Discovering a motif discovering appproach

LCG_BEII 2019

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Goal of the exercise

The goal of this exercise is to get an intuition of a motif discovery approach relying on the detection of over-represented oligonucleotides.

Our approach will be pragmatic.

We retrieved the upstream non-coding sequences of the genes involved in methionine biosynthesis and sulfur assimilation, and counted the occurrences of each hexanucleotide.

We also computed

- the relative frequencies (occurrences of each oligo / sum of all oligo occurrences) in the sequence of interest (the promoters of methionine-associated genes)
- the relative frequencies of ach hexanucleotide in the whole set of yeast promoters.

We would like to know if some 6nt are over-represented in promoters of methionine-associated genes relative to the occurrences that would be expected from a random selection of yeast promoters.

Create a workspace for this practical

• In your home directory, create a work directory for this practical (for example ~/LCG_BEII/practical_motif_discovery,

```
workdir <- "~/LCG_BEII/practical_motif_discovery"
dir.create(workdir, showWarnings = FALSE, recursive = TRUE)
setwd(workdir)</pre>
```

Loading the data table

1. Download the oligonucleotide count table. Scerevisiae MET-genes oligos-6nt-2str-noov occ-freq.tsv

```
oligo.url <- "http://jvanheld.github.io/LCG_BEII/practicals/motif_discovery/data/Scerevisiae_MET-genes_oligo.file <- basename(oligo.url) ## Suppress the URL path and keep only the file name for local storag download.file(oligo.url, destfile = oligo.file)
```

- 2. In **R**, open a new script or R markdown file.
- 3. Load the data table, print the 5 top rows and the 5 bottom rows.

```
oligo.table <- read.delim(oligo.file, header = 1, row.names = 1)
# View(oligo.table)</pre>
```

head(oligo.table, n = 5)

```
        obs_freq
        exp_freq
        occ
        exp_occ

        aaaaaa|ttttt
        0.004592808
        0.004896299
        41
        43.71

        aaaaac|gtttt
        0.001120197
        0.001998518
        10
        17.84

        aaaaag|ctttt
        0.003696651
        0.003604251
        33
        32.18

        aaaaat|atttt
        0.004032710
        0.004160627
        36
        37.14

        aaaaca|tgttt
        0.001344237
        0.001932479
        12
        17.25
```

tail(oligo.table, n = 5)

```
    obs_freq
    exp_freq
    occ
    exp_occ

    ttccaa|ttggaa
    0.0008961577
    0.0008428396
    8
    7.52

    ttcgaa|ttcgaa
    0.0001120197
    0.0003224542
    1
    2.88

    ttgaaa|tttcaa
    0.0019043352
    0.0019087053
    17
    17.04

    ttgcaa|ttgcaa
    0.0001120197
    0.0004030214
    1
    3.60

    tttaaa|tttaaa
    0.0005600986
    0.0009379354
    5
    8.37
```

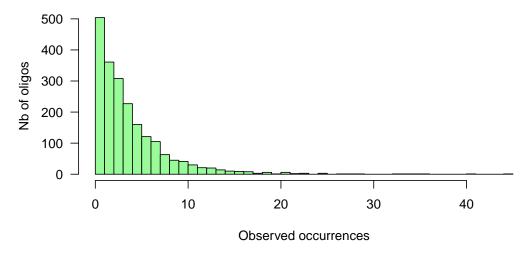
Exploring observed and expected counts

4. Draw an histogram of the observed occurrences and evaluate the spread of counts.

```
x <- oligo.table$occ
range(x)</pre>
```

[1] 0 45

Distribution of oligonucelotide occurrences



5. Draw a scatter plot comparing the observed and expected occurrences for each hexanucleotide.

Observed vs expected occurrences

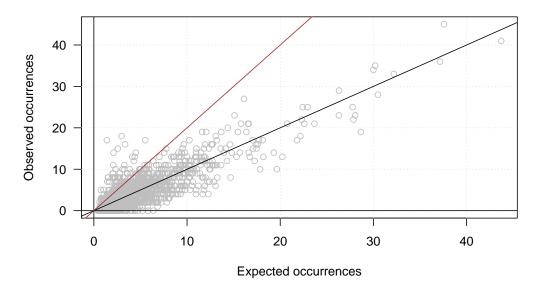


Figure 1: **Scatter plot of observed versus expected occurrences.** The black diagonal corresponds to the null hypothesis, the brown line denotes an arbitrary threshold on fold-change > 2.

6. Compute the ratio of observed / expected occurrences, and draw a scatter plot with this ratio (Y) as a function of the expected occurrences (X).

6. Compute the log-ratio of observed / expected occurrences, and draw a scatter plot with this log-ratio (Y) as a function of the expected occurrences (X).

