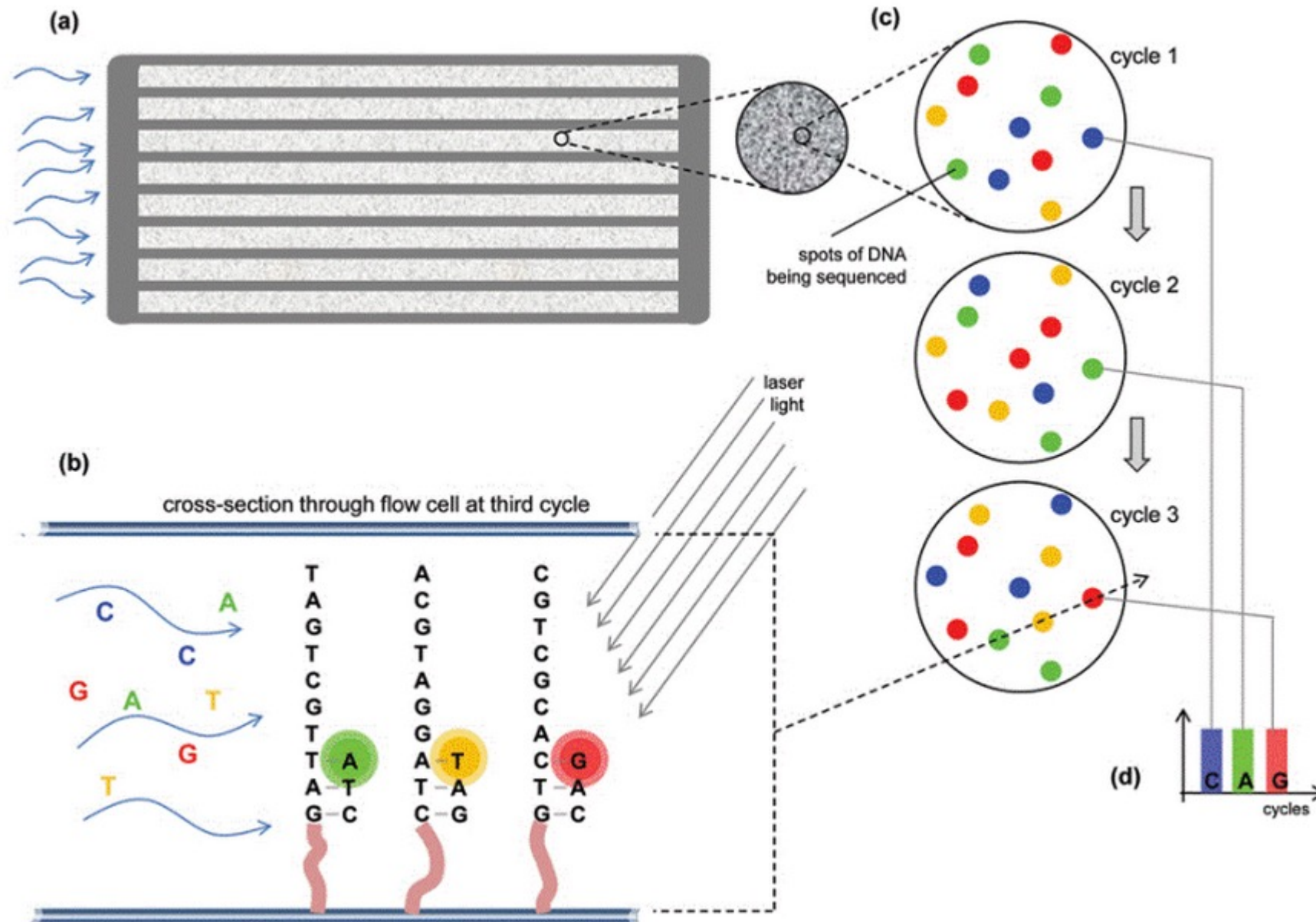


## Sequencing by synthesis



# FASTQ files

Line1: Sequence identifier

