

Class08 Mini-project

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Background

The goal of today's mini-project is to perform principal component analysis (PCA) and hierarchical clustering on a dataset containing features of breast cancer tumors.

Data import

```
wisc.df <- read.csv("WisconsinCancer.csv", row.names = 1)
wisc.data <- wisc.df[, -1]
```

```
diagnosis <- factor(wisc.df$diagnosis)
```

Exploratory data analysis

```
dim(wisc.df)
```

```
[1] 569 31
```

```
sum(diagnosis == "M")
```

```
[1] 212
```

```
sum(diagnosis == "B")
```

```
[1] 357
```

```
length(grep("_mean$", colnames(wisc.data)))
```

```
[1] 10
```

Q1. 569 observations

Q2. 212 observations with a malignant diagnosis

Q3. 10 features

Principal Component Analysis

The main function in base R for PCA is called `prcomp()`. An optional argument `scale` should nearly always be set to `TRUE` for this function.

```
colMeans(wisc.data)
```

radius_mean	texture_mean	perimeter_mean
1.412729e+01	1.928965e+01	9.196903e+01
area_mean	smoothness_mean	compactness_mean
6.548891e+02	9.636028e-02	1.043410e-01
concavity_mean	concave.points_mean	symmetry_mean
8.879932e-02	4.891915e-02	1.811619e-01
fractal_dimension_mean	radius_se	texture_se
6.279761e-02	4.051721e-01	1.216853e+00
perimeter_se	area_se	smoothness_se
2.866059e+00	4.033708e+01	7.040979e-03
compactness_se	concavity_se	concave.points_se
2.547814e-02	3.189372e-02	1.179614e-02
symmetry_se	fractal_dimension_se	radius_worst

```

2.054230e-02      3.794904e-03      1.626919e+01
texture_worst      perimeter_worst     area_worst
2.567722e+01      1.072612e+02      8.805831e+02
smoothness_worst   compactness_worst   concavity_worst
1.323686e-01      2.542650e-01      2.721885e-01
concave.points_worst symmetry_worst    fractal_dimension_worst
1.146062e-01      2.900756e-01      8.394582e-02

```

```
apply(wisc.data, 2, sd)
```

radius_mean	texture_mean	perimeter_mean
3.524049e+00	4.301036e+00	2.429898e+01
area_mean	smoothness_mean	compactness_mean
3.519141e+02	1.406413e-02	5.281276e-02
concavity_mean	concave.points_mean	symmetry_mean
7.971981e-02	3.880284e-02	2.741428e-02
fractal_dimension_mean	radius_se	texture_se
7.060363e-03	2.773127e-01	5.516484e-01
perimeter_se	area_se	smoothness_se
2.021855e+00	4.549101e+01	3.002518e-03
compactness_se	concavity_se	concave.points_se
1.790818e-02	3.018606e-02	6.170285e-03
symmetry_se	fractal_dimension_se	radius_worst
8.266372e-03	2.646071e-03	4.833242e+00
texture_worst	perimeter_worst	area_worst
6.146258e+00	3.360254e+01	5.693570e+02
smoothness_worst	compactness_worst	concavity_worst
2.283243e-02	1.573365e-01	2.086243e-01
concave.points_worst	symmetry_worst	fractal_dimension_worst
6.573234e-02	6.186747e-02	1.806127e-02

```
wisc.pr <- prcomp(wisc.data, center = TRUE, scale. = TRUE)
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624

Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335
	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966
	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997
	PC29	PC30					
Standard deviation	0.02736	0.01153					
Proportion of Variance	0.00002	0.00000					
Cumulative Proportion	1.00000	1.00000					

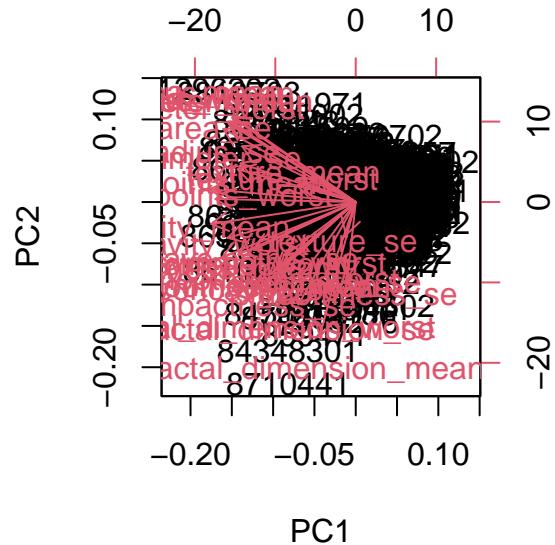
Q4. 44.27% of the variance is captured by PC1

Q5. 3 PC required

Q6. 7 PC required

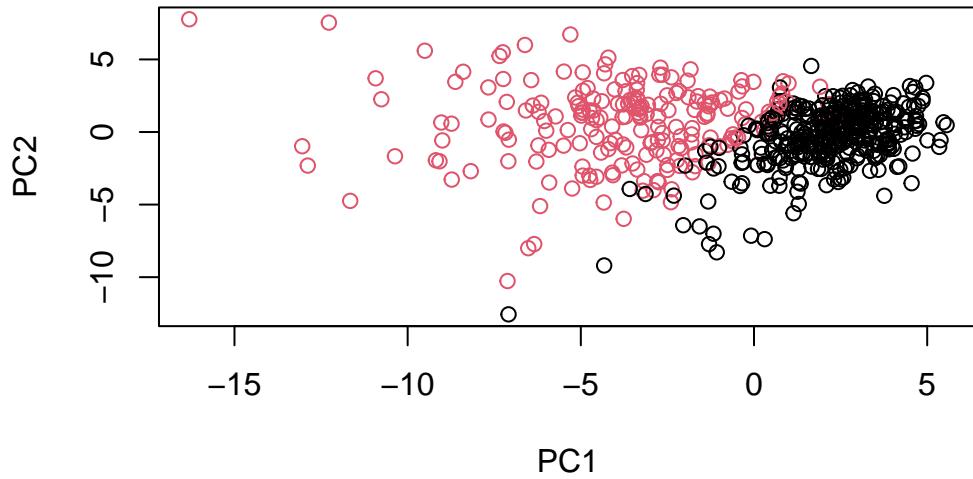
Interpreting PCA results

