

ggNetView manual documentation

Yue Liu

2026-01-09

Contents

1 ggNetView manual documentation	7
1.1 Introduction	7
1.2 Installation	8
1.3 Citation	8
1.4 Source Code	8
1.5 Contact	8
2 Build graph object	9
2.1 Build graph from matrix	9
2.2 Build graph from data frame	12
2.3 Build graph from data frame with module	15
2.4 Build graph from adjacency matrix	16
2.5 Build graph from double matrix	18
3 Random Matrix Theory (RMT)-based random network	21
3.1 Random Matrix Theory (RMT)	21
3.2 Example data	22
3.3 Compute RMT threshold	22
3.4 Build graph from matrix based on RMT-threshold	34
4 Get network information	37
4.1 Full-network information	37
4.2 Sub-network (modularity) information	37
4.3 Sub-network (sample) information	37

5 Extract subgraph	39
5.1 Extract subgraph by module	39
5.2 Extract subgraph by sample	39
6 Network layout	41
6.1 Gephi layout	42
6.2 Fruchterman–Reingold force-directed layout	42
6.3 Diamond layout	42
6.4 Kk layout	42
6.5 Nicley layout	42
6.6 Multrings layout	42
6.7 Petal layout	42
6.8 Circle layout	42
6.9 Circle outline layout	42
6.10 Diamond outline layout	42
6.11 Grid layout	42
6.12 Heart_centered layout	42
6.13 Lgl layout	42
6.14 Randomly layout	42
6.15 Rectangle layout	42
6.16 Rightiso layout	42
6.17 Square layout	42
6.18 Square outline layout	42
6.19 Star layout	42
6.20 Star_concentric layout	42
6.21 Stress layout	42
7 Network topology information	43
7.1 Get network topology information	43
7.2 Get network topology information with matrix	43
7.3 Get network topology information by parallel	43
7.4 Get network topology information with matrix by parallel	43

CONTENTS	5
8 Network comparison	45
8.1 Subgraph comparisopn	45
8.2 Comparison of multi-sample networks	45
9 Network & Environment	47
9.1 Network Environment	47
10 Multi-omics network analysis	49
10.1 Multi-omics	49

Chapter 1

ggNetView manual documentation

1.1 Introduction

Network analysis has been widely applied in the **life sciences, microbiology, ecology, and agronomy** to quantify interactions among **genes, proteins, metabolites, microorganisms, and environmental factors**. These interactions collectively shape the functioning of biological and ecological systems. Despite its broad adoption, existing tools often face limitations in flexibility, customization, reproducibility, and the generation of publication-ready figures.

To address these challenges, **ggNetView** was developed as an **R package** that integrates **ggplot2**, **ggraph**, and **tidygraph** within the **Grammar of Graphics framework**, enabling fully reproducible and highly customizable network visualizations. The package provides deterministic layout algorithms, comprehensive topological analyses, modular coloring schemes, hierarchical annotations, and consistent theming, ensuring standardized and interpretable graphical output.

Although initially designed for applications in **soil science and microbial ecology**, **ggNetView** is broadly applicable to network analyses in **molecular biology**, including **WGCNA** and **protein–protein interaction (PPI) networks**. By lowering technical barriers in network construction and visualization, **ggNetView** enables researchers across disciplines to efficiently produce reproducible, publication-quality network figures.

1.2 Installation

First, install the required dependencies

```
# install.packages("BiocManager")
BiocManager::install("WGCNA")

# install.packages("remotes")
remotes::install_github("alserglab/mascarade")
remotes::install_github("zdk123/SpiecEasi")
```

and then install ggNetView.

```
# install.packages("devtools")
devtools::install_github("Jiawang1209/ggNetView")

# install.packages("pak")
pak::pak("Jiawang1209/ggNetView")
```

1.3 Citation

If you use ggNetView in your research, please cite:

Yue Liu, Chao Wang (2025). ggNetView: An R package for complex biological and ecological network visualization. <https://github.com/Jiawang1209/ggNetView>

1.4 Source Code

The source code for ggNetView is available in the ggNetView repository.

