

the device used in our study (the Baxter Infusor) states that the flow rate increases by about 0.5% for every 1-in rise in head height and by about 2.3% (due to a reduction in solution viscosity) for every 1 °C rise in temperature.¹ Also, the flow rate depends on the type of solution used. For example, the flow rate for 0.9% sodium chloride injection is about 10% higher than that for

5% dextrose injection. For temperature control, we recommend taping the flow-restrictor housing to the skin. An internal study found that this is a very effective method for minimizing the effect of ambient temperature variation.

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Assays for biological agents

We read with concern the study by Goldspiel and colleagues,¹ which evaluated the stability of alemtuzumab after storage at room temperature. The authors used high-performance liquid chromatography (HPLC) to assess the loss of alemtuzumab over time. It is essential to recognize that the pharmacologic properties of proteins, monoclonal antibodies, and other large molecules depend on molecular configuration and that varying storage conditions can alter that configuration and render the molecule biologically inactive. In order to assess the effect of varying storage conditions on these types of products for the purpose of extending expiration dating, it is essential that biological activity—not simply concentration—be assayed. HPLC may assess the quantity of a particular agent in a sample, but it will not detect differences in molecular configuration that can determine activity.² In our study,³ referenced by Goldspiel et al., we found that alemtuzumab retained its biological activity over a wide range of conditions. It is entirely likely that storage at room temperature will not reduce activity, but until a confirmatory study is conducted using a measure of biological activity (e.g., flow cytometry), the findings of Goldspiel et al. should be viewed with caution.

The publication of a biological stability study using HPLC raises an important issue that we believe should be addressed by *AJHP*. Goldspiel et al. followed *AJHP*'s recommendations for evaluating drug

stability, as put forth by Trissel⁴ in 1983, before the widespread use of modern analytic techniques designed to detect biological activity. While Trissel's advice related to study design remains current and important, the analytic methods he recommended in 1983 are outdated and not applicable to biological products. The current standard for analytic assays of monoclonal antibodies are functional—most commonly immunoassays, including enzyme-linked immunosorbent assays.^{2,5}

Biological agents represent a large proportion of hospital drug expenditures and typically have short expiration dates, resulting in waste and unnecessary expenditures.⁶ Our previous work in establishing extended expiration dating for biological agents suggests that substantial savings are possible through waste reduction.^{2,7-9} Given the increase in the number and cost of biological products and the unique challenges that they pose when attempting to demonstrate extended expiration dating, it is imperative that *AJHP* pursue new standards and guidelines for this work.

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■ Letters

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We thank Dr. Kolesar and Mr. Vermeulen for their observation about potential differences in determining the stability of biological compounds versus simple chemical molecules. We agree that guidelines should be developed for conducting stability studies for biological agents that include a standardized method for determining biological activity and a definition of activity level that would be considered equivalent over time.

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Editor's note: The AJHP editors are investigating the establishment of standards for stability studies involving biological agents.

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