```
biplot(wisc.pr)
```

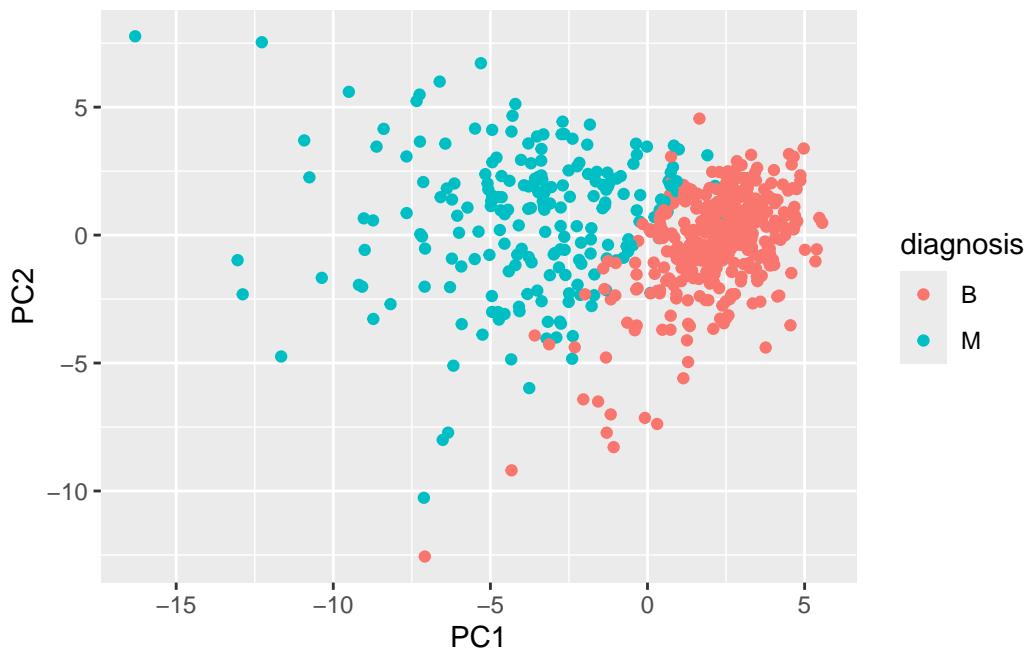


Q7. The variables are all over the place, making it extremly hard to understand visually.

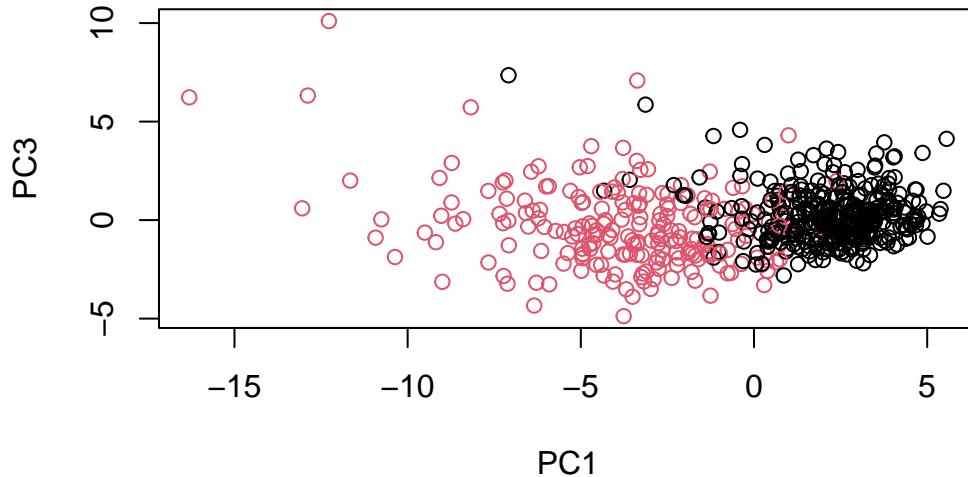
```
plot(wisc.pr$x[, 1:2], col = diagnosis ,  
      xlab = "PC1", ylab = "PC2")
```



