

Assignment 5

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Question 12

(1)

$$P(t + dt) = P(dt)P(t) = (I + Rdt)P(t) \quad (1)$$

Therefore we can show that

$$\frac{dP(t)}{dt} = RP(t) \quad (2)$$

(2)

As the Markov chain is homogeneous Markov chain and π is the ergodic stationary distribution,

$$t \rightarrow \infty \quad P(t)\pi = \pi \quad (3)$$

$$\text{multiply } R, \quad RP(t)\pi = R\pi \quad (4)$$

$$\frac{dP(t)}{dt}\pi = R\pi \quad (5)$$

$$(6)$$

Given that when $t \rightarrow \infty$, $P(t)$ reach steady state, $\frac{dP(t)}{dt} = 0$. Hence $R\pi = 0$.

Question 13

(1) The joint probability of the tree is

$$P(X, Z|T) = P(Z_4) * P(X_5|Z_4) * P(Z_3|Z_4) * P(Z_2|Z_3) * P(Z_1|Z_3) \quad (7)$$

$$* P(X_4|Z_2) * P(X_3|Z_2) * P(X_2|Z_1) * P(X_1|Z_1) \quad (8)$$

(2) To do the naive calculation of $P(X|T)$ via brute-force marginalization over the hidden nodes Z ,

for each node X , we have to marginalize out all the internal nodes Z that it is dependent on. Here is an example with X_4 .

$$P(X_4|T) = \sum_Z P(X_4, Z|T) = \sum_{Z_2, Z_3, Z_4} P(X_4|Z_2)P(Z_2|Z_3)P(Z_3|Z_4)P(Z_4) \quad (9)$$

$$(10)$$

Given each Z represent a nucleotide and can take 4 values(A,T,G,C), then the total number of cases for X_4 is $4^3 = 64$. Hence 64 summation operations are needed for X_4 .

Similarly, for X_1, X_2, X_3 , they all have 64 cases and 64 summation operations, and for X_5 we have 4 summation cases as $P(X_5, Z) = \sum_{Z_4} P(X_5|Z_4)P(Z_4)$ and it only has 4 cases.

Hence in total the number of summation operation is $64 \times 4 + 4 = 260$.

(3)

$$P(X, Z|T) = P(Z_4) * P(X_5|Z_4) * P(Z_3|Z_4) * P(Z_2|Z_3) * P(Z_1|Z_3) \quad (11)$$

$$* P(X_4|Z_2) * P(X_3|Z_2) * P(X_2|Z_1) * P(X_1|Z_1) \quad (12)$$

By rearranging the expression in such way we only need to do 4 summations.

Question 14

(1)

```
if(!require("phangorn")) {
  install.packages("phangorn")
}
```

```
## Loading required package: phangorn
```

```
## Loading required package: ape
```

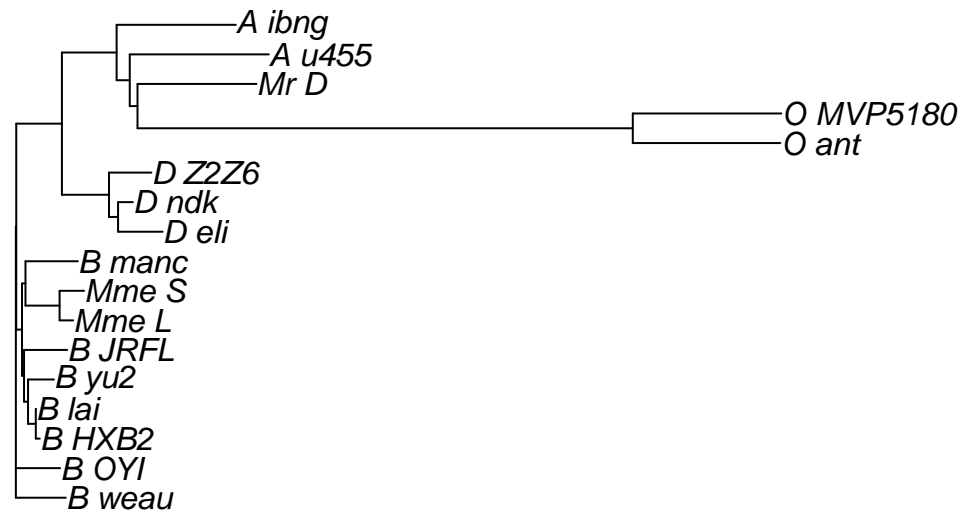
```
if(!require("ape")) {
  install.packages("ape")
}
```

```
library(phangorn)
library(ape)
```

```
ParisRT <- read.dna("ParisRT.txt")
```

(2)

```
distParis <- dist.dna(ParisRT)
initTree <- NJ(distParis)
plot(initTree)
```



(3)

```
kimura <- pml(tree= initTree, data= phyDat(ParisRT), model= "K80")
kimura$logLik
```

```
## [1] -3003.487
```

The log likelihood of the fitted model is -3003.487.