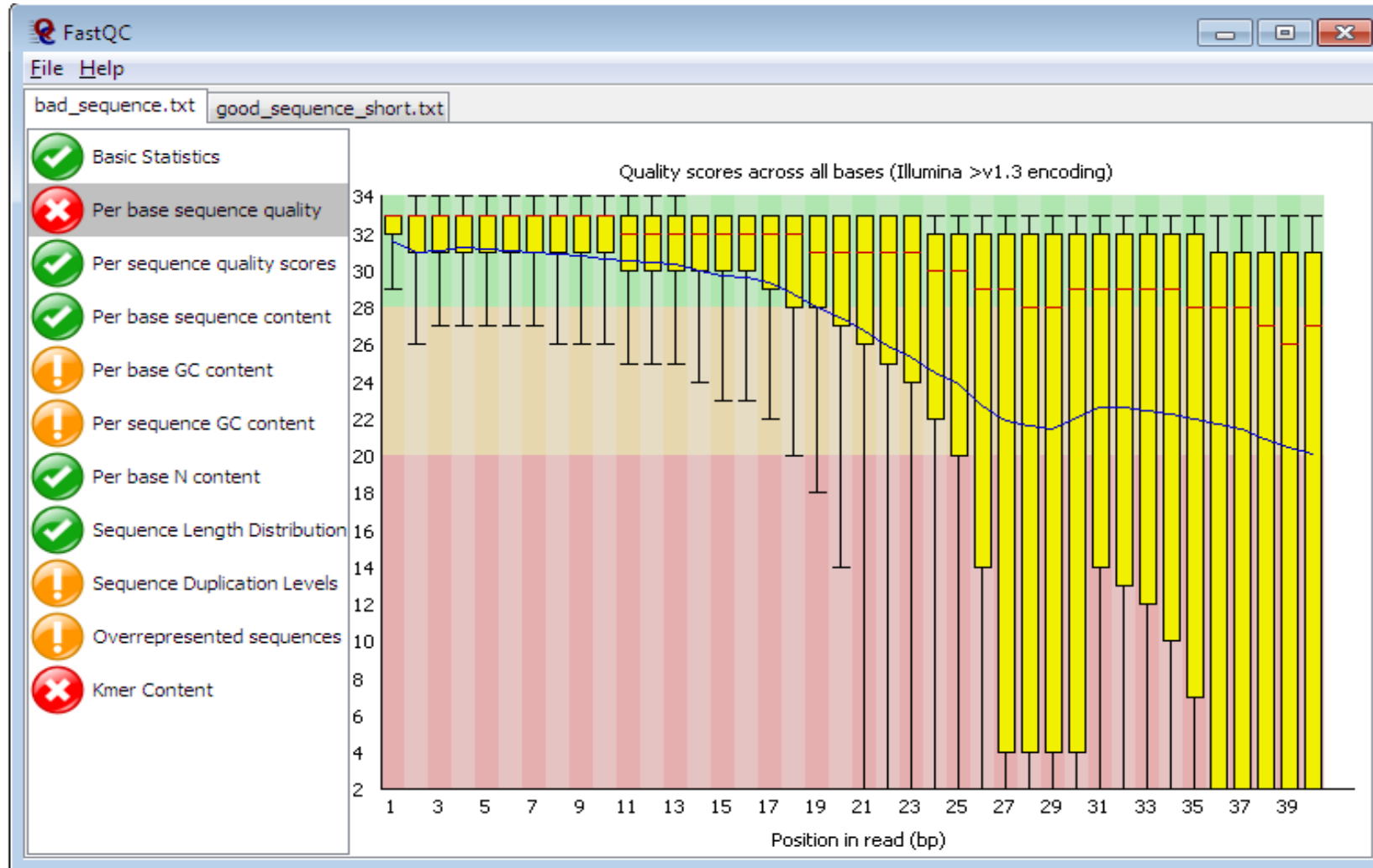
Line2: Raw sequence

Line3: meaningless

Line4: quality values for the sequence

[illegible]