```
library(ggplot2)
ggplot(wisc.pr$x[, 1:2], aes(x=PC1, y=PC2, color=diagnosis)) + geom_point()
```

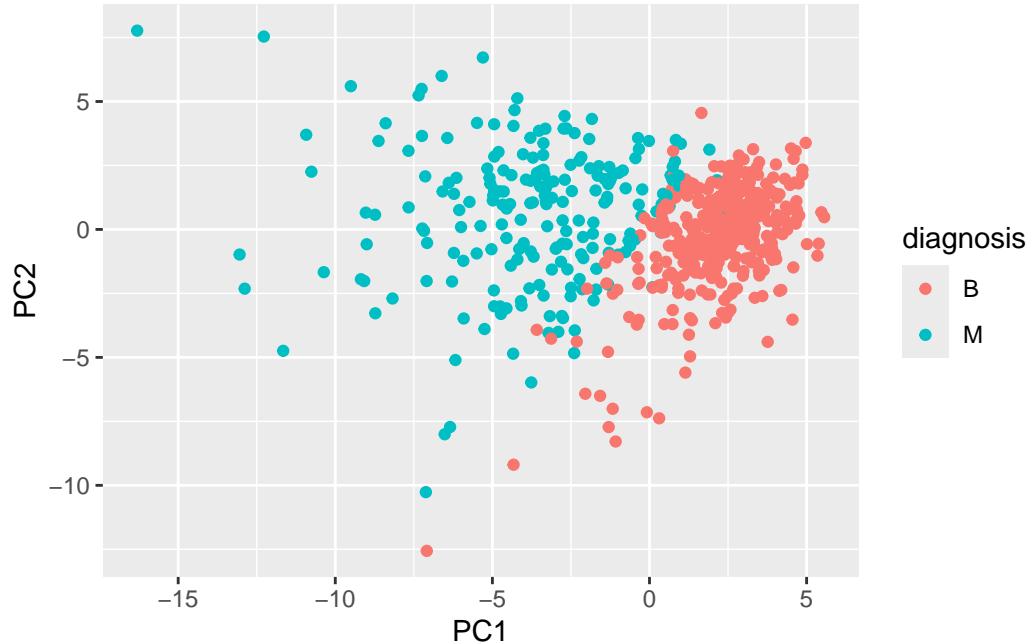


```
plot(wisc.pr$x[, c(1,3)], col = diagnosis ,  
     xlab = "PC1", ylab = "PC3")
```



Q8. Black dots are clustered together while red dots are more spread out, indicating that benign tumors have more similarity within the group.

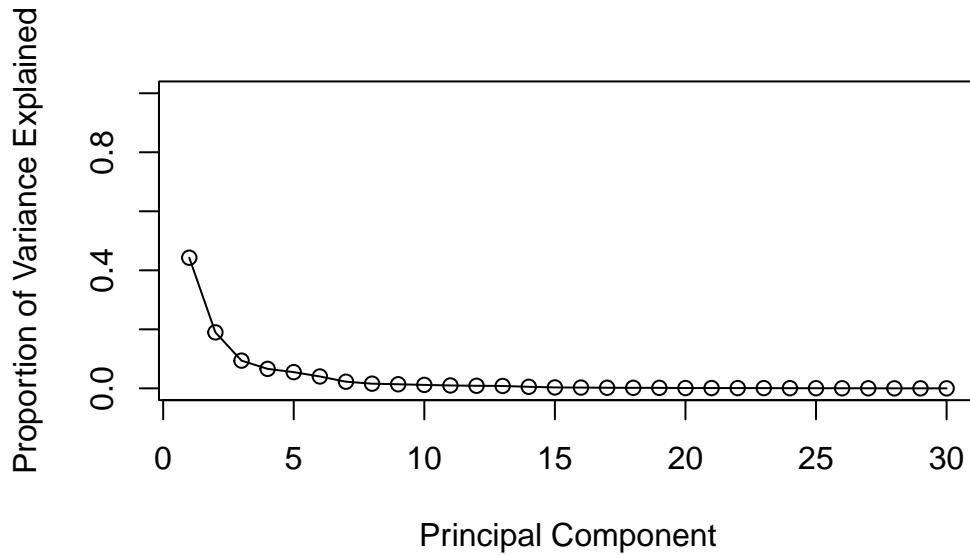
```
# Create a data.frame for ggplot  
df <- as.data.frame(wisc.pr$x)  
df$diagnosis <- diagnosis  
  
