class07

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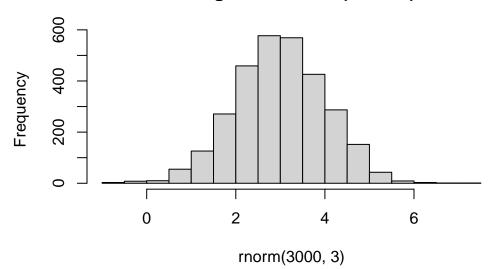
Today we will explore unsupervised machine learning methods including clustering and dimensional reduction methods.

Let's start by making up some data (where we know there are clear groups/clusters) that we can use to test out different clustering methods

we can use the rnorm() function to help us here:

hist(rnorm(3000, 3))

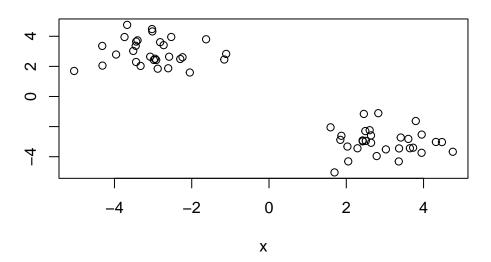
Histogram of rnorm(3000, 3)



Make data with two "clusters"

x [1,] -2.608804 1.873155 [2,] -4.316237 3.358419 [3,] -3.075171 2.642164 [4,] -1.628504 3.798793 [5,] -2.297184 2.492298 [6,] -3.019553 4.319745

plot(z)



How big is **z**?

nrow(z)

[1] 60

```
ncol(z)
```

[1] 2

K-means clustering

The main function in "base" R for K-means clustering is called kmeans()

```
k <- kmeans(z, 2)
k
```

K-means clustering with 2 clusters of sizes 30, 30

Cluster means:

X

1 -3.007059 2.952554

2 2.952554 -3.007059

Clustering vector:

Within cluster sum of squares by cluster:

```
[1] 43.88827 43.88827
```

(between_SS / total_SS = 92.4 %)

Available components:

```
[1] "cluster" "centers" "totss" "withinss" "tot.withinss"
```

[6] "betweenss" "size" "iter" "ifault"

```
attributes(k)
```

\$names

```
[1] "cluster" "centers" "totss" "withinss" "tot.withinss"
```

[6] "betweenss" "size" "iter" "ifault"

\$class

[1] "kmeans"

Q. How many points lie in each cluster?

k\$size

[1] 30 30

Q. What component of our results tells us about the cluster membership (i.e. which point lie in which cluster)?

k\$cluster

Q. Center of each cluster?

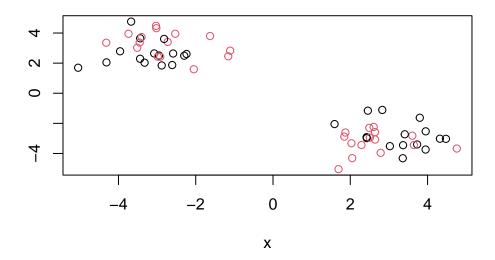
k\$centers

x 1 -3.007059 2.952554 2 2.952554 -3.007059

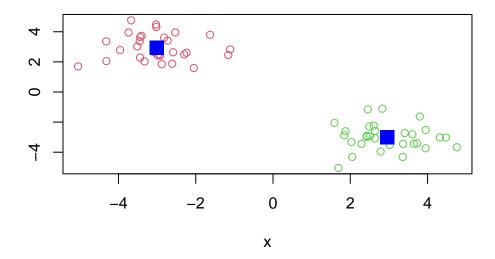
Q. Put this reult info together and make a little "base R" plot of our clustering reult. Also add the cluster center points to this plot.

