Example – Olive Oil data set

Heirarchical Clustering

- > olive<-read.table("olive.txt",h=T)</pre>
- > newolive<-olive[,3:10]

(1) agnes

Let us try first the "single" linkage:

- > library(cluster)
- > x<-daisy(newolive)
- > agn<-agnes(x,metric="euclidean",stand=FALSE,method="single")

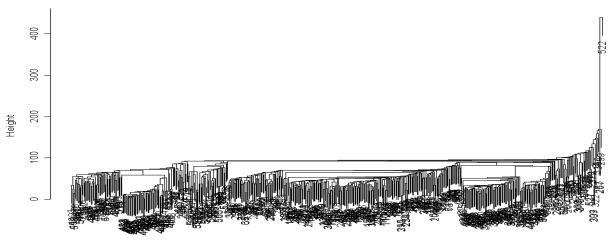
Use the following interactive command for both the "dedrogram" and "banner plot":

> plot(agn,ask=T)

or use the following command for only a dendrogram:

> plot(agn,which.plots=2)





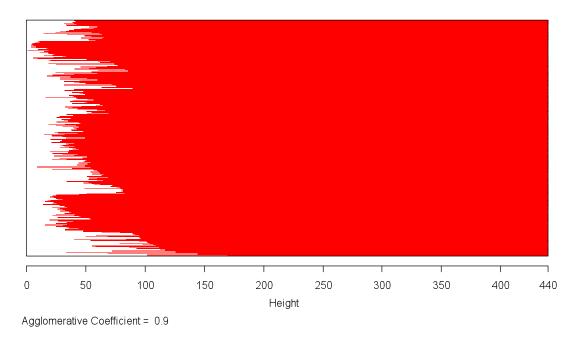
Agglomerative Coefficient = 0.9

If you use

> plot(agn,which.plots=1)

for a "banner plot", you are not able to get a clear plot since we have more than 500 objects (this is just a horizontal version of the dendrogram).

Banner of agnes(x = x, metric = "euclidean", stand = FALSE, method = "single")



However, from the output the AC (Agglomerative Coefficient) is derived to be 0.9. You can also check out the AC (Agglomerative Coefficient) using

> agn\$ac
[1] 0.8980742

This shows a pretty good clustering structure.

Check that if the resulting grouping agrees with the original "Regions":

```
> olive[,1][agn$order]
[556] 3 1 2 3 3 1 2 1 1 1 1 1 1 1 1 1 3
```

Check that if the resulting grouping agrees with the original "Areas":

> olive[,2][agn\$order]

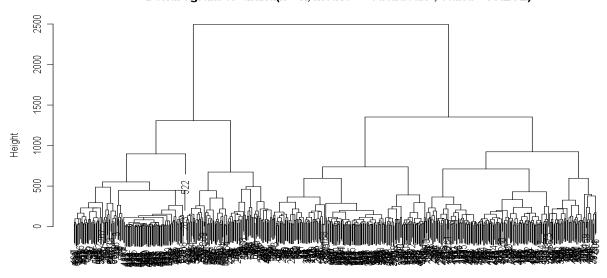
```
[519] 4 3 4 4 4 4 4 4 7 1 8 2 2 2 2 3 3 3 2 7 4 8 3 3 4 7 8 3 3 3 3 3 3 3 3 3 8 4
[556] 8 2 5 8 8 4 6 3 3 3 1 3 3 4 4 4 7
```

Q: How about using other linkages?

(2) Diana – divisive method (splitting method)

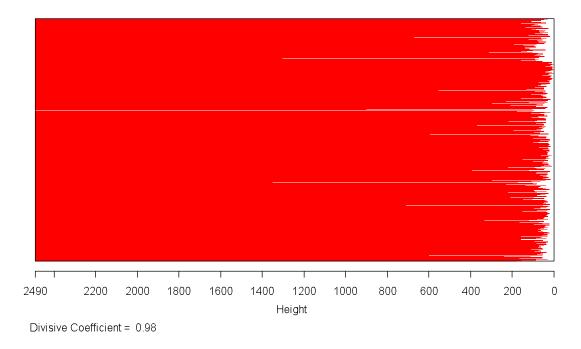
- > di<-diana(x,metric="euclidean",stand=FALSE)
- > print(di)
- > plot(di, which.plots=2)

Dendrogram of diana(x = x, metric = "euclidean", stand = FALSE)



Divisive Coefficient = 0.98

Banner of diana(x = x, metric = "euclidean", stand = FALSE)



You can also check out the DC (Divisive Coefficient) using:

> di\$dc [1] 0.977154

Note that DC=0.977154 shows a pretty strong clustering. Check that if the resulting grouping agrees with the original "Regions":

```
> olive[,1][di$order]
[186] 3 1 1 3 1 1 3 1 3 1 1 1 1 1 1 1 1 3 1 1 1 1 3 3 3 3 3 3 3 3 3 3 3 1 1 1 1 1
```

Check that if the resulting grouping agrees with the original "Areas":

```
> olive[,2][di$order]
[556] 3 3 3 3 3 4 3 3 3 3 3 3 3 3 3 3 3 3
```

(3) mona() – can be used only for binary data!!

