

# DNA methylation Haplotypes

*Jiantao Shi*

19 May 2018

**Abstract**

DNA methylation haplotypes represent methylation status of cytosines along single DNA molecules. Few published tools can extract DNA methylation haplotypes conveniently. Here we present a Java tool that could extract DNA methylation haplotypes from BAM files generated by popular aligners (BSMAP, BISMARK and MAQ) for bisulfite sequences.

## Contents

1	Input files . . . . .	2
1.1	BAM files. . . . .	2
1.2	CpG location files. . . . .	2
1.3	Interval. . . . .	2
2	Usage . . . . .	2
3	output files. . . . .	3
3.1	Text file. . . . .	3
3.2	Tabix indexed file . . . . .	3

# 1 Input files

---

`mHaplotype` requires indexed BAM files and CpG location files to run.

## 1.1 BAM files

Sorted and indexed BAM files are standard output of most pipelines for bisulfite sequences. Currently, `mHaplotype` mainly support BAM files generated by BSMAP, BISMARK and MAQ.

1. **BSMAP** is one of the fastest aligner for bisulfite sequences. Please use `-R` option when running BSMAP, which generates tag `ZS:Z` in resulting BAM file, in which `++` or `+-` for reads from watson strand and `-+` or `--` for reads from crick strand. You may refer to BSMAP publication for details.
2. **BISMARK** is another aligner for bisulfite sequences with rich QC information. `mHaplotype` check SAM flag of each read. Reads with flag 99 or 147 will be parsed as watson strand, and 83 or 163 as crick strand.
3. **MAQ** is not designed for bisulfite sequences but has been used by some groups. If the aligner is specified as MAQ, then strand information is inferred from [SAM flags](#). Specifically, if `read reverse strand` is detected, the read is parsed as crick strand, otherwise watson strand.

## 1.2 CpG location files

`mHaplotype` require a folder with CpG location files. They must be named as `chr14.txt`. Each file contains two columns, second column must be CpG location.

## 1.3 Interval

`mHaplotype` process one interval at a time. An interval, in the format of `14:57248000-57293348`, is needed in command line.

# 2 Usage

---

`mHaplotype` is designed to capture standard input so that it could work with `samtools`, which could be used to filter out low quality reads using option `-F 3840`, such as `not primary alignment`, `read fails platform/vendor quality checks`, `read is PCR or optical duplicate` and `supplementary alignment`. When running `mHaplotype` without any option, help will be printed, as shown below.

```
java -Xmx4g -jar haplotype.jar
  -T bam2haplotype
  -A [BSMAP, MAQ, BISMARK]
  -C CpG position folder
  -i Interval String
```

## DNA methylation Haplotypes

```
-O Output file name
```

We have included example BAM file in folder `exampleData` and one CpG position file in folder `CpG/hg19`. An typical command looks like this:

```
samtools view -F 3840 exampleData/GE0_OTX2_Cancer.bam | java -Xmx4g -jar mHaplotype.jar -A BSMAP -T bam2haplot
```

## 3 output files

---

### 3.1 Text file

`mHaplotype` output a text file with three columns: Genomic interval, Haplotype, Counts. Genomic interval is defined as the first and last CpG site position for each haplotype. The above command generate output file `GE0_OTX2_Cancer.txt`. The first few lines of this file is listed below.

```
14:57263949-57264186    1111111111 1
14:57279428-57279463    00  38
14:57279428-57279463    01  2
```

### 3.2 Tabix indexed file

The haplotype file could very large especially for WGBS data. Fortunately, it is a genomic position-based file and could be indexed by [Tabix](#) for fastq query. The bash script below convert unsorted haplotype file to indexed haplotype file.

```
cat GE0_OTX2_Cancer.txt | sed 's/[: -]/\t/g' | sort -k1,1 -k2,2n | bgzip > GE0_OTX2_Cancer.gz
tabix -b 2 -e 3 -p bed GE0_OTX2_Cancer.gz
```