<https://github.com/Jiawang1209/ggNetView>

1.5 Contact

- Email: Jiawang1209@163.com

Chapter 2

Build graph object

Basic workflow of `ggNetView`

1. Build a **graph object**
 2. Understand and manipulate the **graph object**
 3. Visualize the network using **layout algorithms**
 4. Retrieve **topology** of network
-

Load R Package

```
library(tidyverse)
library(ggNetView)
```

2.1 Build graph from matrix

Example data

```
# Access built-in example datasets in ggNetView

# Raw ASV or OTU table
data("otu_tab")
dim(otu_tab)
```

```

## [1] 2859    18

otu_tab[1:5, 1:5]

##      K01   K02   K03   K04   K05
## ASV_1 1113 1968  816 1372 1062
## ASV_2 1922 1227 2355 2218 2885
## ASV_3  568  460  899  902 1226
## ASV_4 1433  400  535  759 1287
## ASV_6  882  673  819  888 1475

# Rarefied ASV or OTU table
data("otu_rare")
dim(otu_rare)

## [1] 2859    18

otu_rare[1:5, 1:5]

##      K01   K02   K03   K04   K05
## ASV_1  992 1636  604 1084  806
## ASV_2 1725 1018 1814 1743 2196
## ASV_3  520  389  687  701  932
## ASV_4 1280  328  425  580 1004
## ASV_6  794  557  633  706 1142

# Relative abundance table of rarefied ASVs or OTUs
data("otu_rare_relative")
dim(otu_rare_relative)

## [1] 2859    18

otu_rare_relative[1:5, 1:5]

##          K01          K02          K03          K04          K05
## ASV_1 0.03306667 0.05453333 0.02013333 0.03613333 0.02686667
## ASV_2 0.05750000 0.03393333 0.06046667 0.05810000 0.07320000
## ASV_3 0.01733333 0.01296667 0.02290000 0.02336667 0.03106667
## ASV_4 0.04266667 0.01093333 0.01416667 0.01933333 0.03346667
## ASV_6 0.02646667 0.01856667 0.02110000 0.02353333 0.03806667

```

```
# Taxonomic annotation table of ASVs or OTUs
data("tax_tab")
dim(tax_tab)

## [1] 2859     8

tax_tab[1:5, 1:5]

## # A tibble: 5 x 5
##   OTUID Kingdom Phylum      Class      Order
##   <chr>   <chr>   <chr>      <chr>      <chr>
## 1 ASV_2   Archaea Thaumarchaeota Unassigned Nitrosphaerales
## 2 ASV_3   Bacteria Verrucomicrobia Spartobacteria Unassigned
## 3 ASV_31  Bacteria Actinobacteria Actinobacteria Actinomycetales
## 4 ASV_27  Archaea Thaumarchaeota Unassigned Nitrosphaerales
## 5 ASV_9   Bacteria Unassigned    Unassigned   Unassigned
```

Build graph object

```
graph_obj <- build_graph_from_mat(
  mat = otu_rare_relative,
  transfrom.method = "none", # based your input data
  r.threshold = 0.7,
  p.threshold = 0.05,
  method = "WGCNA",
  cor.method = "pearson",
  proc = "Bonferroni",
  module.method = "Fast_greedy",
  node_annotation = tax_tab,
  top_modules = 15,
  seed = 1115
)

graph_obj

## # A tbl_graph: 213 nodes and 844 edges
## #
## # An undirected simple graph with 29 components
## #
## # Node Data: 213 x 14 (active)
##   name    modularity modularity2 modularity3 Modularity Degree Strength Kingdom
##   <chr>    <fct>      <ord>      <chr>      <ord>      <dbl>    <dbl> <chr>
## 1 ASV_649 5          5          5          5          27       26.5 Bacter~
```

```

##  2 ASV_705 5      5      5      5    27  26.5 Bacter-
##  3 ASV_12~ 5     5      5      5    27  26.5 Bacter-
##  4 ASV_13~ 5     5      5      5    27  26.5 Bacter-
##  5 ASV_14~ 5     5      5      5    27  26.5 Bacter-
##  6 ASV_14~ 5     5      5      5    27  26.5 Bacter-
##  7 ASV_24~ 5     5      5      5    27  26.5 Bacter-
##  8 ASV_25~ 5     5      5      5    27  26.4 Bacter-
##  9 ASV_28~ 5     5      5      5    27  26.5 Bacter-
## 10 ASV_28~ 5    5      5      5    27  26.5 Bacter-
## # i 203 more rows
## # i 6 more variables: Phylum <chr>, Class <chr>, Order <chr>, Family <chr>,
## #   Genus <chr>, Species <chr>
## #
## # Edge Data: 844 x 5
##   from      to weight correlation corr_direction
##   <int> <int> <dbl>      <dbl> <chr>
## 1   194    195  0.959      0.959 Positive
## 2   185    208  0.954      0.954 Positive
## 3   185    213  0.957      0.957 Positive
## # i 841 more rows

```

2.2 Build graph from data frame

Example data

```
# Access built-in example datasets in ggNetView
data("ppi_example")
df = ppi_example$ppi
head(df)
```

```

##   from      to weight
## 1   A1 D40  9.306533
## 2   A2 D39 11.783920
## 3   A3 D38 23.005025
## 4   A4 D37  7.412060
## 5   A5 D36 18.778894
## 6   A6 D35 16.592965
```

```
node_annotation = ppi_example$annotation
head(node_annotation)
```

```
##   node group
## 1   A1    A
## 2   A2    A
## 3   A3    A
## 4   A4    A
## 5   A5    A
## 6   A6    A
```

