



Technical Update • March 2013

Cleveland Clinic Laboratories is dedicated to keeping you updated and informed about recent testing changes. That's why we are happy to provide this technical update on a monthly basis.

Recently changed tests are **bolded**, and could include revisions to methodology, reference range, days performed or CPT code. For your convenience, tests are listed alphabetically and the order and billing codes are provided.

If you wish to compare the new information with previous test demographics, refer to the Test Directory, which can be accessed at clevelandcliniclabs.com.

Deleted tests and new tests are listed separately. Please update your database as necessary. For additional detail, contact Client Services at 216.444.5755 or 800.628.6816 or via email at clientservices@ccf.org.

Test Update Page #	Summary of Changes by Test Name	Order Code	Billing Code	Name Change	New Test	Test Discontinued	Specimen Requirement	Special Information	Component Change	Methodology	Reference Range	Days Performed/Reported	CPT
2	Acylglycines, Quantitative Urine												
2	Alpha Galactosidase, Leukocytes												
2	Arsenic, Hair												
2	Arylsulfatase A, Leukocytes												
7	Bacterial Antigen Detection												
3	Blood Culture												
3	Bordetella PCR												
3	Complement C7 Functional												
3	Complement C9 Functional												
3	Cyanide, Blood												
7	FISH for Cutaneous Melanoma												
4	Fungal Blood Culture												
4	Galactokinase												
4	Granulocyte Antibodies												
4	Hexagonal Phase Phospholipid Neutralization												
4	Hexosaminidase A & Total, WBC												
5	Inhibin B												
5	Iodine, Random Urine												
5	Iodine, Urine 24 hours												
5	Measles IgG Antibody												
5	MTHFR by PCR												
5	MTHFR Gene Analysis												
5	Mumps IgG Antibody												
5	Pancreatic Polypeptide												

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5	Phospholipids, Serum											
5	Porphyrins, Serum Total											
5	PTH, Intact											
6	Synthetic Glucocorticoid Screen, Serum											
6	Synthetic Glucocorticoid Screen, Urine											
7	TRH Stimulation											
7	TRH Stimulation of Prolactin											
6	Trichinella IgG Antibody											
6	Tricyclic Antidepressant ID											
6	Vaginal Pathogens DNA Direct Probes											
6	Varicella-Zoster IgG Ab											

Test Changes

Test Name	Order Code	Billing Code	Change	Effective Date
Acylglycines, Quantitative Urine	UACYLG	83755	Special Information: Patient's age is required; Include family history, clinical condition (asymptomatic or acute episode), diet and drug therapy information with the sample. Includes: Ethylmalonic Acid, UR 2-Methylsuccinic Acid, UR Glutaric Acid, UR Isobutyrylglycine, UR n-Butyrylglycine, UR 2-Methylbutyrylglycine, UR Isovalerylglycine, UR n-Hexanoylglycine, UR n-Octanoylglycine, UR 3-Phenylpropionylglycine, UR Suberylglycine, UR trans-cinnamoylglycine, UR Dodecanedioic Acid, UR (12 DCA) Tetradecanedioic Acid, UR (14 DCA) Hexadecanedioic Acid, UR (16 DCA) Test build may need to be modified Specimen Requirement: 10 mL random urine in a clean container; Do not use preservative; Frozen	2/6/2013
Alpha Galactosidase, Leukocytes	AGAL	87795	Special Information: Specimens must arrive at the performing lab within 72 hours of collection. Specimens received after 72 hours could have falsely normal results. Do not draw specimen on the day before a holiday.	3/28/2013
Arsenic, Hair	ARSHR	82687	Reference Range: 0 - 15 years: Not established 16 - 99 years: 0.0 - 0.9 mcg/g hair Days Performed: Tuesday, Friday Reported: 3 - 8 days	4/1/2013
Arylsulfatase A, Leukocytes	ARYLA	87813	Special Information: Specimen must arrive at the performing laboratory within 48 hours of collection. Methodology: Colorimetric Enzyme Assay Days Performed: Thursday Reported: 9 - 16 days	3/28/2013

