# EEG Recording Checklist

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### Prior to arrival

- 1. Print:
  - Consent form
  - Information sheets
  - Payment sheet
  - Relevant questionnaires
- 2. Fill syringes with gel
- 3. Assign participant number
- 4. If relevant, run stimShuffle script
- 5. Launch stimulus presentation software (E-Prime, Matlab, or Python) and enter information for the current subject
- 6. Connect EEG system to fiber optic cable
- 7. Confirm sampling rate at 512 Hz (1/32) (Decimation)
- 8. Set ActiView config file as BILSWED
- 9. Confirm settings:
  - High pass at 0.05 Hz
  - Low pass at 100 Hz
  - Sampling rate at 512 (Decimation 1/32)
  - Reference: None (raw)
- 10. Set up Swedish keyboard
- 11. Wash hands

### Participant arrival and setup

- 1. Welcome participant
- 2. Consent form
- 3. Give participant Language History Questionnaire and a writing utensil

# Capping

- 1. Measure head size around and ear-to-ear
- 2. Gel and place mastoid electrodes
- 3. Place cap on participant and put gel in each hole
- 4. Attach electrodes to cap
- 5. Place additional electrodes:
  - (a) Mastoids: Left = EX1, Right = EX2
  - (b) Outer canthi of both eyes (2 electrodes): Left = EX3, Right = EX4
  - (c) Above (EX5) AND below (EX6) left eye (2 electrodes)
- 6. Attach electrodes to system
- 7. Check high and low bandpass (.05 100 Hz)
- 8. Check sampling rate (512 Hz (1/32))
- 9. Press "START" in ActiView
- 10. Resolve electrode issues
  - Make sure electrode offset is close to zero for all attached electrodes
  - Make sure offset remains constant when participant moves
  - Check monopolar display to make sure everything looks good
  - In case of issues, remove electrode from cap, scrub gel off the end of it, and replace
  - Add additional gel to electrodes as needed
- 11. Press "START FILE" in Actiview
- 12. Enter unique subject identifier in "Enter Local Subject Identification"

- 13. Select "A1 B32" under "Save subset:"
- 14. Make sure to check "Add 8 EX electrodes"
- 15. Save file in the data folder for your experiment using the unique subject identifier as the filename

### Experimental session

- 1. Seat participant in chair
- 2. Double check instruction file for subject number
- 3. Remind:
  - Don't blink your eyes during the sentences
  - Leave your hands on the keys
  - $\bullet\,$  Try to move as little as possible
  - There will be pauses every about every X minutes
  - If you need a break in addition to these, press "P" during appropriate window (depending on the experiment)
- 4. Press the pink "Paused" button in ActiView to begin recording
- 5. Launch experiment in Matlab, Python, or E-Prime and put in subject number
- 6. Double-check to make sure you actually started recording!

# After the task is done

- 1. Stop recording in ActiView
- 2. Remove cap
- Perform any follow-up tasks or questionnaires as needed
- 4. Debrief
- 5. Show participant to the shower room, and give them a towel, shampoo, etc.
- 6. Copy ID card back/front
- 7. Pay money and sign form

#### Afterwards

- 1. Copy EEG data to backup from BIOSEMI computer
- 2. Copy experimental output files to backup from EXPERIMENT computer – results as well as stimuli for all tasks
- 3. If EEG system battery is low, charge it before putting materials away
- 4. Record any equipment problems in Calpendo

### Clean up

- 1. Put remaining gel from syringes back in the tube
- 2. Wash syringes inside and out
- 3. Wash flat electrodes use toothbrush to get gel off of the sensor
- 4. Wash scalp electrodes
- Remove chinstrap and clean the cap. Make sure no gel remains in any of the holes. Rinse well.
- 6. Hang cap and electrodes to dry