

# Class09\_Structural1

AUTHOR

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## PDB statistics

```
PDBstat <- read.csv("Data_Export_Summary.csv", row.names = 1)
PDBstat
```

	X.ray	EM	NMR	Integrative	Multiple.methods	
Protein (only)	176,204	20,299	12,708	342	218	
Protein/Oligosaccharide	10,279	3,385	34	8	11	
Protein/NA	9,007	5,897	287	24	7	
Nucleic acid (only)	3,066	200	1,553	2	15	
Other	173	13	33	3	0	
Oligosaccharide (only)	11	0	6	0	1	
	Neutron	Other	Total			
Protein (only)	83	32	209,886			
Protein/Oligosaccharide	1	0	13,718			
Protein/NA	0	0	15,222			
Nucleic acid (only)	3	1	4,840			
Other	0	0	222			
Oligosaccharide (only)	0	4	22			

```
# Convert numeric-looking columns from character to numeric
PDBstat[] <- lapply(PDBstat, function(x) as.numeric(gsub(","," ",x)))

# Check structure
str(PDBstat)
```

```
'data.frame':   6 obs. of  8 variables:
 $ X.ray      : num  176204 10279 9007 3066 173 ...
 $ EM         : num  20299 3385 5897 200 13 ...
 $ NMR        : num  12708 34 287 1553 33 ...
 $ Integrative: num  342 8 24 2 3 0
 $ Multiple.methods: num  218 11 7 15 0 1
 $ Neutron     : num  83 1 0 3 0 0
 $ Other       : num  32 0 0 1 0 4
 $ Total       : num  209886 13718 15222 4840 222 ...
```

```
(sum(PDBstat$X.ray) + sum(PDBstat$EM))/sum(PDBstat$Total) * 100
```

[1] 93.69604

Q1. 93.7% of the structures in the PDB were determined by X-ray and EM.

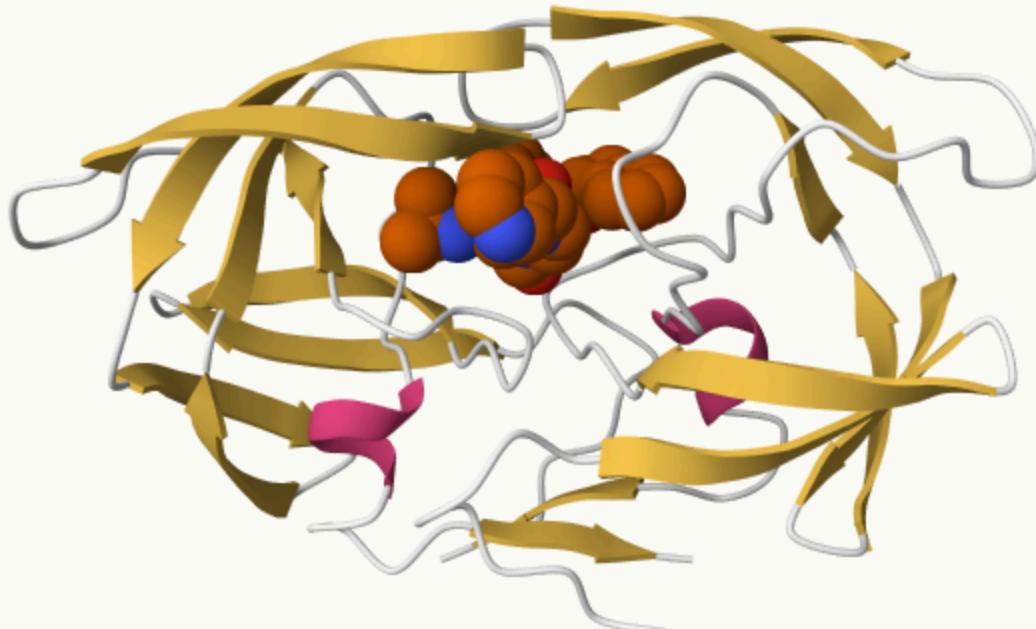
```
sum(PDBstat[c("Protein (only)", "Protein/Oligosaccharide", "Protein/NA"), "Total"])/sum(P
```

[1] 97.91562

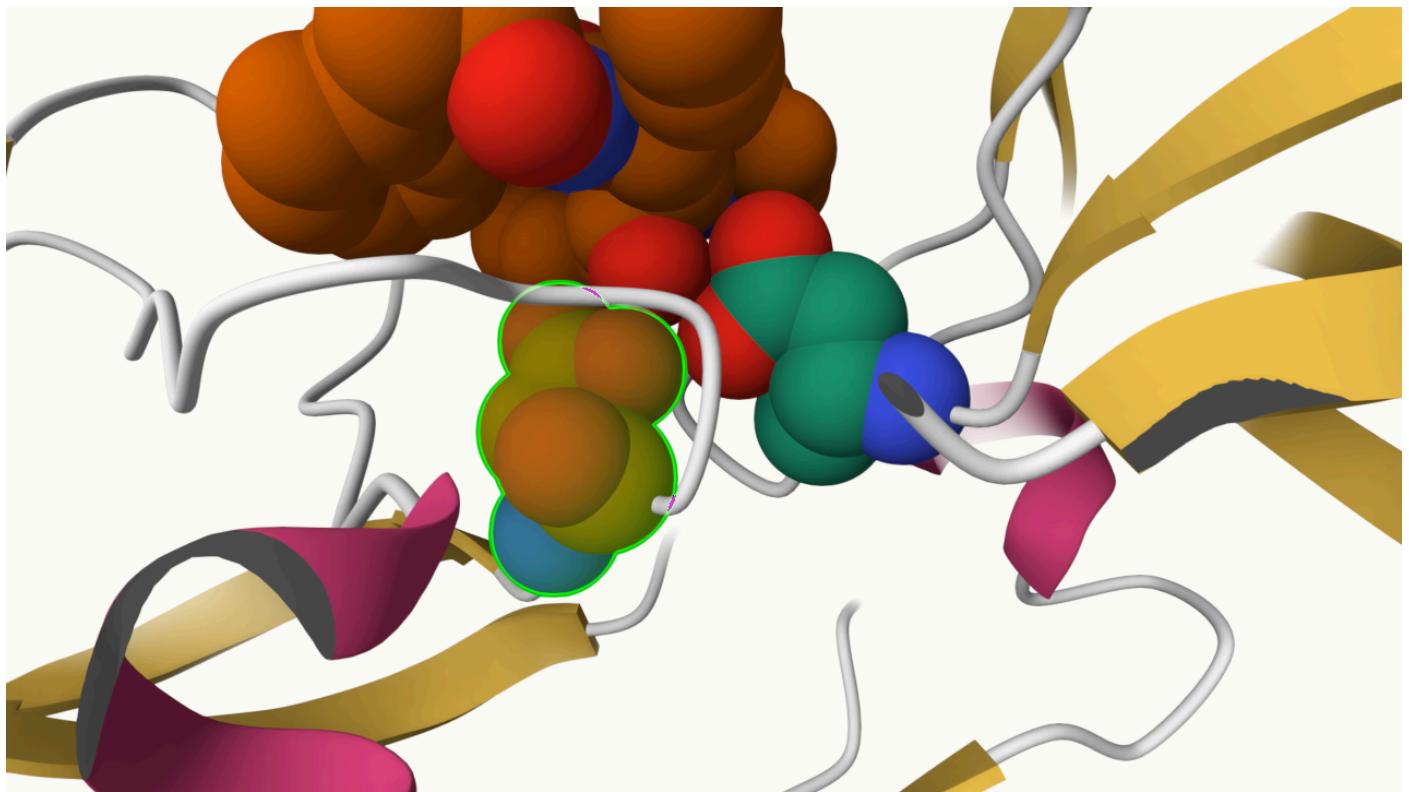
Q2. 97.9% of the structures in the PDB contain protein.

