

FRAILTY MODEL INCORPORATING ASCERTAINMENT CORRECTION WITH MISSING
DATA IN FAMILY-BASED STUDY

by

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Abstract

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This is an abstract section...

To my mother, Kai Hua, for all the upbringings.
To my father, Guangmin Bi, for teaching me not to give up.
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List of Symbols

i	Individual index
j	Family (Cluster) index
p	Proband index
d_j	Number of events in family j
t	Some time
a	Some Time for the proband
T	Event Time
δ_{ij}	Event indicator for individual i in family j
w	The observed survival data (t, δ)
n	Number of individuals
J	Number of Families (Clusters)
m	Index of the sampled completed dataset in the MCEM
M	Number of the sampled completed dataset in the MCEM
z	Frailty term
q	q -th element of Gauss Hermite Quadrature
ω	q -th weight of Gauss Hermite Quadrature
y_q	q -th node of Gauss Hermite Quadrature
N_q	Total number of quadratures
$h(\cdot)$	Hazard fuction
$h_0(\cdot)$	Baseline hazard function
$H(\cdot)$	Cumulative hazard fuction
$S(\cdot)$	Survival fuction
$A_j(\cdot)$	Ascertainment of family j into the study
$L(\cdot)$	Likelihood function
$\ell(\cdot)$	Log-likelihood function
$\mathcal{L}(\cdot)$	Laplace transform
\mathbf{x}	Covariates
$\boldsymbol{\beta}$	Model coefficients vector
$\boldsymbol{\theta}$	Parameter vector
Λ	The combination of $(\boldsymbol{\beta}, \lambda, \alpha)$

λ	Weibull shape parameter
α	Weibull scale parameter
v	General form of the parameter in an undefined frailty distribution
k	Gamma shape and rate parameters
σ^2	Log-Normal variance parameter
ψ	Missing data distribution parameters

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Chapter 1

Introduction

1.1 Background

The Breast Cancer type 1/2, usually referred as BRCA1/2, are proteins that consists of genes that code for BRCA1 in humans. BRCA1/2 are human tumor suppressor genes, that are responsible for repairing the DNA [1]. When the mutation exists on these genes may cause the impairments of proper functions, which can lead to the possibility of capturing the breast, ovarian, or other specific cancers [2, 3]. Inheriting one of these mutations does not guarantee developing cancer disease, but the mutation can increase the risk of getting those cancers.

In the field of medicine, these cancer types are classified as Hereditary Breast and Ovarian Cancer Syndrome (HBOC). The average life expectancy of individuals with BRCA1, without any interventions, is approximately 4.2 years shorter than that of non-carriers of the BRCA1 gene [4]. Significant advancements have been made in the medical and statistical modeling of breast cancer risk among BRCA1/2 carriers. These include the application of competing risk survival analysis based on breast and ovarian cancer outcomes developed by Choi et al. [5], as well as various clinical trials investigating risk-reducing treatment approaches for breast cancer patients [6]. Despite these efforts, it remains crucial to ensure statistical validity across these studies especially when missing data exists.

From a statistical perspective, the study is centered on a specific disease, which may introduce selection bias due to the sampling process. This bias arises from the selection criteria based on specific probands in each family. To mitigate this sampling bias, an ascertainment correction should be applied to the likelihood calculation, conditioning on the proband information. To accurately capture the heterogeneity between families in the context of time-to-cancer outcomes,

the use of a frailty model is recommended. There are various choices for frailty distributions in survival analysis, including the Gamma distribution and the log-Normal distribution.

1.2 Motivation

Although numerous studies on risk assessment in susceptible populations and statistical advancements in dynamic prediction have significantly contributed to understanding BRCA1/2 families, the issue of missing data remains a substantial challenge, particularly in the context of survival outcomes. Over the past decade, several methodologies have been proposed to address missing data, including the Expectation-Maximization (EM) algorithm, the Monte-Carlo EM algorithm for cases where the E-step lacks a closed form, and Multiple Imputation (MI). However, when applying frailty models, which incorporate random effects in survival analysis, the literature addressing missing data is relatively sparse.

In genetic epidemiology, research is typically conducted on a family-wise basis. Therefore, considering the family structure when addressing statistical problems is both essential and unavoidable. Moreover, existing techniques for handling missing data must be carefully adapted, as the clustered nature of the dataset introduces additional complexity. Within the genetic framework, many variables, such as genetic information and polygenic risk scores (PRS), are not independent between individuals. Traditional methodologies often fail to account for family correlations and ascertainment bias. Given that families are selected based on a proband, it is crucial to apply ascertainment correction to minimize the selection bias. This situation presents an opportunity to further investigate and develop adequate methods for handling missing data, taking into account family correlations and ascertainment bias.

In this project, we aim to investigate the current implementation of Multiple Imputation (MI) methods for frailty models. Additionally, we propose a novel MI method that explicitly incorporates the kinship matrix during the imputation of genetically related variables. This proposed method will be evaluated by comparing it to existing MI methods and Complete Case Analysis (CCA).