(obs/exp) ratio

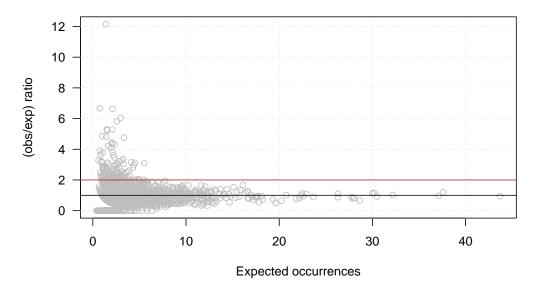


Figure 2: **Scatter plot of observed versus expected occurrences.** The black diagonal corresponds to the null hypothesis, the brown line denotes an arbitrary threshold on fold-change > 2.

$$lr = log(x/ < X >)$$

7. Compute the log-likelihood ratio (llr), defined below, and draw a scatter plot with this llr as a function of the expected occurrences.

$$llr = f \cdot log(x/ < X >)$$

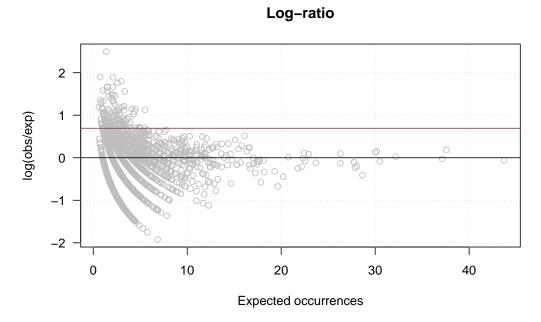


Figure 3: **Scatter plot of observed versus expected occurrences.** The black diagonal corresponds to the null hypothesis, the brown line denotes an arbitrary threshold on fold-change > 2.

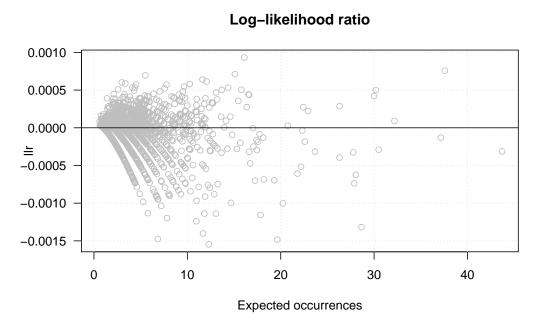


Figure 4: **Scatter plot of log-likelihood ratio (llr) versus expected occurrences.** The black line corresponds to the null hypothesis, the brown line denotes an arbitrary threshold on fold-change > 2.

```
# abline(h = log(2), col = "brown")
```

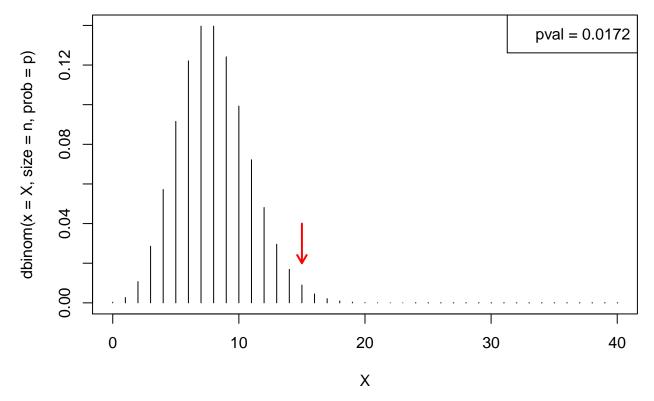
Computing over-representation significance

8. Draw a binomial distribution with parameters n = 8000, p = 0.0001.

```
n <- 8000
p <- 0.001
x <- 15# Number of successes
X <- 0:40 ## values to display

plot(X, dbinom(x = X, size = n, prob = p), type = "h")
arrows(x, 0.04, x, 0.02, lwd = 2, length = 0.1, angle = 30, col = "red")

pval <- pbinom(q = x - 1, size = n, prob = p, lower.tail = FALSE)
legend("topright", legend = paste("pval =", signif(digits = 3, pval)))</pre>
```



8. Use the binomial distribution to compute the P-value of the observed occurrences.

$$P = T(X \ge x)$$

```
x <- oligo.table$obs_freq ## Nuumber of successes
n <- sum(x) ## Number of trials
p <- oligo.table$exp_freq ## Success probability</pre>
```

- 9. Draw an histogram with the P-values of all hexanucleotides, with 20 bins.
- 10. Draw a scatter plot with the P-value (Y) as a function of the log-ratio (X).

11. Compute the E-value, and the significance.

$$E = P \cdot N$$
$$sig = -log_{10}(E)$$

- 12. Draw a Volcano plot, with the significance as a function of the log-ratio.
- 13. Compute the P-value using the Poisson distribution as approximation of the binomial. Are we in suitable conditions for this approximation? Draw a plot comparing the P-values obtained by the binomial and Poisson distributions.
- 14. Compute the P-value using a normal approximation of the binomial distribution.
 - a. Are we in suitable conditions to approximate a binomial with a normal?
 - b. Compare the P-values obtained with the binomial and norma distributions, resp.