# Load the ggplot2 package  
library(ggplot2)  
  
# Make a scatter plot colored by diagnosis  
ggplot(df) + aes(PC1, PC2, col=diagnosis) + geom_point()
```



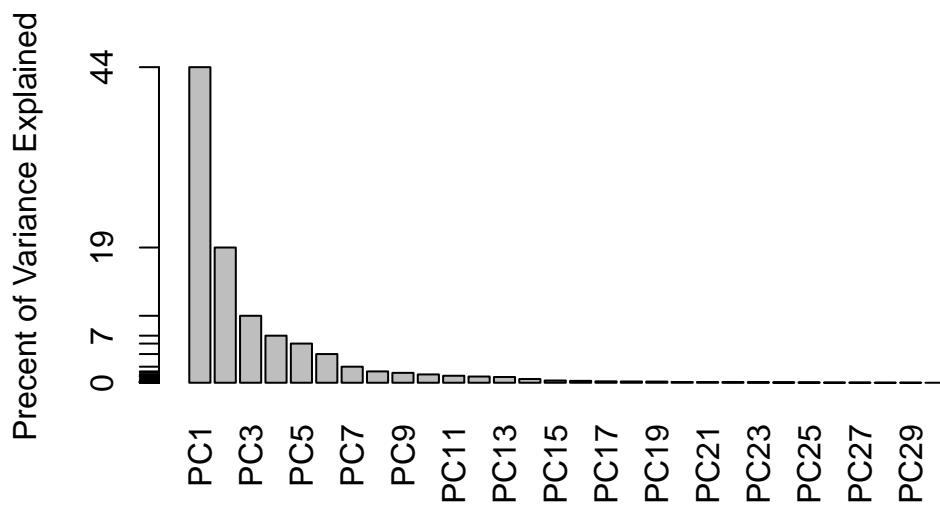
```
pr.var <- wisc.pr$sdev^2
head(pr.var)
```

```
[1] 13.281608 5.691355 2.817949 1.980640 1.648731 1.207357
```

```
pve <- pr.var/sum(pr.var)
plot(pve, xlab = "Principal Component",
      ylab = "Proportion of Variance Explained",
      ylim = c(0, 1), type = "o")
```



```
barplot(pve, ylab = "Percent of Variance Explained",
        names.arg=paste0("PC",1:length(pve)), las=2, axes = FALSE)
axis(2, at=pve, labels=round(pve,2)*100 )
```



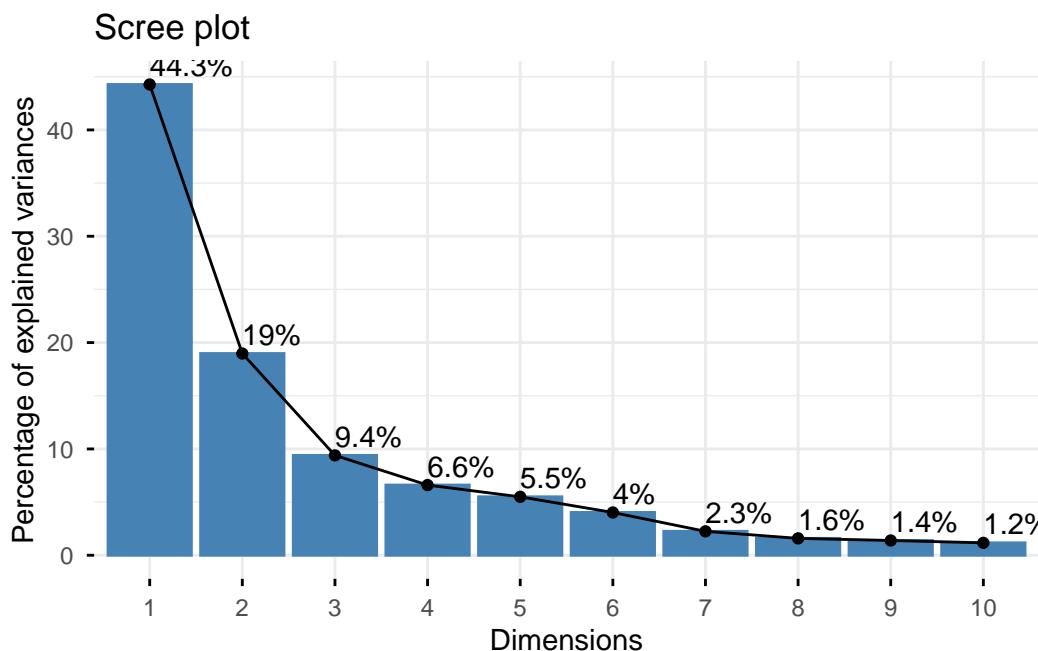
```
#Optional CRAN package:
```

```
library(factoextra)
```

Welcome! Want to learn more? See two factoextra-related books at <https://goo.gl/ve3WBa>

```
fviz_eig(wisc.pr, addlabels = TRUE)
```

Warning in geom_bar(stat = "identity", fill = barfill, color = barcolor, :
Ignoring empty aesthetic: `width`.



Q9. -0.2608538. High concave point usually points to low PC1 value.

