
scVI enables batch-effect and noise correction and highlights differences in HPSC transcriptional profiles of Aplastic Anemia patients and healthy donors

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Abstract

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1 Introduction

Aplastic anemia (AA) is a condition in which a patient's bone marrow fails to form enough red blood cells, white blood cells, and platelets, resulting in pancytopenia (Young 2018). Pathophysiological mechanisms of this disease include direct damage to bone marrow (commonly from chemotherapy), germ line loss-of-function mutations that interfere with blood-cell precursor DNA repair pathways, and autoimmune attack (Young 2018).

Current options for treating AA fall within three categories, each of which attempts to address pancytopenia in a different way. The first, bone marrow transplantation, aims to replace failing bone marrow with healthy donor marrow. This is a curative treatment, but is limited by the prevalence of graft-versus-host disease and by its reliance on tissue donors (Young 2018). The second, immunosuppression, aims to eliminate immune-mediated AA. This category includes treatment with antilymphocyte globulin (combined with cyclosporine), which has a mild lymphocyte-depleting effect. This treatment leads to improved blood production in about 66% of patients (Bacigalupo 2017), however 30% to 60% of these patients experience relapse that requires years of continued cyclosporine therapy (Scheinberg et al. 2012). Despite the success of immunosuppressant treatments, the mechanisms by which the immune system damages bone marrow remain unknown; the strongest evidence for immune-mediation of aplastic anemia is the effectiveness of immunosuppressant treatments (Young 2018). The third treatment category, stem-cell stimulation, aims to promote stem-cell regeneration within the patient directly. One such treatment involves administering eltrombopag, a synthetic mimic of thrombopoietin (a hormone that stimulates the production of platelets). This treatment has been shown in limited clinical trials to improve blood production in 80% of treated patients, though the mechanism of this treatment remains unknown (Young 2018).

Despite the severity of this disease, the mechanisms underlying its onset and the treatment options remain obscure due to limitations in experimental techniques. It is thought that T cells may target hematopoietic stem and progenitor cells (HSPCs), which are blood cell precursor cells, in patients with immune-mediated AA (Tonglin et al. 2022). However HSPCs are severely depleted in AA patients, making them a difficult target to study (Zhu et al. 2021). Recent studies like Tonglin et al. (2022) and Zhu et al. (2021) have used single-cell RNA-sequencing (scRNA-seq) to perform differential expression analysis between healthy and AA patient HSPCs. These studies have thus far used PCA to reduce dataset dimensionality, followed by graph-based clustering and likelihood ratio tests to identify significantly differentially expressed genes between clusters (Zhu et al. 2021). These studies have successfully identified differentially expressed genes between healthy and AA HSPCs. However, their accounting for noise and batch effects is limited to their initial dimension reduction

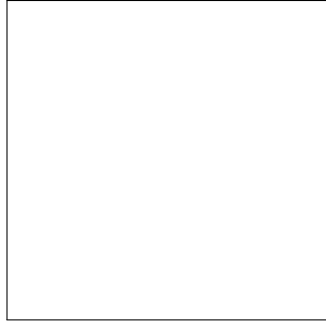


Figure 1: Sample figure caption.

Table 1: Sample table title

Part		
Name	Description	Size (μm)
Dendrite	Input terminal	~ 100
Axon	Output terminal	~ 10
Soma	Cell body	up to 10^6

and this initial dimension reduction assumes a generalized linear model is sufficient for accurately mapping onto the low-dimensional manifold underlying the data.

1.1 Citations within the text

Of note is the command `\citet`, which produces citations appropriate for use in inline text. For example,

```
\citet{giammarco_transplant_2018} investigated\dots
```

produces

Giammarco et al. (2018) investigated...

1.2 Figures

Note that publication-quality tables *do not contain vertical rules*. We strongly suggest the use of the `booktabs` package, which allows for typesetting high-quality, professional tables:

<https://www.ctan.org/pkg/booktabs>

This package was used to typeset Table 1.

1.3 Margins in L^AT_EX

Most of the margin problems come from figures positioned by hand using `\special` or other commands. We suggest using the command `\includegraphics` from the `graphicx` package. Always specify the figure width as a multiple of the line width as in the example below:

```
\usepackage[pdftex]{graphicx} ...
\includegraphics[width=0.8\linewidth]{myfile.pdf}
```

See Section 4.4 in the `graphics` bundle documentation (<http://mirrors.ctan.org/macros/latex/required/graphics/grfguide.pdf>)

A number of width problems arise when L^AT_EX cannot properly hyphenate a line. Please give LaTeX hyphenation hints using the `\-` command when necessary.

Acknowledgments and Disclosure of Funding

References

References follow the acknowledgments. Use unnumbered first-level heading for the references. Any choice of citation style is acceptable as long as you are consistent. It is permissible to reduce the font size to small (9 point) when listing the references. Note that the Reference section does not count towards the page limit.

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A Appendix

Optionally include extra information (complete proofs, additional experiments and plots) in the appendix. This section will often be part of the supplemental material.