# FastQC



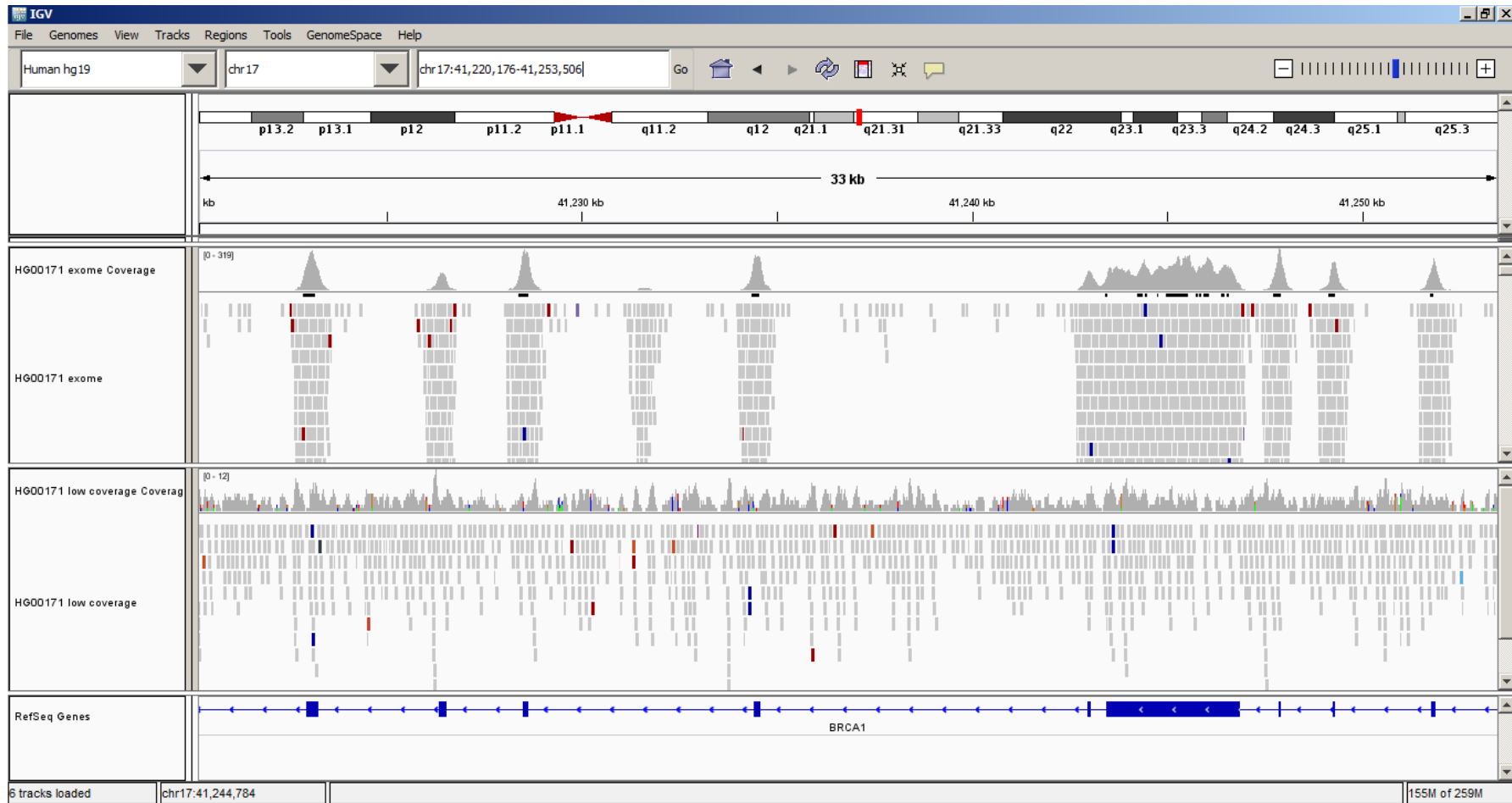
<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>