Q3. HIV currently has 4865 structure files in PDB, HIV-1 protease has 1150 structure files in PDB, which is about 23.6% of the total HIV structure files.

```
knitr::include_graphics("1HSG.png")
```



```
knitr::include_graphics("1HSG (1).png")
```

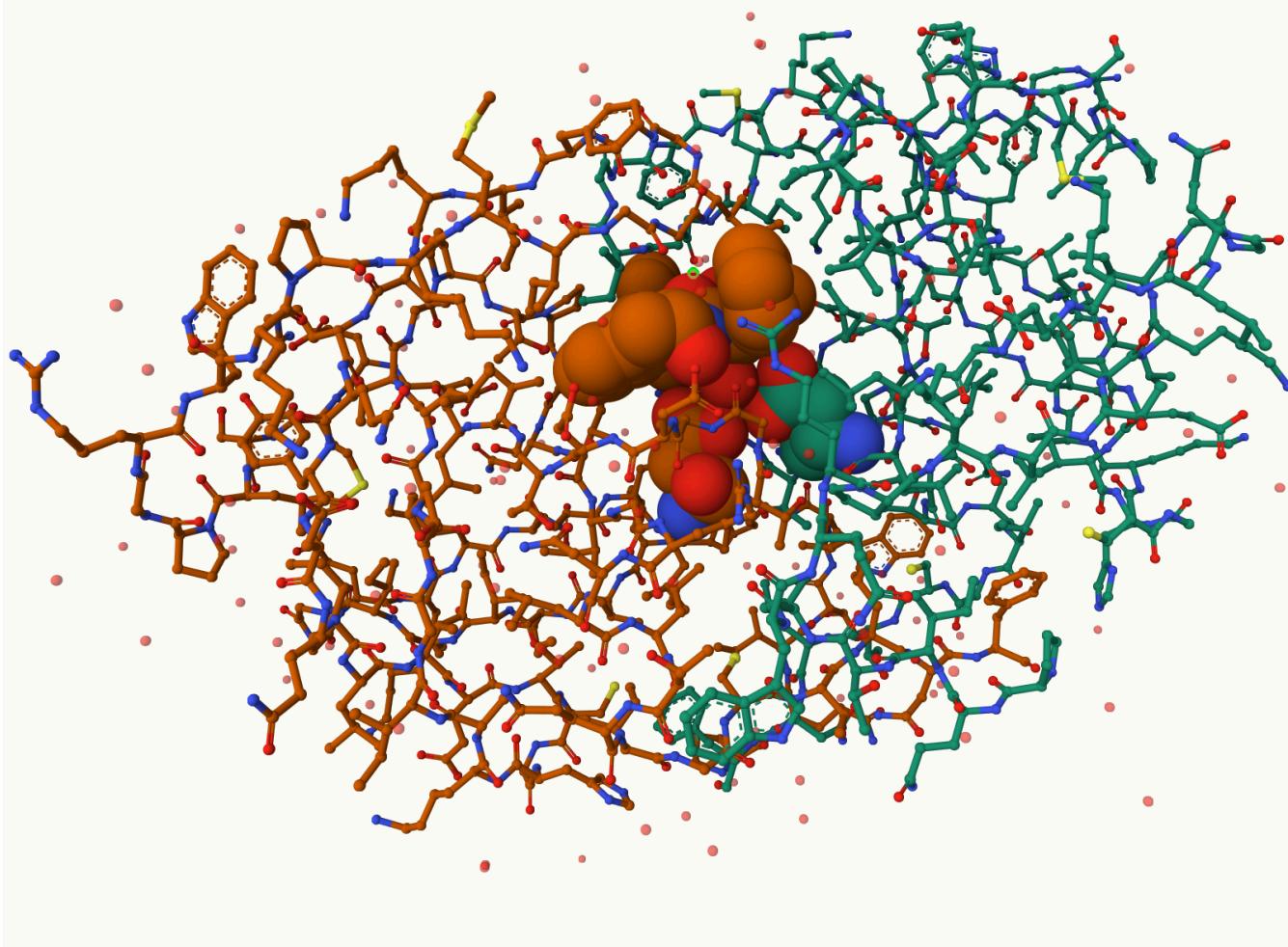


Q4. We can only see the oxygen atom from a water molecule because of the resolution of the structure. Hydrogen atom is not visible at this resolution.