Partitioning Method

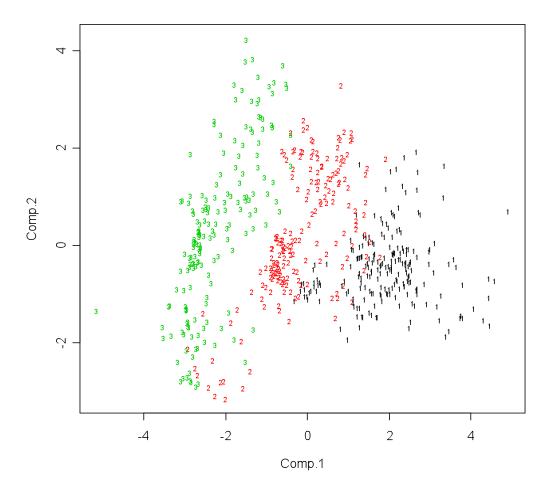
(1) K-means clustering:

Perform K-means clustering considering the number of partitions to be 3:

> km<-kmeans(newolive,3,20)

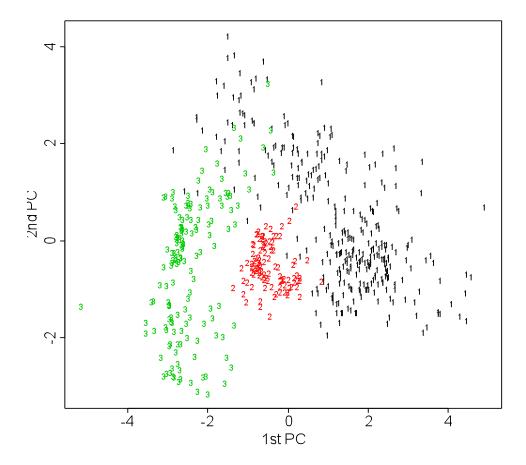
We show the clustering result on the 2-D plane of PC1 vs PC2:

- > pca.newolive<-princomp(scale(newolive,scale=TRUE,center=TRUE),cor=FALSE)
- > pcs.newolive<-predict(pca.newolive)
- > plot(pcs.newolive[,1:2], type="n")
- > text(pcs.newolive,as.character(km\$cluster),col=km\$cluster,cex=0.6)



For comparison, a similar plot can be derived from PCA:

- > plot(pcs.newolive[,1:2],type="n",xlab='1st PC',ylab='2nd PC') > text(pcs.newolive[,1:2],as.character(olive\$Region),col=olive\$Region,cex=0.6)



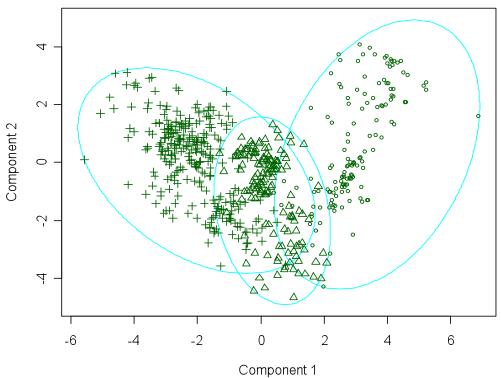
From these two plots, we found that the original regions (shown in PCA) somehow disagree with the K-means clustering, especially on the overlap of "region 1" and "region 2", the overlap of "region 1" and "region 3".

(2) pam

- > pa<-pam(daisy(newolive,stand=T),3,diss=T)
- > plot(pa,ask=T)

The clustering result (which takes a few seconds) is projected on a 2-D PC or MDS space:

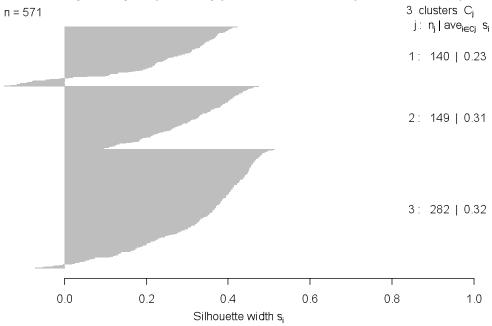
clusplot(pam(x = daisy(newolive, stand = T), k = 3, diss = T))



These two components explain 62.28 % of the point variability.