# SAM format

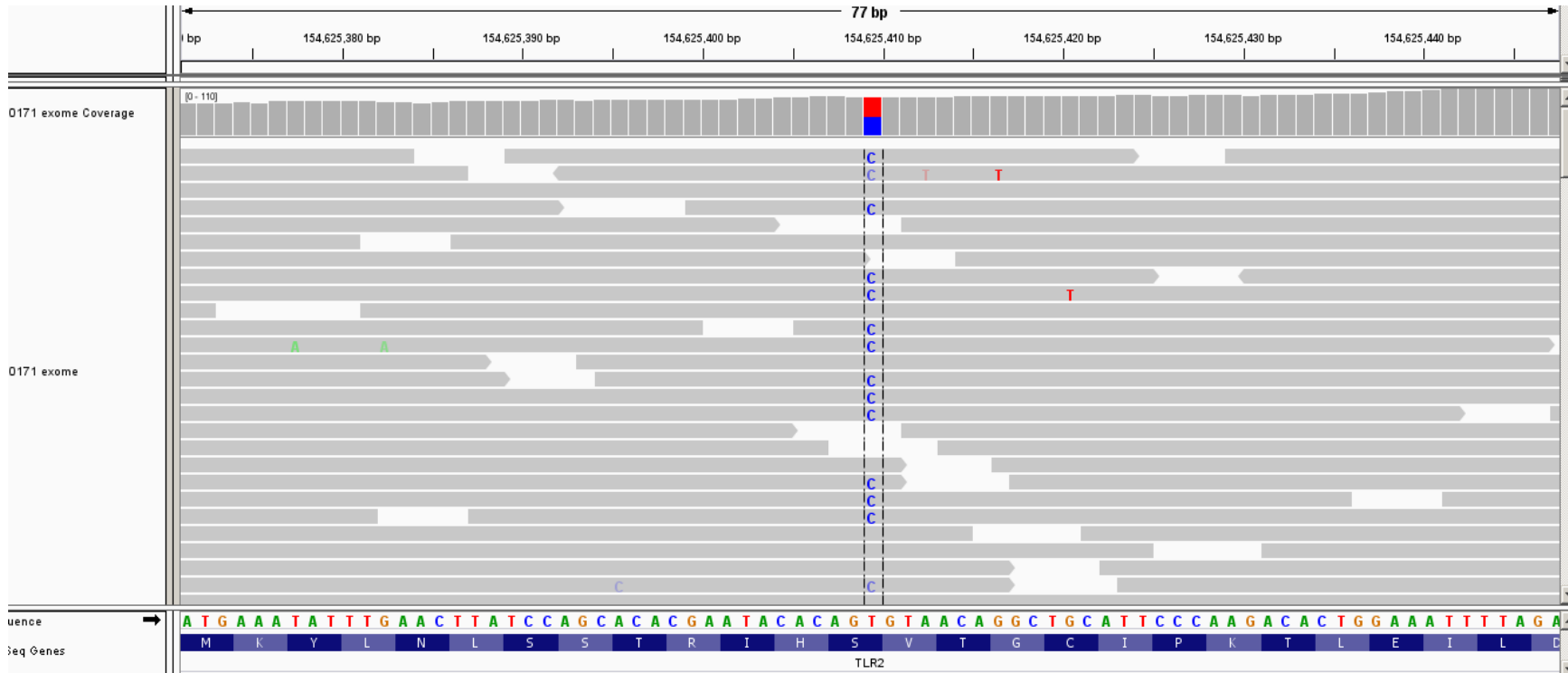
- Standard format for short-read alignments.

```
HWI-ST151_106137860:1:67:20248:73945#0    129      chr17    98508    255      40M      =
      98849    378      AGGGGTTGGCGGGGCAAGGTGGCTCACGCCTGTCATCCCA
      @B@B@:8A?8>@@80DCCDA@85,C7>7>>AB#####
```