Q5. HOH 308

Q6.

```
knitr::include_graphics("1HSG (2).png")
```



## Introduction to Bio3D in R

```
library(bio3d)
pdb <- read.pdb("1HSG")
```

Note: Accessing on-line PDB file

```
 pdb
```

```
Call: read.pdb(file = "1HSG")
```

```
Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

Non-protein/nucleic Atoms#: 172 (residues: 128)  
 Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPKMIGGGIGGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

+ attr: atom, xyz, seqres, helix, sheet,  
 calpha, remark, call

Q7. 198 amino acid residues

Q8. HOH and MK1

Q9. 2 protein chains

**attributes(pdb)**

```
$names
[1] "atom"    "xyz"     "seqres"   "helix"    "sheet"    "calpha"   "remark"   "call"
```

```
$class
[1] "pdb" "sse"
```

**head(pdb\$atom)**

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40
		segid	elesy	charge									
1	<NA>		N	<NA>									
2	<NA>		C	<NA>									
3	<NA>		C	<NA>									
4	<NA>		O	<NA>									
5	<NA>		C	<NA>									
6	<NA>		C	<NA>									

**adk <- read.pdb("6S36")**

Note: Accessing on-line PDB file  
 PDB has ALT records, taking A only, rm.alt=TRUE

`adk`

```
Call: read.pdb(file = "6S36")
```

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIMDAGKLVT  
DELVIALVKERIAQEDCRNGFLDGFPRTIPQADAMKEAGINVYVLEFDVPDELIVDKI  
VGRRVHAPSGRKYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKAEAGNTKYAKVDGTPVAEVRADLEKILG
```

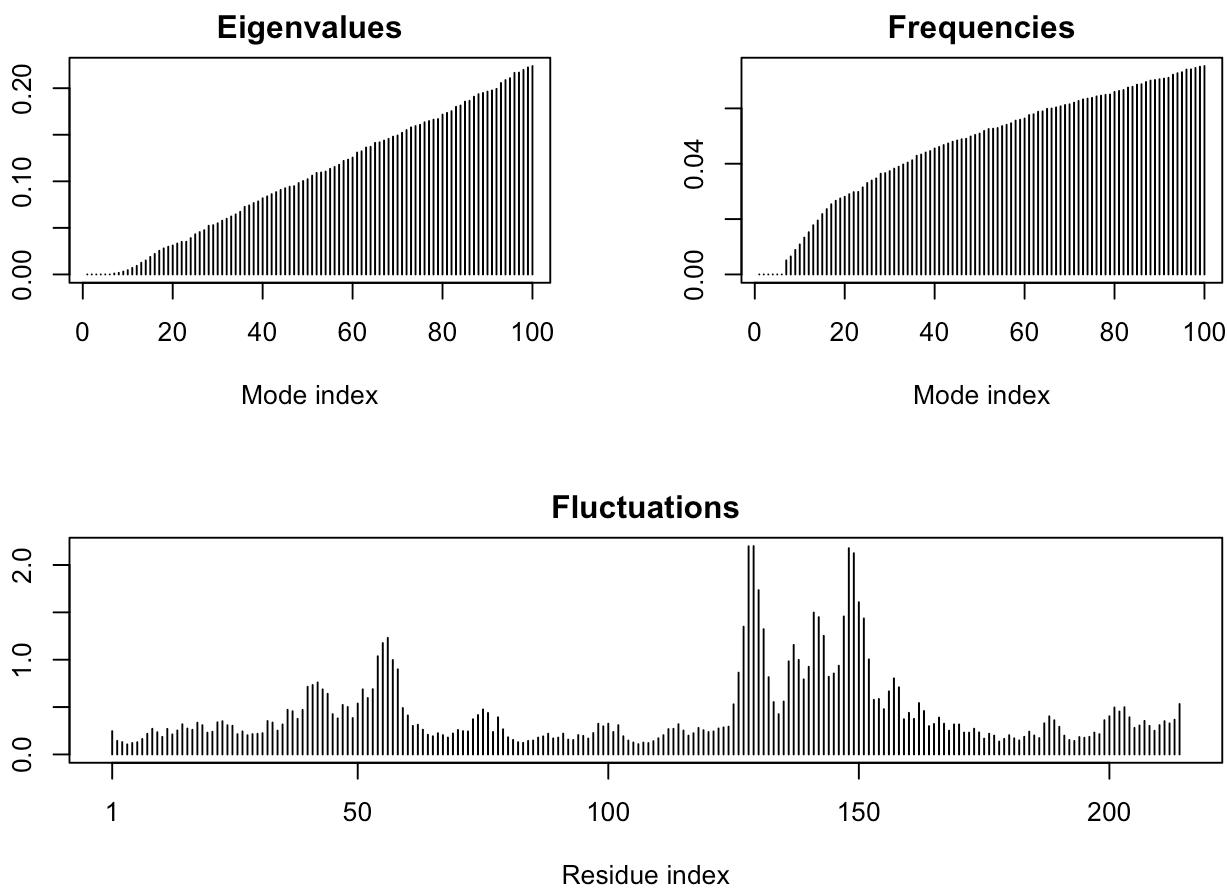
```
+ attr: atom, xyz, seqres, helix, sheet,  
       calpha, remark, call
```

`m <- nma(adk)`

```
Building Hessian... Done in 0.012 seconds.
```

```
Diagonalizing Hessian... Done in 0.263 seconds.
```

`plot(m)`



```
mktrj(m, file="adk_m7.pdb")
```

## Comparative struture analysis of Adenylate Kinase

Q10. msa

Q11. bio3d-view

Q12. True

```
library(bio3d)
aa <- get.seq("1ake_A")
```

Warning in get.seq("1ake\_A"): Removing existing file: seqs.fasta

Fetching... Please wait. Done.

aa

	1	.	.	.	.	.	.	60
pdb   1AKE   A	MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIDMAGKLVT							
	1	.	.	.	.	.	.	60
	61	.	.	.	.	.	.	120
pdb   1AKE   A	DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVVDYVLEFDVPDELIVDRI							
	61	.	.	.	.	.	.	120
	121	.	.	.	.	.	.	180
pdb   1AKE   A	VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG							
	121	.	.	.	.	.	.	180
	181	.	.	.	214			
pdb   1AKE   A	YYSKAEAGNTKYAKVDGTPVAEVRADLEKILG							
	181	.	.	.	214			

Call:

```
read.fasta(file = outfile)
```

Class:

fasta

## Alignment dimensions:

1 sequence rows; 214 position columns (214 non-gap, 0 gap)