You can color by number.

$$plot(z, col=c(1,2))$$



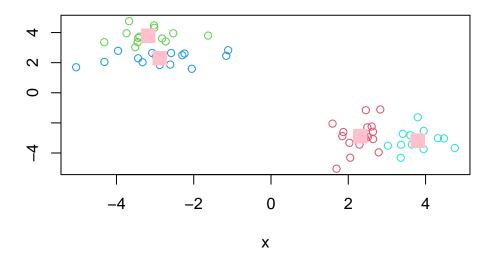
```
plot(z, col = k$cluster + 1)
points(k$centers, col = "blue", pch = 15, cex = 2)
```



Q. Run Kmeans on our input **z** and define 4 clusters making the same result visualization as above (plot of z collored by cluster membership)

```
p <- kmeans(z, 4)

plot(z, col = p$cluster + 1)
points(p$centers, col = "pink", pch = 15, cex = 2)</pre>
```



How to tell if the k is good:

```
p$tot.withinss
```

[1] 53.55179

k\$tot.withinss

[1] 87.77654

Hierarchical Clustering

The main function in base R for this called hclust() it will take as input a distance matrix (key point is that you can't just give your "raw" data as input - you have to first calculate a distance matrix from your data).

```
d <- dist(z)
hc <- hclust(d)
hc</pre>
```

Call:

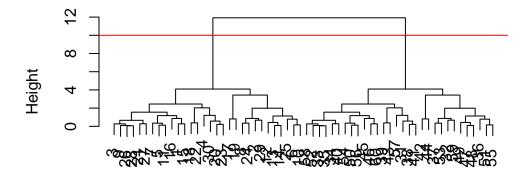
hclust(d = d)

Cluster method : complete
Distance : euclidean

Number of objects: 60

```
plot(hc)
abline(h=10, col = "red")
```

Cluster Dendrogram

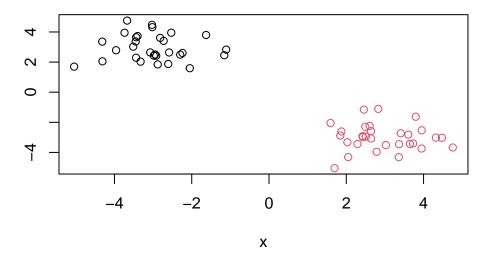


d hclust (*, "complete")

Once I inspect the "tree" I can "cut" the tree to yield my groupings or clusters. The function to do this is called cutree()

```
grps <- cutree(hc, h = 10)
grps</pre>
```

```
plot(z, col=grps)
```



Hands on with Principal Component Analysis (PCA)

Lets do a 17 dimensional data analysis of EU eating habits

Data import

```
url <- "https://tinyurl.com/UK-foods"
x <- read.csv(url, row.names = 1)
head(t(x))</pre>
```

	Cheese	Carcass_	meat	Other	_meat	Fish	Fats_and	_oils	Sugars
England	105		245		685	147		193	156
Wales	103		227		803	160		235	175
Scotland	103		242		750	122		184	147
N.Ireland	66		267		586	93		209	139
	Fresh_p	otatoes	Fresl	n_Veg	Other	_Veg	Processed	d_potat	toes
England		720	1	253		488			198
Wales		874	:	265		570			203
Scotland		566	i	171		418			220
N.Ireland		1033		143		355			187
	Process	sed_Veg	Fresh	fruit	Cerea	als :	Beverages	Soft_d	drinks
England		360		1102	2 :	1472	57		1374
Wales		365		1137	7 :	1582	73		1256
Scotland		337		95	7 :	1462	53		1572
N.Ireland		334		674	l :	1494	47		1506
	Alcohol	lic_drink	s Cor	nfectio	onery				
England		3	75		54				
Wales		4	75		64				
Scotland		4	58		62				
N.Ireland		1	35		41				

Q1. How many rows and columns are in your new data frame named x? What R functions could you use to answer this questions?

nrow(x)

[1] 17

ncol(x)

[1] 4

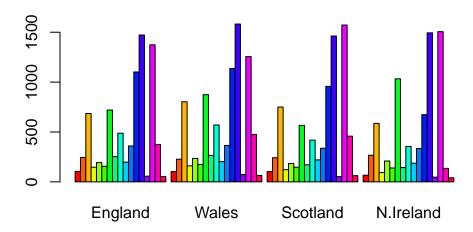
dim(x)

[1] 17 4

Q2. Which approach to solving the 'row-names problem' mentioned above do you prefer and why? Is one approach more robust than another under certain circumstances?

