Statistical fine-mapping in genetics association studies

Contact: Gao Wang (wang.gao@columbia.edu)

Overview

This fine-mapping tutorial was developed using simulated data shipped with susieR package. The simulated data contains simulated phenotype matrix Y in $N \approx 600$ individuals. The goal is to identify from the genotype matrix $X_{N\times P}$ (P=1000) the genetic variables that directly cause changes in the phenotypic values.

The simulated data-set is available here, as part of the susieR package. In this tutorial we will first explore the data with conventional univariate association analysis, then perform preliminary fine-mapping analysis under simplified assumptions to illustrate the idea of statistical fine-mapping, and finally use the R package susieR to perform formal Bayesian fine-mapping analysis. We will also discuss fine-mapping using association summary statistics.

Explore the association data-set

Please execute the following code cell to load the simulated data,

```
library(susieR)
data(N3finemapping)
attach(N3finemapping)
ls()
```

'N3finemapping'

The loaded dataset, N3finemapping, contains regression data *X* and *Y*, along with some other relevant attributes in the context of genetic studies. It also contains the "true" regression coefficients the data is simulated from.

names(N3finemapping)

- 1. 'X'
- 2. 'chrom'
- 3. 'pos'
- 4. 'true coef'
- 5. 'residual variance'
- 6. 'Y'
- 7. 'allele freg'
- 8. 'V

The genotype matrix has 574 individuals and 1001 genetic variants,

dim(X)

- 1. 574
- 2. 1001

The phenotype matrix contains values for the 574 individuals for 2 traits,

dim(Y)

- 1. 574
- 2. 2

We focus on the first trait, let

```
y = Y[,1]
b = true_coef[,1]
```

There are 3 "causal" variants for this trait,

which(b != 0)

- 1. 403
- 2. 653
- 3. 773

Question: Please use R function plot to visualize the true effects vector. What are the effect sizes for these 3 "causal" variants?

Note: in this tutorial we will avoid the phrase "causal", and use "effect variable" to refer to the variants having true non-zero effects on the phenotype.

Association testing via simple linear regression

univariate_regression function in susieR can be used to perform association testing by fitting univariate simple regression for each variable separately. The results are $\hat{\beta}$ for effect size estimates, and their standard errors $SE(\hat{\beta})$.

```
sumstats <- univariate regression(X, y)</pre>
```

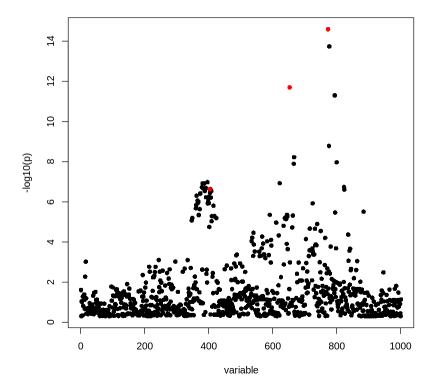
Question: Please use R function plot to compare the true effects variable b with the estimated effect sizes sumstats\$betahat. Are they consistent, and why?

z-scores can be derived from these quantities; $-log_{10}$ p-values can then becomputed from z-scores,

```
z_scores <- sumstats$betahat / sumstats$sebetahat
log10p <- -log10(pchisq(z_scores^2,1,lower.tail=F))</pre>
```

Question: How many variables have p-values smaller than 10^{-8} ? Would you report all of them as significant association findings, and if not, which variable(s) will you likely report, based on univariate association analysis?

susieR package provides a plot function to generate Manhattan plot for the association test results. Notice that the true effect variables are labelled in red,



As shown on this plot, the "lead" SNP (the one with smallest p-value) is indeed one of the true effect variables. However, the 2nd most significant SNP is not one of the other two effect variables, and there are many other SNPs having p-values smaller than 10^{-8} , a widely accepted significant level threshold for genetic association studies.