1.3 Objectives

With the proposed MI method and the BRCA1 data, the objectives of this thesis are designed as follows:

1. To adapt the kinship correlations into the imputation step
2. To incorporate the ascertainment correction into the likelihood while considering that not all probands are affected
3. To assess the novel MI method via the calculation of the estimations, biases, and precisions through the simulation study
4. To apply the novel MI method and adjusted likelihood to model the BRCA1 family data

1.4 Organizations of the Thesis

Chapter 2

Literature Reviews

2.1 Survival Analysis

Survival analysis is a robust statistical methodology used to analyze time-to-event data, where the focus is on the time until an event of interest occurs. It has been extensively applied in medical research, particularly in studies involving cancer, where events such as death or relapse are critical endpoints. The literature identifies several key methods and models that form the backbone of survival analysis. Kaplan and Meier [7] introduced the Kaplan-Meier estimator, a nonparametric statistic used to estimate survival functions from incomplete observations, which remains widely used due to its simplicity and effectiveness in handling censored data. Cox [8] proposed the proportional hazards model, which allows for the inclusion of covariates and has become a standard technique for assessing the effect of explanatory variables on survival. Collett [9], Machin et al. [10], and Kleinbaum and Klein [11] provide detailed expositions on survival analysis methods, including parametric and nonparametric approaches. Recent advancements have addressed complex issues such as interval censoring and competing risks, expanding the applicability and precision of survival analysis. These methodological developments have significantly enhanced the ability to make informed inferences from survival data, contributing to more accurate prognostic assessments and treatment evaluations in clinical research. In the survival analysis, there are several key functions that will contribute essentially to nearly all relevant scientific work.

The survival function $S(t)$ measures the probability of study subject's entry age t is less than the event time T ,

$$S(t) = P(T > t) \tag{2.1}$$

The cumulative distribution function $F(t)$ represents the probability that the event has occurred by time t . This is the complement of the survival function $S(t)$,

$$F(t) = P(T \leq t) = 1 - S(t) \quad (2.2)$$

The probability density function $f(t)$ is the likelihood of the event occurring at an exact time t . It is the derivative of the cumulative distribution function (CDF) or the negative derivative of the survival function.

$$f(t) = \frac{d}{dt}F(t) = -\frac{d}{dt}S(t) = h(t)S(t) \quad (2.3)$$

The hazard function $h(t)$ measures the instantaneous rate at which the event occurs, given that the individual has survived up to the time t . It is the probability that an event occurs in a very small time interval, given survival until the beginning of the interval.

$$h(t) = \lim_{\Delta t \rightarrow 0} \frac{P(t \leq T < t + \Delta t | T \geq t)}{\Delta t} = \frac{f(t)}{S(t)} \quad (2.4)$$

The accumulated risk of experiencing the event up to time t can be expressed using the cumulative hazard function. It is the integral of the hazard function over time, providing a cumulative measure of risk.

$$H(t) = \int_0^t h(u)du = -\log S(t) \quad (2.5)$$

Whenever we can define one of above functions, it is straightforward to derive the rest.

2.1.1 Frailty Model for Family Based Study

Parametric survival analysis methods assume that the time-to-event data follow a specific probability distribution. This approach provides a more detailed and flexible framework for modeling survival data, allowing for more precise estimates and interpretations of survival functions and hazard rates. Hosmer Jr et al. [12] has comprehensively summarized most of parametric baseline hazard

Chapter 3

Methods

3.1 Introduction

Missing data is a common and unavoidable topic for many studies that involved with data. Multiple imputation is a well-established method for dealing with the missing data in various statistical analyses as well. Until today, to our best knowledge, the multiple imputation in the context of frailty models is rarely discussed. The oversight is even more pronounced within the field of genetic epidemiology, such that the frailty model is added a layer of ascertainment correction term to account for selection bias that could be introduced in the data sampling stage. Although it is important that many research may encounter the missing data, the current existing literature lacks comprehensive discussions and methodological developments that integrate multiple imputation with the uniqueness posed by frailty models within the genetic epidemiology area. Addressing the gap and proposing the method will enhance the validity of findings for this field.

When imputing continuous variables, a common approach is to assume a conditionally normal distribution, which can be estimated from a linear regression. The original multiple imputation framework, introduced by Rubin [13], has been widely adopted by researchers across various scientific disciplines using different models. There are many softwares that implement the multiple imputations, such as Blimp software [14], MICE package in R [15], and jomo package in R [16]. Apparently, due to space limitations, this thesis cannot discuss all the well-designed software available. The development of Multiple Imputation within certain specific models remains incomplete, and many details have not yet been comprehensively addressed in the current published literature. In genetic epidemiology, studies often involve family clusters, yet current

multiple imputation methods do not account for genetic correlations. Additionally, the application for multiple imputation combined with the frailty model with ascertainment correction has not been thoroughly explored. Therefore, this research aims to develop a computationally efficient multiple imputation method that accounts for kinship correlations and applies it to frailty models with ascertainment correction.