Test Changes

Test Name	Order Code	Billing Code	Change	Effective Date
Blood Culture	BLCUL	79134	Special Information: Number of Blood Cultures: Each general blood culture request will generate two sets of separately drawn blood cultures. The second blood culture can be drawn immediately after the first is drawn, provided the two blood cultures are collected separately, i.e. from different sites/arms. If the same arm must be used, collect the second culture 30 minutes after the first. A maximum of four blood culture sets is permitted in a 24 hour period. Patient Skin Preparation: Select vein, swab with Chloraprep Sepp (2% chlorhexidine gluconate with 70% alcohol applicator). Saturate the applicator tip by gently pressing it against the skin, apply the solution in a back and forth motion for 30 seconds, completely wetting the area. Allow the prepped area to dry completely. Do not repalpate the vein. If the venipuncture is unsuccessful, re-prep the vein as above. Venipuncture and Inoculation: Perform venipuncture using a sterile syringe (20 mL for adults, 1-20 mL for children depending on weight of the patient). Wipe the rubber stopper of each blood culture bottle with a new Chloraprep. Saturate the applicator tip by gently pressing it against the stopper, apply the solution in concentric circles, completely wetting the stopper; allow the prepped area to dry completely. Always inoculate the aerobic bottle first; optimal volume is 10 mL/bottle. Do not overfill. Invert all tubes/bottles several times to ensure thorough mixture. Label the blood culture bottle with the collect date and time, collection site, set #1 or set #2, initials of individual drawing the blood, patient's name and patient medical record number. Transport: Blood culture specimens should be transported promptly at room temperature to Cleveland Clinic Laboratories on the day collection along with the requisition.	2/12/2013
Bordetella PCR	BORPCR	82511	Includes: Bordetella pertussis PCR Bordetella parapertussis PCR Test build may need to be modified Specimen Requirement: 1 Nasopharyngeal swab in M4 or Universal Transport Media (UTM); Refrigerated Reference Range: Bordetella pertussis PCR: Negative Bordetella parapertussis PCR: Negative CPT: 87798x2	3/7/2013
Complement C7 Functional	C7FUN	87793	Specimen Requirement: 1 mL serum from a red top tube; Patient should be fasting; Do not use serum separator tube; Place specimen on ice after collection; Separate serum from cells and freeze immediately ; Frozen	4/1/2013
Complement C9 Functional	C9FUN	87809	Specimen Requirement: 1 mL serum from a red top tube; Patient should be fasting; Do not use serum separator tube; Place specimen on ice after collection; Separate serum from cells and freeze immediately ; Frozen	4/1/2013
Cyanide, Blood	CYANID	82600	Methodology: Colorimetric Days Performed: Monday, Wednesday, Friday Reported: 2 - 6 days	4/1/2013

Test Changes (cont.)

Test Name	Order Code	Billing Code	Change	Effective Date
Fungal Blood Culture	HISTCL	79133	Special Information: Number of Blood Cultures: Each general blood culture request will generate two sets of separately drawn blood cultures. The second blood culture can be drawn immediately after the first is drawn, provided the two blood cultures are collected separately, i.e. from different sites/arms. If the same arm must be used, collect the second culture 30 minutes after the first. A maximum of four blood culture sets is permitted in a 24 hour period. The alternate specimen type is a bone marrow. Ordering/Drawing a Fungal Blood Culture: Order a Fungal Blood Culture (HISTCL) to rule out Histoplasma. For blood specimens, an Isolator tube should be drawn; 10 mL optimal, 5 mL minimum. For bone marrow specimens, use a Sodium or Lithium heparinized syringe to draw the specimen; 4 mL optimal, 1 mL minimum. Patient Skin Preparation for Blood Draw: Select vein, swab with Chloraprep Sepp (2% chlorhexidine gluconate with 70% alcohol applicator). Saturate the applicator tip by gently pressing it against the skin, apply the solution in a back and forth motion for 30 seconds, completely wetting the area. Allow the prepped area to dry completely. Do not repalpate the vein. If the venipuncture is unsuccessful, re-prep the vein as above. Venipuncture and Inoculation: Perform venipuncture using a sterile syringe (10 mL for adults, 1-10mL for children depending on weight of the patient). Wipe the rubber stopper of the Isolator tube with a new Chloraprep. Saturate the applicator tip by gently pressing it against the stopper, apply the solution in concentric circles, completely wetting the stopper; allow the prepped area to dry completely. Inoculate the blood into the Isolator tube; 10 mL is optimal, minimum volume 5 mL. Invert tube several times to ensure thorough mixture. Label the Isolator tube with the collect date and time, collection site, set #1 or set #2, initials of individual drawing the blood, patient's name and patient medical record number. Transport: Blood culture and bone marrow specimens should be transported promptly at room temperature to Cleveland Clinic Laboratories on the day of collection along with the requisition.	2/7/2013
Galactokinase	GALKIN	82759	Specimen Requirement: 5 mL whole blood in a sodium heparin green top tube; Refrigerated Days Performed: Wednesday Reported: 10 - 17 days	3/28/2013
Granulocyte Antibodies	NEUTR	76364	Specimen Requirement: 2 mL serum from a red top tube; Do not use serum separator tube; Refrigerated Days Performed: Monday, Wednesday, Friday Reported: 8 - 16 days	3/28/2013
Hexagonal Phase Phospholipid Neutralization	STACLT	82741	Reference Range: Hexagonal Phase Screen: 53.8 - 72.4 sec Hexagonal Phase Confirm: 54.3 - 68.5 sec Hexagonal Phase Delta: < 7.0	3/1/2013
Hexosaminidase A & Total, WBC	TAYSAC	82868	Reference Range: Total Hexosaminidase: ≤ 15 years: ≥ 20 nmol/min/mg ≥ 16 years: 16.4 - 36.2 nmol/min/mg Hexosaminidase A ≤ 15 years: 20 - 80% of total ≥ 16 years: 63 - 75% of total Days Performed: Monday, Thursday, Friday Reported: 5 - 9 days	2/21/2013