"One causal variant" assumption for fine-mapping

Under the assumption that there is only one true effect in the region of interest, fine-mapping does not have to use linkage disequilibrium (LD) information between variables. It can be performed by simply calculating the likelihood of the association model for each variable separately, then computing the probability for a variable being the true effect as the ratio of its likelihood over the sum of likelihoods over all variables. We use linear regression 1m in R to compute the log-likelihood,

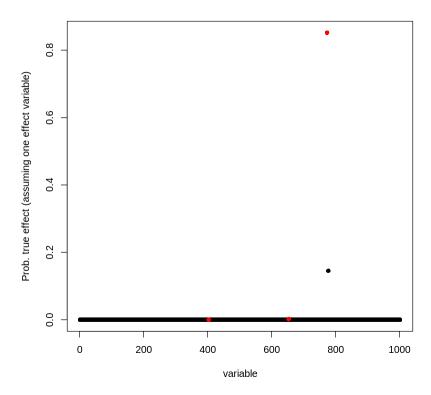
```
lliks = sapply(1:ncol(X), function(i) logLik(lm(y~X[,i])))
```

Then compute the probability for each variable being the true effect variable:

```
lliks = lliks - max(lliks) # To avoid taking exp on large numbers in the next
line, thus improving numerical stability
probs = exp(lliks)/sum(exp(lliks))
```

We use susie_plot function to plot these probabilities computed, and color label the true effect variables:

susie_plot(probs, y="PIP", b=b, ylab = "Prob. true effect (assuming one
effect variable)")



As expected, under the "one causal variant" assumption, only the lead SNP is identified with over 80% probability of having a non-zero effect. The other two true effect variables in red are not identified.

95% credible set for single effect

95% credible set (CS) is defined as a set of variables with 95% probability of containing a true effect. By visually examine the plot above, we can see that the lead SNP has over 80% probability being the true effect, and another SNP near it has almost 20% probability of being the true effect. The 95% credible set should thus contain both of them in order for

the coverage to exceed 95%. Indeed, we can formally verify this using an internal function in_CS to assess if a variable is in a single effect CS. We report the indices of these variables,

```
which(susieR:::in_CS(t(probs), coverage = 0.95) != 0)
```

- 1. 773
- 2. 777

These variables indeed correspond to the two non-zero probability variables on the plot above.

Question: Show the p-values and compute the correlation between these two variables. What do you think is the reason that the zero-effect variable 777 shows a significant evidence of association?

Fine-mapping with individual level data using Sum of Single Effects model

When multiple true effects are expected, formal statistical fine-mapping via Bayesian Variable Selection should be performed. We fit a Sum of Single Effects (SuSiE) model implemented in susieR package, assuming at most 10 causal variables (although SuSiE is robust to the choice of L).

```
fitted <- susie(X, y, L = 10)
```

95% credible sets for each single effect

```
By default, susie function computes 95% CS each containing one effect variable,
```

```
print(fitted$sets)
$cs
$cs$L2
[1] 653
$cs$L1
[1] 773 777
$cs$L3
[1] 362 365 372 373 374 379 381 383 384 386 387 388 389 391 392 396 397 398
[20] 400 401 403 404 405 407 408 415
$purity
   min.abs.corr mean.abs.corr median.abs.corr
L2
      1.0000000
                    1.0000000
                                    1.0000000
L1
      0.9815726
                    0.9907863
                                    0.9907863
```

```
L3 0.8686309 0.9653503 0.9739578

$cs_index
[1] 2 1 3

$coverage
[1] 0.9998236 0.9988858 0.9539811
```

\$requested_coverage
[1] 0.95

The 3 true signals have been captured by the 3 CS reported here. The 3rd CS contains many variables, including the true causal variable 403. The minimum absolute correlation is 0.86.