Build graph object

```
graph_obj_from_df <- build_graph_from_df(
  df = df,
  node_annotation = node_annotation,
  directed = F,
  module.method = "Fast_greedy",
  top_modules = 15,
  seed = 1115
)

graph_obj_from_df

## # A tbl_graph: 100 nodes and 50 edges
## #
## # An unrooted forest with 50 trees
## #
## # Node Data: 100 x 9 (active)
##   name group modularity modularity2 modularity3 Modularity Degreee Segree
##   <chr> <chr> <fct>      <fct>      <chr>      <fct>      <dbl> <dbl>
## 1 C13  C     1          1          1          1          1          1
## 2 C28  C     1          1          1          1          1          1
## 3 C2   C    10         10         10         10          1          1
## 4 D9   D    10         10         10         10          1          1
## 5 A3   A    11         11         11         11          1          1
## 6 D38  D    11         11         11         11          1          1
## 7 B12  B    12         12         12         12          1          1
## 8 D19  D    12         12         12         12          1          1
## 9 A1   A    13         13         13         13          1          1
## 10 D40 D    13         13         13         13          1          1
## # i 90 more rows
## # i 1 more variable: Strength <dbl>
## #
## # Edge Data: 50 x 4
##   from    to weight correlation
##   <int> <int> <dbl>      <dbl>
## 1     9    10    45.2       45.2
```

```

## 2      15     16    50.6      50.6
## 3      5      6    37.8      37.8
## # i 47 more rows

graph_obj_from_df2 <- build_graph_from_df(
  df = df,
  node_annotation = NULL,
  directed = F,
  module.method = "Fast_greedy",
  top_modules = 15,
  seed = 1115
)

graph_obj_from_df2

## # A tbl_graph: 100 nodes and 50 edges
## #
## # An unrooted forest with 50 trees
## #
## # Node Data: 100 x 8 (active)
##   name modularity modularity2 modularity3 Modularity Degree Segree Strength
##   <chr> <fct>      <fct>      <chr>      <fct>      <dbl> <dbl>   <dbl>
## 1 C13  1          1          1          1          1       1       1    26.7
## 2 C28  1          1          1          1          1       1       1    26.7
## 3 C2   10         10         10         10         10      1       1    37.4
## 4 D9   10         10         10         10         10      1       1    37.4
## 5 A3   11         11         11         11         11      1       1    37.8
## 6 D38  11         11         11         11         11      1       1    37.8
## 7 B12  12         12         12         12         12      1       1    41.3
## 8 D19  12         12         12         12         12      1       1    41.3
## 9 A1   13         13         13         13         13      1       1    45.2
## 10 D40 13         13         13         13         13      1       1    45.2
## # i 90 more rows
## #
## # Edge Data: 50 x 4
##   from    to weight correlation
##   <int> <int>  <dbl>      <dbl>
## 1 9     10    45.2      45.2
## 2 15    16    50.6      50.6
## 3 5     6     37.8      37.8
## # i 47 more rows

```

2.3 Build graph from data frame with module

Example data

```
# Access built-in example datasets in ggNetView
data("ppi_module")
df = ppi_module$ppi
head(df)
```

```
##   from   to    weight
## 1 A1 D40 9.306533
## 2 A2 D39 11.783920
## 3 A3 D38 23.005025
## 4 A4 D37 7.412060
## 5 A5 D36 18.778894
## 6 A6 D35 16.592965
```

```
node_annotation = ppi_module$annotation
head(node_annotation)
```

```
##   node Modularity
## 1 A1      A
## 2 A2      A
## 3 A3      A
## 4 A4      A
## 5 A5      A
## 6 A6      A
```

Build graph object

```
graph_obj_from_module <- build_graph_from_module(
  df = df,
  node_annotation = node_annotation,
  directed = F,
  top_modules = 15,
  seed = 1115
)
graph_obj_from_module
```

```
## # A tbl_graph: 100 nodes and 50 edges
## #
## # An unrooted forest with 50 trees
```

```

## #
## # Node Data: 100 x 7 (active)
##   name  Modularity modularity2 modularity3 Degree Segree Strength
##   <chr> <ord>      <ord>      <chr>      <dbl> <dbl>    <dbl>
## 1 D1     D         D         D           1     1     37.1
## 2 D2     D         D         D           1     1     63.9
## 3 D3     D         D         D           1     1     54.6
## 4 D4     D         D         D           1     1     61.7
## 5 D5     D         D         D           1     1     36.1
## 6 D6     D         D         D           1     1     71.0
## 7 D7     D         D         D           1     1     27.9
## 8 D8     D         D         D           1     1     34.6
## 9 D9     D         D         D           1     1     37.4
## 10 D10   D         D         D           1     1     63.2
## # i 90 more rows
## #
## # Edge Data: 50 x 4
##   from      to weight correlation
##   <int> <int>  <dbl>      <dbl>
## 1     40     91    45.2      45.2
## 2     39     92    50.6      50.6
## 3     38     93    37.8      37.8
## # i 47 more rows