```
wisc.pr$rotation["concave.points_mean", 1]
```

```
[1] -0.2608538
```

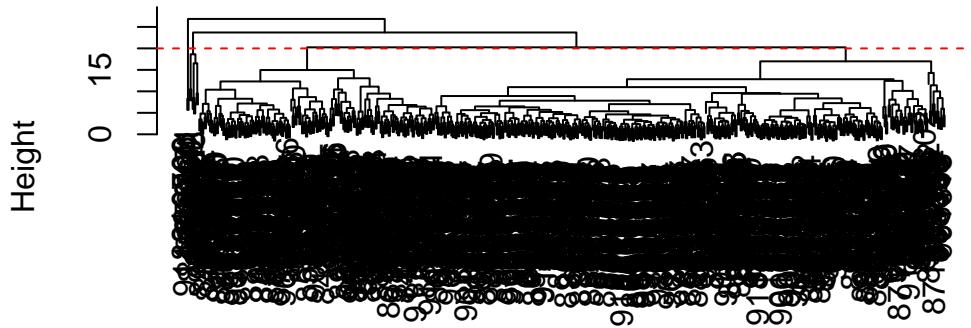
Hierarchical Clustering

```

data.scaled <- scale(wisc.data)
data.dist <- dist(data.scaled)
wisc.hclust <- hclust(data.dist)
plot(wisc.hclust)
abline(h = 20, col = "red", lty = 2)

```

Cluster Dendrogram



data.dist
 hclust (*, "complete")

Q10. h=20

```

wisc.hclust.clusters <- cutree(wisc.hclust, k = (4))
table(wisc.hclust.clusters, diagnosis)

```

	diagnosis	
wisc.hclust.clusters	B	M
1	12	165
2	2	5
3	343	40
4	0	2

```

for(i in 2:10){
  wisc.hclust.clusters.i <- cutree(wisc.hclust, k = (i))
  print(table(wisc.hclust.clusters.i, diagnosis))
}

```

```

diagnosis
wisc.hclust.clusters.i   B   M
    1 357 210
    2   0   2
diagnosis
wisc.hclust.clusters.i   B   M
    1 355 205
    2   2   5
    3   0   2
diagnosis
wisc.hclust.clusters.i   B   M
    1   12 165
    2   2   5
    3 343  40
    4   0   2
diagnosis
wisc.hclust.clusters.i   B   M
    1   12 165
    2   0   5
    3 343  40
    4   2   0
    5   0   2
diagnosis
wisc.hclust.clusters.i   B   M
    1   12 165
    2   0   5
    3 331  39
    4   2   0
    5   12   1
    6   0   2
diagnosis
wisc.hclust.clusters.i   B   M
    1   12 165
    2   0   3
    3 331  39
    4   2   0
    5   12   1
    6   0   2
    7   0   2
diagnosis
wisc.hclust.clusters.i   B   M
    1   12  86
    2   0   79

```

```

      3   0   3
      4 331  39
      5   2   0
      6  12   1
      7   0   2
      8   0   2
      diagnosis
wisc.hclust.clusters.i  B   M
      1  12  86
      2   0  79
      3   0   3
      4 331  39
      5   2   0
      6  12   0
      7   0   2
      8   0   2
      9   0   1
      diagnosis
wisc.hclust.clusters.i  B   M
      1  12  86
      2   0  59
      3   0   3
      4 331  39
      5   0  20
      6   2   0
      7  12   0
      8   0   2
      9   0   2
      10  0   1

```

Q11. Either 4 or 5 clusters seem to provide the best separation.

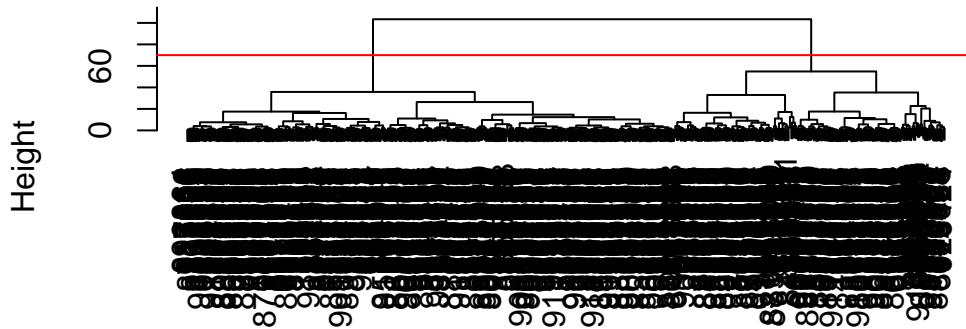
Q12. No method stands out as my favorite as the plot all appears messy with large data set. Results wise, ward.D2 seems to perform slightly better than other methods. TP=179 FP=24

```

d <- dist(wisc.pr$x[,1:3]) #Finding distance between the first 3 columns of the PCA result.
wisc.pr.hclust <- hclust(d, method="ward.D2")
plot(wisc.pr.hclust)
abline(h=70, col="red")

```

Cluster Dendrogram



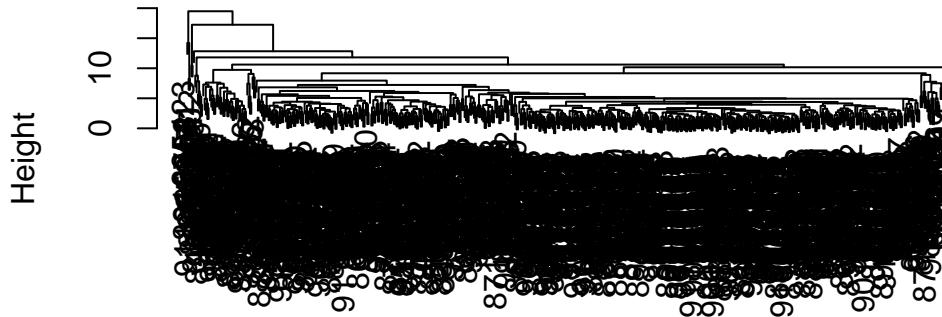
```
d  
hclust (*, "ward.D2")
```

```
grps <- cutree(wisc.pr.hclust, k=2)  
table(grps, diagnosis)
```

diagnosis		
grps	B	M
1	24	179
2	333	33

```
wisc.hclust.average <- hclust(data.dist, method="average")  
plot(wisc.hclust.average)
```

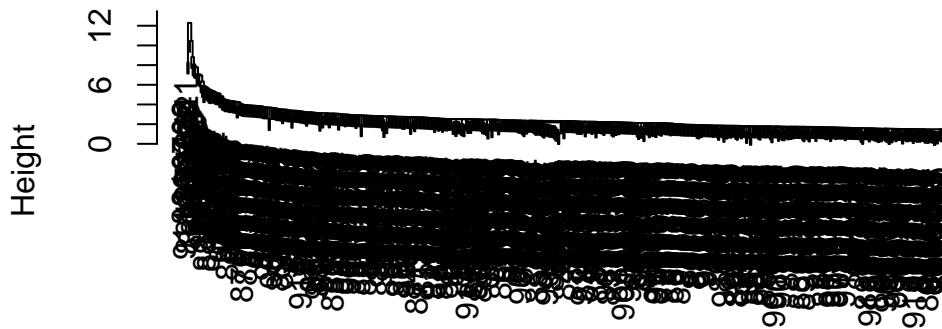
Cluster Dendrogram



```
data.dist  
hclust (*, "average")
```

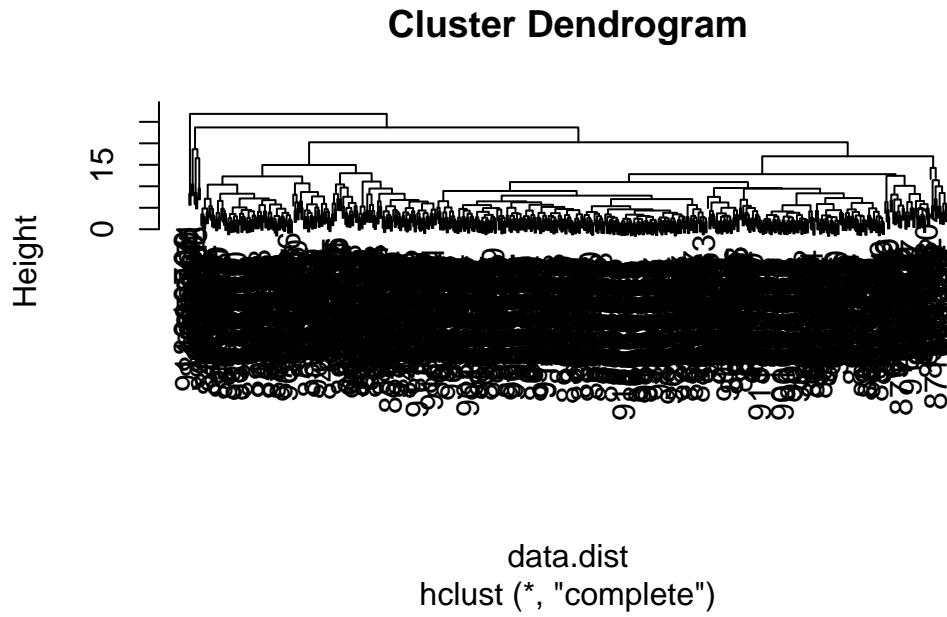
```
wisc.hclust.single <- hclust(data.dist, method="single")  
plot(wisc.hclust.single)
```

Cluster Dendrogram



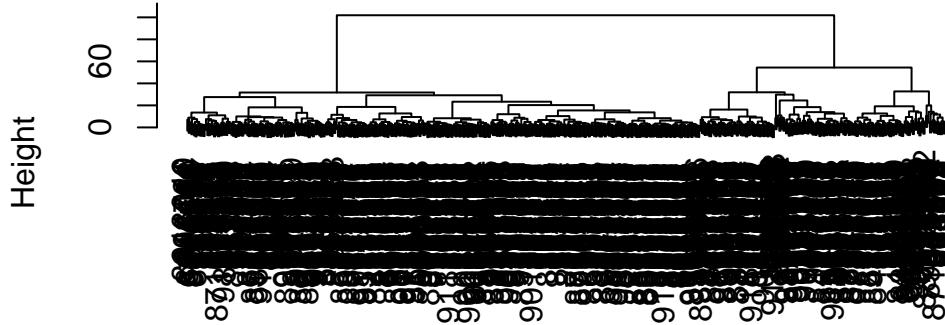
```
data.dist  
hclust (*, "single")
```

```
wisc.hclust.complete <- hclust(data.dist,method="complete")
plot(wisc.hclust.complete)
```



```
wisc.pr.hclust <- hclust(data.dist,method="ward.D2")
plot(wisc.pr.hclust)
```

Cluster Dendrogram



```
data.dist  
hclust (*, "ward.D2")
```

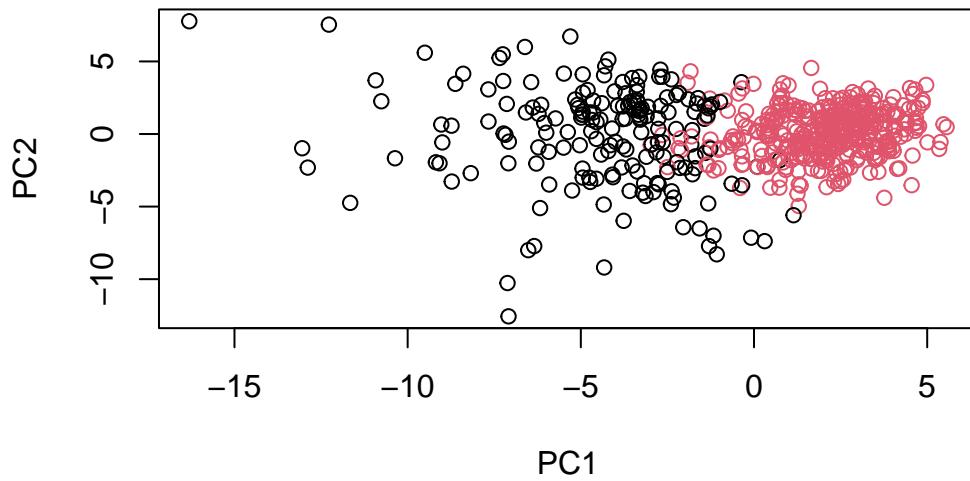
```
grps <- cutree(wisc.pr.hclust, k=2)  
table(grps)
```

```
grps  
 1   2  
184 385
```

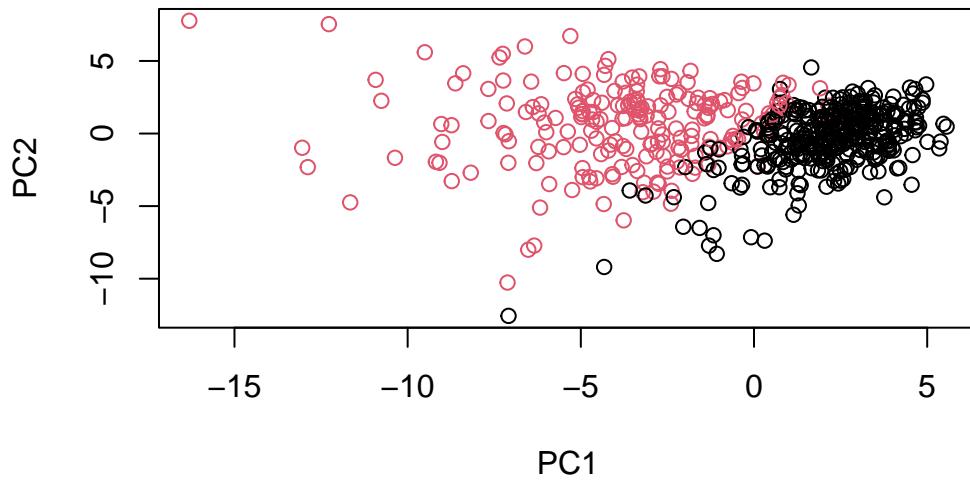
```
table(grps, diagnosis)
```

grps	B	M
1	20	164
2	337	48

```
plot(wisc.pr$x[,1:2], col=grps)
```



```
plot(wisc.pr$x[,1:2], col=diagnosis)
```



Q13. This method doesn't work well with four clusters.