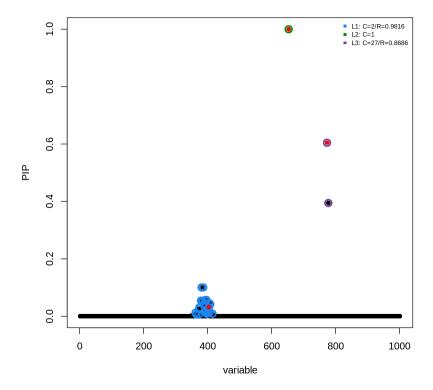
If we request 90% coverage for credible sets, we still capture the 3 signals, but the minimum absolute correlation of the 3rd CS is now 0.91 and size of the CS is also a bit smaller.

```
sets <- susie_get_cs(fitted, X = X, coverage = 0.9)</pre>
print(sets)
$cs
$cs$L2
[1] 653
$cs$L1
[1] 773 777
$cs$L3
[1] 373 374 379 381 383 384 386 387 388 389 391 392 396 398 399 400 401 403
404
[20] 405 407 408
$purity
  min.abs.corr mean.abs.corr median.abs.corr
L2
     1.0000000
                   1.0000000
                                    1.0000000
L1
     0.9815726
                   0.9907863
                                   0.9907863
L3
     0.9119572
                   0.9738724
                                  0.9768163
$cs_index
[1] 2 1 3
$coverage
[1] 0.9998236 0.9988858 0.9119917
$requested_coverage
[1] 0.9
```

Question: why is the size for the 3rd CS smaller and minimum absolute correlation higher, when requesting 90% coverage compared to previous result of requesting 95%?

Posterior inclusion probabilities

Here we summarize the fine-mapping results by posterior inclusion probability (PIP): susie_plot(fitted, y="PIP", b=b, add_legend=T)



The true effect variables are colored red, and the three 95% CS identified are circled in different colors.

Question: Report the PIP for variable 403. Why is the PIP relatively low for this true **effect variable?** You can use the code below to compare $-log_{10}$ p-value and PIP for the CS containing variable 403:

```
i <- fitted$sets$cs$L3
z3 <- cbind(i,log10p[i],fitted$pip[i])
colnames(z3) <- c('position', '-log10 pvalue', 'PIP')
z3[order(z3[,2], decreasing = TRUE),]
A matrix: 27 × 3 of type dbl
position -log10 pvalue PIP</pre>
```

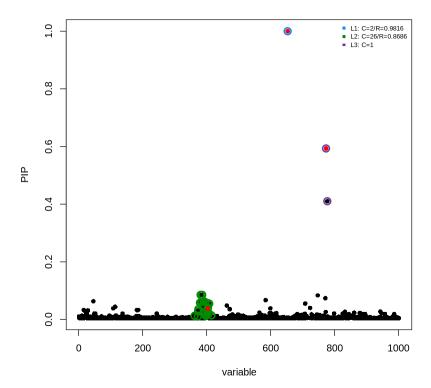
396	6.676732	0.056704331
381	6.618518	0.100360243
386	6.618518	0.100360243
379	6.417587	0.054179507
391	6.396640	0.055952118
383	6.370810	0.052896918
384	6.370810	0.052896918
389	6.360493	0.042161265
405	6.331181	0.045761975
403	6.322861	0.031992848
387	6.272121	0.041041505
388	6.237010	0.039650079
408	6.230055	0.041551961
404	6.139356	0.032013339
374	6.125189	0.030571484
373	6.094780	0.023577221
362	6.005750	0.012145481
399	5.932096	0.026454056
392	5.922454	0.019741011
407	5.906902	0.014699313
400	5.859007	0.021659443
365	5.762214	0.006263425
398	5.694203	0.012907848
401	5.634175	0.014858460
397	5.620728	0.008690915
415	5.506796	0.009003129
372	5.335809	0.005886458

Question: Please rerun the susie analysis setting L=1 to detect a maximum of only one effect across the region. How does the result compare to the previous section of "one causal effect" fine-mapping using likelihoods from univariate analysis?