```

2.4 Build graph from adjacency matrix

Example data

```
data("adjacency_matrix_example")
dim(adjacency_matrix_example)
```

```

## [1] 2859 2859

adjacency_matrix_example[1:5, 1:5]

##          ASV_1      ASV_2 ASV_3 ASV_4      ASV_6
## ASV_1      0 0.0000000    0      0 0.0000000
## ASV_2      0 0.0000000    0      0 0.8947427
## ASV_3      0 0.0000000    0      0 0.0000000
## ASV_4      0 0.0000000    0      0 0.0000000
## ASV_6      0 0.8947427    0      0 0.0000000

```

```

data("tax_tab")
dim(tax_tab)

## [1] 2859     8

tax_tab[1:5, 1:5]

## # A tibble: 5 x 5
##   OTUID  Kingdom Phylum      Class      Order
##   <chr>   <chr>   <chr>      <chr>      <chr>
## 1 ASV_2   Archaea Thaumarchaeota Unassigned Nitrososphaerales
## 2 ASV_3   Bacteria Verrucomicrobia Spartobacteria Unassigned
## 3 ASV_31  Bacteria Actinobacteria Actinobacteria Actinomycetales
## 4 ASV_27  Archaea Thaumarchaeota Unassigned Nitrososphaerales
## 5 ASV_9   Bacteria Unassigned     Unassigned   Unassigned

```

Build graph object

```

graph_obj_adj <- build_graph_from_adj_mat(
  adjacency_matrix = adjacency_matrix_example,
  module.method = "Fast_greedy",
  node_annotation = tax_tab,
  top_modules = 15,
  seed = 1115
)

graph_obj_adj

## # A tbl_graph: 2049 nodes and 9602 edges
## #
## # An undirected simple graph with 100 components
## #
## # Node Data: 2,049 x 14 (active)
##   name    modularity modularity2 modularity3 Modularity Degree Strength Kingdom
##   <chr>   <fct>       <ord>      <chr>      <ord>      <dbl>    <dbl> <chr>
## 1 ASV_916 1           1          1          1          58        50.5 Bacter-
## 2 ASV_777 1           1          1          1          58        48.7 Bacter-
## 3 ASV_606 1           1          1          1          55        45.8 Bacter-
## 4 ASV_740 1           1          1          1          54        47.2 Bacter-
## 5 ASV_14~ 1           1          1          1          54        44.5 Bacter-
## 6 ASV_23~ 1           1          1          1          54        47.4 Bacter-
## 7 ASV_15~ 1           1          1          1          52        45.3 Bacter-
## 8 ASV_24~ 1           1          1          1          52        43.0 Bacter-

```

```

##  9 ASV_19~ 1          1          1          1          52      43.0 Bacter~
## 10 ASV_568 1          1          1          1          51      45.1 Bacter~
## # i 2,039 more rows
## # i 6 more variables: Phylum <chr>, Class <chr>, Order <chr>, Family <chr>,
## #   Genus <chr>, Species <chr>
## #
## # Edge Data: 9,602 x 5
##   from    to weight correlation corr_direction
##   <int> <int> <dbl>      <dbl> <chr>
## 1 1771 1825 0.793      0.793 Positive
## 2 594  597  0.895      0.895 Positive
## 3 588  597  0.864      0.864 Positive
## # i 9,599 more rows

```

2.5 Build graph from double matrix

Example data

```

data("BASV_tab")
dim(BASV_tab)

## [1] 50 10

BASV_tab[1:5, 1:5]

##           Sample1   Sample2   Sample3   Sample4   Sample5
## BASV1  7.769249 14.676091  6.822246 11.806957 12.086254
## BASV2 10.771846  7.749385 14.966987  7.486507 12.645461
## BASV3 11.746937 10.023547 10.025906 11.967562  8.748440
## BASV4  8.210859  4.408296 10.070862  8.954202 14.639298
## BASV5  9.833271  9.776280 10.465463  8.380012  8.905738

```

```

data("FASV_tab")
dim(FASV_tab)

```

```

## [1] 50 10

FASV_tab[1:5, 1:5]