+ attr: id, ali, call

Q13. 214 amino acid residues

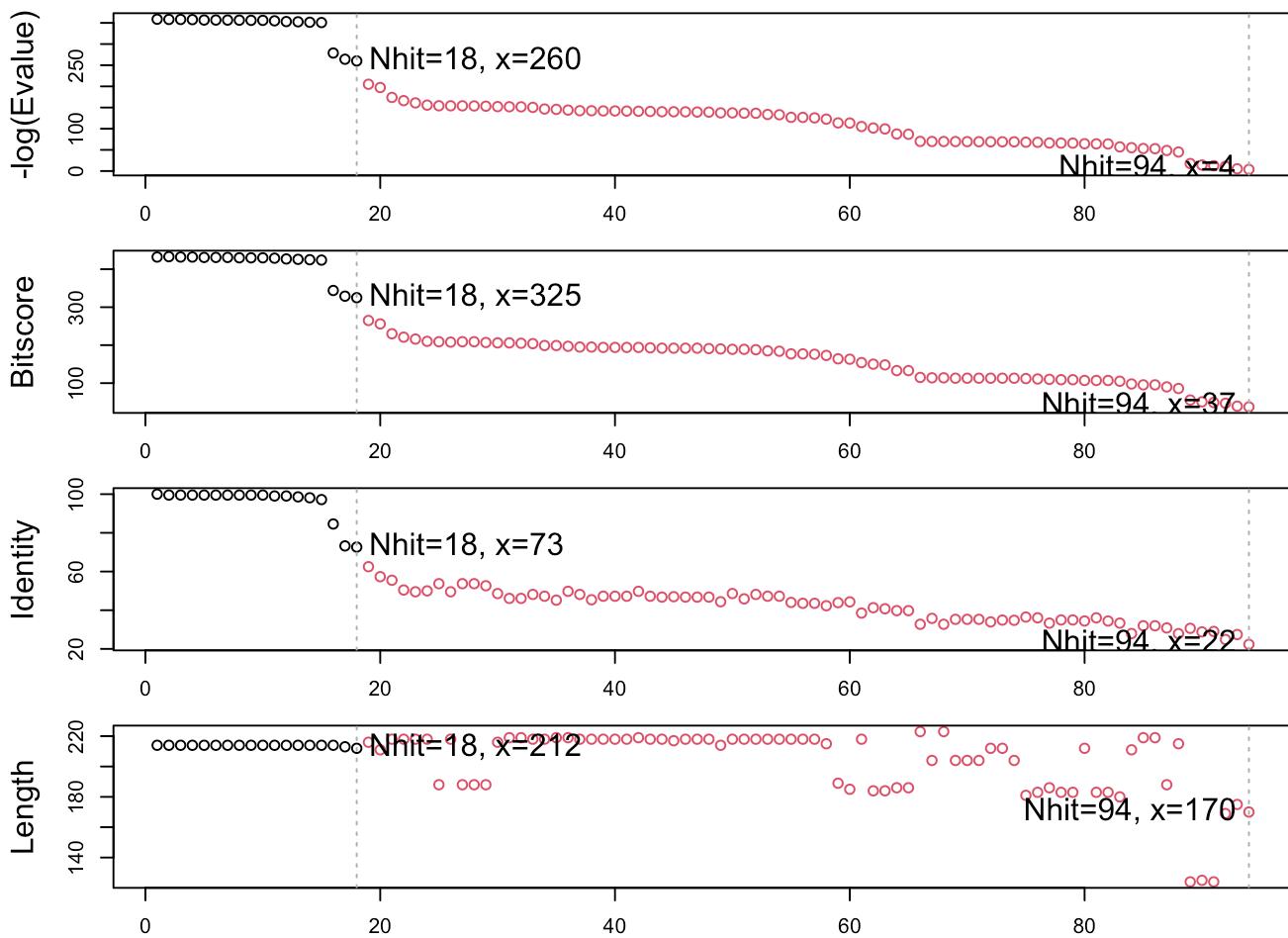
```
# Blast or hmmer search  
b <- blast.pdb(aa)
```