In this chapter, a comprehensive guideline and adjusted multiple imputation formulas are provided. We explicitly demonstrate how the kinship matrix functions in the imputation step and how ascertainment correction is applied in the analysis step of this research. Moreover, the specialized imputation model is introduced, taking into account genetic variance and residual variance (also known as environmental variance). Then the variance estimation using Rubin's Rule is presented, along with the confidence interval based on the completed data following the proposed multiple imputation method.

3.2 Kinship Matrix

The kinship matrix, also known as the relatedness matrix, is a fundamental concept in statistical genetics, particularly in the context of quantitative genetics and genetic epidemiology. It quantifies the genetic relatedness between individuals based on pedigree information [17]. This matrix is essential for estimating heritability and for controlling for familial relatedness in genetic association studies. The elements of the kinship matrix represent the probability that a randomly chosen allele from one individual is identical by descent (IBD) to a randomly chosen allele from another individual. Technically, the kinship matrix can be generated either for each family or for all individuals in a study, once the relationships between each individual in a family or study are known [18]. For example, the kinship matrix K for a family of four individuals may look like this:

$$K = \begin{bmatrix} 1 & 0.5 & 0.25 & 0 \\ 0.5 & 1 & 0.5 & 0 \\ 0.25 & 0.5 & 1 & 0 \\ 0 & 0 & 0 & 1 \end{bmatrix}$$

Here, the rows and columns represent the same individuals. The diagonal of the kinship matrix has a value of 1, indicating the correlation of an individual with themselves. Specifically,

1. In K_{11} , the individual index 1 and himself is fully related to himself.
2. In $K_{12} = 0.5$, the first and second individuals are half-related (e.g., Siblings).
3. In $K_{13} = 0.25$, the first and third individuals are a quarter-related (e.g., half-siblings or grandparent-grandchild).
4. In $K_{14} = 0$, the first and the fourth individuals are not related (e.g., spousal relationship, or lawful adoption).

One advantage of using the kinship matrix is that whether the data is partitioned by family or considered globally for all individuals from different families, the results are mostly equivalent. Below shows another example of the kinship matrix that how it looks like on a global setting for three families.

1. For family 1, A is the parent, B is the child, C is the sibling of A, and D is the cousin of B.
2. For family 2, E is the parent, F is the child, G is the grandparent of F, H is the half-sibling of F
3. For family 3, I is the parent, J is the child, K is the aunt or uncle of J.

Among these three families, the matrix can be partitioned into three segments to represent the kinship correlations. In practice, this is often unnecessary since unrelated individuals will have a correlation value of 0 in the matrix. Here is an example matrix combining all families:

$$K = \begin{matrix} & \begin{matrix} A & B & C & D & E & F & G & H & I & J & K \end{matrix} \\ \begin{matrix} A \\ B \\ C \\ D \\ E \\ F \\ G \\ H \\ I \\ J \\ K \end{matrix} & \begin{pmatrix} 1 & 0.5 & 0.5 & 0.125 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0.5 & 1 & 0.25 & 0.0625 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0.5 & 0.25 & 1 & 0.0625 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0.125 & 0.0625 & 0.0625 & 1 & 0 & 0 & 0 & 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 & 1 & 0.5 & 0.25 & 0.125 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 & 0.5 & 1 & 0.5 & 0.25 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 & 0.25 & 0.5 & 1 & 0.125 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 & 0.125 & 0.25 & 0.125 & 1 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 & 0 & 0 & 0 & 0 & 1 & 0.5 & 0.25 \\ 0 & 0 & 0 & 0 & 0 & 0 & 0 & 0 & 0.5 & 1 & 0.25 \\ 0 & 0 & 0 & 0 & 0 & 0 & 0 & 0 & 0.25 & 0.25 & 1 \end{pmatrix} \end{matrix}$$

The kinship matrix is meant to be symmetric, yielding real eigenvalues and orthogonal eigenvectors, which is beneficial in principal component analysis (PCA) often conducted in genome-wide association studies (GWAS) to reduce dimensionality [19]. Also, this symmetric property ensures the genetic relatedness between any two individuals is consistent regardless of their order in a dataset, which further ensures an accurate interpretation in the the statistical inference and valid and unbiased statistical tests. In statistical analyses, the kinship matrix is not typically estimated; rather, it is derived from a precise understanding of familial relationships. This direct acquisition ensures an accurate representation of genetic relatedness, which is essential for the integrity of subsequent genetic analyses.

In some variables within a familial study, the structure could be highly dependent on the kinship matrix. For example, the polygenic risk score (PRS) may be used as a covariate when modeling certain disease risks. Particularly in genetic studies with missing data, neglecting the kinship correlation structure can lead to biased parameter inference. Current imputation models face the challenge of not adequately handling kinship correlations. Thus, the following sections demonstrate the comparisons when conducting proper multiple imputation with and without ignoring pedigree information.