```

```
##      Sample1   Sample2   Sample3   Sample4   Sample5
## FASV1  8.195641  7.715472 13.247401 10.419618 11.961410
## FASV2 10.032701 12.427388  7.693471 10.711077 12.157527
## FASV3 10.109549 11.891917  9.607849  8.256637  9.989264
## FASV4  9.817056  8.038730  5.519155  7.601228 12.688531
## FASV5 13.370263  9.795554 13.639041 11.071635  8.176218
```

```
data("double_mat_node_df")
dim(double_mat_node_df)
```

```
## [1] 100    2
```

```
head(double_mat_node_df)
```

```
##      name      type
## 1 BASV1 Bacterial
## 2 BASV2 Bacterial
## 3 BASV3 Bacterial
## 4 BASV4 Bacterial
## 5 BASV5 Bacterial
## 6 BASV6 Bacterial
```

Build graph object

```
graph_obj_double_mat <- build_graph_from_double_mat(
  mat1 = BASV_tab,
  mat2 = FASV_tab,
  module.method = "Fast_greedy",
  node_annotation = double_mat_node_df,
  top_modules = 15,
  seed = 1115
)
```

```
## The max module in network is 4 we use the 4 modules for next analysis
```

```
graph_obj_double_mat
```

```
## # A tbl_graph: 100 nodes and 2500 edges
## #
## # A bipartite simple graph with 1 component
## #
## # Node Data: 100 x 9 (active)
```

```

##   name    type    modularity modularity2 modularity3 Modularity Degree Segree
##   <chr>  <chr>    <fct>     <fct>      <chr>    <fct>     <dbl>  <dbl>
## 1 BASV3 Bacterial 1          1          1          1          1          50      50
## 2 BASV6 Bacterial 1          1          1          1          1          50      50
## 3 BASV8 Bacterial 1          1          1          1          1          50      50
## 4 BASV13 Bacterial 1         1          1          1          1          50      50
## 5 BASV17 Bacterial 1         1          1          1          1          50      50
## 6 BASV19 Bacterial 1         1          1          1          1          50      50
## 7 BASV25 Bacterial 1         1          1          1          1          50      50
## 8 BASV26 Bacterial 1         1          1          1          1          50      50
## 9 BASV27 Bacterial 1         1          1          1          1          50      50
## 10 BASV31 Bacterial 1        1          1          1          1          50      50
## # i 90 more rows
## # i 1 more variable: Strength <dbl>
## #
## # Edge Data: 2,500 x 4
##   from    to weight correlation
##   <int> <int> <dbl>      <dbl>
## 1 15    57    0.338     -0.338
## 2 57    90    0.648      0.648
## 3 16    57    0.162      0.162
## # i 2,497 more rows

```

Chapter 3

Random Matrix Theory (RMT)–based random network

When a numerical matrix is used as input, it is necessary to filter and screen the resulting correlation matrix. Empirical thresholds are commonly adopted, usually between 0.6 to 0.85; however, these thresholds are largely subjective.

Compared with empirical thresholding, random matrix theory (RMT) provides a data-driven and objective approach for determining correlation thresholds, effectively separating true biological signals from random noise and improving the robustness and comparability of ecological networks.

3.1 Random Matrix Theory (RMT)

Load R Package

```
library(tidyverse)
library(ggNetView)
```

3.2 Example data

Example data

```
# Access built-in example datasets in ggNetView

# Relative abundance table of rarefied ASVs or OTUs
data("otu_rare_relative")
dim(otu_rare_relative)

## [1] 2859    18

otu_rare_relative[1:5, 1:5]

##          K01        K02        K03        K04        K05
## ASV_1 0.03306667 0.05453333 0.02013333 0.03613333 0.02686667
## ASV_2 0.05750000 0.03393333 0.06046667 0.05810000 0.07320000
## ASV_3 0.01733333 0.01296667 0.02290000 0.02336667 0.03106667
## ASV_4 0.04266667 0.01093333 0.01416667 0.01933333 0.03346667
## ASV_6 0.02646667 0.01856667 0.02110000 0.02353333 0.03806667

# Taxonomic annotation table of ASVs or OTUs
data("tax_tab")
dim(tax_tab)

## [1] 2859    8

tax_tab[1:5, 1:5]

## # A tibble: 5 x 5
##   OTUID Kingdom Phylum      Class      Order
##   <chr>  <chr>   <chr>      <chr>      <chr>
## 1 ASV_2  Archaea Thaumarchaeota Unassigned Nitrososphaerales
## 2 ASV_3  Bacteria Verrucomicrobia Spartobacteria Unassigned
## 3 ASV_31 Bacteria Actinobacteria Actinobacteria Actinomycetales
## 4 ASV_27 Archaea Thaumarchaeota Unassigned Nitrososphaerales
## 5 ASV_9  Bacteria Unassigned     Unassigned   Unassigned
```

