target: An R package to Predict Combined Function of Transcription Factors

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MahShaaban/target_pkg_talk

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Outline

Background & problem motivation

Model & implementation

Case study

Summary

Predicting the interaction of two factors using independent datasets of binding and expression

- ► The binding of a transcription factor to a genomic region (e.g. gene promoter) induces or represses its expression [Rougemont and Naef, 2012].
- Transcription factors share their binding sites with other factor, co-factors and/or DNA-binding proteins. The DNA-binding proteins may form complexes which bind to the DNA as one units.
- ► The integration of the overlapping binding sites and the effect of the gene expression of perturbed factors can be used to infer their combined function; cooperative or competitive.

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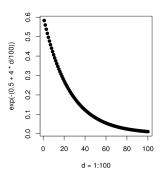
Summary

Modeling the binding sites as the discounted distances of the ChIP peaks

Peak Score (S_p): is the distance (Δ) from transcription start site (TSS) relative to a 100 kb [Wang et al., 2013].

$$S_p = e^{-(0.5+4\Delta)}$$

➤ The shape of the function approximate empirical observations
[Tang et al., 2011].

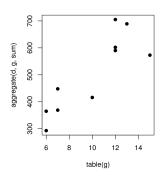


Modeling the regulatory potential as the sum of the weighted peaks

Gene Score (S_g) : is the sum of the scores (S_p) of the k nearby peaks from the TSS.

$$S_g = \sum_{i=1}^k S_{pi}$$

 Regulatory potential increases with the number of binding sites
 [Tang et al., 2011].

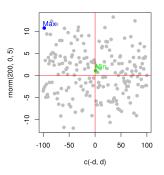


Integrating the factor binding and the expression information

Rank Product (RP_g) : the gene score (R_{gb}) is multiplied by the gene statistics (R_{ge}) from differential expression [Breitling et al., 2004].

$$RP_g = \frac{R_{gb} \times R_{ge}}{n^2}$$

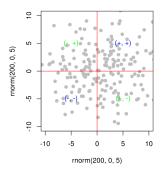
► Integrate the binding events and the functional effect [Tang et al., 2011].



Modeling the interaction of two factors using independent perturbations

Regulatory Interaction (RI): is the product of the gene statistics from differential expression of the perturbation of the two factors (X and Y) separately.

$$RI_g = x_{ge} \times y_{ge}$$
 and, $RP_g = \frac{R_{gb} \times RI_{ge}}{n^2}$

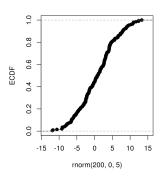


Aggregating the effect of the binding events

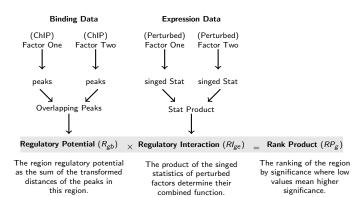
Empirical Cumulative Distribution Function (ECDF):

the proportion of genes in a category (up- or down-regulated genes) that are ranked at or better than the x-axis (regulatory potential value) value [Tang et al., 2011].

Aggregate the effect of the factor perturbation in relation to its regulatory potential.



Workflow for integrating binding and expression data



Functions in the target R package

```
#' Predict associated peaks
#'
   Select overlapping peaks
   get distances between
   peaks and regions, score
#'
   peaks
#'
   Oparam peaks A GRanges
#'
   Oparam regions A GRanges
   Oparam ... Other
#'
   Oreturn A GRanges with
   added metadata columns
#'
   @export
associated_peaks <-
    function (peaks,
              regions,
              . . . )
```

```
#' Predict direct targets
#'
#' Select overlapping peaks
#' get distances between
#' peaks and regions, score
   peaks and regions and
   calculate rank products
#'
   Oparam peaks A GRanges
   Oparam regions A GRanges
   Oparam ... Other
   Oreturn A GRanges with
   added metadata columns
#' @export
direct_targets <-
    function (peaks,
             regions,
```

Functions in the target R package

```
Plot ranks by group
#'
   Plot the cumulative
   distribution of the
   ranks grouped by
   a factor
#'
   Oparam rank A numeric
   Oparam group A factor
   Oparam ... Other
   @export
plot_predicitons <-
  function (rank,
            group,
            . . . )
```

```
#' Test ranks by groups
#' Test whether functions
#' are from the same
#' distribution
   Oparam rank A numeric
   Oparam group A factor
   Oparam compare A
   character of two
   Oparam ... Other
#' @export
test_predicitons <-
    function (rank,
              group,
              compare,
              . . . )
```

Comparison with existing R packages

- ► rTRM attempts to identify the transcriptional regulatory modules (TRMs) which are complexes of transcription factors and co-factors by integrating ChIP, gene expression and protein-protein interactions [Diez et al., 2013]
- ► TFEA.ChIP takes the approach of curating large quantities of data from different sources and using this data to build a model or database where queries of transcription factor targets can be constructed [Puente-Santamaria et al., 2019].
- ▶ transcriptR integrates ChIP- and RNA-Seq data for an entirely different purpose [Karapetyan AR, 2019]. It uses the ChIP data to *denovo* identify transcripts which are then used to quantify the expression in the RNA-Seq data.

Limitations of target

- ► Comparable sets of data for the two factors are required; binding data using ChIP and gene expression data under factor perturbation (overexpression or knockdown).
- Assume that the interaction between two DNA-binding proteins is linear which may not be the case always.
- Cannot detect assisted binding.