The row.names=1 argument setting is my preferred method of solving this issue. I would say this method is much more robust that using x[-1] every time I need to skip the first row.

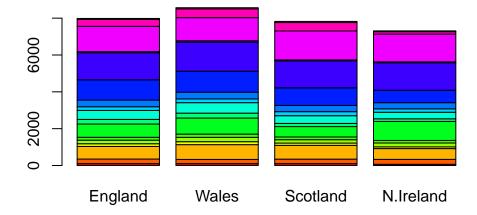
barplot(as.matrix(x), beside=T, col=rainbow(nrow(x)))



Q3: Changing what optional argument in the above barplot() function results in the following plot?

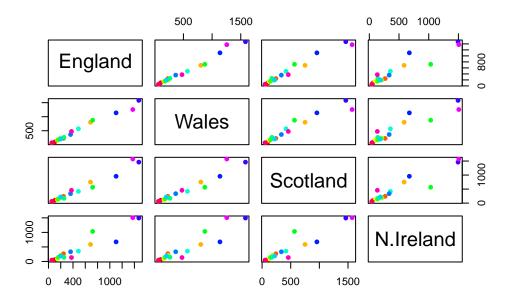
The beside = F argument stacks the bars within each country ontop of each other.

barplot(as.matrix(x), beside=F, col=rainbow(nrow(x)))



Q5: Generating all pairwise plots may help somewhat. Can you make sense of the following code and resulting figure? What does it mean if a given point lies on the diagonal for a given plot?

pairs(x, col=rainbow(nrow(x)), pch=16)



Q6. What is the main differences between N. Ireland and the other countries of the UK in terms of this data-set?

North Ireland seems to consume less fish and more

Looking at these types of "pairwise plots" can be helpful but does not scale well and sucks! There must be a better way...

PCA to the Rescue!

The main function for PCA in base R is called prcomp(). This function wants the transpose of our input data - i.e. the important food categories in columns and the countries as rows.

```
pca <- prcomp(t(x))
summary(pca)</pre>
```

Importance of components:

	PC1	PC2	PC3	PC4
Standard deviation	324.1502	212.7478	73.87622	3.176e-14
Proportion of Variance	0.6744	0.2905	0.03503	0.000e+00
Cumulative Proportion	0.6744	0.9650	1.00000	1.000e+00

Lets see what is in our PCA result object pca

attributes(pca)

```
$names
[1] "sdev" "rotation" "center" "scale" "x"
$class
[1] "prcomp"
```

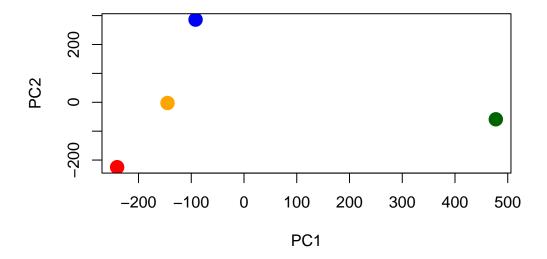
The pca\$x result object is where we will focus first as this details how the countries are related to each other in terms of our new "axis" (a.k.a. "PCs", "eigenvectors", etc.)

head(pca\$x)

```
PC1 PC2 PC3 PC4
England -144.99315 -2.532999 105.768945 -4.894696e-14
Wales -240.52915 -224.646925 -56.475555 5.700024e-13
Scotland -91.86934 286.081786 -44.415495 -7.460785e-13
N.Ireland 477.39164 -58.901862 -4.877895 2.321303e-13
```