3.3 Compute RMT threshold

```
out <- ggNetView_RMT(mat = otu_rare_relative,
                      transfrom.method = "none",
                      method = "WGCNA",
                      cor.method = "pearson",
                      unfold.method = "gaussian",
                      bandwidth = "nrd0",
                      nr.fit.points = 51,
                      discard.outliers = TRUE,
                      discard.zeros = TRUE,
                      min.mat.dim = 40,
                      max.ev.spacing = 3,
                      save_plots = FALSE,
                      out_dir = "RMT_plots",
                      verbose = TRUE,
                      seed = 1115
)
```

```
## [Info] Matrix dimension: 2859 x 2859

## [Info] #non-zeros: 8173831 | Sparseness: 0.0000

## [Scan] 1/51 | threshold = 0

## Effective dimension: 2859

## [Scan] 2/51 | threshold = 0.02

## Effective dimension: 2859

## [Scan] 3/51 | threshold = 0.04

## Effective dimension: 2859

## [Scan] 4/51 | threshold = 0.06

## Effective dimension: 2859

## [Scan] 5/51 | threshold = 0.08

## Effective dimension: 2859
```

24CHAPTER 3. RANDOM MATRIX THEORY (RMT)-BASED RANDOM NETWORK

```
## [Scan] 6/51 | threshold = 0.1
##           Effective dimension: 2859
## [Scan] 7/51 | threshold = 0.12
##           Effective dimension: 2859
## [Scan] 8/51 | threshold = 0.14
##           Effective dimension: 2859
## [Scan] 9/51 | threshold = 0.16
##           Effective dimension: 2859
## [Scan] 10/51 | threshold = 0.18
##           Effective dimension: 2859
## [Scan] 11/51 | threshold = 0.2
##           Effective dimension: 2859
## [Scan] 12/51 | threshold = 0.22
##           Effective dimension: 2859
## [Scan] 13/51 | threshold = 0.24
##           Effective dimension: 2859
## [Scan] 14/51 | threshold = 0.26
##           Effective dimension: 2859
## [Scan] 15/51 | threshold = 0.28
##           Effective dimension: 2859
## [Scan] 16/51 | threshold = 0.3
```

```
##           Effective dimension: 2859

## [Scan] 17/51 | threshold = 0.32

##           Effective dimension: 2859

## [Scan] 18/51 | threshold = 0.34

##           Effective dimension: 2859

## [Scan] 19/51 | threshold = 0.36

##           Effective dimension: 2859

## [Scan] 20/51 | threshold = 0.38

##           Effective dimension: 2859

## [Scan] 21/51 | threshold = 0.4

##           Effective dimension: 2859

## [Scan] 22/51 | threshold = 0.42

##           Effective dimension: 2859

## [Scan] 23/51 | threshold = 0.44

##           Effective dimension: 2859

## [Scan] 24/51 | threshold = 0.46

##           Effective dimension: 2859

## [Scan] 25/51 | threshold = 0.48

##           Effective dimension: 2859

## [Scan] 26/51 | threshold = 0.5

##           Effective dimension: 2859
```

26CHAPTER 3. RANDOM MATRIX THEORY (RMT)-BASED RANDOM NETWORK

```
## [Scan] 27/51 | threshold = 0.52
##           Effective dimension: 2859
## [Scan] 28/51 | threshold = 0.54
##           Effective dimension: 2859
## [Scan] 29/51 | threshold = 0.56
##           Effective dimension: 2859
## [Scan] 30/51 | threshold = 0.58
##           Effective dimension: 2859
## [Scan] 31/51 | threshold = 0.6
##           Effective dimension: 2859
## [Scan] 32/51 | threshold = 0.62
##           Effective dimension: 2859
## [Scan] 33/51 | threshold = 0.64
##           Effective dimension: 2857
## [Scan] 34/51 | threshold = 0.66
##           Effective dimension: 2855
## [Scan] 35/51 | threshold = 0.68
##           Effective dimension: 2842
## [Scan] 36/51 | threshold = 0.7
##           Effective dimension: 2800
## [Scan] 37/51 | threshold = 0.72
```

```
##           Effective dimension: 2705

## [Scan] 38/51 | threshold = 0.74

##           Effective dimension: 2546

## [Scan] 39/51 | threshold = 0.76

##           Effective dimension: 2333

## [Scan] 40/51 | threshold = 0.78

##           Effective dimension: 2049

## [Scan] 41/51 | threshold = 0.8

##           Effective dimension: 1735

## [Scan] 42/51 | threshold = 0.82

##           Effective dimension: 1396

## [Scan] 43/51 | threshold = 0.84

##           Effective dimension: 1082

## [Scan] 44/51 | threshold = 0.86

##           Effective dimension: 825

## [Scan] 45/51 | threshold = 0.88

##           Effective dimension: 600

## [Scan] 46/51 | threshold = 0.9

##           Effective dimension: 454

## [Scan] 47/51 | threshold = 0.92

##           Effective dimension: 324
```