Test Changes (cont.)

Test Name	Order Code	Billing Code	Change	Effective Date
Inhibin B	INHIBB	82819	Specimen Requirement: 1.5 mL serum from a serum separator tube; Frozen Methodology: Enzyme Linked Chemiluminescent Assay (CLIA) Reference Range: Adult Females, follicular phase: 10 - 290 pg/mL Postmenopausal women: <10 pg/mL Girls 0 - 6 years: <10 - 73 pg/mL 6 - 10 years: <10 - 130 pg/mL 10 - 11 years: <10 - 103 pg/mL 11 - 12 years: <10 - 186 pg/mL 12 - 18 years: <10 - 360 pg/mL Healthy adult males: 10 - 225 pg/mL Boys 0 - 1 years: 70 - 630 pg/mL 1 - 2 years: 90 - 400 pg/mL 2 - 6 years: 40 - 270 pg/mL 6 - 10 years: 35 - 170 pg/mL 10 - 11 years: 50 - 300 pg/mL 11 - 12 years: 105 - 450 pg/mL 12 - 18 years: 75 - 475 pg/mL Days Performed: 1 day per week Reported: 6 - 13 days	2/28/2013
Iodine, Random Urine	UIODR	87648	Specimen Requirement: 1 mL random urine in a clean container; Container should not have a metal cap or glued insert; Refrigerated Days Performed: Monday, Tuesday, Wednesday, Friday Reported: 2 - 4 days	3/28/2013
Iodine, Urine 24 hours	UIOD24	75084	Specimen Requirement: 10 mL urine in a clean container from a well-mixed 24 hour collection; Refrigerate during collection; Do not use preservative; Refrigerated Days Performed: Monday, Tuesday, Wednesday, Friday Reported: 2 - 4 days	4/4/2013
Measles IgG Antibody	MEASLG	75399	Methodology: Immunochemilluminometric Assay (ICMA) Reference Range: Negative: < 25.0 AU/mL Equivocal: 25.0 - 30.0 AU/mL Positive: > 30.0 AU/mL	3/26/2013
MTHFR by PCR	MTHFR	81692	Special Information: The MTHFR by PCR will not be discontinued as announced in the February Technical Update. We will continue to offer testing without any changes to our current specimen requirements or test build. We apologize for any extra work this may have caused.	3/1/2013
MTHFR Gene Analysis	MTHF	81692	Special Information: Due to circumstances beyond our control, the MTHFR Gene Analysis announced in the February Technical Update will not be added as a new test as planned. We apologize for any extra work this may have caused.	3/1/2013
Mumps IgG Antibody	MUMPSG	75398	Methodology: Immunochemilluminometric Assay (ICMA) Reference Range: Negative: < 9 AU/mL Equivocal: 9 - 11 AU/mL Positive: > 11 AU/mL	3/26/2013
Pancreatic Polypeptide	PANC	75675	Specimen Requirement: 3 mL plasma from an EDTA lavender top tube; Place specimen on ice after collection; Separate plasma and freeze ASAP; Frozen	3/28/2013
Phospholipids, Serum	PHOLIP	84090	Methodology: Enzymatic Colorimetric	3/28/2013
Porphyrins, Serum Total	SPORPH	77116	Specimen Requirement: 2 mL serum from a red top tube; Protect specimen from light; Separate serum from cells within 1 hour of collection; Frozen	2/21/2013
PTH, Intact	PTHI	83970	Specimen Requirement: 1 mL plasma from an EDTA lavender top tube; Ambient Methodology: Electro Chemiluminescence Immunoassay (ECLIA) Reference Range: 15 - 64 pg/mL	3/4/2013