(4)

The values of the optimised rate matrix can be found below.

```
optimParam <- optim.pml(kimura, optQ= TRUE)
```

```
## optimize edge weights: -3003.487 --> -2992.981
```

```
## optimize rate matrix: -2992.981 --> -2863.264
## optimize edge weights: -2863.264 --> -2862.477
## optimize rate matrix: -2862.477 --> -2862.456
## optimize edge weights: -2862.456 --> -2862.455
## optimize rate matrix: -2862.455 --> -2862.455
## optimize edge weights: -2862.455 --> -2862.455
## optimize rate matrix: -2862.455 --> -2862.455
## optimize edge weights: -2862.455 --> -2862.455
```

```
optimParam
```

```
## model: K80
## loglikelihood: -2862.455
## unconstrained loglikelihood: -2098.897
##
## Rate matrix:
##      a      c      g      t
## a 0.000000 2.431848 6.857165 1.118323
## c 2.431848 0.000000 0.6119506 7.262319
## g 6.857165 0.6119506 0.0000000 1.000000
## t 1.118323 7.2623187 1.0000000 0.000000
##
## Base frequencies:
##      a      c      g      t
## 0.25 0.25 0.25 0.25
```

(5)

After optimizing with respect to branch lengths, nucleotide substitution rates, and tree topology, the results are as below.

```
optimParam2 <- optim.pml(kimura, optQ = TRUE, optNni = TRUE, optEdge = TRUE)
```

```
## optimize edge weights: -3003.487 --> -2992.981
## optimize rate matrix: -2992.981 --> -2863.264
## optimize edge weights: -2863.264 --> -2862.477
## optimize topology: -2862.477 --> -2849.886 NNI moves: 5
## optimize rate matrix: -2849.886 --> -2849.791
## optimize edge weights: -2849.791 --> -2849.789
## optimize topology: -2849.789 --> -2849.789 NNI moves: 0
## optimize rate matrix: -2849.789 --> -2849.789
## optimize edge weights: -2849.789 --> -2849.789
## optimize rate matrix: -2849.789 --> -2849.789
## optimize edge weights: -2849.789 --> -2849.789
```

```
optimParam2$logLik
```

```
## [1] -2849.789
```

The log likelihood is -2849.789.

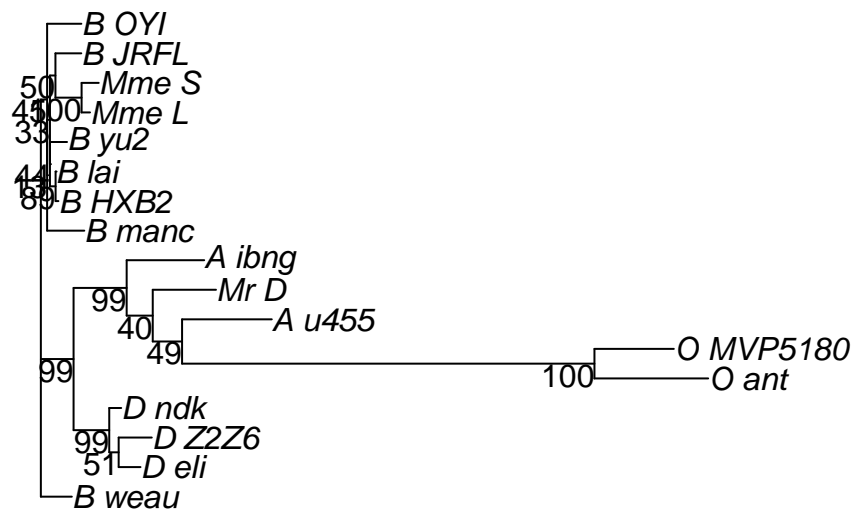
(6)

```
bootPML <- bootstrap.pml(optimParam2, optNni= TRUE)
```

In this bootstrapping function, we are resampling the sequences that we are using to construct a phylogenetic tree, and seeing if the same branch is observed even when generating a new tree based on bootstrapped data, which would indicate confidence in the observed branch.

(7)

```
plotBS(tree= optimParam2$tree, BStreets = bootPML, type= "phylogram")
```



Judging from this plot, it is more likely that Mme_S was more likely to have infected the patient Mme_L.