Availability

- ▶ target is available as an open source R/Bioconductor package
- ► An interactive application can be invoked locally through R or accessed directly on the web

```
(https://mahshaaban.shinyapps.io/target-app/)
```

The source code for the package and the interactive application are available at

```
(https://github.com/MahShaaban/target).
```

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Case Study: of two evolutionary and functionally related transcription factors YY1 & YY2

Yin And Yang 1 (YY1)

- Belong to transcription factor GLI- Kruppel class of zinc finger proteins
- Involved in repressing and activating a diverse number of promoters
- Direct histone deacetylases and histone acetyltransferases to a promoter

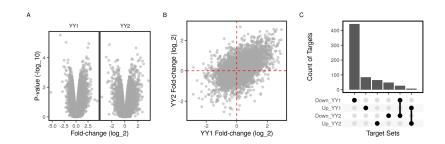
Yin And Yang 2 (YY2)

- Arisen by retrotransposition of the related YY1 gene on chromosome 14
- Exhibit positive and negative control on a large number of genes
- Antagonize YY1 and function in development and differentiation.

Datasets of YY1 and YY2 ChIP-Seq and microarray knockdown in HeLa cells

Dataset	Factor	Type	Source	Ref.
GSE31417 GSE96878 GSE14964 GSE14964	YY2 YY1-kd	ChIP-Seq ChIP-Seq Microarray Microarray	ChIP-Atlas ChIP-Atlas KnockTF KnockTF	[Michaud et al., 2013] [Wu et al., 2017] [Chen et al., 2010] [Chen et al., 2010]

Differential expression of YY1 and YY2 in knockdown vs control HeLa cells

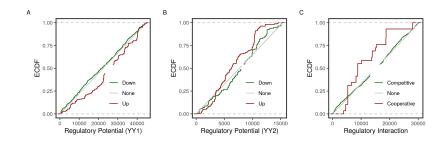


Both factors induce and repress large number of genes.

The effect of the factors knockdown on gene expression is correlated.

Each factor has a large number of specific targets, the two share a small set of targets

Predicted functions of YY1 and YY2 on specific and shared targets in HeLa cells



The knockdown of YY1 & YY2 has opposing effects on their specific targets

YY1 & YY2 cooperate on most shared targets, except for a few strong targets where they compete

Testing YY1 and YY2 combined functions

Factor	Test	Statistic	P Value
YY1	Down vs Up	0.79	0e+00
YY2	Up vs Down	0.41	5e-13
Two Factors	Cooperate vs Compete	0.97	0e + 00

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- We provided a fast and flexible implementation of the BETA algorithm for predicting direct targets of transcription factors from binding and expression data.
- We extended the method to determine the combined function of two factors on the same region.
- ► The algorithm is available as an R package and an interactive web application.
- We applied the method to ChIP-Seq and knockdown microarrays of YY1 & 2 in HeLa cells. We found that the two factors cooperate on most shared targets

Find out more

Ahmed, M., Min, D. & Kim, D. Integrating binding and expression data to predict transcription factors combined function. BMC Genomics 21, 610 (2020).

https://doi.org/10.1186/s12864-020-06977-1

Resources:

- ► Bioconductor page
- ► Package source code
- ► Talk source code
- ► Paper source code

References I



Breitling, R., Armengaud, P., Amtmann, A., and Herzyk, P. (2004).

Rank products: A simple, yet powerful, new method to detect differentially regulated genes in replicated microarray experiments.

FERS Letters.



Chen, L., Shioda, T., Coser, K. R., Lynch, M. C., Yang, C., and Schmidt, E. V. (2010).

Genome-wide analysis of YY2 versus YY1 target genes.

Nucleic Acids Research, 38(12):4011-4026.



Diez, D., Hutchins, A. P., and Miranda-Saavedra, D. (2013).

Systematic identification of transcriptional regulatory modules from protein-protein interaction networks. Nucleic Acids Research.



Karapetyan AR (2019).

An Integrative Tool for ChIP- And RNA-Seq Based Primary Transcripts Detection and Quantification.



Michaud, J., Praz, V., James Faresse, N., Jnbaptiste, C. K., Tyagi, S., Schütz, F., and Herr, W. (2013). HCFC1 is a common component of active human CpG-island promoters and coincides with ZNF143, THAP11, YY1, and GABP transcription factor occupancy.

Genome research, 23(6):907-16.



Puente-Santamaria, L., Wasserman, W. W., and del Peso, L. (2019).

TFEA.ChIP: a tool kit for transcription factor binding site enrichment analysis capitalizing on ChIP-seq datasets.

Bioinformatics.



Rougemont, J. and Naef, F. (2012).

Computational Analysis of Protein-DNA Interactions from ChIP-seq Data.

In Gene Regulatory Networks, pages 263-273. Springer.

References II



Tang, Q., Chen, Y., Meyer, C., Geistlinger, T., Lupien, M., Wang, Q., Liu, T., Zhang, Y., Brown, M., and Liu, X. S. (2011).

A comprehensive view of nuclear receptor cancer cistromes. Cancer research, 71(22):6940–7.



Wang, S., Sun, H., Ma, J., Zang, C., Wang, C., Wang, J., Tang, Q., Meyer, C. A., Zhang, Y., and Liu, X. S. (2013).

Target analysis by integration of transcriptome and ChIP-seq data with BETA.

Nature Protocols.



Wu, X.-N., Shi, T.-T., He, Y.-H., Wang, F.-F., Sang, R., Ding, J.-C., Zhang, W.-J., Shu, X.-Y., Shen, H.-F., Yi, J., Gao, X., and Liu, W. (2017).

Methylation of transcription factor YY2 regulates its transcriptional activity and cell proliferation. Cell discovery, 3:17035.