Test Changes (cont.)

Test Name	Order Code	Billing Code	Change	Effective Date
Synthetic Glucocorticoid Screen, Serum	SGLUCO	88378	Days Performed: Tuesday, Friday Reported: 3 - 6 days	4/1/2013
Synthetic Glucocorticoid Screen, Urine	UGLUCO	88379	Days Performed: Wednesday Reported: 4 - 10 days	4/1/2013
Trichinella IgG Antibody	TRICH	75621	Specimen Requirement: 0.5 mL serum from a serum separator tube; Frozen Days Performed: Tuesday, Thursday Reported: 2 - 8 days	3/28/2013
Tricyclic Antidepressant ID	TAID	89249	Order Code: Previously TADID Billing Code: Previously 79193 Specimen Requirement: 1 mL serum from a red top tube; Do not use serum separator tube; Separate serum from cells within 2 hours of collection; Refrigerated CPT: 80152, 80182, 80174, 80160, 80166, 80299x4	3/21/2013
Vaginal Pathogens DNA Direct Probes	VAGDNA	88354	Special Information: Specimens must be collected and transported using the BD Affirm VPIII Ambient Temperature Transport System. Specimens must be received within 72 hours of collection. Vaginal specimens are the only acceptable specimen type. This test is for use in women with symptoms of vaginitis. Other tests for the evaluation of vaginitis are available on vaginal fluid or vaginal swab specimens: If only bacterial vaginosis is suspected, the clinician should order a miscellaneous Gram stain. If only candidiasis is suspected, the clinician should order a fungal smear. If a vaginal specimen is submitted in the BD Affirm Ambient Temperature Sample Collection tube, the Vaginal Pathogens DNA Direct Probe test (VAGDNA) will be performed. Methodology: Nucleic acid probe, qualitative Reference Range: <i>Candida</i> species DNA Probe: Negative <i>Gardnerella vaginalis</i> DNA Probe: Negative <i>Trichomonas vaginalis</i> DNA Probe: Negative	3/5/2013
Varicella-Zoster IgG Ab	VZVG	75622	Methodology: Immunochemiluminometric Assay (ICMA) Reference Range: Negative: < 135 index Equivocal: 135 - 165 index Positive: > 165 index	3/26/2013

New Tests

Test Name	Order Code	Billing Code	Test Information	Effective Date
FISH for Cutaneous Melanoma	CMFISH	89257	Specimen Requirement: One paraffin block containing representative tissue OR six unstained 4 um tissue sections on electrostatically charged slides containing representative tumor. Tissue must be fixed in formalin; Ambient Methodology: Fluorescent In-Situ Hybridization (FISH) Reference Range: Positive for genotype associated with malignant melanoma Negative for genotype associated with malignant melanoma Days Performed: 1 day per week Reported: 7 days CPT: 88368x4 Price: \$1,245 (non-discountable)	3/26/2013

Discontinued Tests

Test Name	Order Code	Billing Code	Test Information	Effective Date
TRH Stimulation	TRHSTM	82058	This test will no longer be available.	2/19/2013
TRH Stimulation of Prolactin	TRHPRO	82066	This test will no longer be available.	2/19/2013

Discontinuation of Bacterial Antigen Detection

Effective March 1, bacterial antigen detection by latex agglutination will no longer be available through the Robert J. Tomsich Pathology & Laboratory Medicine Institute. The test was introduced in the 1980s to evaluate patients with suspected bacterial meningitis and is no longer recommended because of limited sensitivity and specificity. We recommend ordering CSF culture with gram stain (routinely prepared by cytocentrifugation) to evaluate patients for bacterial meningitis.

Questions may be directed to Sandra Richter, MD (216.444.6519) or Marianne Cline (216.445.0420).