

Cell Differentiation, Cell Proliferation and Cell Cycle

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1 Introduction

In developmental biology, **cell differentiation** is the process by which a less specialized cell becomes a more specialized cell type. Cell differentiation produces variety of cells rooted from fertilized egg. These differences are largely due to highly-controlled modifications in gene expression, which almost never change DNA sequence itself. In mammals, only the zygote and early embryonic cells are able to differentiate into all cell types, which is hard to isolate. Adult stem cells only partially differentiate. We will induce U-937 cell to adhesion cell or macrophage by PMA as following.

The U-937 cell line is a human hematopoietic cell line from a 39 years old man with histiocytic lymphoma, which is an oncogenic human monocyte cell line. Phorbol 12-Myristate 13-Acetate (PMA) is a phorbol ester used to convert monocytes into macrophages. With addition of the PMA, the non-adherent U-937 cell up-regulate their adhesion molecules to attach to the flask surface and differentiate. PMA enhances the expression of CD11b and PKC, while increases $\beta - 2$ integrins to help monocytes to attach to flask surface. Then PMA activates calcium and phospholipid dependent isoforms of PKC and stimulates cyclic AMP metabolism, causing maturation into a macrophage-like morphology. We will observe morphology change during differentiation time.

Not only cell differentiation, but also cell proliferation is the fundamental of living organisms. Cell cycle is the series of events that take place in a cell leading to its division and duplication of its DNA to produce two daughter cells.[2] Normally, cell cycle could be separated into 3 stages:

- Interphase: the cell grows, proteins are made, the number of organelles increase, DNA replication. This stage contains 3 phases: G_1 phase, S phase, G_2 phase. The amount of genetic information (in human) will double to $4n$ from $2n$, but keep same chromosome number ($2n$). G_1 cells have half of DNA content of the G_2 cell.
- Division of the nucleus (mitosis or meiosis): In mitosis, this stage called M phase, when chromosome separated. M phase can be separated to 4 subphases. The amount of genetic information (in human) will change to $2n$ from $4n$ after M phase and cytokinesis.

- Cytokinesis: the cytoplasm divides between the new daughter nuclei.

So, G_1 , S, and G2/M cells can be distinguished based on their DNA content. In this Lab, we will use the flow cytometry to character cell cycle based on the propidium iodide(PI) stain for nucleic acids. PI bind to DNA and RNA by interaction with base.

Flow cytometry can handle larger population of cells for obtaining more data than other method, which has 3 main components in the system to operate cell and collect data:

1. Fluidics System: Hydrodynamic focusing
2. Optics System: produce excited light, collect emission fluorescence(also called side-scattered light) and forward-scattered light.
3. Electronics System: generate electrical pulse, transform light signal to electrical signal and store data.

Flow cytometry data could be analysed in 3 features: A(area), W(width),H(height). We apply width and height to deduce cell cycle as following:

- If compared to normal cells(2n), area is twice more, width doesn't change, then it should be a 4n cell
- If compared to normal cells(2n), both area and width are twice more, then it may be a double 2n cell together.

2 Materials & Methods

U-937 cells, PI, Flow cytometry(Beckman Coulter Cytoflex), PBS(phosphate buffer saline), PMA(Santa Cruz, SC-3576, MV6161.83g/mol, dissolved DMSO, 10mM, stock solution), AccutaseTM(sigma A6964), 70% alcohol(-20 precool), PI staining solution(50 μ g/ml PI, 100 μ g/ml RnaA, 0.2% TrionX-100 in PBS), RMI1640 supplied with 10%FBS(cell culture medium)

Centrifuge, Inverted Microscopy, CO₂ incubator, hood, pipettes and tip, cell density counter, 60mm tissue culture dish, 1.5mL centrifuge tube, 15ml centrifuge tube, Pasteur pipette, Flow cytometer, refrigerator, 5mL FCM tube

2.1 Cell difference

Transfer cell suspension to 15ml tube centrifuge at 120xg for 3min → Discard supernant, resuspend the cells with 1mL RPMI 1640 → count cell and take 10^6

cell to 60mm dish with 4mL 1640 → add 4 μ L PMA to experimental group + add 4 μ L DMSO to control group → culture in CO₂ incubator for 24hr, 48hr, 72hr.