3.3 Multiple Imputation without Kinship Matrix

Multiple imputation is an advantageous method for addressing missing data under the missing at random (MAR) mechanism, as it preserves the inherent variability of the data and produces valid statistical inferences. The imputation step appropriately accounts for uncertainties that may be caused by the missing data. This method has been proven to be robust for common statistical analyses, especially in contexts such as clinical trials or psychological observational studies where genetic elements are not present. Existing multiple imputation methods for continuous variables often assume a conditionally normal distribution. Suppose y is the variable containing missing values, and \mathbf{x} are other fully observed variables. We assume there are total p parameters being estimated in this linear regression. The conditionally normal model (linear regression) can be written as

$$y|\mathbf{x}, \boldsymbol{\beta} \sim N(\boldsymbol{\beta}\mathbf{x}, \sigma^2) \quad (3.1)$$

where $\boldsymbol{\beta}$ here represents the true parameter value that brings a linearly associated relationship between observed \mathbf{x} and y . The σ is the standard error introduced in this imputation model.

In the linear regression setting,

$$y = \mathbf{x}\boldsymbol{\beta} + \epsilon, \quad \epsilon \sim N(0, \sigma^2) \quad (3.2)$$

This simply corresponds to the likelihood

$$f(y|\mathbf{x}, \boldsymbol{\beta}, \sigma^2) \propto (\sigma^2)^{-n/2} \exp\left(-\frac{1}{2\sigma^2}(y - \mathbf{x}\boldsymbol{\beta})^\top (y - \mathbf{x}\boldsymbol{\beta})\right) \quad (3.3)$$

because of the conditional normality of y . From the likelihood 3.3, we can solve that

$$\hat{\boldsymbol{\beta}} = (\mathbf{x}^\top \mathbf{x})^{-1} \mathbf{x}^\top y \quad (3.4)$$

From 3.3, note that $(y - \mathbf{x}\boldsymbol{\beta})^\top (y - \mathbf{x}\boldsymbol{\beta})$ can be written as

$$(y - \mathbf{x}\boldsymbol{\beta})^\top (y - \mathbf{x}\boldsymbol{\beta}) = ((y - \mathbf{x}\boldsymbol{\beta}) + (\mathbf{x}\hat{\boldsymbol{\beta}} - \mathbf{x}\boldsymbol{\beta}))^\top ((y - \mathbf{x}\boldsymbol{\beta}) + (\mathbf{x}\hat{\boldsymbol{\beta}} - \mathbf{x}\boldsymbol{\beta})) \quad (3.5)$$

$$= (y - \mathbf{x}\boldsymbol{\beta})^\top (y - \mathbf{x}\boldsymbol{\beta}) + (\boldsymbol{\beta} - \hat{\boldsymbol{\beta}})^\top (\mathbf{x}^\top \mathbf{x})(\boldsymbol{\beta} - \hat{\boldsymbol{\beta}}) + 2(\mathbf{x}\hat{\boldsymbol{\beta}} - \mathbf{x}\boldsymbol{\beta})^\top (y - \mathbf{x}\boldsymbol{\beta}) \quad (3.6)$$

$$= (y - \mathbf{x}\boldsymbol{\beta})^\top (y - \mathbf{x}\boldsymbol{\beta}) + (\boldsymbol{\beta} - \hat{\boldsymbol{\beta}})^\top (\mathbf{x}^\top \mathbf{x})(\boldsymbol{\beta} - \hat{\boldsymbol{\beta}}) \quad (3.7)$$

So Equation 3.3 will become

$$f(y|\mathbf{x}, \boldsymbol{\beta}, \sigma^2) \propto \underbrace{(\sigma^2)^{-v/2} \exp\left(-\frac{vs^2}{2\sigma^2}\right)}_{f(\sigma)} \underbrace{(\sigma^2)^{-\frac{n-v}{2}} \exp\left(-\frac{1}{2\sigma^2}(\boldsymbol{\beta} - \hat{\boldsymbol{\beta}})^\top (\mathbf{x}^\top \mathbf{x})(\boldsymbol{\beta} - \hat{\boldsymbol{\beta}})\right)}_{f(\boldsymbol{\beta}|\sigma)} \quad (3.8)$$

where $vs^2 = (\mathbf{y} - \mathbf{x}\hat{\boldsymbol{\beta}})^\top (\mathbf{y} - \mathbf{x}\hat{\boldsymbol{\beta}}) = SSE$ where SSE is the Error Sum of Squares by definition, such that $v = n_{obs} - p$, n_{obs} is the number of observed observations in the data. Then $f(\sigma^2)$ can be written as a proportional density to the inverse gamma distribution,

$$f(\sigma^2) \propto (\sigma^2)^{-\frac{v}{2}-1} \exp\left(-\frac{vs^2}{2\sigma^2}\right) = (\sigma^2)^{-\frac{v}{2}-1} \exp\left(-\frac{SSE}{2\sigma^2}\right) \quad (3.9)$$