```
d <- dist(wisc.pr$x[, 1:7], method = "euclidean")
wisc.pr.hclust <- hclust(d, method="ward.D2")
wisc.pr.hclust.clusters.2 <- cutree(wisc.pr.hclust, k=2)
table(wisc.pr.hclust.clusters.2, diagnosis)
```

```
diagnosis
wisc.pr.hclust.clusters.2   B   M
                           1 28 188
                           2 329 24
```

```
wisc.pr.hclust <- hclust(d, method="ward.D2")
wisc.pr.hclust.clusters.4 <- cutree(wisc.pr.hclust, k=4)
table(wisc.pr.hclust.clusters.4, diagnosis)
```

```
diagnosis
wisc.pr.hclust.clusters.4   B   M
                           1   0  45
                           2   2  77
                           3  26  66
                           4 329  24
```

```
table(wisc.hclust.clusters, diagnosis)
```

```
diagnosis
wisc.hclust.clusters   B   M
                           1 12 165
                           2  2  5
                           3 343 40
                           4  0  2
```

```
table(wisc.pr.hclust.clusters.2, diagnosis)
```

```
diagnosis
wisc.pr.hclust.clusters.2   B   M
                           1 28 188
                           2 329 24
```

```
table(wisc.pr.hclust.clusters.4, diagnosis)
```

diagnosis		
wisc.pr.hclust.clusters.4	B	M
1	0	45
2	2	77
3	26	66
4	329	24

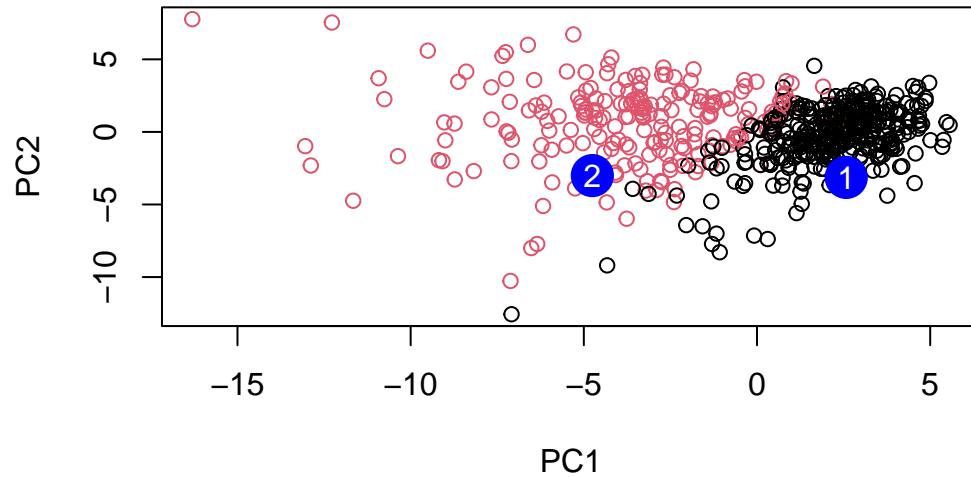
Q14. I think the hierarchical cluster performed with ward.D2 method and 2 clusters is the best, as it identifies most of the malignant tumors correctly while keeping a low number of benign tumors misclassified.

Prediction

```
#url <- "new_samples.csv"
url <- "https://tinyurl.com/new-samples-CSV"
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
npc
```

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
[1,]	2.576616	-3.135913	1.3990492	-0.7631950	2.781648	-0.8150185	-0.3959098
[2,]	-4.754928	-3.009033	-0.1660946	-0.6052952	-1.140698	-1.2189945	0.8193031
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
[1,]	-0.2307350	0.1029569	-0.9272861	0.3411457	0.375921	0.1610764	1.187882
[2,]	-0.3307423	0.5281896	-0.4855301	0.7173233	-1.185917	0.5893856	0.303029
	PC15	PC16	PC17	PC18	PC19	PC20	
[1,]	0.3216974	-0.1743616	-0.07875393	-0.11207028	-0.08802955	-0.2495216	
[2,]	0.1299153	0.1448061	-0.40509706	0.06565549	0.25591230	-0.4289500	
	PC21	PC22	PC23	PC24	PC25	PC26	
[1,]	0.1228233	0.09358453	0.08347651	0.1223396	0.02124121	0.078884581	
[2,]	-0.1224776	0.01732146	0.06316631	-0.2338618	-0.20755948	-0.009833238	
	PC27	PC28	PC29	PC30			
[1,]	0.220199544	-0.02946023	-0.015620933	0.005269029			
[2,]	-0.001134152	0.09638361	0.002795349	-0.019015820			

```
plot(wisc.pr$x[,1:2], col = diagnosis ,  
      xlab = "PC1", ylab = "PC2")  
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)  
text(npc[,1], npc[,2], c(1,2), col="white")
```



Q16. Patient 2 might need a prioritized check-up as it is clustered with malignant tumors, indicating the possibility of a malignant tumor for this patient.