2.2 Characteristic cell cycle with FACS

Induce U-937 cell with PMA (final solution 10mM) for 72hr → harvest cell. For control group, directly transfer to 15ml tube. For experiment group cell: wash dish with 2ml PBS, transfer to tube; digest residual cell with 1ml Accutase solution in 37°C incubator; terminate 2ml complete medium; finally transfer to 15ml tube. → Spin down cell, centrifuge 120xg, 5min → discard supernatant → Flick the tube → wash cell with 3ml cold PBS → centrifuge 120Xg, 5min, 4°C → Discard supernatant, flick tube → Add 2ml cold 70% ethanol **slowly** → Fix cells at -20°C for 1 hr.

Centrifuge 250xg, 5min → Discard ethanol → Wash with 3ml of cold PBS → Flick the tube, avoid clumping of cells → Suspend with 3ml PBS, add PI staining solution to working concentration. Stain at dark, RT, 30min → Transfer cells through the cell strainer to 5ml FCM tube. → Run flow cytometry

3 Result

- PMA successfully induce the differentiation of U-937 cell. Differentiated U-937 tend to attach to surface of dish. (Figure 3,4)
- The longer U-937 was treated, more cell differentiated. (Figure 1,3 and Figure 2,4)
- 52.6% of untreated U-937 cell is in G_1 phase, 16% of untreated cell is in G_2 or M phase (Figure 7). Almost all PMA treated cell stayed on G_1 phase. (Figure 8)

4 Discussion

How PMA effect cell cycle?

As we known, protein kinase C (PKC) play an important role in vascular endothelial cell proliferation. We found following phenomena from our and other data:

1. when added to G_2 cells, PMA inhibited subsequent cell division; (our flow cytometer data Figure 7, 8)
2. these growth-arrested cells did not show morphological features of mitotic cells[1]

3. PMA did not interrupt mitosis in cells released from nocodazole-induced M phase arrest.[1]

To solve these problem, other expriment showed that,

1. 1-Oleoyl-2-acetyl-sn-glycerol (OAG) added repeatedly from G2 also inhibited mitosis. The activation of cdc2 kinase around the G2/M transition was suppressed by PMA and OAG.
2. Although cdc2 was expressed in the presence of PMA, dephosphorylation of its tyrosine residue was inhibited by PMA.
3. In parallel, the expression of cdc25B was suppressed by PMA. The total and the cdc2-associated amount of cyclin B were both reduced by PMA.

Sumary up, experiment data suggested that the PKC pathway negatively regulates the G2/M transition and that the inhibition of cdc2 kinase by the reduction in the levels of cdc25B and cyclin B may contribute to this effect.[1]

5 Figures

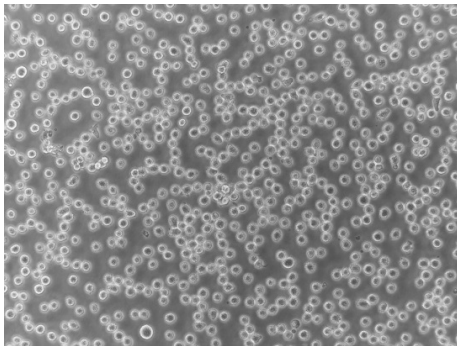


Figure 1. No treated U-937 after 24hr(20x)

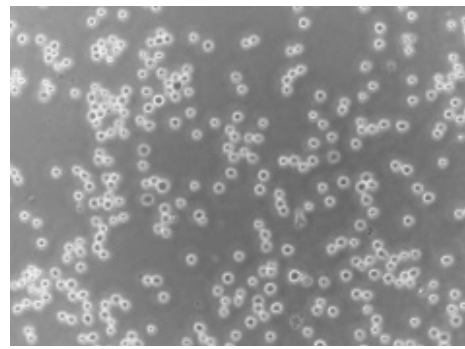


Figure 2. PMA treated U-937 after 24hr

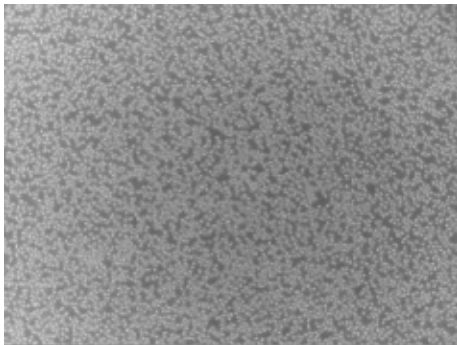


Figure 3. No Treated U-937 after 72hr(20x)