In this $\sigma^2 \sim \text{Inverse-Gamma}(\alpha, \phi)$, we have $\alpha = \frac{v}{2} = \frac{n_{obs}-p}{2}$ and $\phi = \frac{1}{2}vs^2 = \frac{SSE}{2}$. From the inverse-gamma property, when

$$\sigma^2 \sim \text{Inverse-Gamma}\left(\frac{n_{obs}-p}{2}, \frac{SSE}{2}\right) \quad (3.10)$$

and assume $\exists \lambda$, such that

$$\sigma^2 = \frac{SSE}{2} / \lambda \quad (3.11)$$

then λ can be transformed to

$$\lambda = \frac{\chi_{n_{obs}-p}^2}{2} \quad (3.12)$$

since $\chi_{df}^2 = \text{Gamma}(\frac{df}{2}, 2)$ where df stands for degrees of freedom, so

$$\sigma^2 = \frac{\frac{SSE}{2}}{\frac{\chi_{n_{obs}-p}^2}{2}} = \frac{SSE}{\chi_{n_{obs}-p}^2} \quad (3.13)$$

Thus, we can sample the standard deviation of the missing data distribution from

$$\sigma^* = \hat{\sigma} \sqrt{\frac{SSE}{\chi_{n_{obs}-p}^2}} = \hat{\sigma} \sqrt{\frac{SSE}{g}} \quad (3.14)$$

and from sampling $g \sim \chi_{n_{obs}-p}^2$. Moreover, we know

$$\text{Var}(\hat{\beta}) = \sigma^2(\mathbf{x}^\top \mathbf{x}^{-1}) = \mathbf{V} \quad (3.15)$$

so the marginal distribution for β is written as

$$\beta \sim N(\hat{\beta}, \sigma^2(\mathbf{x}^\top \mathbf{x}^{-1})) \quad (3.16)$$

Also note that when a random variable $T \sim N(\mathbf{m}, \mathbf{c})$, then T can be generated from a standard normal variable \mathbf{u} with

$$T = \mathbf{m} + \mathbf{L}\mathbf{u} \quad (3.17)$$

where \mathbf{L} is the cholesky decomposition of \mathbf{c} such that $\mathbf{c} = \mathbf{L}\mathbf{L}^\top$. For β , from 3.16, it can be derived as

$$\beta = \hat{\beta} + \sigma(\mathbf{x}^\top \mathbf{x})^{-1/2} \mathbf{u}_1 \quad (3.18)$$

$$= \hat{\beta} + \mathbf{u}_1 \mathbf{V}^{-1/2} \quad (3.19)$$

where $\mathbf{V}^{-1/2}$ is the cholesky decomposition of \mathbf{V} . Adjusting for σ^* to make sure $\boldsymbol{\beta}$ matches the variability implied by the random draw of σ^* ,

$$\boldsymbol{\beta}^* = \hat{\boldsymbol{\beta}} + \frac{\sigma^*}{\hat{\sigma}} \mathbf{u}_1 \mathbf{V}^{-1/2} \quad (3.20)$$

such that \mathbf{u}_1 is a row vector of p independent draws from a standard normal distribution, $u_{1k} \stackrel{iid}{\sim} N(0, 1)$, and $k = 1, \dots, p$. The imputation for y_i^* is then computed as

$$y_i^* = \boldsymbol{\beta}^* \mathbf{x}_i, \quad (3.21)$$

Therefore, the comprehensive steps of the multiple imputation for continuous missing data can be summarized as follows:

1. Calculate $\hat{y} = \hat{\boldsymbol{\beta}} \mathbf{x}$ using y_{obs} , and $\hat{\boldsymbol{\beta}}$ can be obtained easily, as well as $\hat{\sigma}$, and $\text{Var}(\hat{\boldsymbol{\beta}}) = \mathbf{V}$
2. Draw $g \sim \chi^2_{n_{obs}-p}$ for one random draw
3. Calculate $\sigma^* = \hat{\sigma} / \sqrt{SSE/g}$
4. Draw a p dimensional vector \mathbf{u}_1 such that $u_{1k} \stackrel{iid}{\sim} N(0, 1)$ and $k = 1, \dots, p$
5. Calculate $\boldsymbol{\beta}^* = \hat{\boldsymbol{\beta}} + \frac{\sigma^*}{\hat{\sigma}} \mathbf{u}_1 \mathbf{V}^{1/2}$ such that $\mathbf{V}^{1/2}$ is the cholesky decomposition of \mathbf{V}
6. Draw $u_{2i} \stackrel{iid}{\sim} N(0, 1)$
7. Impute $y_{mis,i} = \boldsymbol{\beta}^* \mathbf{x}_i$
8. (Option: PMM) Match the imputed value to the nearest observed value.
9. Repeat 2. to 7. (2. to 8. for PMM) for M times to obtain M complete datasets

The Predictive Mean Matching (PMM) in the above imputation steps is corresponding to one of imputation options proposed by Rubin [20], then further discussed by Little [21]. This option serves as an algorithm that matches the imputed value to the nearest observed value. Mathematically, when we have obtained the $\hat{y}_{mis,i}$ from step 7, we then replace $\hat{y}_{mis,i}$ to $y_{obs,j}$ where $j = \arg \min_{j \in \{1, \dots, n_{obs}\}} |\hat{y}_{mis,i} - y_{obs,j}|$. This option arguably preserved the distribution of the observed data because it imputed missing values with actual observed values rather than the predicted ones. This helps maintain the variability and distributional property in certain

ways of the data. But this may suffer from lack of uncertainty and potential biases when the missing proportion is high.