Searching ... please wait (updates every 5 seconds) RID = FZ968N0M015

Reporting 94 hits

```
#Plot a summary of search results  
hits <- plot(b)
```

```
* Possible cutoff values:    260 3  
                           Yielding Nhits: 18 94  
  
* Chosen cutoff value of:   260  
                           Yielding Nhits: 18
```



```
# List out some 'top hits'
head(hits$pdb.id)
```

```
[1] "1AKE_A" "8BQF_A" "4X8M_A" "6S36_A" "8Q2B_A" "8RJ9_A"
```

```
# Download related PDB files
files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1AKE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6S36.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6RZE.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3HPR.pdb.gz exists. Skipping download
```

```
Warning in get.pdb(hits$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4V.pdb.gz exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/5EJE.pdb.gz exists. Skipping download

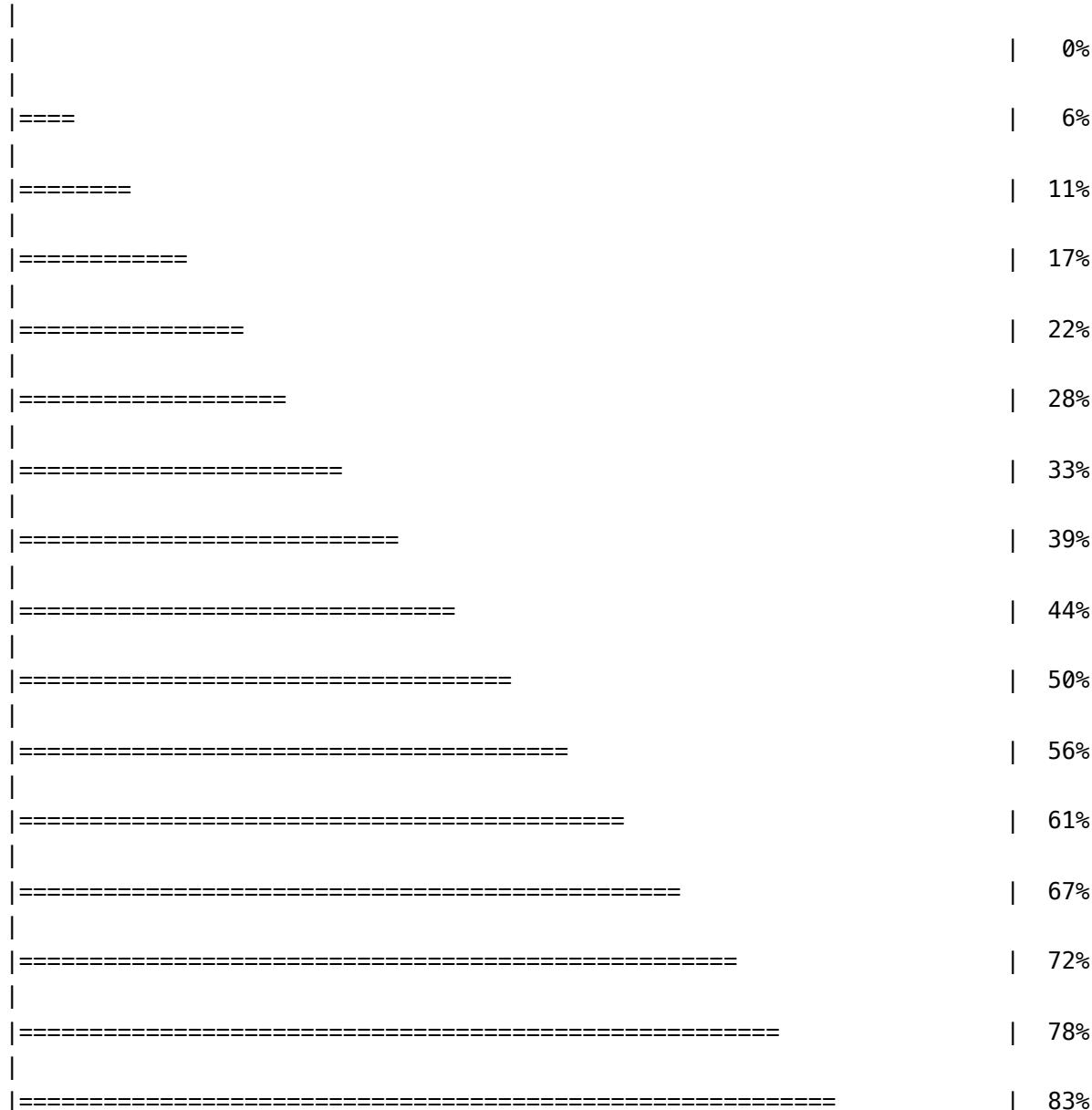
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/1E4Y.pdb.gz exists. Skipping download

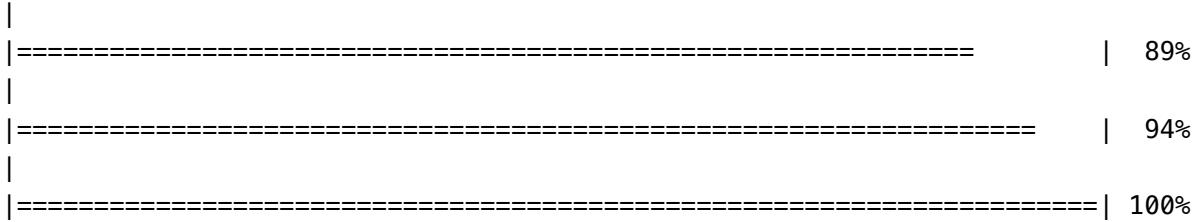
Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/3X2S.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/6HAP.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):  
pdbs/4K46.pdb.gz exists. Skipping download





```
# Align releted PDBs
pdbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdbs/split_chain/1AKE_A.pdb
pdbs/split_chain/8BQF_A.pdb
pdbs/split_chain/4X8M_A.pdb
pdbs/split_chain/6S36_A.pdb
pdbs/split_chain/8Q2B_A.pdb
pdbs/split_chain/8RJ9_A.pdb
pdbs/split_chain/6RZE_A.pdb
pdbs/split_chain/4X8H_A.pdb
pdbs/split_chain/3HPR_A.pdb
pdbs/split_chain/1E4V_A.pdb
pdbs/split_chain/5EJE_A.pdb
pdbs/split_chain/1E4Y_A.pdb
pdbs/split_chain/3X2S_A.pdb
pdbs/split_chain/6HAP_A.pdb
pdbs/split_chain/6HAM_A.pdb
pdbs/split_chain/8PVW_A.pdb
pdbs/split_chain/4K46_A.pdb
pdbs/split_chain/4NP6_A.pdb
```

```
PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..
..
```

Extracting sequences

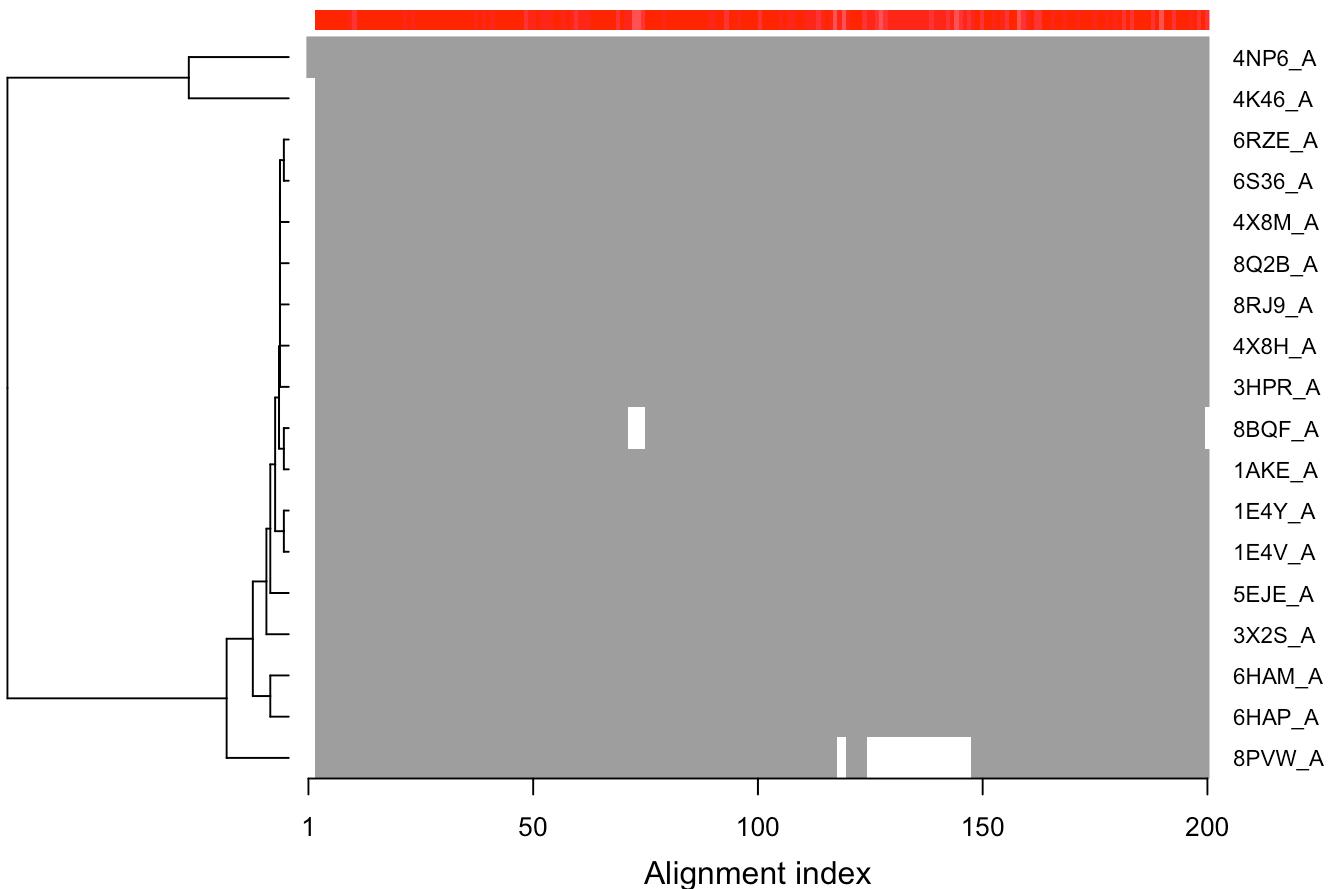
```
pdb/seq: 1  name: pdbs/split_chain/1AKE_A.pdb
          PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2  name: pdbs/split_chain/8BQF_A.pdb
          PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 3  name: pdbs/split_chain/4X8M_A.pdb
pdb/seq: 4  name: pdbs/split_chain/6S36_A.pdb
```