28 CHAPTER 3. RANDOM MATRIX THEORY (RMT)-BASED RANDOM NETWORK

```
## [Scan] 48/51 | threshold = 0.94

##          Effective dimension: 215

## [Scan] 49/51 | threshold = 0.96

##          Effective dimension: 157

## [Scan] 50/51 | threshold = 0.98

##          Effective dimension: 79

## [Scan] 51/51 | threshold = 1

##          Effective dimension: 2

##          Too small. Stop scanning.

out$chosen_threshold

## [1] 0.82

out2 <- ggNetView_RMT(mat = otu_rare_relative,
                      transfrom.method = "none",
                      method = "cor",
                      cor.method = "pearson",
                      unfold.method = "gaussian",
                      bandwidth = "nrd0",
                      nr.fit.points = 51,
                      discard.outliers = TRUE,
                      discard.zeros = TRUE,
                      min.mat.dim = 40,
                      max.ev.spacing = 3,
                      save_plots = FALSE,
                      out_dir = "RMT_plots",
                      verbose = TRUE,
                      seed = 1115
)

## [Info] Matrix dimension: 2859 x 2859

## [Info] #non-zeros: 8173761 | Sparseness: 0.0000
```

```
## [Scan] 1/51 | threshold = 0
##           Effective dimension: 2859
## [Scan] 2/51 | threshold = 0.02
##           Effective dimension: 2859
## [Scan] 3/51 | threshold = 0.04
##           Effective dimension: 2859
## [Scan] 4/51 | threshold = 0.06
##           Effective dimension: 2859
## [Scan] 5/51 | threshold = 0.08
##           Effective dimension: 2859
## [Scan] 6/51 | threshold = 0.1
##           Effective dimension: 2859
## [Scan] 7/51 | threshold = 0.12
##           Effective dimension: 2859
## [Scan] 8/51 | threshold = 0.14
##           Effective dimension: 2859
## [Scan] 9/51 | threshold = 0.16
##           Effective dimension: 2859
## [Scan] 10/51 | threshold = 0.18
##           Effective dimension: 2859
## [Scan] 11/51 | threshold = 0.2
```

30CHAPTER 3. RANDOM MATRIX THEORY (RMT)-BASED RANDOM NETWORK

```
##           Effective dimension: 2859

## [Scan] 12/51 | threshold = 0.22

##           Effective dimension: 2859

## [Scan] 13/51 | threshold = 0.24

##           Effective dimension: 2859

## [Scan] 14/51 | threshold = 0.26

##           Effective dimension: 2859

## [Scan] 15/51 | threshold = 0.28

##           Effective dimension: 2859

## [Scan] 16/51 | threshold = 0.3

##           Effective dimension: 2859

## [Scan] 17/51 | threshold = 0.32

##           Effective dimension: 2859

## [Scan] 18/51 | threshold = 0.34

##           Effective dimension: 2859

## [Scan] 19/51 | threshold = 0.36

##           Effective dimension: 2859

## [Scan] 20/51 | threshold = 0.38

##           Effective dimension: 2859

## [Scan] 21/51 | threshold = 0.4

##           Effective dimension: 2859
```

```
## [Scan] 22/51 | threshold = 0.42
##           Effective dimension: 2859
## [Scan] 23/51 | threshold = 0.44
##           Effective dimension: 2859
## [Scan] 24/51 | threshold = 0.46
##           Effective dimension: 2859
## [Scan] 25/51 | threshold = 0.48
##           Effective dimension: 2859
## [Scan] 26/51 | threshold = 0.5
##           Effective dimension: 2859
## [Scan] 27/51 | threshold = 0.52
##           Effective dimension: 2859
## [Scan] 28/51 | threshold = 0.54
##           Effective dimension: 2859
## [Scan] 29/51 | threshold = 0.56
##           Effective dimension: 2859
## [Scan] 30/51 | threshold = 0.58
##           Effective dimension: 2859
## [Scan] 31/51 | threshold = 0.6
##           Effective dimension: 2859
## [Scan] 32/51 | threshold = 0.62
```