This imputation method is widely applied to most of missing continuous variable problems. However, in genetic epidemiology, when clustered data are in a special form - some continuous variable may be correlated among individuals within one family, this method may yield a biased estimate. Because this assumes an independent structure when adding the uncertainty to the imputed value, which ignores the potential correlations that may be due to the genetic nature. Therefore, in next section, we modified this imputation approach by adding the kinship correlations to the imputation step, and derived the conditional expectation of the missing value for individual i given other individuals $-i$.

3.4 Multiple Imputation with Kinship Matrix

In order to account for the kinship correlations, the conditional normal distribution needs to be adjusted where the variable contains the missing components are said to be multivariate. Denote \mathbf{y} as the continuous variable vector that is subject to missing, \mathbf{x} is other covariates that formed a design matrix that determined to be eligible to be part of the imputation model, and β is simply the parameter of the imputation model.

$$\mathbf{y}|\mathbf{x}, \beta \sim MVN(\beta\mathbf{x}, \sigma_g^2 K + \sigma_e^2 I) \quad (3.22)$$

where in the linear mixed effect regression form with flexible covariance matrix, the model can be written as

$$\mathbf{y} = \mathbf{x}\beta + \mathbf{u} + \mathbf{e} \quad (3.23)$$

such that two random parts of the model have two multivariate normal distributions,

$$\mathbf{u} \sim MVN(0, \sigma_g^2 K), \text{ and } \mathbf{e} \sim MVN(0, \sigma_e^2 I)$$

where σ_g^2 introduces the genetic variances, σ_e^2 introduces the residual variances. K is the kinship matrix with the diagonal of 1, and I is the identity matrix. Note that the flexible covariance matrix can be written as

$$\Sigma = \sigma_g^2 K + \sigma_e^2 I \quad (3.24)$$

So in this multivariate version of linear mixed effects model, the kinship matrix is being adapted. With unknown mean and fully unknown covariance matrix, the prior of the covariance matrix can be selected as an Inverse Wishart distribution with degrees of freedom and the scale parameter in Bayesian sense. However, in this case, the covariance matrix is not fully unknown. The only unknown parts of this covariance matrix is introduced by simply σ_g^2 and σ_e^2 . Deriving the Bayesian prior of these two parameters is challenging, especially these are incorporated into the partially known multivariate parameter Σ . Therefore, prior definition steps are adjusted to empirical estimates of these two variances. Although, in a normal model, the inverse-gamma distribution can be an option as a conjugate prior for variance component. The conjugacy simplifies the mathematical derivation, but there are two variance components when the flexible covariance matrix for this designed imputation model, which defers from the regular choice. With the above preliminary settings before the imputation step, the adjusted multiple imputation can be concluded into following steps:

1. Obtain the kinship matrix K among all individuals
2. Calculate the estimates of $\hat{y} = \mathbf{x}\beta$, obtain estimates of $\hat{\beta}$, $\hat{\sigma}_g^2$, $\hat{\sigma}_e^2$, $\text{Var}(\hat{\beta}) = \mathbf{V}$. In this step, naturally, $\hat{\Sigma}$ is obtained.
3. Suppose there are p predictors in the imputation model, draw p -dimensional vector \mathbf{w}_1 such that $w_{1k} \stackrel{iid}{\sim} N(0, 1)$ where $k = 1, \dots, p$
4. Calculate $\beta^* = \hat{\beta} + w_1 \mathbf{V}^{1/2}$ such that $\mathbf{V}^{1/2}$ is the cholesky decomposition of \mathbf{V}
5. Obtain $\mu_i^* = \beta^* \mathbf{x}_i$
6. Obtain the conditional expectations

$$E(y_{mis,i} | \mathbf{y}_{-i}) = \mu_i^* + \hat{\Sigma}_{i,-i} \hat{\Sigma}_{-i,-i}^{-1} (\mathbf{y}_{-i} - \boldsymbol{\mu}_{-i}^*) \quad (3.25)$$

where $\boldsymbol{\mu}_{-i}^*$ is the observed mean vector for other individuals than i .

7. Impute $y_{mis,i} = E(y_{mis,i} | \mathbf{y}_{-i})$
8. (Option: PMM) Match the imputed value to the nearest observed value
9. Repeat 3. to 7. (3. to 8. for PMM) for M times to obtain M completed datasets.