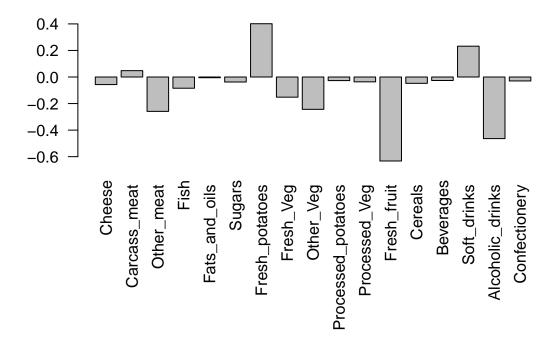
Q7. Complete the code below to generate a plot of PC1 vs PC2. The second line adds text labels over the data points.

Q8. Customize your plot so that the colors of the country names match the colors in our UK and Ireland map and table at start of this document.



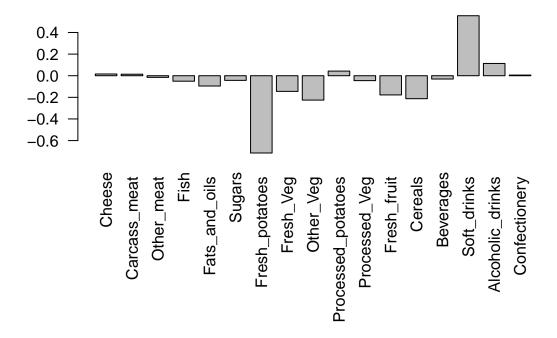
We can look at the so-called PC "loadings" result object to see how the original foods contribute to our new PCs (i.e. how the original variables contribute to our new better variables).

```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,1], las=2 )
```



Q9: Generate a similar 'loadings plot' for PC2. What two food groups feature prominantely and what does PC2 maniply tell us about?

```
par(mar=c(10, 3, 0.35, 0))
barplot( pca$rotation[,2], las=2 )
```



The two food groups that are the most prominently featured are, Fresh_potatoes, and Alcoholic drinks. PC2 is the second principal component which is a measure of directional variance in a perpendicular direction to PC1.

PCA of RNA-seq Data

```
url2 <- "https://tinyurl.com/expression-CSV"
rna.data <- read.csv(url2, row.names=1)
head(rna.data)</pre>
```

```
wt4 wt5 ko1 ko2 ko3 ko4 ko5
       wt1 wt2
                 wt3
       439 458
                 408
                      429 420
                               90
                                    88
                                        86
                                            90
                                                 93
gene1
       219 200
                 204
gene2
                      210 187 427 423 434 433 426
gene3 1006 989
                1030 1017 973 252
                                  237 238 226 210
gene4
       783 792
                 829
                      856 760 849 856 835 885 894
gene5
       181 249
                 204
                      244 225 277 305 272 270 279
       460 502
                 491
                      491 493 612 594 577 618 638
gene6
```

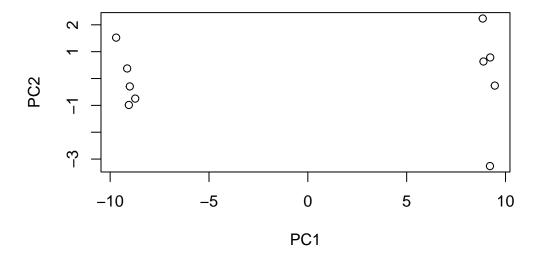
Q10: How many genes and samples are in this data set?

nrow(rna.data)

[1] 100

```
## Again we have to take the transpose of our data
pca <- prcomp(t(rna.data), scale=TRUE)

## Simple un polished plot of pc1 and pc2
plot(pca$x[,1], pca$x[,2], xlab="PC1", ylab="PC2")</pre>
```



```
## Variance captured per PC
pca.var <- pca$sdev^2

## Percent variance is often more informative to look at
pca.var.per <- round(pca.var/sum(pca.var)*100, 1)
pca.var.per</pre>
```

[1] 92.6 2.3 1.1 1.1 0.8 0.7 0.6 0.4 0.4 0.0