```
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5    name: pdbs/split_chain/8Q2B_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 6    name: pdbs/split_chain/8RJ9_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 7    name: pdbs/split_chain/6RZE_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 8    name: pdbs/split_chain/4X8H_A.pdb
pdb/seq: 9    name: pdbs/split_chain/3HPR_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 10   name: pdbs/split_chain/1E4V_A.pdb
pdb/seq: 11   name: pdbs/split_chain/5EJE_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 12   name: pdbs/split_chain/1E4Y_A.pdb
pdb/seq: 13   name: pdbs/split_chain/3X2S_A.pdb
pdb/seq: 14   name: pdbs/split_chain/6HAP_A.pdb
pdb/seq: 15   name: pdbs/split_chain/6HAM_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 16   name: pdbs/split_chain/8PVW_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 17   name: pdbs/split_chain/4K46_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 18   name: pdbs/split_chain/4NP6_A.pdb
```

```
# Vector containing PDB codes for figure axis
ids <- basename.pdb(pdfs$id)

# Draw schematic alignment
plot(pdfs, labels=ids)
```

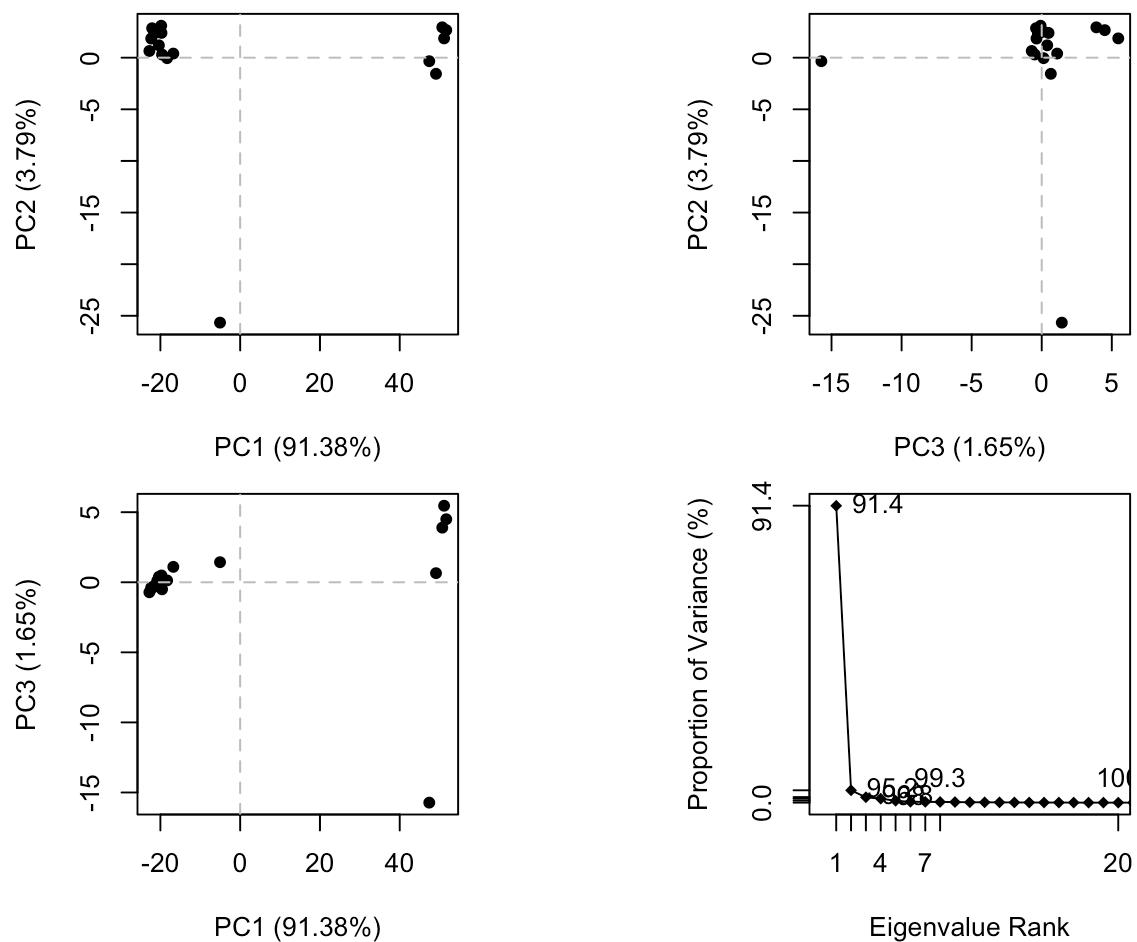
## Sequence Alignment Overview