The Equation 3.25 holds with the following proof:

Proof. Consider a random vector $\mathbf{x} \sim MVN(\boldsymbol{\mu}, \boldsymbol{\Sigma})$, where $\mathbf{x} \in \mathbb{R}^n$, $\boldsymbol{\mu} \in \mathbb{R}^n$ is the mean vector, and $\boldsymbol{\Sigma} \in \mathbb{R}^{n \times n}$ is the covariance matrix. We partition \mathbf{x} into x_i and \mathbf{x}_{-i} , where x_i is a scalar and \mathbf{x}_{-i} is the $(n-1)$ -dimensional vector:

$$\mathbf{x} = \begin{pmatrix} x_i \\ \mathbf{x}_{-i} \end{pmatrix}, \boldsymbol{\mu} = \begin{pmatrix} \mu_i \\ \boldsymbol{\mu}_{-i} \end{pmatrix}, \quad \boldsymbol{\Sigma} = \begin{pmatrix} \Sigma_{ii} & \boldsymbol{\Sigma}_{i,-i} \\ \boldsymbol{\Sigma}_{-i,i} & \boldsymbol{\Sigma}_{-i,-i} \end{pmatrix}$$

Here Σ_{ii} is a scalar, which is a variance of x_i , $\boldsymbol{\Sigma}_{i,-i}$ is the covariance vector between x_i and \mathbf{x}_{-i} , and $\boldsymbol{\Sigma}_{-i,-i}$ is the covariance matrix of \mathbf{x}_{-i} . With the property of the multivariate normal distribution, such that the conditional distribution of a subset of a multivariate normal vector given another subset is still normal, with mean and covariance that can be computed from the parameters of the joint distribution. We will only focus on the conditional expectation in this case. The joint density of (x_i, \mathbf{x}_{-i}) is given by:

$$f \left(\begin{pmatrix} x_i \\ \mathbf{x}_{-i} \end{pmatrix} \right) = \frac{1}{(2\pi)^{n/2} |\boldsymbol{\Sigma}|^{1/2}} \exp \left(-\frac{1}{2} \begin{pmatrix} x_i - \mu_i \\ \mathbf{x}_{-i} - \boldsymbol{\mu}_{-i} \end{pmatrix}^T \boldsymbol{\Sigma}^{-1} \begin{pmatrix} x_i - \mu_i \\ \mathbf{x}_{-i} - \boldsymbol{\mu}_{-i} \end{pmatrix} \right)$$

Now we compute the inverse of the covariance matrix $\boldsymbol{\Sigma}$, using the block matrix inversion formula:

$$\boldsymbol{\Sigma}^{-1} = \begin{pmatrix} \Sigma_{ii} & \boldsymbol{\Sigma}_{i,-i} \\ \boldsymbol{\Sigma}_{-i,i} & \boldsymbol{\Sigma}_{-i,-i} \end{pmatrix}^{-1} = \begin{pmatrix} A & B \\ C & D \end{pmatrix}$$

where the submatrices A , B , C , and D are given by:

$$A = (\Sigma_{ii} - \boldsymbol{\Sigma}_{i,-i} \boldsymbol{\Sigma}_{-i,-i}^{-1} \boldsymbol{\Sigma}_{-i,i})^{-1}$$

$$B = -A \boldsymbol{\Sigma}_{i,-i} \boldsymbol{\Sigma}_{-i,-i}^{-1}$$

$$C = -\boldsymbol{\Sigma}_{-i,-i}^{-1} \boldsymbol{\Sigma}_{-i,i} A$$

$$D = \boldsymbol{\Sigma}_{-i,-i}^{-1} + \boldsymbol{\Sigma}_{-i,-i}^{-1} \boldsymbol{\Sigma}_{-i,i} A \boldsymbol{\Sigma}_{i,-i} \boldsymbol{\Sigma}_{-i,-i}^{-1}$$

The quadratic form in the exponent of the joint density is

$$\begin{pmatrix} x_i - \mu_i \\ \mathbf{x}_{-i} - \boldsymbol{\mu}_{-i} \end{pmatrix}^T \boldsymbol{\Sigma}^{-1} \begin{pmatrix} x_i - \mu_i \\ \mathbf{x}_{-i} - \boldsymbol{\mu}_{-i} \end{pmatrix} \quad (3.26)$$

Expanding 3.26, and using the symmetric of the covariance matrix that we know $C = B^\top$, we can get

$$\begin{aligned} & \begin{pmatrix} x_i - \mu_i \\ \mathbf{x}_{-i} - \boldsymbol{\mu}_{-i} \end{pmatrix}^T \boldsymbol{\Sigma}^{-1} \begin{pmatrix} x_i - \mu_i \\ \mathbf{x}_{-i} - \boldsymbol{\mu}_{-i} \end{pmatrix} \\ &= (x_i - \mu_i)^T A(x_i - \mu_i) + (x_i - \mu_i)^T B(\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i}) + \\ &+ (\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i})^T C(x_i - \mu_i) + (\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i})^T D(\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i}) \\ &= (x_i - \mu_i)^T A(x_i - \mu_i) + 2(x_i - \mu_i)^T B(\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i}) + (\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i})^T D(\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i}) \end{aligned}$$

The conditional mean comes directly from the term $2(x_i - \mu_i)^T B(\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i})$, leading to the linear adjustment term $\boldsymbol{\Sigma}_{i,-i} \boldsymbol{\Sigma}_{-i,-i}^{-1} (\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i})$, where

$$E(x_i | \mathbf{x}_{-i}) = \mu_i + \boldsymbol{\Sigma}_{i,-i} \boldsymbol{\Sigma}_{-i,-i}^{-1} (\mathbf{x}_{-i} - \boldsymbol{\mu}_{-i})$$

□

Using the conditional expectation captures the genetic correlations when imputing the missing variable. In fact, once the correlation matrix is known, even broader scope than the genetic epidemiology, can still apply this proposed multiple imputation method.

Generally, when there is no preliminary knowledge on the dataset, one should use as many variables as possible for the imputation model covariate [22]. Also, in the survival analysis, there has been always an argument on whether the $\log(T)$ or $\log(H_0(T))$ should be used within the imputation model [23]. In this thesis, we have used both $\log(T)$ and $\log(H_0(T))$ in the simulation study to determine the differences to avoid potential biases caused by misspecification of the model.

3.5 Pooling Step and Variance Estimation

After multiple imputation, Rubin's rules are applied to combine the results from the multiple datasets. These rules are used to derive overall estimates and associated variances that incorporate the uncertainty due to missing data. Specifically, Rubin's rule for variance estimation involves calculating within-imputation variance and between-imputation variance. The total variance is the sum of the average within-imputation variance and the between-imputation variance adjusted by a factor related to the number of imputations. This approach ensures that the variability due to missing data is properly reflected in the final statistical inferences. Rubin's rule is advantageous compared to empirical variance estimation because it accounts for both within-imputation and between-imputation variances, thus providing a more accurate reflection of the uncertainty due to missing data. This dual consideration enhances the validity of statistical inferences by incorporating the variability introduced through the imputation process. In contrast, empirical variance estimation may underestimate this uncertainty, leading to overly optimistic conclusions [22].

Based on the Rubin's rule, the pooling step of the analysis is defined as

$$\bar{\theta} = \frac{1}{M} \sum_{i=1}^M \theta_i \quad (3.27)$$

where θ_i is the parameter estimate that we are making the inference in a study for i -th imputation after M imputations. So $\bar{\theta}$ is the pooled parameter estimates. Based on Barnard and Rubin [24], the variance estimation is also defined in a two-level structure, which are within imputation variance and between imputation variance. The within imputation variance is similar to the pooled parameter estimation, where

$$V_W = \frac{1}{M} \sum_{i=1}^m SE_i^2 \quad (3.28)$$

such that V_W determines the within-imputation variance, and it's simply "pooled" variance among all analyses from the imputed dataset i . The between-imputation variance, on the other hand, accounts for the extra variances caused by the missing data. The between-imputation variance V_B is defined as

$$V_B = \frac{\sum_{i=1}^M (\theta_i - \bar{\theta})^2}{M - 1} \quad (3.29)$$

where V_B is the unbiased estimation. Note that θ is estimated using the only finite M imputed datasets, according to Van Buuren [25], V_B is the approximation when $M \rightarrow \infty$. Therefore, the total variance can be written as

$$V_{\text{Total}} = V_W + V_B + \frac{V_B}{M} \quad (3.30)$$

when M is large enough, V_{Total} tends to have the only two components of within and between variances. The test statistic (Wald statistic) is

$$\frac{(\bar{\theta} - \theta_0)^2}{V_{\text{Total}}} \sim F_{1, df_{adj}} \quad (3.31)$$

which

$$\frac{\bar{\theta} - \theta_0}{\sqrt{V_{\text{Total}}}} \sim t_{df_{adj}, \frac{1-\alpha}{2}} \quad (3.32)$$

such that α is the significance level. Define

$$\rho = \frac{V_B + \frac{V_B}{M}}{V_{\text{Total}}} \quad (3.33)$$

and

$$r = \frac{V_B + \frac{V_B}{M}}{V_W} \quad (3.34)$$

where ρ is the fraction of missing information, which quantifies the proportion of the total variance that is attributable to the fact that the data have been imputed M times. Also, r is the relative increase in variance due to non-responses (missing data), which measures the increase in variance due to the missing data compared to if there were no missing data. In the interpretation, ρ gives the proportion of the total variance that is due to the uncertainty introduced by the missing data. A higher ρ indicates a larger fraction of the total uncertainty comes from the fact that the data were imputed. A higher r indicates that the missing data have a larger impact on the overall variance. In the first MI literature by Rubin [13], the degrees of freedom was defined as

$$df_{old} = (M - 1) \times \left(1 + \frac{1}{r}\right)^2 \quad (3.35)$$

The estimated degrees of freedom for the observed data, adjusted for the missing information,

are

$$df_{obs} = \frac{(n - k) + 1}{(n - k) + 3} (n - k)(1 - \rho) \quad (3.36)$$

where n is the number of observations in each imputed data, and k is the number of parameters to be estimated. Barnard and Rubin [24] further alternated the calculation of the degrees of freedom by combining Equations 3.35 and 3.36

$$df_{adj} = \frac{df_{old}df_{obs}}{df_{old} + df_{obs}} \quad (3.37)$$

When conducting the statistical tests on the pooled estimates, df_{adj} is used for the t-distribution degrees of freedom. The confidence interval is straightforward, where we obtain

$$SE_{pooled} = \sqrt{V_{Total}} \quad (3.38)$$

then we simply calculate

$$CI = \bar{\theta} \pm t_{df_{adj}, \frac{1-\alpha}{2}} \times SE_{pooled} \quad (3.39)$$

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