## Visualization



## Visualization



# VCF format

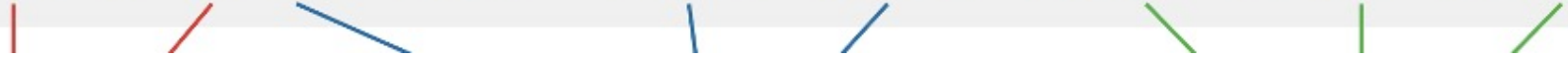
```
#fileformat=VCFv4.0
##FILTER=<ID=ABFilter,Description="AB > 0.75">
##FILTER=<ID=HRunFilter,Description="HRun > 5.0">
##FILTER=<ID=LowQual,Description="Low quality">
##FILTER=<ID=QDFilter,Description="QD < 5.0">
##FILTER=<ID=QUALFilter,Description="QUAL < 30.0">
##FILTER=<ID=SBFilter,Description="SB > -0.10">
##FILTER=<ID=SnpcCluster,Description="SNPs found in clusters">
##FORMAT=<ID=AD,Number=.,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read Depth (only filtered reads used for calling)">
##FORMAT=<ID=GQ,Number=1,Type=Float,Description="Genotype Quality">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=PL,Number=3,Type=Float,Description="Normalized, Phred-scaled likelihoods for AA,AB,BB genotypes where A=ref and B=alt; not applicabl
##INFO=<ID=AB,Number=1,Type=Float,Description="Allele Balance for hets (ref/(ref+alt))">
##INFO=<ID=AC,Number=.,Type=Integer,Description="Allele count in genotypes, for each ALT allele, in the same order as listed">
##INFO=<ID=AF,Number=.,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##INFO=<ID=BaseQRankSum,Number=1,Type=Float,Description="Phred-scaled p-value From Wilcoxon Rank Sum Test of Alt Vs. Ref base qualities">
##INFO=<ID=DB,Number=0,Type=Flag,Description="dbSNP Membership">
##INFO=<ID=DS,Number=0,Type=Flag,Description="Were any of the samples downsampled?">
##INFO=<ID=Dels,Number=1,Type=Float,Description="Fraction of Reads Containing Spanning Deletions">
##INFO=<ID=HRun,Number=1,Type=Integer,Description="Largest Contiguous Homopolymer Run of Variant Allele In Either Direction">
##INFO=<ID=HaplotypeScore,Number=1,Type=Float,Description="Consistency of the site with at most two segregating haplotypes">
##INFO=<ID=MQ,Number=1,Type=Float,Description="RMS Mapping Quality">
##INFO=<ID=MQ0,Number=1,Type=Integer,Description="Total Mapping Quality Zero Reads">
##INFO=<ID=MQRankSum,Number=1,Type=Float,Description="Phred-scaled p-value From Wilcoxon Rank Sum Test of Alt Vs. Ref read mapping qualities">
##INFO=<ID=QD,Number=1,Type=Float,Description="Variant Confidence/Quality by Depth">
##INFO=<ID=ReadPosRankSum,Number=1,Type=Float,Description="Phred-scaled p-value From Wilcoxon Rank Sum Test of Alt Vs. Ref read position bias">
##INFO=<ID=SB,Number=1,Type=Float,Description="Strand Bias">
##UnifiedGenotyper="analysis_type=UnifiedGenotyper input_file=[bamFiles.list] sample_metadata=[] read_buffer_size=null phone_home=STANDARD read_f
##VariantFiltration="analysis_type=VariantFiltration input_file=[] sample_metadata=[] read_buffer_size=null phone_home=STANDARD read_filter=[] in
##GATK_version=1.0.5614
##Reference_Sequence=<url=https://grc-aspera-public:Asppass1@aspera.gs.washington.edu/aspera/user/?B=%2Fhuman_refseq, file=hg19_genome_reference.
##DBSNP_ROD=<url=https://grc-aspera-public:Asppass1@aspera.gs.washington.edu/aspera/user/?B=%2Fdbsnp, file=chr_order.sorted.dbsnp_131_hg19.rod.gz
##Exome_Target=<url=https://grc-aspera-public:Asppass1@aspera.gs.washington.edu/aspera/user/?B=%2Fexome_targets, file=nimblegen_solution_uwrefseq
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT 1796 1797 1798 1799 1800 1801 1802 1803 1804 1
1 69270 . A G 752.59 QDFilter;SBFilter AC=32;AF=0.667;AN=48;BaseQRankSum=53.792;Dels=0.00;HRun=0;HaplotypeScore=
1 69428 rs71245814 T G 2215.91 PASS AB=0.61;AC=4;AF=0.0263;AN=152;BaseQRankSum=200.000;DB;DS;Dels=0.00;HRun=0;Haplotype
1 69511 rs2691305 A G 43693.52 PASS AB=0.61;AC=110;AF=0.7857;AN=140;BaseQRankSum=19.099;DB;DS;Dels=0.00;HRun=
1 69680 . G A 406.53 SBFilter AC=2;AF=0.0149;AN=134;BaseQRankSum=50.877;Dels=0.00;HRun=0;HaplotypeScore=0.3804;
1 69897 rs75758884 T C 340.36 SBFilter AC=22;AF=0.846;AN=26;BaseQRankSum=21.543;DB;Dels=0.00;HRun=1;HaplotypeSco
1 865584 . G A 245.91 PASS AB=0.56;AC=2;AF=0.0105;AN=190;BaseQRankSum=9.661;Dels=0.00;HRun=0;HaplotypeScore=0.3829;M
1 865628 rs41285790 G A 383.75 PASS AB=0.57;AC=1;AF=0.0053;AN=190;BaseQRankSum=43.179;DB;Dels=0.00;HRun=0;HaplotypeSc
1 865694 rs9988179 C T 7122.84 PASS AB=0.53;AC=14;AF=0.0737;AN=190;BaseQRankSum=200.000;DB;Dels=0.00;HRun=0;Haplotype
1 865700 . C T 646.66 PASS AB=0.50;AC=2;AF=0.0105;AN=190;BaseQRankSum=9.945;Dels=0.00;HRun=0;HaplotypeScore=1.3140;M
1 866422 . C T 1045.98 PASS AB=0.59;AC=1;AF=0.0053;AN=190;BaseQRankSum=200.000;DS;Dels=0.00;HRun=1;HaplotypeScore=3.7
1 866438 . G A 2011.24 PASS AB=0.48;AC=1;AF=0.0053;AN=190;BaseQRankSum=3.012;DS;Dels=0.00;HRun=0;HaplotypeScore=2.887
. . . . .
```