32CHAPTER 3. RANDOM MATRIX THEORY (RMT)-BASED RANDOM NETWORK

```
##           Effective dimension: 2859

## [Scan] 33/51 | threshold = 0.64

##           Effective dimension: 2857

## [Scan] 34/51 | threshold = 0.66

##           Effective dimension: 2855

## [Scan] 35/51 | threshold = 0.68

##           Effective dimension: 2842

## [Scan] 36/51 | threshold = 0.7

##           Effective dimension: 2800

## [Scan] 37/51 | threshold = 0.72

##           Effective dimension: 2705

## [Scan] 38/51 | threshold = 0.74

##           Effective dimension: 2546

## [Scan] 39/51 | threshold = 0.76

##           Effective dimension: 2333

## [Scan] 40/51 | threshold = 0.78

##           Effective dimension: 2049

## [Scan] 41/51 | threshold = 0.8

##           Effective dimension: 1735

## [Scan] 42/51 | threshold = 0.82

##           Effective dimension: 1396
```

```
## [Scan] 43/51 | threshold = 0.84

## Effective dimension: 1082

## [Scan] 44/51 | threshold = 0.86

## Effective dimension: 825

## [Scan] 45/51 | threshold = 0.88

## Effective dimension: 600

## [Scan] 46/51 | threshold = 0.9

## Effective dimension: 454

## [Scan] 47/51 | threshold = 0.92

## Effective dimension: 324

## [Scan] 48/51 | threshold = 0.94

## Effective dimension: 215

## [Scan] 49/51 | threshold = 0.96

## Effective dimension: 157

## [Scan] 50/51 | threshold = 0.98

## Effective dimension: 79

## [Scan] 51/51 | threshold = 1

## Effective dimension: 10

## Too small. Stop scanning.
```

```
out2$chosen_threshold
```

```
## [1] 0.82
```

3.4 Build graph from matrix based on RMT-threshold

Build graph object

```
graph_obj <- build_graph_from_mat(
  mat = otu_rare_relative,
  transfrom.method = "none", # based your input data
  r.threshold = out$chosen_threshold,
  p.threshold = 0.05,
  method = "WGCNA",
  cor.method = "pearson",
  proc = "Bonferroni",
  module.method = "Fast_greedy",
  node_annotation = tax_tab,
  top_modules = 15,
  seed = 1115
)
```

```
graph_obj
```

```
## # A tbl_graph: 213 nodes and 844 edges
## #
## # An undirected simple graph with 29 components
## #
## # Node Data: 213 x 14 (active)
##   name      modularity modularity2 modularity3 Modularity Degree Strength Kingdom
##   <chr>    <fct>      <ord>      <chr>      <ord>      <dbl>    <dbl> <chr>
##   1 ASV_649 5          5          5          5          27       26.5 Bacter-
##   2 ASV_705 5          5          5          5          27       26.5 Bacter-
##   3 ASV_12~ 5          5          5          5          27       26.5 Bacter-
##   4 ASV_13~ 5          5          5          5          27       26.5 Bacter-
##   5 ASV_14~ 5          5          5          5          27       26.5 Bacter-
##   6 ASV_14~ 5          5          5          5          27       26.5 Bacter-
##   7 ASV_24~ 5          5          5          5          27       26.5 Bacter-
##   8 ASV_25~ 5          5          5          5          27       26.4 Bacter-
##   9 ASV_28~ 5          5          5          5          27       26.5 Bacter-
##  10 ASV_28~ 5         5          5          5          27       26.5 Bacter-
```

```
## # i 203 more rows
## # i 6 more variables: Phylum <chr>, Class <chr>, Order <chr>, Family <chr>,
## #   Genus <chr>, Species <chr>
## #
## # Edge Data: 844 x 5
##   from      to weight correlation corr_direction
##   <int> <int>  <dbl>      <dbl> <chr>
## 1   194    195  0.959      0.959 Positive
## 2   185    208  0.954      0.954 Positive
## 3   185    213  0.957      0.957 Positive
## # i 841 more rows
```


Chapter 4

Get network information

Once the graph object is constructed, basic network information can be retrieved, and subnetworks can be extracted.

Load R Package

```
library(tidyverse)
library(ggNetView)
```

4.1 Full-network information

4.2 Sub-network (modularity) information

4.3 Sub-network (sample) information

Chapter 5

Extract subgraph

5.1 Extract subgraph by module

5.2 Extract subgraph by sample

Chapter 6

Network layout

6.1 Gephi layout

6.2 Fruchterman–Reingold force-directed layout

6.3 Diamond layout

6.4 Kk layout

6.5 Nicley layout

6.6 Multrings layout

6.7 Petal layout

6.8 Circle layout

6.9 Circle outline layout

6.10 Diamond outline layout

6.11 Grid layout

6.12 Heart_centered layout

6.13 Lgl layout

6.14 Randomly layout

6.15 Rectangle layout

Chapter 7

Network topology information

- 7.1 Get network topology information**
- 7.2 Get network topology information with matrix**
- 7.3 Get network topology information by parallel**
- 7.4 Get network topology information with matrix by parallel**

Chapter 8

Network comparison

8.1 Subgraph comparisopn

8.2 Comparison of multi-sample networks

Chapter 9

Network & Environment

9.1 Network Environment

Chapter 10

Multi-omics network analysis

10.1 Multi-omics