Plot the silhouette plot:

Silhouette plot of pam(x = daisy(newolive, stand = T), k = 3, diss = T)



Average silhouette width: 0.3

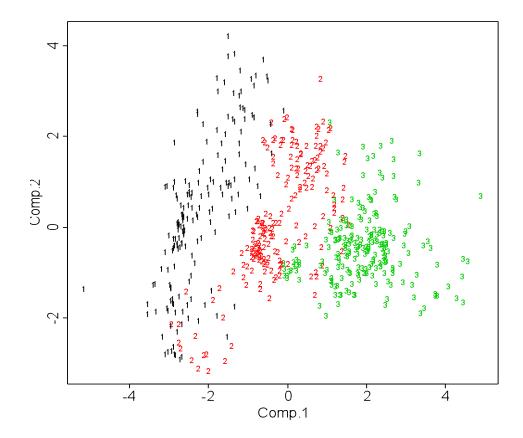
The SC (Silhouette Coefficient) is derived to be 0.3, which shows a weak structure of clustering.

We can use the following command to see if the clustering recovers the original groups of "Regions":

```
> pa$clustering
                                        12
                                           13
                                                14
                                                                 18
                                        32
                                                    35
                                                                 38
                                                                     39
                                                                         40
    22 23 24 25 26 27 28
                            29
                                30
                                    31
                                            33
                                                34
                                                        36
                                                             37
                                         3
                                             2
                                                     2
                                                              2
561 562 563 564 565 566 567 568 569 570 571 572
                              2
```

We can also compare this result with PCA:

- > plot(pcs.newolive[,1:2], type="n")
- > text(pcs.newolive,as.character(pa\$clustering),col=pa\$clustering,cex=0.6)



Q: Does the resulting grouping recover the original groups of "Regions"?

- **Q:** Change the number of clusters, is the result better?
- (3) Try clara(), which is faster for large data.

Self-Organizing Maps (SOM)

- > install.packages('som')
- > library(som)
- > n.newolive<-normalize(newolive) /* Standardize variables*/
- > install.packages('kohonen')
- > library(kohonen)

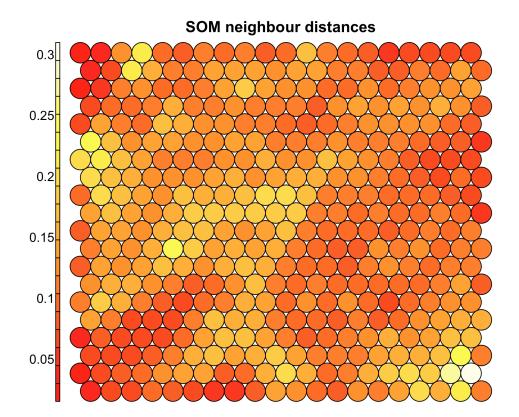
Make a plot with 20x20=400 grids (neurons):

> olive.som<-som(n.newolive,grid = somgrid(20, 20, "hexagonal"))

We first mark the labels of "Region" in the resulting SOM:

> plot(olive.som,type="mapping",labels=olive[,1])

Q: Does the resulting SOM recover the grouping based on "Region"?

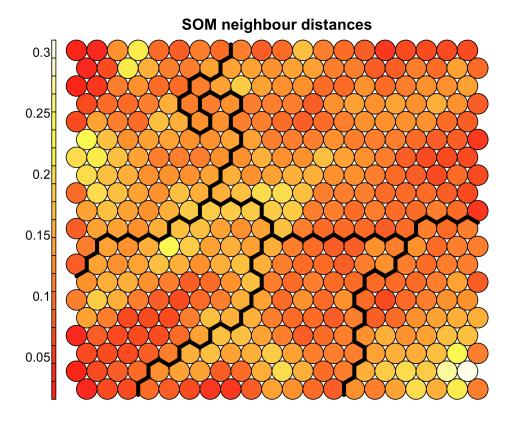


We can use the **agglomerative hierachical clustering** to help us cluster the codebook/weight vectors (the object "olive.som\$codes") and then add the boundaries of clusters in the plot:

> som.hc <- cutree(hclust(dist(olive.som\$codes)), 5)

Here we ask for the display of 5 clusters, which is based on the highest SC of pam(). Adding the boundaries in the plot:

> add.cluster.boundaries(olive.som,som.hc)



Observe the detailed clustering for each object: