# **CIAlign**

CIAlign is a command line tool which performs various functions to parse and analyse a multiple sequence alignment (MSA).

The tool is designed to be highly customisable, allowing users to specify exactly which functions to run and which settings to use. It is also transparent, generating a clear log file and diagram showing exactly how the alignment has changed.

This allows the user to:

- Remove sources of noise from their MSA
- Crop of poorly aligned sequence ends
- Remove of insertions which are not present in the majority of sequences
- Remove of sequences below a threshold number of bases or amino acids
- Remove columns containing only gaps
- Remove sequences above a threshold level percentage of divergence from the majority
- Generate consensus sequences
- Visualise alignments
- Generate image files showing the alignment before and after parsing and showing which columns and rows have been removed
- Draw sequence logos
- Visualise coverage at each postiion in the alignment
- Analyse alignment statistics
- Generate a similarity matrix showing the percentage identity between each sequence pair

## Requirements

- python >= 3.6
- matplotlib >= 2.1.1
- numpy >= 1.16.3
- scipy >= 1.3.0

# Installation

? conda

The current release of CIAlign can be downloaded directly using this link Add the CIAlign directory to your PATH environment variable as described here

# Usage

## Basic Usage

CIAlign --infile INFILE --outfile\_stem STEM --inifile cialign.ini

#### **Parameters**

| Parameter        | Description   | Default |
|------------------|---|---------|
| -infile          | Path to input alignment FASTA file                                  | None    |
| -inifile         | Path to ini file  | None    |
| $-outfile\_stem$ | Prefix for output files, including the path to the output directory | CIAlign |
| -silent          | Do not print progress to the screen                                 | False   |

### **Functions**

Specify which functions to run by adding the following optional arguments to the command

## Parsing an MSA

Each of these steps will be performed sequentially in the order specified in the table below.

The parsed alignment after all steps have been performed will be saved as OUTFILE\_STEM\_parsed.fasta

| Parameter             | Description  | Default<br>Value |
|-----------------------|--|------------------|
| $-{ m crop\_ends}$    | Crop the ends of sequences if they are poorly aligned                            | False            |
| $-crop\_ends\_mingap$ | minimum gap size to consider<br>when classifying a sequence as<br>poorly aligned | 10               |

| Parameter                              | Description   | Default<br>Value |
|--|---|------------------|
| -remove_badlyaligned                   | Remove sequences with $\leq$ N proportion of positions at which the most common base / amino acid in the alignment is present | False            |
| $-remove\_\ badly a ligned\_\ minperc$ | Minimum proportion of positions which should be identical to the most common base / amino acid in order to be preserved       | 0.9              |
| $-{ m remove\_insertions}$             | Remove insertions found in $\leq$ 50% of sequences from the alignment   | False            |
| $-insertion\_min\_size$                | Only remove insertions >= this number of residues   | 3                |
| $-insertion\_max\_size$                | Only remove insertions <= this number of residues   | 300              |
| $-insertion\_min\_flank$               | Minimum number of bases on<br>either side of an insertion to<br>classify it as an insertion                                   | 5                |
| $-$ remove $\_$ short                  | Remove sequences $\leq$ N bases / amino acids from the alignment  | False            |
| $-remove\_minlength$                   | Minimum number of non-gap<br>residues in a sequence to be<br>preserved  | 50               |
| -remove_gaponly                        | Remove gap only columns from the alignment  | True             |

# Generating a Consensus Sequence

This step generates a consensus sequence based on the parsed alignment. If no parsing functions are performed, the consensus will be based on the input alignment.

# Output files:

- $\bullet$   $\mathbf{OUTFILE\_STEM\_consensus.fasta}$  the consensus sequence only
- $\bullet$  OUTFILE\_STEM\_with\_consensus.fasta the parsed alignment plus the consensus

| Parameter              | Description  | Default   |
|------------------------|--|-----------|
| -make_consensus        | Make a consensus sequence based on<br>the parsed alignment   | False     |
| $-consensus\_type$     | Type of consensus sequence to make - can be majority, to use the most common character at each position in the consensus, even if this is a gap, or majority_nongap, to use the most common non-gap character at each position | majority  |
| $-consensus\_keepgaps$ | If there are gaps in the consensus (if majority_nongap is used as consensus_type), should these be included in the consensus (True) or should this position in the consensus be deleted (False)                                | False     |
| -consensus_name        | Name to use for the consensus sequence in the output fasta file  | consensus |

# Visualising Alignments

Each of these functions produces some kind of visualisation of your alignment.

### Mini Alignments

These functions produce "mini alignments" - images showing a small representation of your whole alignment, so that gaps and poorly aligned regions are clearly visible.

### Output files:

- $\bullet$  OUTFILE\_STEM\_input.png (or svg, tiff, jpg) the input alignment
- OUTFILE\_STEM\_output.png (or svg, tiff, jpg) the parsed output alignment
- OUTFILE\_STEM\_markup.png (or svg, tiff, jpg) the input alignment with deleted rows and columns marked

| Parameter               | Description                                     | Default |
|-------------------------|---|---------|
| $-\mathrm{plot\_input}$ | Draws a mini alignment for the input FASTA file | False   |

| Parameter                         | Description   | Default |
|-----------------------------------|---|---------|
| $-$ plot $\_$ output              | Draws a mini alignment for the output FASTA file  | False   |
| $-\mathrm{plot}\_\mathrm{markup}$ | Draws the input alignment but with the columns and rows which have been removed by each function marked | False   |
| $-plot\_dpi$                      | DPI for mini alignments   | 300     |
| $-plot\_format$                   | Image format for mini alignments - can be png, svg, tiff or jpg   | png     |
| $-plot\_width$                    | Mini alignment width in inches  | 5       |
| $-plot\_height$                   | Mini alignment height in inches   | 3       |

## Sequence logos

These functions draw sequence logos representing your output (parsed) alignment. If no parsing functions are specified, the logo will be based on your input alignment.

## $Output\_files:$

- $\bullet$  OUTFILE \_STEM \_logo \_bar.png (or svg, tiff, jpg) the alignment represented as a bar chart
- OUTFILE\_STEM\_logo\_text.png (or svg, tiff, jpg) the alignment represented as a standard sequence logo using text

| Parameter                       | Description   | Default   |
|---------------------------------|---|-----------|
| -make_sequence_logo             | Draw a sequence logo  | False     |
| $-sequence\_logo\_type$         | Can be bar, to draw the logo as a<br>bar chart, text, to draw a standard<br>sequence logo using text, or both, to<br>draw both    | bar       |
| $-sequence\_logo\_dpi$          | DPI for sequence logo   | 300       |
| $-sequence\_logo\_font$         | font (see NB below) for bases / amino acids in a text based sequence logo   | monospace |
| $-sequence\_logo\_nt\_per\_row$ | number of bases / amino acids to<br>show per row in the sequence logo,<br>where the logo is too large to show<br>on a single line | 50        |

| Parameter               | Description   | Default |
|-------------------------|---|---------|
| -sequence_logo_filetype | Image file type to use for the sequence logo - can be png, svg, tiff or jpg | png     |

NB: to see a vailable fonts on your system, run CIAlign <code>-list\_fonts\_only</code> and view <code>CIAlign\_fonts.png</code>

### Coverage Plots

This function plots the number of non-gap residues at each postion in the alignment.

Output file:

- OUTFILE\_STEM\_input\_coverage.png (or svg, tiff, jpg) image showing the input alignment coverage
- OUTFILE\_STEM\_output\_coverage.png (or svg, tiff, jpg) image showing the parsed alignment coverage

| Parameter                    | Description  | Default |
|------------------------------|--|---------|
| -plot_coverage_input         | Plot the coverage of the multiple sequence alignment     | False   |
| $-plot\_coverage\_output$    | Plot the coverage of the multiple sequence alignment     | False   |
| $-plot\_coverage\_dpi$       | DPI for coverage plot                                    | 300     |
| $-plot\_coverage\_height$    | Height for coverage plot (inches)                        | 3       |
| $-plot\_coverage\_width$     | Width for coverage plot (inches)                         | 5       |
| $-plot\_coverage\_colour$    | Colour for coverage plot (hex code or name)              | #007bf5 |
| $-plot\_coverage\_file type$ | File type for coverage plot (can be png, svg, tiff, jpg) | png     |

# **Analysing Alignment Statistics**

These functions provide additional analyses you may wish to perform on your alignment.

# Similarity Matrices

Generates a matrix showing the proportion of identical bases / amino acids between each pair of sequences in the MSA.

| Parameter                       | Description   | Default |
|---------------------------------|---|---------|
|                                 | make a similarity matrix for<br>the input alignment   | False   |
| _ make_similarity_matrix_output | make a similarity matrix for the output alignment   | False   |
| $-make\_simmatrix\_keepgaps$    | Include positions with gaps in<br>either or both sequences in the<br>similarity calculation         | False   |
| $-make\_simmatrix\_dp$          | Number of decimal places to<br>display in the similarity matrix<br>output file                      | 4       |
| $-make\_simmatrix\_minoverlap$  | Minimum overlap between two<br>sequences to have non-zero<br>similarity in the similarity<br>matrix | 1       |