```
anno <- pdb.annotate(ids)
unique(anno$source)
```

```
[1] "Escherichia coli"
[2] "Escherichia coli K-12"
[3] "Escherichia coli 0139:H28 str. E24377A"
[4] "Escherichia coli str. K-12 substr. MDS42"
[5] "Photobacterium profundum"
[6] "Vibrio cholerae O1 biovar El Tor str. N16961"
```

```
# Perform PCA
pc.xray <- pca(pdbs)
plot(pc.xray)
```

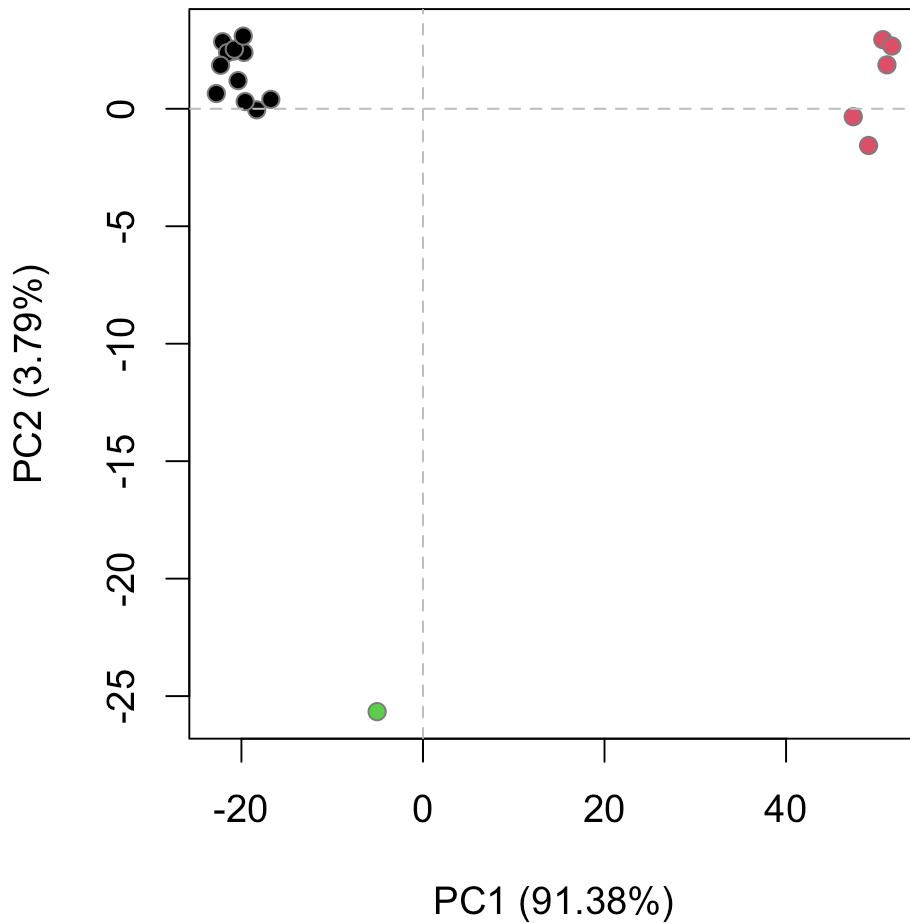


```
# Calculate RMSD
rd <- rmsd(pdbs)
```

Warning in rmsd(pdbs): No indices provided, using the 182 non NA positions

```
# Structure-based clustering
hc.rd <- hclust(dist(rd))
grps.rd <- cutree(hc.rd, k=3)

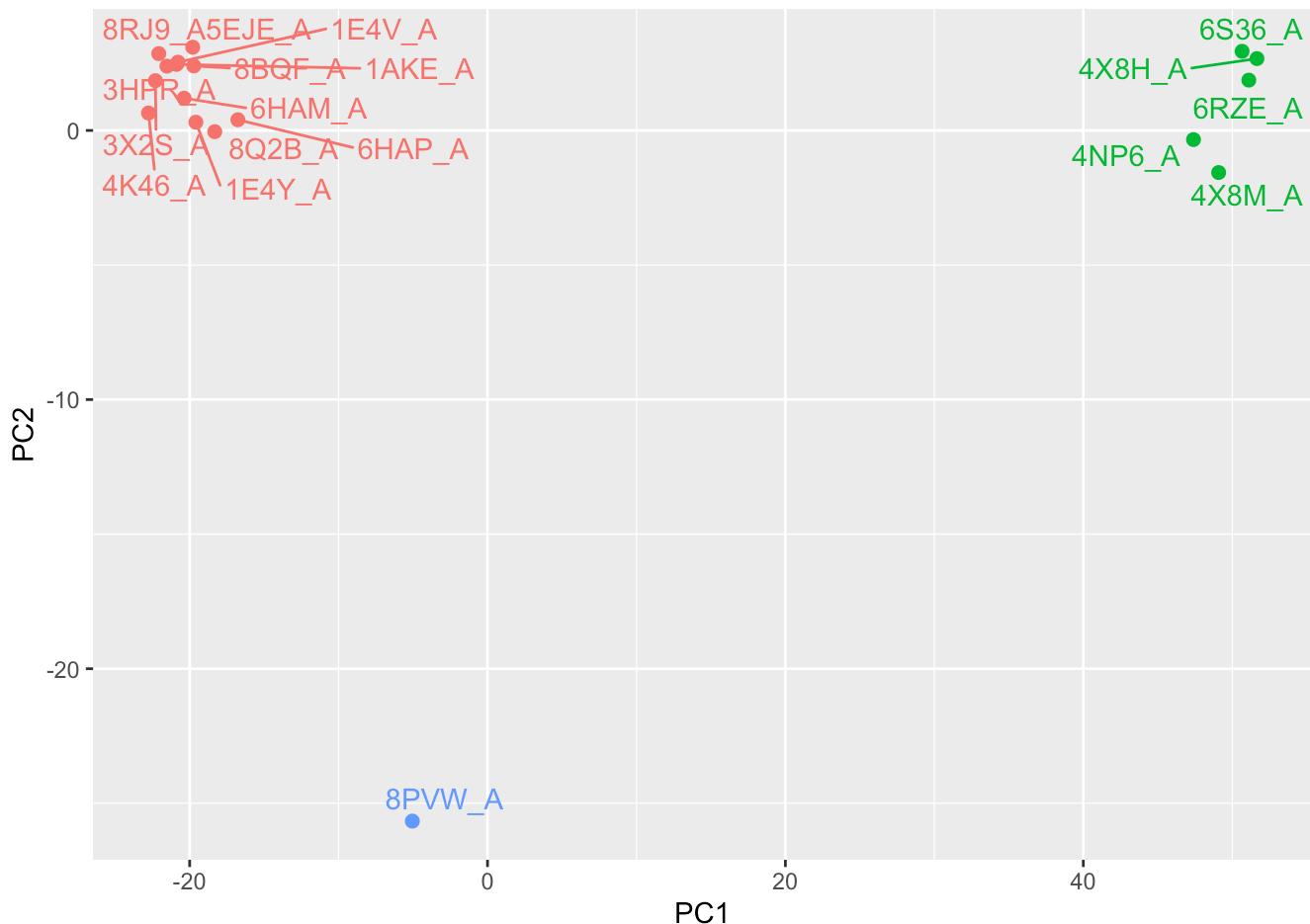
plot(pc.xray, 1:2, col="grey50", bg=grps.rd, pch=21, cex=1)
```



