

# Mitochondrial DNA - Cheek Cell Extraction

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## Procedure

1. Pour 10 ml of the saline solution (0.9% NaCl) into mouth and vigorously swish for 1 minute.
2. Expel saline solution into a paper cup.
3. Swirl to mix cells in the cup and transfer 1 ml (1000  $\mu$ l) of the liquid to 1.5 ml tube.
4. Place your sample tube, together with other student samples, in a balanced configuration in a microcentrifuge, and spin for 1.5 minute.
5. Carefully pour off supernatant into paper cup or sink. Be careful not to disturb the cell pellet at the bottom of the test tube. A small amount of saline will remain in the tube.
6. Resuspend cells in remaining saline by pipetting in and out. (If needed, 30  $\mu$ l of saline solution may be added to facilitate resuspension.)
7. Withdraw 30  $\mu$ l of cell suspension, and add to tube containing 100  $\mu$ l of Chelex. Vortex to mix.
8. Boil cell sample for 10 minutes. Use boiling water bath, heat block, or program thermal cycler for 10 minutes at 99°C. Then, cool tube briefly on ice (optional).
9. After boiling, mix tube. Place in a balanced configuration in a microcentrifuge, and spin for 30 sec.

The sample can be stored in the refrigerator until PCR. When using the DNA-containing supernatant, be sure not to touch/transfer any chelex beads with the pipette.