## VCF

```
##fileformat=VCFv4.2
##contig=<ID=2,length=51304566>
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
```

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	SAMPLE1	SAMPLE2	SAMPLE3	SAMPLE4	SAMPLE5	SAMPLE6	SAMPLE7
2	81170	.	C	T	.	.	AC=9;AN=7424	GT:DP:GQ	0/0:4:12	0/0:3:9	0/1:1:3	0/1:9:24	1/0:4:12	0/0:5:15	0/0:4:12
2	81171	.	G	A	.	.	AC=6;AN=7446	GT:DP:GQ	0/1:4:12	0/0:3:9	0/0:1:3	0/0:9:24	0/1:4:12	0/1:5:15	0/0:4:12
2	81182	.	A	G	.	.	AC=5;AN=7506	GT:DP:GQ	0/0:5:15	0/0:4:12	0/0:5:15	0/0:9:24	0/0:4:12	0/0:4:12	0/0:4:12
2	81204	.	T	G	.	.	AC=2;AN=7542	GT:DP:GQ	1/0:5:15	0/0:9:27	0/0:10:30	0/0:15:39	0/0:9:27	1/0:13:39	0/1:14:42





## Primary, secondary and tertiary analysis

### Primary analysis

Happens in the instrument, checks during the run for quality control.  
Consists of image alignment, color and quality value calls. **FASTQ files**

### Secondary analysis

A reference sequence is converted into color space and the data are aligned to the reference sequence.  
A consensus sequence may then be constructed from the sequencing reads.  
Comparison of the consensus sequence to a reference genome enables the identification of SNPs and structural variations. **SAM/BAM files**

### Tertiary analysis

Data analysis that takes place after the reads are mapped.  
Visualization of data. Annotation of variants. **VCF files**