Choice of prior effect size

In SuSiE, the prior effect size is parameterized by the percentage of variance explained by a variable, h_j^2 . By default, SuSiE estimates h_j^2 and uses it as the prior (an empirical Bayes approach). Here we demonstrate fixing the prior effect size to 0.2, i.e., we believe a non-zero effect variable can explain 20% variation in quantitative trait values:

```
fitted2 = susie(X, y, L = 10, estimate_prior_variance = FALSE,
scaled_prior_variance = 0.2)
susie_plot(fitted2, y='PIP', b=b, add_legend=T)
```



The result is very similar to when prior effects are estimated.

Question: Please change scaled_prior_variance to different numbers between 0 and 1, and examine the results. What happens if you set it to 0.001, and why?

Fine-mapping with summary statistics via <code>susie_rss</code>

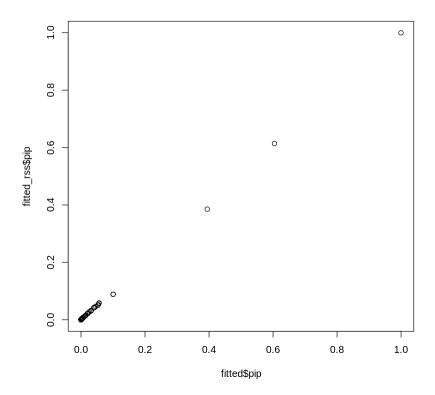
Summary statistics of genetic association studies typically contain effect size ($\hat{\beta}$ coefficient from regression), p-value and minor allele frequencies. These information can be used to perform fine-mapping with given an additional input of correlation matrix between variables. The correlation matrix in genetics is typically referred to as LD matrix (LD for linkage disequilibrium). One may use external reference panels to estimate it when this matrix cannot be obtained from samples directly. There is a potential problem using LD from reference panel, and we demonstrated it in the next section. Caution that LD matrix here has to be correlation matrix r, not r^2 or abs(r).

We have previously computed z-scores for this data-set. The correlation matrix can be computed directly from X matrix,

```
R \leftarrow cor(X)
```

SuSiE regression with summary statistics is implemented as susie_rss function, fitted_rss <- susie_rss(z_scores, R, L = 10)

We compare the PIP obtained with previous analysis using individual level data, plot(fitted\$pip, fitted_rss\$pip, ylim=c(0,1))



The results very much agree. This is expected when the summary statistics are indeed derived from the individual level data.

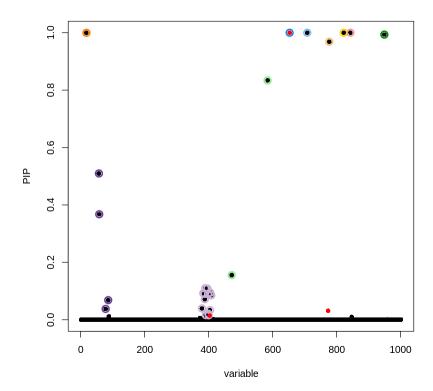
Use LD from external reference data

When original genotype information is not available, one may use reference panel to estimate LD matrix. False discoveries may occur if the LD pattern from the reference panel does not agree well with the original sample.

To illustrate, we randomly generated 500 samples from N(0, R) and treated them as reference panel genotype matrix X_ref .

```
set.seed(1234)
tmp = matrix(rnorm(500*1001), 500, 1001)
eigenR = eigen(R)
eigenR$values[eigenR$values < 1e-10] = 0</pre>
```

Using susie_rss function with LD from reference panel,



Question: Please compare and comment on the CS reported using reference LD panel versus using the correct LD computed from genotype data. Are there any false discoveries?