```
#Plotting results with ggplot2
library(ggplot2)
library(ggrepel)

df <- data.frame(PC1=pc.xray$z[,1],
                  PC2=pc.xray$z[,2],
                  col=as.factor(grps.rd),
                  ids=ids)

p <- ggplot(df) +
  aes(PC1, PC2, col=col, label=ids) +
  geom_point(size=2) +
  geom_text_repel(max.overlaps = 20) +
  theme(legend.position = "none")
p
```



## Normal mode analysis

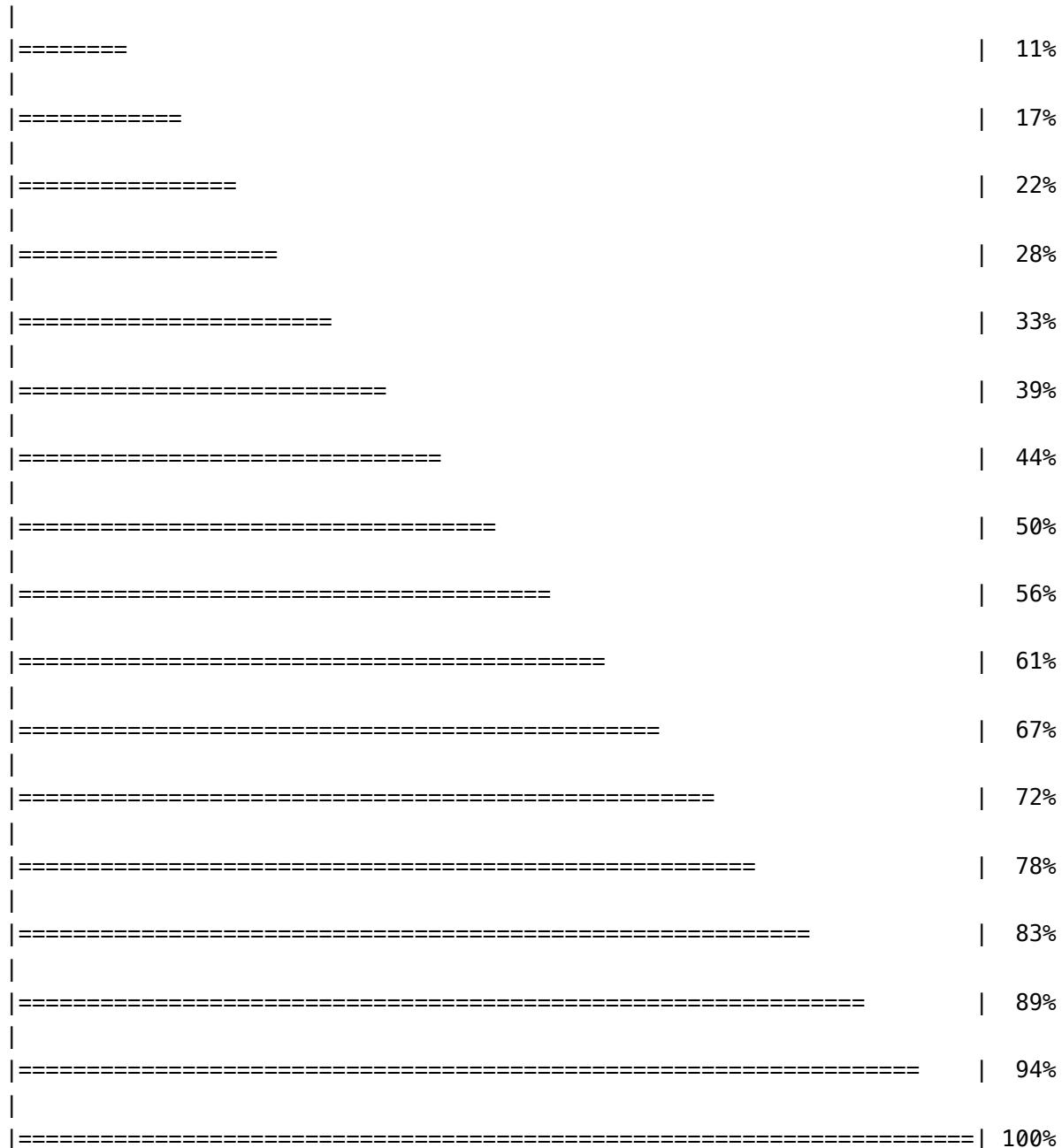
```
# NMA of all structures
modes <- nma(pdbs)
```

Warning in nma.pdbs(pdbs): 8BQF\_A.pdb might have missing residue(s) in structure:  
Fluctuations at neighboring positions may be affected.

Details of Scheduled Calculation:

- ... 18 input structures
- ... storing 540 eigenvectors for each structure
- ... dimension of x\$U.subspace: ( 546x540x18 )
- ... coordinate superposition prior to NM calculation
- ... aligned eigenvectors (gap containing positions removed)
- ... estimated memory usage of final 'eNMA' object: 40.6 Mb

      ====	0%   6%
---------------------	------------



```
plot(modes, pdbs, col=grps.rd)
```

Extracting SSE from pdbs\$sse attribute