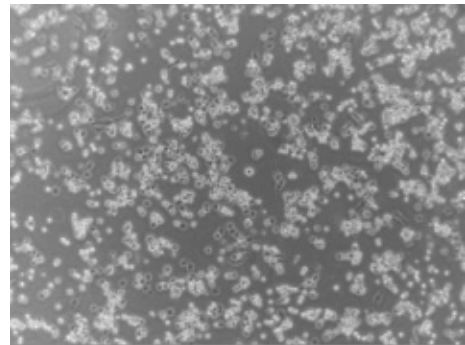


Figure 4. PMA treated U-937 after 72hr

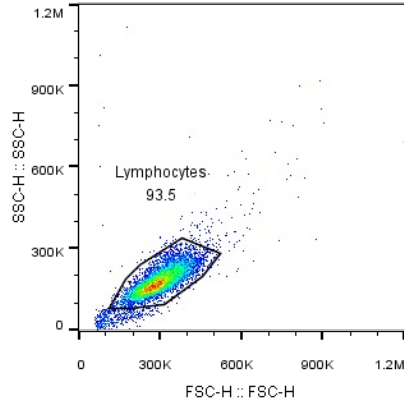


Figure 5. No treated U-937 cell data(all cell)

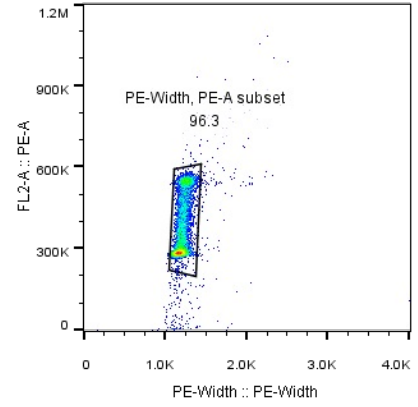


Figure 6. No treated U-937 cell data(gated)

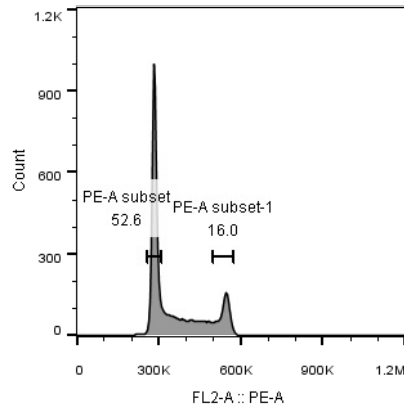


Figure 7. No treated U-937 cell fluorescence distribution

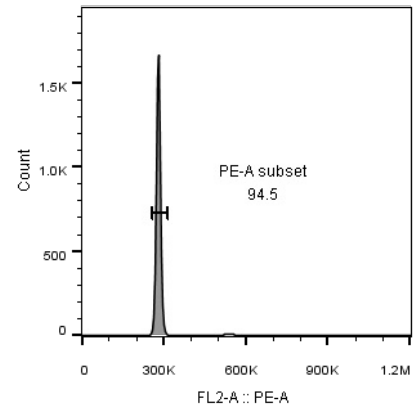


Figure 8. PMA treated U-937 cell fluorescence distribution

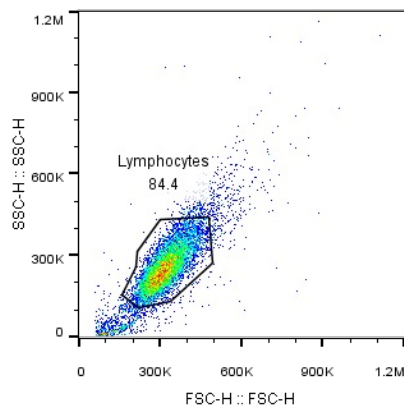


Figure 9. PMA treated U-937 data

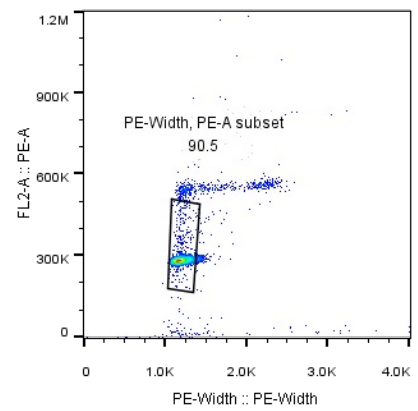


Figure 10. (gated)PMA treated U-937 data

6 Reference

1. Cell cycle arrest in the G2 phase induced by phorbol ester and diacylglycerol in vascular endothelial cells C. Kosaka, T. Sasaguri, A. Ishida, and J. Ogata. American Journal of Physiology-Cell Physiology 1996 270:1, C170-C178
2. https://en.wikipedia.org/wiki/Cell_cycle

7 Contribution

Wenhao zhang & I finished these work together. Thanks to Teacher Jia and Zhao, who parepare materials and teach the experiment pricinple