We recommend the following modification to the LD matrix when reference panel (R_{ref}) are used,

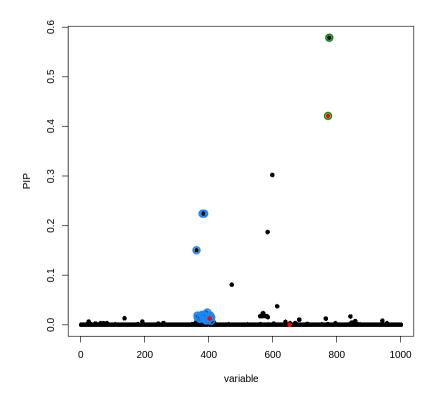
$$\tilde{R} = \text{cov2cor}((1 - w)R_{ref} + wzz^{\mathsf{T}})),$$

where z is the original summary statistics. We suggest using $w=1/n_{ref}$, n_{ref} is the sample size from the reference panel. The modified LD matrix becomes

$$\tilde{R} = \operatorname{cov2cor}(\frac{n_{ref} - 1}{n_{ref}} R_{ref} + \frac{1}{n_{ref}} z z^{\mathsf{T}})) = \operatorname{cov2cor}(\frac{1}{n_{ref}} (X_{ref}^{\mathsf{T}} X_{ref} + z z^{\mathsf{T}})),$$

where X_ref is column-standardized. This modification is implemented in susie_rss with parameter z_ld_weight:

```
n_ref = 500
fitted_rss_ref_corrected <- susie_rss(z_scores, R_ref, z_ld_weight = 1/n_ref,
L = 10)
susie_plot(fitted_rss_ref_corrected, y="PIP", b=b)</pre>
```



Question: Please compare and comment on the results using corrected reference LD panel. Is the result improved?

Exercise: fine-mapping analysis on a different data-set

Please load and analyze another data-set provided in susieR package:

data(N2finemapping)

Please use the same R code for N3finemapping data to analyze this new data-set, similarly focusing on the first column of the phenotype matrix as the phenotype of interest. This data-set contains two true effect variables. However, as you are soon to find out in univariate linear regression analysis, the lead SNP via univariate analysis is neither of the two effect variables.

Question: Perform fine-mapping under "one causal variant" assumption. Does it capture any true effect? Does fine-mapping with SuSiE capture all true effects?

sessionInfo()

R version 3.6.2 (2019-12-12)

Platform: x86_64-pc-linux-gnu (64-bit) Running under: Ubuntu 18.04.2 LTS

Matrix products: default

BLAS: /usr/lib/x86_64-linux-gnu/atlas/libblas.so.3.10.3 LAPACK: /usr/lib/x86_64-linux-gnu/atlas/liblapack.so.3.10.3

locale:

- [1] LC_CTYPE=en_US.UTF-8 LC_NUMERIC=C
- [3] LC_TIME=en_US.UTF-8 LC_COLLATE=en_US.UTF-8 LC_MONETARY=en_US.UTF-8
- [7] LC_PAPER=en_US.UTF-8 LC_NAME=C
- [9] LC_ADDRESS=C LC_TELEPHONE=C
- [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

attached base packages:

[1] stats graphics grDevices utils datasets methods base

other attached packages:

[1] susieR_0.10.0

loaded via a namespace (and not attached):

- [1] Rcpp_1.0.6 magrittr_2.0.1 tidyselect_0.2.5 munsell_0.5.0
- [5] uuid_0.1-2 colorspace_2.0-0 lattice_0.20-38 R6_2.5.0
- [9] rlang_0.4.10 plyr_1.8.6 dplyr_0.8.3 tools_3.6.2
- [13] grid 3.6.2 gtable 0.3.0 htmltools 0.4.0 ellipsis 0.3.1
- [17] assertthat_0.2.1 digest_0.6.27 tibble_3.0.5 lifecycle_0.2.0
- [21] crayon 1.3.4 Matrix 1.2-18 IRdisplay 0.7.0 purrr 0.3.3
- [25] ggplot2_3.3.3 repr_1.0.2 vctrs_0.3.6 base64enc_0.1-3
- [29] IRkernel_1.1 glue_1.4.2 evaluate_0.14 pbdZMQ_0.3-3
- [33] compiler_3.6.2 pillar_1.4.7 scales_1.1.1 reshape_0.8.8
- [37] jsonlite_1.7.2 pkgconfig_2.0.3